

Journal Pre-proofs

Ethylenic conjugated coumarin thiazolidinediones as new efficient antimicrobial modulators against clinical methicillin-resistant *Staphylococcus aureus*

Chun-Fang Hu, Peng-Li Zhang, Yan-Fei Sui, Jing-Song Lv, Mohammad Fawad Ansari, Narsaiah Battini, Shuo Li, Cheng-He Zhou, Rong-Xia Geng

PII: S0045-2068(19)31439-7
DOI: <https://doi.org/10.1016/j.bioorg.2019.103434>
Reference: YBIOO 103434

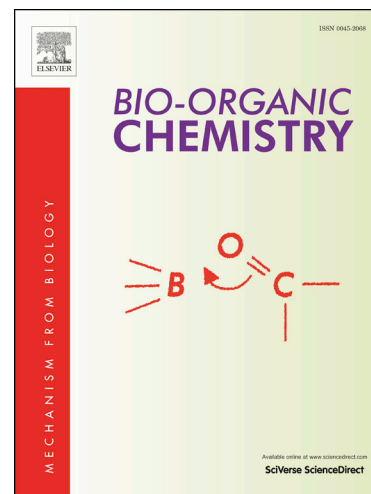
To appear in: *Bioorganic Chemistry*

Received Date: 31 August 2019
Revised Date: 30 October 2019
Accepted Date: 11 November 2019

Please cite this article as: C. Hu, P. Zhang, Y. Sui, J. Lv, M. Fawad Ansari, N. Battini, S. Li, C. Zhou, R. Geng, Ethylenic conjugated coumarin thiazolidinediones as new efficient antimicrobial modulators against clinical methicillin-resistant *Staphylococcus aureus*, *Bioorganic Chemistry* (2019), doi: <https://doi.org/10.1016/j.bioorg.2019.103434>

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2019 Published by Elsevier Inc.



Title:

Ethylenic conjugated coumarin thiazolidinediones as new efficient antimicrobial modulators against clinical methicillin-resistant *Staphylococcus aureus*

Author Names and Affiliations:

Chun-Fang Hu^a, Peng-Li Zhang^a, Yan-Fei Sui^a, Jing-Song Lv^a, Mohammad Fawad Ansari^{a,†}, Narsaiah Battini^{a,‡}, Shuo Li^{b,*}, Cheng-He Zhou^{a,*} and Rong-Xia Geng^{a,*}

^a Institute of Bioorganic & Medicinal Chemistry, Key Laboratory of Luminescent and Real-Time Analytical Chemistry (Southwest University), Ministry of Education, School of Chemistry and Chemical Engineering, Southwest University, Tiansheng street 2, Beibei district, Chongqing 400715, PR China.

^b School of Chemical Engineering, Chongqing University of Technology, Chongqing 400054, PR China.

Corresponding Address:

* Tel.: +86-23-68254967; Fax: +86-23-68254967; E-mail: zhouch@swu.edu.cn (Cheng-He Zhou); geng0712@swu.edu.cn (Rong-Xia Geng); Lishuo@cqut.edu.cn (Shuo Li).

† Postdoctoral fellow from Jamia Millia Islamia, Delhi, India

‡ Postdoctoral fellow from CSIR-Indian Institute of Integrative Medicine, Jammu, India

Abstract:

In an effort for the development of novel antimicrobial agents, ethylenic conjugated coumarin thiazolidinediones as potential multi-targeting new antimicrobial compounds were synthesized through convenient procedures from commercially available resorcinol and were evaluated for their antimicrobial potency. Bioactive evaluation revealed that some of the prepared compounds showed strong antimicrobial activities towards the tested microorganisms including clinically drug-resistant strains. Especially, propargyl derivative **12b** exhibited effective anti-MRSA potency with MIC value of 0.006 $\mu\text{mol/mL}$, which was highly advantageous over clinical antibacterial drug norfloxacin. Compound **12b** showed rapid killing effect, low toxicity against hepatocyte LO2 cells line, and no obvious drug resistance development against MRSA. Preliminary exploration of action mechanism manifested that molecule **12b** acted upon MRSA through forming stable supramolecular complex with bacterial DNA which might impede DNA replication. Molecular docking showed that compound **12b** could bind with DNA-gyrase through hydrogen bonds.

Keywords: Coumarin, Thiazolidinedione, Antimicrobial, MRSA, DNA

1. Introduction

In recent years, antibiotic-resistant pathogens have spread rapidly since its emergence, which is one of the greatest threats to modern medicinal development and mankind [1]. *Staphylococcus aureus* (*S. aureus*), notorious for its ability to produce a series of virulence factors, is categorized as kind of fatal pathogen responsible for community- and hospital-acquired infections [2,3]. Particularly, the complicated treatments of methicillin-resistant *S. aureus* (MRSA) infections have increased the morbidity and mortality of patients [4]. Isolated strains of MRSA have developed resistance to a series of antibacterial drugs, including β -lactams, macrolides, fluoroquinolones, glycopeptides and oxazolidinones [5]. However, the emergence of multi-resistant MRSA strain and complicated treatments mean that it is urgent for researchers to explore new treatment strategies [6].

Poor penetration of compounds across the microbial membrane is the greatest challenge for the discovery of novel antimicrobial drugs. Natural products have been used to treat disease for thousands of years and play an essential role in the discovery and development of drugs [7]. Some natural products, which can break through the penetration barriers, are historically significant as lead compounds for antimicrobials [8]. Coumarins are a vast class of natural products widely occurring as secondary plant metabolites [9]. More than 1800 different natural and synthetic coumarins have been discovered and applied to date, and many of them exhibited high level of biological activities including insecticidal, antioxidant, anticancer, anti-HIV, antimicrobial, and antibiotic [10–12]. Series of naturally occurring coumarin based antibiotics such as novobiocin, chlorobiocin and coumermycin A1 have been used successfully in clinic (Fig. 1). Especially, novobiocin has a good inhibitory effect on DNA gyrase, which exerts antibacterial ability *via* inhibition of ATPase activity of bacterial DNA gyrase in competition with ATP binding to the B subunit of the enzyme [13,14]. In addition, some investigators have focused on structural modification of the coumarin skeleton in the benzene ring, lactone ring or both, and found that the resulted coumarin derivatives showed good antimicrobial potential [15]. Consequently, the in-depth researches toward coumarin compounds for potential antimicrobial agents have gained much more attentions.

Fig. 1

Thiazolidinedione is an important heterocyclic fragment in design and development of novel medicinal agents concerning biological activity, including antihyperglycemic, anticancer, antiarthritic, antiinflammatory and antimicrobial [16,17]. The unique five membered structure of thiazolidinedione ring with carbonyl groups at 2- and 4-positions which acts as electron-accepting group along with NH and –S– as electron-donating groups is favorable to effectively interact with biologically important species such as DNA, enzymes and receptors [18]. The representative exploration of clinical thiazolidinedione drugs like pioglitazone and its analogues as antidiabetic agents, and GSK1059615 as first clinical PI3K inhibitor [19,20], have been motivating medicinal chemists to develop more effective bioactive molecules from the thiazolidinedione moiety. Especially, the substitution at 5-position of thiazolidinedione brings out the greatest change in structure and properties of thiazolidinedione with potent antimicrobial activity [21]. Furthermore, the introduction of different substituents on

NH group of thiazolidinedione, which might be beneficial to improve solubility and interaction with biomolecular targets, has been an important and prevalent strategy to modify thiazolidinedione analogues for the potential clinical drugs [22].

Molecular hybridization strategy is to integrate two or more pharmacophoric units into one molecular scaffold with possibly different modes of action [23]. The multifunctional attributes of these hybrids offer multiple biological activities, high selectivity and favorable pharmacokinetics, which renders hybrid molecules a rationally attractive source for current drug discovery [24]. In our previous work, we found that the introduction of thiazolidinedione to berberine showed great capacity to treat microorganisms, especially MRSA. The antibacterial mechanism showed that the suppression was accounted for dual roles of blocking bacterial DNA replication and intercalating into DNA with excellent antimicrobial activity [25]. In view of the above factors, it is quite interesting for us to combine the thiazolidinedione moiety and coumarin ring *via* an ethylenic bond bridge to develop a new structural framework of potentially antimicrobial agents (Fig. 2).

Fig. 2

The design of target ethylenic conjugated coumarin thiazolidinediones were mainly based on the following three considerations: (1) the ethylenic bond-bridged new π -conjugated skeleton expanded the conjugated system of coumarin ring and helpfully enhanced the DNA binding affinity of target molecules; (2) the different coupled positions of coumarin ring with thiazolidinedione were considered with aim to evaluate their contribution to antimicrobial activities; (3) various alkyl and unsaturated hydrocarbon chains with different lengths were introduced in order to regulate the binding affinity as well as lipid-water partition coefficient, while the substituents like chloro- and fluoro- on phenyl groups were considered to affect the pharmacological properties by modulating lipophilicity and cell membrane permeability, thus influencing the rate of transport and absorption of drugs.

All the synthesized ethylenic conjugated coumarin thiazolidinediones were characterized by ^1H NMR, ^{13}C NMR and HRMS spectral analysis, and were screened for their antibacterial and antifungal activities *in vitro*. Compound **12b** was selected to be further investigated for its interaction with DNA to explore the DNA binding efficacy and antimicrobial mechanism. To identify the effect of **12b** on cell viability, the cytotoxicity was tested against human hepatocyte LO2 cell line. Bacterial membrane permeabilization was implemented to evaluate drug uptake. Further experiments including bactericidal kinetic assay, drug resistance development, and theoretical investigation of molecular docking were also performed to verify the prospect of the most active compound.

2. Results and discussion

2.1. Chemistry

The commercial resorcinol, malic acid, ethylacetoacetate and 4-chloroacetoacetate were employed as starting materials to synthesize the target ethylenic conjugated coumarin thiazolidinediones, which were prepared *via* multistep reactions including cyclization, formylation, hydrolysis, oxidation, electrophilic substitution and Knoevenagel condensation reaction, as outlined in **Schemes 1** and **2**. Condensation of resorcinol **1** with malic

acid, ethylacetoacetate or 4-chloroacetoacetate ethyl ester in concentrated sulfuric acid at 0 °C respectively afforded coumarin derivatives **2a–b** and **5** with high yields (81.2–90.0%). Compounds **2a–b** were easily formylated by hexamethylenetetramine (HMTA) using glacial acetic acid as solvent to produce the corresponding coumarin aldehydes **3a–b** in yields of 10.0–24.0%. The hydrolysis of 4-chloromethyl-7-hydroxycoumarin **5** in boiling water produced 7-hydroxy-4-(hydroxymethyl) coumarin **6** with 92.0% yield, which was further oxidized by manganese dioxide to give 4-formyl-7-hydroxycoumarin **7** in yield of 41.0%. Intermediates **3a–b** and **7** underwent condensation reaction with thiazolidinedione in anhydrous ethanol using piperidine as base to afford the target compounds **4a–b** and **8** with moderate yields of 51.9–66.0%.

The antibacterial activities of all the ethylenic conjugated coumarin thiazolidinediones (**4a–b** and **8**) were tested, and we found that 4-methyl-8-thiazolidinedionyl coumarin derivative **4b** showed more efficient antimicrobial ability than compounds **4a** and **8**. On the purpose of investigating the essential influence of various substituents on the antimicrobial activity, the structure of compound **4b** was further modified by various substituents including aliphatic bromides, benzyl halides and 2-bromoethanol. Much effort in modifying the 7-hydroxyl group of compound **4b** is confronted with the side reaction of NH substitution. According to previous researches, NH group should be preserved to increase aqueous solubility and enhance DNA affinity. Therefore, another synthetic route was adopted starting from compound **3b** as shown in **Scheme 2**. The 7-hydroxyl group of compound **3b** was substituted by a variety of saturated or unsaturated alkyl bromides using potassium carbonate as base in *N,N*-dimethylformamide (DMF) at 80 °C to give corresponding aliphatic derivatives **9a–g** and **11a–b** with good yields (73.1–85.8%). They were further reacted with thiazolidinedione to produce the various desirable ethylenic conjugates of coumarin thiazolidinediones **10a–g** and **12a–b** in 48.6–75.4% yields, respectively. A series of target phenyl coumarin thiazolidinediones derivatives **15a–e** were also synthesized under similar reaction conditions from intermediate **3b** by reacting with the substituted benzyl chlorides and then condensing with thiazolidinedione. In order to investigate whether the substituents at NH group would decrease, retain or improve the antimicrobial activity, the target compounds **13a–b** were effectively prepared by similar reaction starting from intermediates **12a–b** with the same aliphatic bromides. All the newly synthesized ethylenic conjugated coumarin thiazolidinediones were confirmed by ¹H NMR, ¹³C NMR and HRMS spectra.

Scheme 1

Scheme 2

2.2. Analysis of configuration (*E* or *Z*)

When thiazolidinedione was reacted with coumarin aldehydes **3**, **7**, **9**, **11** and **14** respectively by the Knoevenagel condensation, theoretically there are two isomers on tautomeric equilibrium. In order to identify the structure of the isomers, the single crystal of the compound **13a** was cultured and the single crystal X-ray diffraction analysis (Fig. 3) showed that the thiazolidinedione fragment and coumarin ring *via* an ethylenic bond bridge generated a new structural type with *Z* configuration. It was possible that the *E* configuration was generally unstable, and it easily converted to the *Z* configuration.

2.3. Biological activity

All newly synthesized ethylenic conjugated coumarin thiazolidinediones were evaluated for *in vitro* antibacterial and antifungal activity against five Gram-positive bacteria, six Gram-negative bacteria and four fungal strains recommended by the Clinical and Laboratory Standard Institute (CLSI) using the referenced antimicrobial drugs norfloxacin and fluconazole as positive controls.

2.3.1. Antibacterial activity

The result for antibacterial activity (Table 1) showed that some of the newly synthesized ethylenic conjugated coumarin thiazolidinediones exhibited potent efficacy comparable to reference drug norfloxacin against the tested bacterial stains.

We sought to establish the structure-activity relationship (SAR) for this class of compounds by introducing thiazolidinedione on the three different coumarin rings, and then chosen the best antimicrobial activity coumarin ring and modified the C-7 hydroxyl group of coumarin skeleton by different length of “C-linker” aliphatic chains, hydroxyethyl fragment and substituted aromatic rings to investigated their antimicrobial effects. Preliminary biological screening showed that 8-thiazolidinedionyl coumarin derivatives **4a** and **4b** possessed slightly better antibacterial efficacy than 4-thiazolidinedionyl coumarin derivative **8**. At the same time, 4-methyl-8-thiazolidinedionyl coumarin **4b** was superior to coumarin **4a** without substitution at the C-4 position of 8-thiazolidinedionyl group in antibacterial activity, especially against Gram-negative bacteria. Most of the 7-hydroxyl thiazolidinedione coumarin derivatives exhibited moderate to good antibacterial activities against MRSA, *K. pneumonia* and *A. baumannii*, demonstrating the rationality of our design strategy.

Compounds **10a–b** and **12a–b** displayed excellent antibacterial activity with MIC (0.006 to 0.012 $\mu\text{mol/mL}$) against MRSA in comparison with norfloxacin with MIC = 0.025 $\mu\text{mol/mL}$. Particularly, compounds **12a** and **12b** were the most potent inhibitors of MRSA (MIC = 0.006 $\mu\text{mol/mL}$), which showed 4-fold higher inhibitory activity than norfloxacin. Ethylenic conjugated coumarin thiazolidinediones **12a–b** with short unsaturated alkyl chain showed better inhibitory activity than short saturated alkyl derivatives **10a** and **10b** against MRSA. Compounds with long saturated alkyl chain (i.e., **10c**, **10d**, **10e**, **10f**) didn't suppress the growth of MRSA effectively. The above results demonstrated that the introduction of the short unsaturated hydrocarbons and aliphatic chains could significantly strengthen antibacterial activity, which might regulate the lipid-water partition coefficient and binding affinity to target enzymes. Furthermore, when the NH position of compounds **12a** and **12b** were substituted by the same unsaturated hydrocarbons, the antibacterial abilities decreased about 56 folds with MIC values of 0.167 and 0.338 $\mu\text{mol/mL}$, indicating that the NH group could strengthen antibacterial ability by its weak interactions with biological targets.

Most of derivatives with MIC (0.047 to 0.387 $\mu\text{mol/mL}$) were found to improve antibacterial activities against *K. pneumonia* in comparison with norfloxacin (MIC = 0.401 $\mu\text{mol/mL}$). Especially, compound **12b** was identified as the most potent inhibitor of *K. pneumonia* (MIC = 0.047 $\mu\text{mol/mL}$), around 9-fold higher inhibitory activity than norfloxacin. Noticeably, coumarin halophenyl derivatives **15b–d** also had higher levels of bacterial growth inhibition activities against *K. pneumonia* with MIC values of 0.311, 0.139 and 0.278 $\mu\text{mol/mL}$, respectively. This

manifested that the lipophilicity and membrane permeability might be increased by the introduction of halogen atoms, which could be helpful for the rate of absorption as well as transportation of drugs to strengthen the antibacterial activities.

The antibacterial evaluation showed that MRSA was sensitive to most of the ethylenic conjugated coumarin thiazolidinediones, especially compounds **12a** and **12b**. Most of target compounds could not inhibit the growth of *S. aureus* ATCC 25923, *S. aureus* ATCC 29213, *P. aeruginosa* ATCC 27853, *E. coli*, *E. faecalis*, *P. aeruginosa*, *E. coli* ATCC 25922 effectively. Modification of compound **4b** indicated that the short unsaturated hydrocarbon and alkyl chain derivatives at the C-7 of coumarin ring were much more potent than the other substituents.

Table 1

2.3.2. Antifungal activity

The data for antifungal activity listed in **Table S1** displayed that most of the target ethylenic conjugated coumarin thiazolidinediones showed comparable or better antifungal activity than reference drug. Compounds **10a–b** and **12a–b** (MIC = 0.006–0.012 $\mu\text{mol/mL}$) exhibited superior inhibition against the growth of *A. fumigates*, at least 139 folds more potent than fluconazole (MIC = 1.672 $\mu\text{mol/mL}$). It was found that the short unsaturated hydrocarbon and alkyl chains significantly strengthened antifungal activity. Similarly, benzyl derivatives had better activities than ethylenic conjugated coumarin thiazolidinediones unsubstituted at C-7 hydroxyl group. In the series of compounds **15a–e**, compound **15e** showed the best anti-*A. fumigates* activity. It implied that strong electron-withdrawing groups on phenyl ring were beneficial to antifungal activities. Notably, *C. tropicalis* was more valuable to compounds **4b**, **10a–b**, **12a–c** and **15a–e** (MICs = 0.023–0.371 $\mu\text{mol/mL}$) than fluconazole (MIC = 0.836 $\mu\text{mol/mL}$).

All the antimicrobial results revealed that the antimicrobial efficacies of ethylenic conjugated coumarin thiazolidinediones is due to not only the modification of thiazolidinedione on coumarin ring but also the substitution at 7-position of coumarin skeleton. In particular, ethylenic conjugated coumarin thiazolidinedione compound **12a** bearing a propenyl chain and compound **12b** with a propinyl chain exhibited unprecedentedly effective inhibition toward MRSA with the same low MIC (0.006 $\mu\text{mol/mL}$), and they could be further studied as potentially antimicrobial agents.

2.4. Bactericidal kinetics

It is well-known that antibacterial effect can be evaluated through the rate of bactericidal activity [26]. To explore the antibacterial potency of compound **12b**, the kinetics of killing MRSA were investigated for compound **12b** bearing a propinyl chain. As shown in Fig. 4, compound **12b** ($4 \times \text{MIC}$) was superior to norfloxacin with 5 log CFU/mL reduction within 3 h, suggesting that it possessed bacteriostatic and bactericidal effects.

Fig. 4

2.5. Development of resistance to compound **12b**

The ever-growing risk of bacterial resistance is a critical concern, and it is the most dreadful that the various

antimicrobial resistant bacterial strains caused serious complications [27]. However, antibacterial agents that induce the death of resistant bacterial strains are deficient [28]. Thus, researches on the resistance of the highly active compound **12b** against the microbial strains were significantly essential. Due to the excellent effect of compound **12b** against MRSA, the susceptible MRSA was chosen as an experimental strain for measuring drug resistance, and norfloxacin was taken as control. The bacterial resistance of compound **12b** remained unchanged after 16 passages, whereas MRSA developed resistance against norfloxacin after six passages, suggesting that compound **12b** was harder to induce drug resistance than norfloxacin (Fig. 5).

Fig. 5

2.6 Cytotoxicity

Cytotoxicity is an important consideration for the development of potential drug candidates. Therefore, compound **12b** was further examined for their cytotoxicity by MTT assay against the growth of human hepatocyte LO2 cells [29]. It was evident from the results (Fig. 6) that the compound **12b** didn't show any appreciable cytotoxicity and cells were significantly alive even after 24 h or 48 h, and more than 95% of cells were viable at the lowest concentration (0.8 μ M). Therefore, compound **12b** showed the low toxicity, implying no obvious harm to the human body.

Fig. 6

2.7. Bacterial membrane permeabilization

Bacterial membrane active agents are topical antibacterial compounds that permeabilize bacterial membranes, thus enhancing drug uptake. While Gram-positive bacteria are surrounded by an inner membrane and a thick layer of cell wall [30], this structure forms a physical barrier that renders Gram-positive bacteria naturally resistant to many antibacterial agents. These facts adequately illustrate the significance of membrane active molecules. Herein, the disruption of bacterial membrane of compound **12b** was evaluated by utilizing propidium iodide (PI) dye. If it can target or interact with cell membrane, compound **12b** will pass through the compromised cell membranes and increase fluorescence with a complex formed by DNA. The fluorescence intensity of experimental group (in the presence of compound **12b**) gradually increased with time and then stabilized, while the control group almost showed no change (Fig. 7). It indicated that compound **12b** could effectively permeate the membrane of MRSA.

Fig. 7

2.8. Interactions of compound **12b** with MRSA DNA

Previous work revealed that coumarins could effectively interact with DNA through its excellent DNA binding affinity [31]. As is well known to us, DNA is used for storing genetic information to continue the life and can also be considered as a multiple-site target for the design and synthesis of efficient antimicrobial drugs [32]. Therefore, to investigate the possible antimicrobial mechanism, the binding behavior of compound **12b** with isolated

drug-resistant MRSA DNA were investigated by UV-vis spectroscopy using spectral probe of neutral red.

2.8.1. Absorption spectra of DNA in the presence of compound **12b**

The UV-Vis absorption in Fig. 8 showed a red-shift peak for the maximum absorption of DNA with proportional increasing concentrations of compound **12b**. It was observed that the absorption value of **12b**-DNA complex was a little smaller than the simply sum of free DNA and free compound **12b**, in line with previous studies [33,34], which suggested the hypochromism of the complex of DNA and compound **12b**. This implied the intercalation of the aromatic chromophore of compound **12b** into the helix and the aromatic chromophore was in close proximity to the DNA bases.

Fig. 8

2.8.2. Absorption spectra of NR interaction with DNA

In order to further explore the mode of action of molecule **12b** and MRSA DNA, this work used neutral red (NR) as spectral probe. As shown in Fig. S2, with the increasing concentration of MRSA DNA, the NR absorption peak around 460 nm gradually decreased and a new band at around 530 nm appeared, which accounted for the formation of DNA-NR complex. At the same time, the formation of DNA-NR complex was also revealed by the signal occurrence of an isosbestic point at 504 nm.

2.8.3. Absorption spectra of competitive interaction of compound **12b** and NR with DNA

The competitive binding of NR and compound **12b** with DNA was shown in Fig. 9. With the increasing concentration of compound **12b**, the maximum absorption peak of DNA-NR complex at 530 nm decreased, which was opposite with the absorption of free NR as the concentrations of DNA increased. This showed that compound **12b** replaced NR and intercalated in the double helices of MRSA DNA. Therefore, it was further manifested that compound **12b** could exert the antibacterial activities by intercalating into DNA to block its replication.

Fig. 9

2.9. Molecular docking

To further rationalize the observed antimicrobial activity, a flexible ligand-receptor docking investigation was undertaken against gyrase-DNA complex (PDB code: 2XCS) [35,36]. Molecular docking (Fig. 10) demonstrated that the carbonyl group of thiazolidinedione moiety could bind with the active site ARG1122 in a non-covalent binding mode. The oxygen atom of carbonyl group in thiazolidinedione ring was adjacent to the ARG1122, forming two hydrogen bonds with distances of 2.8 and 3.3 Å, respectively. The hydrogen bonds could be helpful for the stability of the **12b**-DNA complex, which might be responsible for the good inhibitory efficacy of compound **12b** against MRSA.

Fig. 10

3. Conclusion

A class of new ethylenic conjugated coumarin thiazolidinediones as potential antimicrobial agents were developed *via* some appropriate and productive synthetic routes. Their structures were confirmed by ^1H NMR, ^{13}C NMR and HRMS spectra. The antimicrobial evaluation *in vitro* revealed that some target compounds manifested better efficacy toward the tested strains of bacteria and fungi in comparison with the reference drugs norfloxacin and fluconazole. Moreover, the SAR study indicated that the type of substituents on the 7-hydroxyl group of coumarin skeleton was an important influencing factor on antimicrobial potency and most of the aliphatic and benzyl substituents exerted relatively potent antimicrobial activities. Especially, compound **12b**, the ethylenic conjugated coumarin thiazolidinediones with propargyl group, exhibited excellent unexpected interference and inhibitory ability toward MRSA (MIC = 0.006 $\mu\text{mol/mL}$) and drug-resistant *A. fumigates* (MIC=0.012 $\mu\text{mol/mL}$). In addition, it showed no toxicity against LO2 cell line and did not prompt resistance development in MRSA. Preliminary mechanism studies demonstrated that the active molecule **12b** might target on cell membrane and DNA by intercalating into MRSA DNA to form a steady **12b**-DNA complex, which might shut off DNA replication to inhibit the growth of MRSA. Moreover, molecular calculation study showed the binding between compound **12b** and gyrase-DNA complex through hydrogen bonds. These results strongly demonstrated that compound **12b** bearing propargyl group at C-7 position of coumarin ring would have enormous possibilities as new bactericidal compound, especially as a multi-targeting anti-MRSA agent.

4. Experimental protocols

4.1. General methods

All commercially available solvents and reagents were purchased without further purification. Melting points were determined with X-6 melting point apparatus. TLC analysis was done using pre-coated silica gel plates. ^1H NMR and ^{13}C NMR spectra were recorded on AVANCE III 600 MHz spectrometer using tetramethylsilane (TMS) as an internal standard. The chemical shifts were reported in parts per million (ppm), the coupling constants (J) were expressed in hertz (Hz) and signals were described as singlet (s), doublet (d), triplet (t), quartet (q) as well as multiplet (m). Mass spectra were recorded on an LCMS-2010A, the high-resolution mass spectra (HRMS) were analyzed by an IonSpec FTICR mass spectrometer with ESI resource.

4.1.1. General procedures for the preparation of hydroxycoumarin and coumarin aldehydes (**2a–b**, **3a–b** and **5–7**)

Compound **2a–b**, **3a–b** and **5–7** were obtained according to the synthetic methods reported in literature [37,38].

4.1.2. General procedures for the preparation of aliphatic and aromatic coumarin derivatives (**9a–f**, **11a–c** and **14a–e**)

Compound **9a–f**, **11a–c** and **14a–e** were obtained through the previously reported methods in literature [39].

4.1.3. Synthesis of 7-ethoxy-4-methyl-2-oxo-2H-chromene-8-carbaldehyde (**9a**)

Compound **3b** (350 mg, 1.7 mmol) and potassium carbonate (355 mg, 2.6 mmol) were stirred in N,N-Dimethylformamide (50 mL) at 80 °C for 1 h, then the bromoethane (280 mg, 2.6 mmol) was added and the

temperature maintained at 80 °C for 6 h. After cooling at room temperature, the reaction mixture was diluted with water (25 mL) and extracted with chloroform (3 × 20 mL). The organic phase was concentrated under vacuum. The crude product was purified by gel column chromatography (eluent, petroleum ether/chloroform (20/1, V/V)) to afford the desired compound **9a** (321 mg, 80.6%) as yellow solid. Mp: 210–212 °C; ¹H NMR (600 MHz, CDCl₃) δ 10.65 (s, 1H, CHO), 7.72 (d, *J* = 8.9 Hz, 1H, coumarin-5-*H*), 6.93 (d, *J* = 8.9 Hz, 1H, coumarin-6-*H*), 6.19 (s, 1H, coumarin-3-*H*), 4.24 (q, *J* = 6.9 Hz, 2H, OCH₂CH₃), 2.41 (s, 3H, coumarin-4-CH₃), 1.52 (t, *J* = 6.9 Hz, 3H, OCH₂CH₃) ppm; ¹³C NMR (151 MHz, CDCl₃) δ 187.1 (CHO), 162.6 (OC=O), 159.6, 155.2, 151.9, 130.6, 113.5, 112.9, 112.6, 108.6, 65.4 (OCH₂), 18.8 (coumarin-4-CH₃), 14.5 (OCH₂CH₃) ppm.

4.1.4. Synthesis of 4-methyl-2-oxo-7-propoxy-2H-chromene-8-carbaldehyde (**9b**)

According to the similar preparation of compound **9a**, compound **3b** (350 mg, 1.7 mmol), 1-bromopropane (320 mg, 2.6 mmol) and potassium carbonate (355 mg, 2.6 mmol) were employed to produce the pure product **9b** (342 mg, 81.7%) as light yellow solid. Mp: 158–159 °C; ¹H NMR (600 MHz, CDCl₃) δ 10.67 (s, 1H, CHO), 7.72 (d, *J* = 9.0 Hz, 1H, coumarin-5-*H*), 6.93 (d, *J* = 9.0 Hz, 1H, coumarin-6-*H*), 6.19 (s, 1H, coumarin-3-*H*), 4.12 (t, *J* = 6.4 Hz, 2H, OCH₂CH₂CH₃), 2.41 (s, 3H, coumarin-4-CH₃), 1.94–1.88 (m, 2H, OCH₂CH₂CH₃), 1.09 (t, *J* = 7.4 Hz, 3H, O(CH₂)₂CH₃) ppm; ¹³C NMR (151 MHz, CDCl₃) δ 187.1 (CHO), 162.8 (OC=O), 159.6, 155.3, 151.8, 130.6, 113.5, 113.0, 112.6, 108.6, 71.1 (OCH₂), 22.4, 18.8 (coumarin-4-CH₃), 10.4 ppm.

4.1.5. Synthesis of 7-butoxy-4-methyl-2-oxo-2H-chromene-8-carbaldehyde (**9c**)

According to the similar preparation of compound **9a**, compound **3b** (350 mg, 1.7 mmol), 1-bromobutane (356 mg, 2.6 mmol) and potassium carbonate (355 mg, 2.6 mmol) were employed to produce the pure product **9c** (380 mg, 85.8%) as yellow solid. Mp: 154–155 °C; ¹H NMR (600 MHz, CDCl₃) δ 10.65 (s, 1H, CHO), 7.72 (d, *J* = 9.0 Hz, 1H, coumarin-5-*H*), 6.93 (d, *J* = 9.0 Hz, 1H, coumarin-6-*H*), 6.18 (s, 1H, coumarin-3-*H*), 4.16 (t, *J* = 6.4 Hz, 2H, OCH₂(CH₂)₂CH₃), 2.41 (s, 3H, coumarin-4-CH₃), 1.89–1.83 (m, 2H, OCH₂CH₂CH₂CH₃), 1.58–1.51 (m, 2H, O(CH₂)₂CH₂CH₃), 1.00 (t, *J* = 7.4 Hz, 3H, O(CH₂)₃CH₃) ppm; ¹³C NMR (151 MHz, CDCl₃) δ 187.1 (CHO), 162.9 (OC=O), 159.6, 155.2, 151.9, 130.6, 113.5, 113.0, 112.6, 108.6, 69.4 (OCH₂), 30.9, 19.1, 18.8 (coumarin-4-CH₃), 13.7 ppm.

4.1.6. Synthesis of 7-(decyloxy)-4-methyl-2-oxo-2H-chromene-8-carbaldehyde (**9f**)

According to the similar preparation of compound **9a**, compound **3b** (350 mg, 1.7 mmol), 1-bromodecane (575 mg, 2.6 mmol) and potassium carbonate (355 mg, 2.6 mmol) were employed to produce the pure product **9f** (476 mg, 81.3%) as yellow solid. Mp: 201–202 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 10.46 (s, 1H, CHO), 7.97 (d, *J* = 9.0 Hz, 1H, coumarin-5-*H*), 7.22 (d, *J* = 9.0 Hz, 1H, coumarin-6-*H*), 6.30 (s, 1H, coumarin-3-*H*), 4.21 (t, *J* = 6.4 Hz, 2H, OCH₂(CH₂)₈CH₃), 2.42 (s, 3H, coumarin-4-CH₃), 1.80–1.74 (m, 2H, OCH₂CH₂(CH₂)₇CH₃), 1.44 (dd, *J* = 15.0, 7.5 Hz, 2H, O(CH₂)₂CH₂(CH₂)₆CH₃), 1.33 (t, *J* = 10.7 Hz, 2H, O(CH₂)₃CH₂(CH₂)₅CH₃), 1.28–1.22 (m, 10H, O(CH₂)₄(CH₂)₅CH₃), 0.85 (t, *J* = 6.9 Hz, 3H, O(CH₂)₉CH₃) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 187.46 (CHO), 163.21 (OC=O), 159.62, 154.08, 153.71, 132.48, 113.64, 112.65, 112.16, 109.88, 69.86 (OCH₂), 31.74, 31.13, 29.37, 29.10, 28.82, 25.77, 22.53, 18.8 (coumarin-4-CH₃), 14.55, 14.38 ppm.

4.1.7. Synthesis of 7-(allyloxy)-4-methyl-2-oxo-2H-chromene-8-carbaldehyde (**11a**)

According to the similar preparation of compound **9a**, compound **3b** (350 mg, 1.7 mmol), 3-bromoprop-1-ene (315 mg, 2.6 mmol) and potassium carbonate (355 mg, 2.6 mmol) were employed to produce the pure product **11a** (327 mg, 78.7%) as yellow solid. Mp: 172–173 °C; ¹H NMR (600 MHz, CDCl₃) δ 10.68 (s, 1H, CHO), 7.73 (d, *J* = 9.0 Hz, 1H, coumarin-5-*H*), 6.94 (d, *J* = 9.0 Hz, 1H, coumarin-6-*H*), 6.20 (s, 1H, coumarin-3-*H*), 6.06 (ddd, *J* = 15.5, 10.2, 4.8 Hz, 1H, CH=CH₂), 5.54 (d, *J* = 17.2 Hz, 1H, CH=CH₂), 5.37 (d, *J* = 10.7 Hz, 1H, CH=CH₂), 4.75 (d, *J* = 4.7 Hz, 2H, OCH₂), 2.42 (s, 3H, coumarin-4-CH₃) ppm; ¹³C NMR (151 MHz, CDCl₃) δ 185.2 (CHO), 160.2 (OC=O), 157.7, 153.7, 150.1, 129.8 (CHC=CH₂), 128.8, 116.8 (CHC=CH₂), 112.0, 111.4, 111.0, 107.3, 68.2 (OCH₂), 17.0 ppm.

4.1.8. Synthesis of 4-methyl-2-oxo-7-(prop-2-yn-1-yloxy)-2H-chromene-8-carbaldehyde (**11b**)

According to the similar preparation of compound **9a**, compound **3b** (350 mg, 1.7 mmol), 3-bromoprop-1-yne (309 mg, 2.6 mmol) and potassium carbonate (355 mg, 2.6 mmol) were employed to produce the pure product **11b** (301 mg, 73.1%) as yellow solid. Mp: 206–207 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 10.45 (s, 1H, CHO), 8.02 (d, *J* = 9.0 Hz, 1H, coumarin-5-*H*), 7.27 (d, *J* = 9.0 Hz, 1H, coumarin-6-*H*), 6.32 (s, 1H, coumarin-3-*H*), 5.08 (s, 2H, OCH₂), 3.70 (s, 1H, C≡CH), 2.43 (s, 3H, coumarin-4-CH₃) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 187.3 (CHO), 161.3 (OC=O), 159.4, 154.2, 153.5, 132.2, 114.4, 113.1, 112.6, 110.3, 79.9 (C≡CH), 78.6 (C≡CH), 57.6 (OCH₂), 18.8 (coumarin-4-CH₃) ppm.

4.1.9. Synthesis of 7-((2-fluorobenzyl)oxy)-4-methyl-2-oxo-2H-chromene-8-carbaldehyde (**14a**)

According to the similar preparation of compound **9a**, compound **3b** (350 mg, 1.7 mmol), 1-(chloromethyl)-2-fluorobenzene (376 mg, 2.6 mmol) and potassium carbonate (355 mg, 2.6 mmol) were employed to produce the pure product **14a** (411 mg, 77.5%) as yellow solid. Mp: 255–257 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 10.51 (s, 1H, CHO), 8.03 (d, *J* = 9.0 Hz, 1H, coumarin-5-*H*), 7.77 (d, *J* = 5.1 Hz, 1H, Ph-4-*H*), 7.53 (d, *J* = 2.8 Hz, 1H, coumarin-6-*H*), 7.44–7.41 (m, 2H, Ph-3,5-2*H*), 7.35 (d, *J* = 9.0 Hz, 1H, Ph-6-*H*), 6.34 (s, 1H, coumarin-3-*H*), 5.43 (s, 2H, OCH₂), 2.43 (s, 3H, coumarin-4-CH₃) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 187.4 (CHO), 162.1 (OC=O), 159.5, 154.4, 153.6, 133.9, 132.8, 132.6, 130.5, 130.3, 129.9, 127.9, 114.3, 112.5, 110.3, 68.8 (OCH₂), 18.8 (coumarin-4-CH₃) ppm.

4.1.10. Synthesis of 7-((4-fluorobenzyl)oxy)-4-methyl-2-oxo-2H-chromene-8-carbaldehyde (**14b**)

According to the similar preparation of compound **9a**, compound **3b** (350 mg, 1.7 mmol), 1-(chloromethyl)-4-fluorobenzene (376 mg, 2.6 mmol) and potassium carbonate (355 mg, 2.6 mmol) were employed to produce the pure product **14b** (424 mg, 79.9%) as yellow solid. Mp: 248–249 °C; ¹H NMR (600 MHz, CDCl₃) δ 10.71 (s, 1H, CHO), 7.72 (d, *J* = 8.9 Hz, 1H, coumarin-5-*H*), 7.47 (dd, *J* = 8.5, 5.4 Hz, 2H, Ph-2,6-2*H*), 7.09 (t, *J* = 8.6 Hz, 2H, Ph-3,5-2*H*), 6.98 (d, *J* = 9.0 Hz, 1H, coumarin-6-*H*), 6.21 (d, *J* = 0.9 Hz, 1H, 1H, coumarin-3-*H*), 5.25 (s, 2H, OCH₂), 2.41 (s, 3H, coumarin-4-CH₃) ppm; ¹³C NMR (151 MHz, CDCl₃) δ

186.9 (CHO), 163.5 (OC=O), 161.9, 161.6, 159.4, 155.7, 151.8, 131.2, 130.6, 128.8, 115.8, 115.7, 114.0, 113.4, 113.0, 109.2, 70.5 (OCH₂), 18.8 (coumarin-4-CH₃) ppm.

4.1.11. Synthesis of 7-((3,4-dichlorobenzyl)oxy)-4-methyl-2-oxo-2H-chromene-8-carbaldehyde (**14c**)

According to the similar preparation of compound **9a**, compound **3b** (300 mg, 1.47 mmol), 1,2-dichloro-4-(chloromethyl)benzene (431 mg, 2.21 mmol) and potassium carbonate (305 mg, 2.21 mmol) were employed to produce the pure product **14c** (230 mg, 43.2%) as white solid. Mp: 248–249 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 10.54 (s, 1H, CHO), 8.02 (d, *J* = 9.0 Hz, 1H, coumarin-5-*H*), 7.85 (s, 1H, Ph-5-*H*), 7.69 (d, *J* = 8.3 Hz, 1H, Ph-2-*H*), 7.53 (d, *J* = 8.2 Hz, 1H, coumarin-6-*H*), 7.27 (d, *J* = 9.0 Hz, 1H, Ph-6-*H*), 6.34 (s, 1H, coumarin-3-*H*), 5.40 (s, 2H, OCH₂), 2.43 (s, 3H, coumarin-4-CH₃) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 187.52 (CHO), 132.52, 131.25, 129.70, 127.94, 112.46, 110.24, 69.46 (OCH₂), 18.82 (coumarin-4-CH₃) ppm.

4.1.12. Synthesis of (Z)-5-((7-hydroxy-2-oxo-2H-chromen-4-yl)methylene)thiazolidine-2,4-dione (**8**)

Compound **7** (200 mg, 1.06 mmol), thiazolidine-2,4-dione (186 mg, 1.59 mmol) and piperidine (15 mg) were stirred in ethanol (20 mL) at 80 °C for 12 h. Upon completion of the reaction, the residue was filtered and washed with ethanol, and the crude product was further purified by column chromatography (eluent, petroleum ether /chloroform (1/1, V/V)) to afford the desired compound **8** (200 mg, 65.3%) as yellow solid. Mp: > 250 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 12.88 (s, 1H, NH), 10.76 (s, 1H, OH), 7.88 (s, 1H, C=CH), 7.64 (d, *J* = 8.8 Hz, 1H, coumarin-5-*H*), 6.83 (dd, *J* = 8.7, 2.3 Hz, 1H, coumarin-6-*H*), 6.78 (d, *J* = 2.3 Hz, 1H, coumarin-8-*H*), 6.22 (s, 1H, coumarin-3-*H*) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 167.44 (SC=O), 166.66 (CC=ONH), 162.46 (OC=O), 160.16, 155.70, 148.07, 133.93, 127.18, 124.29, 113.85, 110.46, 109.99, 103.25 ppm; HRMS (ESI) calcd. for C₁₃H₇NO₅S [M-H]⁺, 287.9967, found, 287.9966.

4.1.13. Synthesis of (Z)-5-((7-hydroxy-2-oxo-2H-chromen-8-yl)methylene)thiazolidine-2,4-dione (**4a**)

According to the similar preparation of compound **8**, compound **3a** (190 mg, 1.0 mmol), thiazolidine-2,4-dione (176 mg, 1.5 mmol) were employed to produce the pure product **4a** (150 mg, 51.9%) as orange solid. Mp: > 250 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 7.89–7.83 (m, 2H, C=CH, coumarin-4-*H*), 7.44 (d, *J* = 8.7 Hz, 1H, coumarin-5-*H*), 6.65 (s, 1H, coumarin-6-*H*), 6.09–6.05 (m, 1H, coumarin-3-*H*) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 171.09 (SC=O), 170.70 (CC=ONH), 165.44 (OC=O), 160.64, 154.85, 145.42, 131.45, 126.48, 123.88, 115.67, 109.25, 108.91 ppm; HRMS (ESI) calcd. for C₁₃H₇NO₅S [M+H]⁺, 290.0123, found, 290.0125; [M+Na]⁺, 311.9943, found, 311.9940.

4.1.14. Synthesis of (Z)-5-((7-hydroxy-4-methyl-2-oxo-2H-chromen-8-yl)methylene)thiazolidine-2,4-dione (**4b**)

According to the similar preparation of compound **8**, compound **3b** (204 mg, 1.0 mmol), thiazolidine-2,4-dione (175 mg, 1.5 mmol) were employed to produce the pure product **4b** (200 mg, 66.0%) as orange solid. Mp: > 250 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 12.09 (s, 1H, NH), 7.93 (s, 1H, C=CH), 7.71 (d, *J* = 8.8 Hz, 1H, coumarin-5-*H*), 6.94 (d, *J* = 8.8 Hz, 1H, coumarin-6-*H*), 6.22 (s, 1H, coumarin-3-*H*), 2.39 (s, 3H, coumarin-4-CH₃) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 169.03 (SC=O), 167.98 (CC=ONH), 159.67 (OC=O), 154.21, 152.96,

128.80, 127.84, 124.27, 113.01, 112.49, 110.95, 108.48, 18.76 (coumarin-4-CH₃) ppm; HRMS (ESI) calcd. for C₁₄H₉NO₅S [M+H]⁺, 304.0280, found, 304.0282.

4.1.15. Synthesis of (Z)-5-((7-ethoxy-4-methyl-2-oxo-2H-chromen-8-yl)methylene)thiazolidine-2,4-dione (**10a**)

According to the similar preparation of compound **8**, compound **9a** (334 mg, 1.44 mmol), thiazolidine-2,4-dione (253 mg, 2.16 mmol) were employed to produce the pure product **10a** (256 mg, 53.7%) as yellow solid. Mp: > 250 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 12.52 (s, 1H, NH), 7.88 (s, 1H, C=CH), 7.84 (d, *J* = 9.0 Hz, 1H, coumarin-5-*H*), 7.18 (d, *J* = 9.0 Hz, 1H, coumarin-6-*H*), 6.29 (d, *J* = 0.8 Hz, 1H, coumarin-3-*H*), 4.31 (d, *J* = 7.0 Hz, 2H, OCH₂CH₃), 2.42 (d, *J* = 0.6 Hz, 3H, coumarin-4-CH₃), 1.42 (t, *J* = 7.0 Hz, 3H, OCH₂CH₃) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 168.60 (SC=O), 167.72 (CC=ONH), 159.49 (OC=O), 159.09, 153.95, 152.32, 128.99, 123.76, 113.77, 111.97, 109.79, 109.16, 65.50 (OCH₂), 18.71 (coumarin-4-CH₃), 14.78 ppm; HRMS (ESI) calcd. for C₁₆H₁₃NO₅S [M+H]⁺, 332.0593, found, 332.0592.

4.1.16. Synthesis of (Z)-5-((4-methyl-2-oxo-7-propoxy-2H-chromen-8-yl)methylene)thiazolidine-2,4-dione (**10b**)

According to the similar preparation of compound **8**, compound **9b** (224 mg, 0.99 mmol), thiazolidine-2,4-dione (175 mg, 1.49 mmol) were employed to produce the pure product **10b** (170 mg, 49.7%) as white solid. Mp: > 250 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 12.53 (s, 1H, NH), 7.88 (s, 1H, C=CH), 7.84 (d, *J* = 9.0 Hz, 1H, coumarin-5-*H*), 7.19 (d, *J* = 9.0 Hz, 1H, coumarin-6-*H*), 6.29 (s, 1H, coumarin-3-*H*), 4.20 (t, *J* = 6.7 Hz, 2H, OCH₂CH₂CH₃), 2.43 (s, 3H, coumarin-4-CH₃), 1.82 (dt, *J* = 14.2, 7.1 Hz, 2H, OCH₂CH₂CH₃), 0.99 (t, *J* = 7.4 Hz, 3H, OCH₂CH₂CH₃) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 168.60 (SC=O), 167.69 (CC=ONH), 159.49 (OC=O), 159.34, 153.97, 152.22, 129.06, 123.79, 113.80, 111.99, 109.90, 109.30, 71.22 (OCH₂), 22.26, 18.73 (coumarin-4-CH₃), 10.68 ppm; HRMS (ESI) calcd. for C₁₇H₁₅NO₅S [M+H]⁺, 346.0749, found, 346.0750.

4.1.17. Synthesis of (Z)-5-((7-butoxy-4-methyl-2-oxo-2H-chromen-8-yl)methylene)thiazolidine-2,4-dione (**10c**)

According to the similar preparation of compound **8**, compound **9c** (128 mg, 0.49 mmol), thiazolidine-2,4-dione (87 mg, 0.74 mmol) were employed to produce the pure product **10c** (105 mg, 59.3%) as light yellow solid. Mp: > 250 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 12.52 (s, 1H, NH), 7.88 (d, *J* = 1.3 Hz, 1H, C=CH), 7.86–7.83 (m, 1H, coumarin-5-*H*), 7.19 (d, *J* = 9.0 Hz, 1H, coumarin-6-*H*), 6.29 (s, 1H, coumarin-3-*H*), 4.24 (t, *J* = 6.6 Hz, 2H, OCH₂(CH₂)₂CH₃), 2.41 (d, *J* = 12.1 Hz, 3H, coumarin-4-CH₃), 1.82–1.75 (m, 2H, OCH₂CH₂CH₂CH₃), 1.7–1.40 (m, 2H, O(CH₂)₂CH₂CH₃), 0.94 (dd, *J* = 12.3, 4.9 Hz, 3H, O(CH₂)₃CH₃) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 168.58 (SC=O), 167.67 (CC=ONH), 159.49 (OC=O), 159.35, 153.99, 152.22, 129.06, 123.82, 113.81, 111.98, 109.90, 109.31, 69.53 (OCH₂), 30.84, 19.08, 18.74 (coumarin-4-CH₃), 14.07 ppm; HRMS (ESI) calcd. for C₁₈H₁₇NO₅S [M+H]⁺, 360.0906, found, 360.0905.

4.1.18. Synthesis of (Z)-5-((4-methyl-2-oxo-7-(pentylloxy)-2H-chromen-8-yl)methylene)thiazolidine-2,4-dione (**10d**)

According to the similar preparation of compound **8**, compound **9d** (217 mg, 0.79 mmol), thiazolidine-2,4-dione (139 mg, 1.19 mmol) were employed to produce the pure product **10d** (175 mg, 59.4%) as yellow solid. Mp: 250 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 12.53 (s, 1H, NH), 7.87 (s, 1H, C=CH), 7.84 (d, *J* =

8.9 Hz, 1H, coumarin-5-*H*), 7.19 (d, $J = 9.0$ Hz, 1H, coumarin-6-*H*), 6.29 (s, 1H, coumarin-3-*H*), 4.22 (t, $J = 6.6$ Hz, 2H, $\text{OCH}_2(\text{CH}_2)_3\text{CH}_3$), 2.42 (s, 3H, coumarin-4- CH_3), 1.84–1.77 (m, 2H, $\text{OCH}_2\text{CH}_2(\text{CH}_2)_2\text{CH}_3$), 1.42–1.31 (m, 4H, $\text{O}(\text{CH}_2)_2(\text{CH}_2)_2\text{CH}_3$), 0.88 (t, $J = 7.2$ Hz, 3H, $\text{O}(\text{CH}_2)_4\text{CH}_3$) ppm; ^{13}C NMR (151 MHz, $\text{DMSO-}d_6$) δ 167.74 (SC=O), 159.52 (CC=ONH), 159.35 (OC=O), 154.02, 152.24, 129.01, 123.81, 113.84, 111.99, 109.97, 109.37, 69.80 (OCH_2), 28.43, 28.00, 22.22, 18.74 (coumarin-4- CH_3), 14.28 ppm; HRMS (ESI) calcd. for $\text{C}_{19}\text{H}_{19}\text{NO}_5\text{S}$ $[\text{M}+\text{Na}]^+$, 366.0882, found, 366.0880.

4.1.19. Synthesis of (Z)-5-((7-(hexyloxy)-4-methyl-2-oxo-2H-chromen-8-yl)methylene)thiazolidine-2,4-dione (**10e**)

According to the similar preparation of compound **8**, compound **9e** (195 mg, 0.68 mmol), thiazolidine-2,4-dione (119 mg, 1.02 mmol) were employed to produce the pure product **10e** (128 mg, 48.6%) as white solid. Mp: 240–242 °C; ^1H NMR (600 MHz, $\text{DMSO-}d_6$) δ 12.52 (s, 1H, NH), 7.87 (s, 1H, C=CH), 7.84 (d, $J = 9.0$ Hz, 1H, coumarin-5-*H*), 7.18 (d, $J = 9.0$ Hz, 1H, coumarin-6-*H*), 6.29 (d, $J = 1.1$ Hz, 1H, coumarin-3-*H*), 4.22 (t, $J = 6.6$ Hz, 2H, $\text{OCH}_2(\text{CH}_2)_4\text{CH}_3$), 2.42 (d, $J = 0.9$ Hz, 3H, coumarin-4- CH_3), 1.82–1.77 (m, 2H, $\text{OCH}_2\text{CH}_2(\text{CH}_2)_3\text{CH}_3$), 1.44–1.38 (m, 2H, $\text{O}(\text{CH}_2)_2\text{CH}_2(\text{CH}_2)_2\text{CH}_3$), 1.33–1.27 (m, 4H, $\text{O}(\text{CH}_2)_3(\text{CH}_2)_2\text{CH}_3$), 0.86 (t, $J = 7.1$ Hz, 3H, $\text{O}(\text{CH}_2)_5\text{CH}_3$) ppm; ^{13}C NMR (151 MHz, $\text{DMSO-}d_6$) δ 168.53 (SC=O), 167.64 (CC=ONH), 159.49 (OC=O), 159.33, 153.97, 152.24, 129.16, 128.98, 123.81, 113.81, 111.99, 109.94, 109.34, 69.80 (OCH_2), 31.72, 29.34, 29.08, 28.70, 25.79, 22.53, 18.73 (coumarin-4- CH_3), 14.36 ppm; HRMS (ESI) calcd. for $\text{C}_{20}\text{H}_{21}\text{NO}_5\text{S}$ $[\text{M}+\text{Na}]^+$, 410.1038, found, 410.1037.

4.1.20. Synthesis of (Z)-5-((7-(decyloxy)-4-methyl-2-oxo-2H-chromen-8-yl)methylene)thiazolidine-2,4-dione (**10f**)

According to the similar preparation of compound **8**, compound **9f** (215 mg, 0.62 mmol), thiazolidine-2,4-dione (109 mg, 0.93 mmol) were employed to produce the pure product **10f** (205 mg, 74.5%) as light yellow solid. Mp: 225–227 °C; ^1H NMR (600 MHz, $\text{DMSO-}d_6$) δ 12.52 (s, 1H, NH), 7.88 (s, 1H, C=CH), 7.84 (d, $J = 8.8$ Hz, 1H, coumarin-5-*H*), 7.19 (d, $J = 8.9$ Hz, 1H, coumarin-6-*H*), 6.29 (s, 1H, coumarin-3-*H*), 4.23 (t, $J = 6.1$ Hz, 2H, $\text{OCH}_2(\text{CH}_2)_8\text{CH}_3$), 2.42 (s, 3H, coumarin-4- CH_3), 1.82–1.76 (m, 2H, $\text{OCH}_2\text{CH}_2(\text{CH}_2)_7\text{CH}_3$), 1.39 (d, $J = 6.6$ Hz, 2H, $\text{O}(\text{CH}_2)_2\text{CH}_2(\text{CH}_2)_6\text{CH}_3$), 1.27 (d, $J = 44.9$ Hz, 12H, $\text{O}(\text{CH}_2)_3(\text{CH}_2)_6\text{CH}_3$), 0.85 (t, $J = 6.5$ Hz, 3H, $\text{O}(\text{CH}_2)_9\text{CH}_3$) ppm; ^{13}C NMR (151 MHz, $\text{DMSO-}d_6$) δ 168.53 (SC=O), 167.64 (CC=ONH), 159.49 (OC=O), 159.33, 153.97, 152.24, 129.16, 128.98, 123.81, 113.81, 111.99, 109.94, 109.34, 69.80 (OCH_2), 31.72, 29.34, 29.08, 28.70, 25.79, 22.53, 18.73 (coumarin-4- CH_3), 14.36 ppm; HRMS (ESI) calcd. for $\text{C}_{24}\text{H}_{29}\text{NO}_5\text{S}$ $[\text{M}+\text{H}]^+$, 444.1845, found, 444.1846.

4.1.21. Synthesis of (Z)-5-((7-(2-hydroxyethoxy)-4-methyl-2-oxo-2H-chromen-8-yl)methylene)thiazolidine-2,4-dione (**10g**)

According to the similar preparation of compound **8**, compound **9g** (51 mg, 0.21 mmol), thiazolidine-2,4-dione (37.5 mg, 0.32 mmol) were employed to produce the pure product **10g** (41 mg, 56.3%) as white solid. Mp: > 250 °C; ^1H NMR (600 MHz, $\text{DMSO-}d_6$) δ 7.71 (d, $J = 8.9$ Hz, 1H, C=CH), 7.32 (s, 1H, coumarin-5-*H*), 7.14 (d, $J = 8.9$ Hz, 1H, coumarin-6-*H*), 6.24 (s, 1H, coumarin-3-*H*), 4.19 (t, $J = 5.2$ Hz, 2H, $\text{OCH}_2\text{CH}_2\text{OH}$), 3.78 (t, $J = 5.2$

Hz, 2H, OCH₂CH₂OH), 2.42 (s, 3H, coumarin-4-CH₃) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 181.36 (SC=O), 175.98 (CC=ONH), 160.11 (OC=O), 159.25, 153.93, 151.65, 141.96, 126.42, 114.71, 113.76, 113.52, 111.75, 109.23, 71.06 (OCH₂), 59.92, 22.91 (coumarin-4-CH₃) ppm; HRMS (ESI) calcd. for C₁₆H₁₃NO₆S [M+H]⁺, 348.0536, found, 348.05301.

4.1.22. Synthesis of (Z)-5-((7-(allyloxy)-4-methyl-2-oxo-2H-chromen-8-yl)methylene)thiazolidine-2,4-dione (**12a**)

According to the similar preparation of compound **8**, compound **11a** (200 mg, 0.82 mmol), thiazolidine-2,4-dione (144 mg, 1.23 mmol) were employed to produce the pure product **12a** (212 mg, 75.4%) as yellow solid. Mp: > 250 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 12.53 (s, 1H, NH), 7.90 (s, 1H, C=CH), 7.85 (d, *J* = 9.0 Hz, 1H, coumarin-5-*H*), 7.18 (d, *J* = 9.0 Hz, 1H, coumarin-6-*H*), 6.31 (d, *J* = 1.1 Hz, 1H, coumarin-3-*H*), 6.13–6.06 (m, 1H, OCH₂CH=CH₂), 5.43 (dd, *J* = 17.3, 1.6 Hz, 1H, CH=CH₂), 5.33 (dd, *J* = 10.6, 1.4 Hz, 1H, CH=CH₂), 4.86 (d, *J* = 5.3 Hz, 2H, OCH₂CH=CH₂), 2.43 (d, *J* = 0.9 Hz, 3H, coumarin-4-CH₃) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 168.63 (SC=O), 167.75 (CC=ONH), 159.46 (OC=O), 158.77, 153.97, 152.22, 133.04 (CCH=CH), 129.28, 128.93, 123.62, 118.94 (CCH=CH), 114.00, 112.12, 110.01, 109.60, 70.06 (OCH₂), 18.74 (coumarin-4-CH₃) ppm; HRMS (ESI) calcd. for C₁₇H₁₃NO₅S [M+H]⁺, 344.0593, found, 344.0595.

4.1.23. Synthesis of (Z)-5-((4-methyl-2-oxo-7-(prop-2-yn-1-yloxy)-2H-chromen-8-yl)methylene)thiazolidine-2,4-dione (**12b**)

According to the similar preparation of compound **8**, compound **11b** (320 mg, 1.32 mmol), thiazolidine-2,4-dione (232 mg, 1.98 mmol) were employed to produce the pure product **12b** (268 mg, 59.5%) as white solid. Mp: > 250 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 12.54 (s, 1H, NH), 7.92 (d, *J* = 9.0 Hz, 1H, coumarin-5-*H*), 7.89 (s, 1H, C=CH), 7.26 (d, *J* = 9.0 Hz, 1H, coumarin-6-*H*), 6.34 (s, 1H, coumarin-3-*H*), 5.12 (d, *J* = 2.1 Hz, 2H, OCH₂), 3.70 (s, 1H, C≡CH), 2.44 (s, 3H, coumarin-4-CH₃) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 168.61 (SC=O), 167.67 (CC=ONH), 159.40 (OC=O), 157.54, 153.94, 152.23, 129.52, 128.87, 123.31, 114.50, 112.43, 110.31, 109.76, 79.94 (C≡CH), 78.56 (C≡CH), 56.99, 49.07, 18.76 (coumarin-4-CH₃) ppm; HRMS (ESI) calcd. for C₁₇H₁₁NO₅S [M+Na]⁺, 364.0256, found, 364.0258.

4.1.24. Synthesis of (Z)-3-allyl-5-((7-(allyloxy)-4-methyl-2-oxo-2H-chromen-8-yl)methylene)thiazolidine-2,4-dione (**13a**)

According to the similar preparation of compound **9a**, compound **12a** (88 mg, 0.26 mmol), 3-bromoprop-1-ene (35 mg, 0.29 mmol) and potassium carbonate (40 mg, 0.29 mmol) were employed to produce the pure product **13a** (50 mg, 50.2%) as yellow solid. Mp: > 250 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 8.06 (s, 1H, C=CH), 7.88 (d, *J* = 9.0 Hz, 1H, coumarin-5-*H*), 7.20 (d, *J* = 9.0 Hz, 1H, coumarin-6-*H*), 6.32 (d, *J* = 0.9 Hz, 1H, coumarin-3-*H*), 6.09 (ddd, *J* = 15.9, 10.6, 5.3 Hz, 1H, OCH₂CH=CH₂), 5.90–5.84 (m, 1H, NCH₂CH=CH₂), 5.43 (dd, *J* = 17.3, 1.5 Hz, 1H, OCH₂CH=CH₂), 5.33 (dd, *J* = 10.6, 1.3 Hz, 1H, OCH₂CH=CH₂), 5.20–5.16 (m, 2H, NCH₂CH=CH₂), 4.88 (d, *J* = 5.3 Hz, 2H, OCH₂CH=CH₂), 4.26 (d, *J* = 5.4 Hz, 2H, NCH₂CH=CH₂), 2.43 (d, *J* = 0.6 Hz, 3H, coumarin-4-CH₃) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 167.69 (SC=O), 165.74 (CC=ONH), 159.40 (OC=O),

158.83, 153.99, 152.34, 133.00, 131.70, 129.27, 126.81, 124.88, 119.03, 117.97, 114.06, 112.17, 109.76, 109.67, 109.53, 70.13, 43.89, 18.76 (coumarin-4-CH₃) ppm; HRMS (ESI) calcd. for C₂₀H₁₇NO₅S [M+H]⁺, 384.0906, found, 384.0906; [M+Na]⁺, 406.0725, found, 406.0726.

4.1.25. *Synthesis of (Z)-5-((4-methyl-2-oxo-7-(prop-2-yn-1-yloxy)-2H-chromen-8-yl)methylene)-3-(prop-2-yn-1-yl)thiazolidine-2,4-dione (13b)*

According to the similar preparation of compound **9a**, compound **12b** (150 mg, 0.44 mmol), 3-bromo-1-propyne (58 mg, 0.484 mmol) and potassium carbonate (61 mg, 0.44 mmol) were employed to produce the pure product **13b** (98 mg, 58.8%) as yellow solid. Mp: > 250 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 8.07 (s, 1H, C=CH), 7.95 (d, *J* = 9.0 Hz, 1H, coumarin-5-*H*), 7.28 (d, *J* = 9.0 Hz, 1H, coumarin-6-*H*), 6.35 (s, 1H, coumarin-3-*H*), 5.14 (d, *J* = 2.1 Hz, 2H, OCH₂CCH), 4.43 (d, *J* = 2.4 Hz, 2H, NCH₂CCH), 3.71 (t, *J* = 2.1 Hz, 1H, OCH₂CCH), 3.33 (t, *J* = 2.4 Hz, 1H, NCH₂CCH), 2.45 (s, 3H, coumarin-4-CH₃) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 167.14 (SC=O), 165.03 (CC=ONH), 159.31 (OC=O), 157.60, 153.95, 152.37, 129.33, 126.67, 125.21, 114.56, 112.49, 110.01, 109.82, 80.04, 78.48, 77.65, 74.98, 57.09, 31.00, 18.76 (coumarin-4-CH₃) ppm; HRMS (ESI) calcd. for C₂₀H₁₃NO₅S [M+H]⁺, 380.0593, found, 380.0594.

4.1.26. *Synthesis of (Z)-5-((7-((2-fluorobenzyl)oxy)-4-methyl-2-oxo-2H-chromen-8-yl)methylene)thiazolidine-2,4-dione (15a)*

According to the similar preparation of compound **8**, compound **14a** (232 mg, 0.74 mmol), thiazolidine-2,4-dione (130 mg, 1.11 mmol) were employed to produce the pure product **15a** (205 mg, 67.4%) as yellow solid. Mp: > 250 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 12.44 (s, 1H, NH), 7.87 (d, *J* = 8.8 Hz, 2H, coumarin-5-*H*, C=CH), 7.55 (dd, *J* = 7.5, 6.4 Hz, 1H, ph-4-*H*), 7.46–7.41 (m, 1H, ph-3-*H*), 7.32 (d, *J* = 9.0 Hz, 1H, coumarin-6-*H*), 7.28–7.22 (m, 2H, ph-5,6-2*H*), 6.30 (s, 1H, coumarin-3-*H*), 5.43 (s, 2H, OCH₂), 2.43 (s, 3H, coumarin-4-CH₃) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 168.54 (SC=O), 167.72 (CC=ONH), 161.72 (OC=O), 160.08, 159.42, 158.85, 153.96, 152.13, 131.23, 129.65, 128.99, 125.05, 123.45, 123.17, 116.02, 115.88, 114.37, 112.30, 110.44, 109.78, 65.59, 18.74 (coumarin-4-CH₃) ppm; HRMS (ESI) calcd. for C₂₁H₁₄FNO₅S [M+H]⁺, 412.0655, found, 412.0653.

4.1.27. *Synthesis of (Z)-5-((7-((4-fluorobenzyl)oxy)-4-methyl-2-oxo-2H-chromen-8-yl)methylene)thiazolidine-2,4-dione (15b)*

According to the similar preparation of compound **8**, compound **14b** (87 mg, 0.27 mmol), thiazolidine-2,4-dione (48 mg, 0.41 mmol) were employed to produce the pure product **15b** (56 mg, 50.5%) as light yellow solid. Mp: > 250 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 12.51 (s, 1H, NH), 7.90 (s, 1H, C=CH), 7.85 (d, *J* = 8.9 Hz, 1H, coumarin-5-*H*), 7.54–7.50 (m, 2H, ph-2,6-2*H*), 7.24 (dd, *J* = 16.5, 8.4 Hz, 3H, ph-3,5-2*H*, coumarin-6-*H*), 6.31 (s, 1H, coumarin-3-*H*), 5.39 (s, 2H, OCH₂), 2.42 (s, 3H, coumarin-4-CH₃) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 168.55 (SC=O), 167.69 (CC=ONH), 163.22 (OC=O), 161.60, 159.43, 158.92, 153.97, 152.14, 132.56, 130.50, 129.48, 128.94, 123.62, 115.91, 115.77, 114.18, 112.21, 110.32, 109.86, 70.38, 18.73 (coumarin-4-CH₃) ppm; HRMS (ESI) calcd. for C₂₁H₁₄FNO₅S [M+Na]⁺, 434.0474, found, 434.0474.

4.1.28. Synthesis of (Z)-5-((7-((2,4-dichlorobenzyl)oxy)-4-methyl-2-oxo-2H-chromen-8-yl)methylene)thiazolidine-2,4-dione (**15c**)

According to the similar preparation of compound **8**, compound **14c** (276 mg, 0.76 mmol), thiazolidine-2,4-dione (134 mg, 1.14 mmol) were employed to produce the pure product **15c** (185 mg, 52.9%) as yellow solid. Mp: > 250 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 12.48 (s, 1H, NH), 7.87 (d, *J* = 7.4 Hz, 2H, ph-3-*H*, C=CH), 7.70 (d, *J* = 2.0 Hz, 1H, coumarin-5-*H*), 7.59 (d, *J* = 8.3 Hz, 1H, ph-5-*H*), 7.48 (dd, *J* = 8.3 Hz, 1H, ph-6-*H*), 7.28 (d, *J* = 9.1 Hz, 1H, coumarin-6-*H*), 6.32 (d, *J* = 1.0 Hz, 1H, coumarin-3-*H*), 5.42 (s, 2H, OCH₂), 2.43 (s, 3H, coumarin-4-CH₃) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 168.47 (SC=O), 167.65 (CC=ONH), 159.38 (OC=O), 158.62, 153.91, 152.09, 134.48, 134.33, 132.83, 132.07, 129.75, 129.53, 129.00, 128.07, 123.44, 114.51, 112.39, 110.52, 109.83, 68.29, 18.73 (coumarin-4-CH₃) ppm; HRMS (ESI) calcd. for C₂₁H₁₃Cl₂NO₅S [M+H]⁺, 461.9970, found, 461.9969.

4.1.29. Synthesis of (Z)-5-((7-((3,4-dichlorobenzyl)oxy)-4-methyl-2-oxo-2H-chromen-8-yl)methylene)thiazolidine-2,4-dione (**15d**)

According to the similar preparation of compound **8**, compound **14d** (230 mg, 0.64 mmol), thiazolidine-2,4-dione (113 mg, 0.96 mmol) were employed to produce the pure product **15d** (178 mg, 60.3%) as yellow solid. Mp: > 250 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 12.46 (s, 1H, NH), 7.85–7.80 (m, 2H, ph-5-*H*, C=CH), 7.74 (d, *J* = 1.7 Hz, 1H, coumarin-5-*H*), 7.66 (d, *J* = 8.3 Hz, 1H, ph-2-*H*), 7.44 (dd, *J* = 8.3, 1.8 Hz, 1H, ph-6-*H*), 7.20 (d, *J* = 9.0 Hz, 1H, coumarin-6-*H*), 6.30 (d, *J* = 0.8 Hz, 1H, coumarin-3-*H*), 5.40 (s, 2H, OCH₂), 2.42 (s, 3H, coumarin-4-CH₃) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 159.62 (SC=O), 158.42 (CC=ONH), 153.91 (OC=O), 151.95, 137.83, 131.65, 131.15, 129.90, 128.10, 114.34, 112.25, 109.78, 69.37, 18.71 (coumarin-4-CH₃) ppm; HRMS (ESI) calcd. for C₂₁H₁₃Cl₂NO₅S [M+H]⁺, 461.9964, found, 461.99619.

4.1.30. Synthesis of (Z)-5-((4-methyl-7-((4-nitrobenzyl)oxy)-2-oxo-2H-chromen-8-yl)methylene)thiazolidine-2,4-dione (**15e**)

According to the similar preparation of compound **8**, compound **14e** (254 mg, 0.75 mmol), thiazolidine-2,4-dione (132 mg, 1.13 mmol) were employed to produce the pure product **15e** (100 mg, 30.4%) was obtained as white solid. Mp: > 250 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 12.49 (s, 1H, NH), 8.28–8.25 (m, 2H, ph-3,5-2*H*), 7.94 (d, *J* = 21.2 Hz, 1H, C=CH), 7.85 (d, *J* = 9.0 Hz, 1H, coumarin-5-*H*), 7.73–7.70 (m, 2H, ph-2,6-2*H*), 7.21 (d, *J* = 9.0 Hz, 1H, coumarin-6-*H*), 6.32 (d, *J* = 1.0 Hz, 1H, coumarin-3-*H*), 5.59 (s, 2H, OCH₂), 2.41 (d, *J* = 6.2 Hz, 3H, coumarin-4-CH₃) ppm; ¹³C NMR (151 MHz, DMSO-*d*₆) δ 159.45 (SC=O), 159.05 (CC=ONH), 158.48 (OC=O), 153.95, 152.11, 147.70, 144.25, 128.76, 128.60, 125.98, 124.09, 114.43, 112.36, 112.02, 109.80, 109.37, 69.79, 18.73 (coumarin-4-CH₃) ppm; HRMS (ESI) calcd. for C₂₁H₁₄N₂O₇S [M+H]⁺, 439.0600, found, 439.0601.

Conflicts of interest

There are no conflicts to declare.

Acknowledgments

This research was supported in part by grants from the National Natural Science Foundation of China (Nos. 21672173, 21971212, 21850410447), China Postdoctoral Science Foundation (2019M653821XB), Chongqing Special Foundation for Postdoctoral Research Proposal (No. XmT2018082).

References

- [1] (a) S.L. Silver, Challenges of antibacterial discovery, *Clin. Microbiol. Rev.* 24 (2011) 71–109. doi: 10.1128/CMR.00030-10; (b) E.D. Brown, G.D. Wright, Antibacterial drug discovery in the resistance era, *Nature* 529 (2016) 336–343. doi: 10.1038/nature17042.
- [2] J.M. Stokes, C.R. MacNair, B. Ilyas, S. French, J.P. Cote, C. Bouwman, E.D. Brown, Pentamidine sensitizes Gram negative pathogens to antibiotics and overcomes acquired colistin resistance, *Nat. Microbiol.* 2 (2017) 17028. doi: 10.1038/nmicrobiol.2017.28
- [3] (a) B. Mishra, G.S. Wang, Ab initio design of potent anti-MRSA peptides based on database filtering technology, *J. Am. Chem. Soc.* 134 (2012) 12426–12429. doi: 10.1021/ja305644e; (b) Z.Z. Li, V.K.R. Tangadanchu, N. Battini, R.R.Y. Bheemanaboina, Z.L. Zang, S.L. Zhang, C.H. Zhou, Indole-nitroimidazole conjugates as efficient manipulators to decrease the genes expression of methicillin-resistant *Staphylococcus aureus*, *Eur. J. Med. Chem.* 179 (2019) 723–735. doi.org/10.1016/j.ejmech.2019.06.093.
- [4] (a) T.L. Harris, R.J. Worthington, C. Melander, Potent small-molecule suppression of oxacillin resistance in methicillin-resistant *Staphylococcus aureus*, *Angew. Chem. Int. Ed.* 51 (2012) 11254–11257. doi: 10.1002/anie.201206911; (b) L.L. Wang, N. Battini, R.R.Y. Bheemanaboina, M.F. Ansari, J.P. Chen, Y.P. Xie, G.X. Cai, S.L. Zhang, C.H. Zhou, A new exploration towards aminothiazolquinolone oximes as potentially multi-targeting antibacterial agents: design, synthesis and evaluation acting on microbes, DNA, HSA and topoisomerase IV, *Eur. J. Med. Chem.* 179 (2019) 166–181. doi.org/10.1016/j.ejmech.2019.06.046.
- [5] N.A. Turner, B.K. Sharma-Kuinkel, S.A. Maskarinec, E.M. Eichenberger, P.P. Shah, M. Carugati, T.L. Holland, V.G. Fowler Jr, Methicillin-resistant *Staphylococcus aureus*: an overview of basic and clinical research, *Nat. Rev. Microbiol.* 17 (2019) 203–218. doi: 10.1038/s41579-018-0147-4.
- [6] S.F. Cui, D. Addla, C.H. Zhou, Novel 3-aminothiazolquinolones: design, synthesis, bioactive evaluation, SARs, and preliminary antibacterial mechanism, *J. Med. Chem.* 59 (2016) 4488–4510. doi: 10.1021/acs.jmedchem.5b01678.
- [7] (a) W. Li, Y.N. Sun, X.T. Yan, S.Y. Yang, E.J. Kim, H.K. Kang, Y.H. Kim, Coumarins and lignans from *Zanthoxylum schinifolium* and their anticancer activities, *J. Agric. Food Chem.* 61 (2013) 10730–10740. doi: 10.1021/jf403479c; (b) Y. Hu, Y.F. Shen, X.H. Wu, X. Tu, G.X. Wang, Synthesis and biological evaluation of coumarin derivatives containing imidazole skeleton as potential antibacterial agents, *Eur. J. Med. Chem.* 143 (2018) 958–969. doi: 10.1016/j.ejmech.2017.11.100.
- [8] (a) L.L. Ling, T. Schneider, A.J. Peoples, A.L. Spoering, I. Engels, B.P. Conlon, A. Mueller, T.F. Schäberle, D.E. Hughes, S. Epstein, M. Jones, L. Lazarides, V.A. Steadman, D.R. Cohen, C.R. Felix, K.A. Fetterman, W.P. Millett, A.G. Nitti, A.M. Zullo, C. Chen, K. Lewis, Erratum: A new antibiotic kills pathogens without detectable resistance, *Nature* 517 (2015) 455–459. doi: 10.1038/nature14303;
- [9] (a) H. Liu, Z.L. Ren, W. Wang, J.X. Gong, M.J. Chu, Q.W. Ma, J.C. Wang, X.H. Lv, Novel coumarin-pyrazole carboxamide derivatives as potential topoisomerase II inhibitors: design, synthesis and antibacterial activity, *Eur. J. Med. Chem.* 157 (2018) 81–87. doi: 10.1016/j.ejmech.2018.07.059; (b) P. Andriollo, C.K. Hind, P. Picconi, K.S. Nahar, S. Jamshidi, A. Varsha, M.

- Clifford, J.M. Sutton, K.M. Rahman, C8-linked pyrrolbenzodiazepine monomers with inverted building blocks show selective activity against multidrug resistant Gram-positive bacteria, *ACS Infect. Dis.* 4 (2018) 158–174. doi: 10.1021/acsinfecdis.7b00130.
- [10] (a) X.M. Peng, G.L.V. Damu, C.H. Zhou, Coumarin-derived azolyl ethanol: synthesis, antimicrobial evaluation and preliminary action mechanism, *Curr. Pharm. Des.* 19 (2013) 3884–3930. doi: 10.1007/s11426-015-0351-0; (b) L. Zhang, X.M. Peng, G.L.V. Damu, R.X. Geng, C.H. Zhou, Comprehensive review in current developments of imidazole-based medicinal chemistry, *Med. Res. Rev.* 34 (2014) 340–437. doi: 10.1002/med.21290.
- [11] (a) Y. Shi, C.H. Zhou, Synthesis and evaluation of a class of new coumarin triazole derivatives as potential antimicrobial agents, *Bioorg. Med. Chem. Lett.* 21 (2011) 956–960. doi:10.1016/j.bmcl.2010.12.059.
- [12] (a) B. Li, R. Pai, M. Di, D. Aiello, M.H. Barnes, M.M. Butler, T.F. Tashjian, N.P. Peet, T.L. Bowlin, D.T. Moir, Coumarin-based inhibitors of bacillus anthracis and staphylococcus aureus replicative DNA helicase: chemical optimization, biological evaluation, and antibacterial activities, *J. Med. Chem.* 55 (2012) 10896–10908. doi: 10.1021/jm300922h; (b) E. Jameel, T. Umar, J. Kumar, N. Hoda, Coumarin: a privileged scaffold for the design and development of antineurodegenerative agents, *Chem. Biol. Drug. Des.* 87 (2016) 21–38. doi.org/10.1111/cbdd.12629.
- [13] (a) X.M. Peng, K.V. Kumar, G.L.V. Damu, C.H. Zhou, Coumarin-derived azolyl ethanol: synthesis, antimicrobial evaluation and preliminary action mechanism study, *Sci. China Chem.* 59 (2016) 878–894. doi: 10.1007/s11426-015-0351-0.
- [14] (a) S.L. Badshah, A. Ullah, New developments in non-quinolone-based antibiotics for the inhibition of bacterial gyrase and topoisomerase IV, *Eur. J. Med. Chem.* 152 (2018) 393–400. doi: 10.1016/j.ejmech.2018.04.059; (b) C.H. Zhou, Y. Wang, Recent researches in triazole compounds as medicinal drugs, *Curr. Med. Chem.* 19 (2012) 239–280. doi: 10.2174/092986712803414213.
- [15] (a) G.L.V. Damu, S.F. Cui, X.M. Peng, Q.M. Wen, G.X. Cai, C.H. Zhou. Synthesis and bioactive evaluation of a novel series of coumarinazoles. *Bioorg. Med. Chem. Lett.* 24 (2014) 3605–3608. doi: 10.1016/j.bmcl.2014.05.029; (b) M.E. Riveiro, K.N. De, A. Moglioni, R. Vazquez, F. Monczor, C. Shayo, C. Davio, Coumarins: old compounds with novel promising therapeutic perspectives, *Curr. Med. Chem.* 17 (2010) 1325–1338. doi: 10.2174/092986710790936284.
- [16] (a) N. Chadha, M.S. Bahia, M. Kaur, O. Silakari, Thiazolidine-2,4-dione derivatives: programmed chemical weapons for key protein targets of various pathological conditions, *Bioorg. Med. Chem.* 23 (2015) 2953–2974. doi: 10.1016/j.bmc.2015.03.071; (b) X.M. Peng, G.X. Cai, C.H. Zhou, Recent developments in azole compounds as antibacterial and antifungal agents, *Curr. Top. Med. Chem.* 13 (2013) 1963–2010. doi: 10.2174/15680266113139990125.
- [17] A.M. Youssef, M.S. White, E.B. Villanueva, I.M. El-Ashmawy, A. Klegeris, Synthesis and biological evaluation of novel pyrazolyl-2,4-thiazolidinediones as anti-inflammatory and neuroprotective agents, *Bioorg. Med. Chem.* 5 (2010) 2019–28. doi: 10.1016/j.bmc.2010.01.021.
- [18] (a) Y. Wu, S. Karna, C.H. Choi, M. Tong, H. Tai, D.H. Na, C.H. Jang, H. Cho, Synthesis and biological evaluation of novel thiazolidinedione analogues as 15-hydroxyprostaglandin dehydrogenase inhibitors, *J. Med. Chem.* 54 (2011) 5260–5264. doi: 10.1021/jm200390u; (b) W.W. Gao, L. Gopala, R.R.Y. Bheemanaboina, G.B. Zhang, S. Li, C.H. Zhou, Discovery of 2-aminothiazolyl berberine derivatives as effectively antibacterial agents toward clinically drug-resistant Gram-negative acinetobacter baumannii, *Eur. J. Med. Chem.* 146 (2018) 15–37. doi: 10.1016/j.ejmech.2018.01.038.
- [19] L. Ma, C.F. Xie, Y.H. Ma, J. Liu, M.L. Xiang, X. Ye, H. Zheng, Z.Z. Chen, Q.Y. Xu, T. Chen, J.Y. Chen, J.C. Yang, N. Qiu,

- G.C. Wang, X.L. Liang, A.H. Peng, S.Y. Yang, Y.Q. Wei, L.J. Chen, Synthesis and biological evaluation of novel 5-benzylidenethiazolidine-2,4-dione derivatives for the treatment of inflammatory diseases, *J. Med. Chem.* 54 (2011) 2060–2080. doi: 10.1021/jm1011534.
- [20] (a) P. Sharma, T.S. Reddy, D. Thummuri, K.R. Senwar, N.P. Kumar, V.G.M. Naidu, S.K. Bhargava, N. Shankaraiah, Synthesis and biological evaluation of new benzimidazole-thiazolidinedione hybrids as potential cytotoxic and apoptosis inducing agents, *Eur. J. Med. Chem.* 124 (2016) 608–621. doi: 10.1016/j.ejmech.2016.08.029; (b) X.F. Liu, C.J. Zheng, L.P. Sun, X.K. Liu, H.R. Piao, Synthesis of new chalcone derivatives bearing 2,4-thiazolidinedione and benzoic acid moieties as potential anti-bacterial agents, *Eur. J. Med. Chem.* 46 (2011) 3469–3473. doi: 10.1016/j.ejmech.2011.05.012.
- [21] K. Metwally, H. Pratsinis, D. Kletsas, Novel 2,4- thiazolidinediones: Synthesis, in vitro cytotoxic activity, and mechanistic investigation, *Eur. J. Med. Chem.* 133 (2017) 340–350. doi: 10.1016/j.ejmech.2017.03.052.
- [22] (a) T. Tomašić, N. Zidar, M.M. Premru, D. Kikelj, L.P. Mašič, Synthesis and antibacterial activity of 5-ylidenethiazolidin-4-ones and 5-benzylidene-4,6-pyrimidinediones, *Eur. J. Med. Chem.* 45 (2010) 1667–1672. doi: 10.1016/j.ejmech.2009.12.030; (b) Y.Y. Hu, R.R.Y. Bheemanaboina, N. Battini, C.H. Zhou, Sulfonamide-derived four-component molecular hybrids as novel DNA-targeting membrane active potentiators against clinical escherichia coli, *Mol. Pharm.* 16 (2019) 1036–1052. doi: 10.1021/acs.molpharmaceut.8b01021.
- [23] M.A. Farha, E.D. Brown, Strategies for target identification of antimicrobial natural products, *Nat. Prod. Rep.* 33 (2016) 668–680. doi: 10.1039/c5np00127g.
- [24] G.B. Zhang, S.K. Maddili, V.K.R. Tangadanchu, L. Gopala, W.W. Gao, G.X. Cai, C.H. Zhou, Discovery of natural berberine-derived nitroimidazoles as potentially multi-targeting agents against drug-resistant escherichia coli, *Sci. China Chem.* 61 (2018) 557–568. doi: 10.1007/s11426-017-9169-4.
- [25] H. Sun, M.F. Ansari, N. Battini, R.R.Y. Bheemanaboina, C.H. Zhou, Novel potential artificial MRSA DNA intercalators: synthesis and biological evaluation of berberine-derived thiazolidinediones, *Org. Chem. Front.* 6 (2019) 319–334. doi: 10.1039/c8qo01180j.
- [26] Y. Cheng, S.R. Avula, W.W. Gao, D. Addla, V.K.R. Tangadanchu, L. Zhang, J.M. Lin, C.H. Zhou, Multi-targeting exploration of new 2-aminothiazolyl quinolones: synthesis, antimicrobial evaluation, interaction with DNA, combination with topoisomerase IV and penetrability into cells, *Eur. J. Med. Chem.* 124 (2016) 935–945. doi: 10.1016/j.ejmech.2016.10.011.
- [27] Z.Z. Li, L. Gopala, V.K.R. Tangadanchu, W.W. Gao, C.H. Zhou, Discovery of novel nitroimidazole enols as pseudomonas aeruginosa DNA cleavage agents, *Bioorg. Med. Chem.* 25 (2017) 6511–6522. doi: 10.1016/j.bmc.2017.10.028.
- [28] P. Jeyakkumar, H.B. Liu, L. Gopala, Y. Cheng, X.M. Peng, R.X. Geng, C.H. Zhou, Novel benzimidazolyl tetrahydroprotoberberines: design, synthesis, antimicrobial evaluation and multi-targeting exploration, *Bioorg. Med. Chem. Lett.* 27 (2017) 1737–1743. doi: 10.1016/j.bmcl.2017.02.071.
- [29] J. Kang, L. Gopala, V.K.R. Tangadanchu, W.W. Gao, C.H. Zhou, Novel naphthalimide nitroimidazoles as multitargeting antibacterial agents against resistant acinetobacter baumannii, *Future Med. Chem.* 10 (2018) 711–724. doi: 10.4155/fmc-2017-0160.
- [30] W.W. Gao, S. Rasheed, V.K.R. Tangadanchu, Y. Sun, X.M. Peng, Y. Cheng, F.X. Zhang, J.M. Lin, C.H. Zhou, Design, synthesis and biological evaluation of amino organophosphorus imidazoles as a new type of potential antimicrobial agents, *Sci. China Chem.* 60 (2017) 769–785. doi: 10.1007/s11426-016-9009-6.

- [31] S.K. Maddili, Z.Z. Li, K.V. Kumar, R.R.Y. Bheemanaboinab, B. Tunikic, V.K.R. Tangadanchua, C.H. Zhou, Azoalkyl ether imidazo[2,1-b]benzothiazoles as potentially antimicrobial agents with novel structural skeleton, *Bioorg. Med. Chem. Lett.* 28 (2018) 2426–2431. doi: 10.1016/j.bmcl.2018.06.016.
- [32] H.B. Liu, W.W. Gao, V.K.R. Tangadanchu, C.H. Zhou, R.X. Geng, Novel aminopyrimidinyl benzimidazoles as potentially antimicrobial agents: design, synthesis and biological evaluation, *Eur. J. Med. Chem.* 143 (2018) 66–84. doi.org/10.1016/j.ejmech.2017.11.027.
- [33] L.L. Wang, N. Battini, R.R.Y. Bheemanaboina, S.L. Zhang, C.H. Zhou, Design and synthesis of aminothiazolyl norfloxacin analogues as potential antimicrobial agents and their biological evaluation, *Eur. J. Med. Chem.* 167 (2019) 105–123. doi.org/10.1016/j.ejmech.2019.01.072.
- [34] Y. Zhang, V.K.R. Tangadanchu, R.R.Y. Bheemanaboina, Y. Cheng, C.H. Zhou, Novel carbazole-triazole conjugates as DNA-targeting membrane active potentiators against clinical isolated fungi, *Eur. J. Med. Chem.* 155 (2018) 579–589. doi: 10.1016/j.ejmech.2018.06.022.
- [35] (a) H.Z. Zhang, P. Jeyakkumar, K.V. Kumar, C.H. Zhou, Synthesis of novel sulfonamide azoles via C–N cleavage of sulfonamides by azole ring and relational antimicrobial study, *New. J. Chem.* 39 (2015) 5776–5796. doi: 10.1039/c4nj01932f; (b) D. Li, R.R.Y. Bheemanaboina, N. Battini, V.K.R. Tangadanchu, X.F. Fang, C.H. Zhou, Novel organophosphorus aminopyrimidines as unique structural DNA-targeting membrane active inhibitors towards drug-resistant methicillin-resistant *Staphylococcus aureus*, *Med. Chem. Commun.* 9 (2018) 1529–1537. doi: 10.1039/c8md00301g.
- [36] Y.Y. Chen, L. Gopala, R.R.Y. Bheemanaboina, H.B. Liu, Y. Cheng, R.X. Geng, C.H. Zhou, Novel naphthalimide aminothiazoles as potential multitargeting antimicrobial agents, *ACS Med. Chem. Lett.* 8 (2017) 1331–1335. doi: 10.1021/acsmchemlett.7b00452.
- [37] W.Y. Lin, L.L. Long, J.B. Feng, B. Wang, C.C. Guo, Synthesis of meso-coumarin-conjugated porphyrins and investigation of their luminescence properties, *Eur. J. Org. Chem.* 26 (2007) 4301–4304. doi: 10.1002/ejoc.200700475.
- [38] C. Krishna, M.V. Bhargavi, Y.J. Rao, G.L.D. Krupadanam, Synthesis of pyrano isoxazoline/isoxazole annulated coumarins *via* intramolecular nitrile oxide cycloaddition and their cytotoxicity, *Russ. J. Gen. Chem.* 87 (2017) 1857–1863. doi: 10.1134/S1070363217080345.
- [39] J. Tang, K.D. Qian, B.N. Zhang, Y. Chen, P. Xia, D.L. Yu, Y. Xia, Z.Y. Yang, C.H. Chen, S.L. Morris-Natschke, K.H. Lee, Anti-AIDS agents 82: synthesis of seco-(3'R,4'R)-3',4'-di-O-(S)-camphanoyl-(+)-cis-khellactone (DCK) derivatives as novel anti-HIV agents, *Bioorg. Med. Chem.* 18 (2010) 4363–4373. doi:10.1016/j.bmc.2010.04.089.

Lists of table and scheme captions

Table 1. Antibacterial data as MIC ($\mu\text{mol/mL}$) for compounds **8**, **4a–b**, **10a–f**, **12a–c**, **13a–b** and **15a–e**.

Figure 1. Drug molecules containing coumarin scaffold.

Figure 2. Design of novel coumarin thiazolidinediones.

Figure 3. The crystal structure of compound **13a**.

Figure 4. Time–kill kinetics of compound **12b** ($4 \times \text{MIC}$) against MRSA.

Figure 5. Drug resistance development of compound **12b** against MRSA.

Figure 6. Cytotoxic assay of target compound **12b** on human hepatocyte LO2 cells tested by MTT methodology.

Figure 7. Membrane permeabilization of resistant bacterial MRSA toward compound **12b** ($12 \times \text{MIC}$).

Figure 8. UV absorption spectra of MRSA DNA with different concentrations of compound **12b** ($\text{pH} = 7.4$, $T = 290 \text{ K}$). Inset: comparison of absorption at 260 nm between the value of compound **12b**-DNA complex and the sum values of free DNA and free compound **12b**. $c(\text{DNA}) = 7.4 \times 10^{-5} \text{ mol/L}$, and $c(\text{compound } \mathbf{12b}) = 0\text{--}2.0 \times 10^{-5} \text{ mol/L}$ for curves *a–i* respectively at an increment 0.25×10^{-5} .

Figure 9. UV absorption spectra of the competitive reaction between **12b** and NR with MRSA DNA. $c(\text{DNA}) = 7.4 \times 10^{-5} \text{ mol/L}$, $c(\text{NR}) = 2 \times 10^{-5} \text{ mol/L}$, and $c(\text{compound } \mathbf{12b}) = 0\text{--}4.5 \times 10^{-5} \text{ mol/L}$ for curves *a–i* respectively at increment 0.5×10^{-5} . (Inset) Absorption spectra of the system with the increasing concentration of **12b** in the wavelength range of 350–600 nm absorption spectra of competitive reaction between compound **12b** and NR with DNA.

Figure 10. Molecular modeling of compound **12b** docked in gyrase-DNA.

Scheme 1 Synthetic route of coumarin thiazolidinediones **1–8**.

Scheme 2 Synthetic route of coumarin thiazolidinediones **9–15e**.

Table 1. *In vitro* antibacterial data as MIC ($\mu\text{mol/mL}$) for target compounds **8**, **4a–b**, **10a–g**, **12a–b**, **13a–b** and **15a–e**.^a

Comps	Gram-positive bacteria ^b					Gram-negative bacteria ^c					
	MRSA	<i>E. f.</i>	<i>S. a.</i>	<i>S. a.</i> 25923	<i>S. a.</i> 29213	<i>K. p.</i>	<i>E. c.</i>	<i>P. a.</i>	<i>A. b.</i>	<i>P. a.</i> 27853	<i>E. c.</i> 25922
8	0.886	0.886	0.886	1.772	1.772	0.886	1.772	0.886	1.772	0.886	0.443
4a	1.772	0.886	0.443	1.772	1.772	1.772	1.772	1.772	0.886	1.772	0.886
4b	0.422	0.106	0.422	0.422	0.422	0.422	0.422	0.211	0.422	0.422	0.422
10a	0.012	0.773	0.193	0.773	0.773	0.387	0.773	0.387	0.387	0.773	0.773
10b	0.012	0.742	0.012	0.742	0.742	0.371	0.742	0.371	0.371	0.371	0.742
10c	1.426	1.426	1.426	1.426	1.426	1.426	1.426	1.426	1.426	1.426	1.426
10d	0.343	0.686	1.372	1.372	0.686	0.343	1.372	0.343	0.686	1.372	1.372
10e	0.331	0.331	0.661	0.331	0.661	0.331	0.331	0.661	0.331	0.331	0.661
10f	1.155	1.155	1.155	1.155	1.155	1.155	1.155	1.155	1.155	1.155	1.155
10g	0.369	0.184	0.369	0.369	0.369	0.369	0.369	0.184	0.184	0.369	0.184
12a	0.006	0.746	0.023	0.746	0.746	0.373	0.746	0.373	0.373	0.373	0.746
12b	0.006	0.751	0.375	0.751	0.047	0.047	0.751	0.188	0.188	0.188	0.375
13a	0.167	0.668	0.334	0.668	0.668	0.334	0.668	0.668	0.334	0.668	0.334
13b	0.338	0.338	0.338	0.675	0.338	0.675	0.675	0.338	0.675	0.675	0.338
15a	0.039	0.156	0.311	0.039	0.311	0.623	0.311	0.311	0.311	0.156	0.311
15b	0.311	0.156	0.311	0.156	0.311	0.311	0.623	0.311	0.156	0.156	0.311
15c	0.278	0.278	0.555	0.278	0.278	0.139	0.278	0.278	0.278	0.278	0.139
15d	0.139	0.278	0.278	0.278	0.278	0.278	0.278	0.278	0.139	0.278	0.139
15e	0.584	0.292	0.292	0.146	0.584	0.292	0.292	0.584	0.292	0.292	0.584
A^d	0.025	0.006	0.025	0.100	0.003	0.401	0.200	0.025	0.802	0.013	0.025

^aMinimum inhibitory concentrations were determined by micro broth dilution method for microdilution plates. ^bMRSA, methicillin-resistant *Staphylococcus aureus* N315; *E. f.*, *Enterococcus faecalis*; *S. a.*, *Staphylococcus aureus*; *S. a.* 25923, *Staphylococcus aureus* ATCC 25923; *S. a.* 29213, *Staphylococcus aureus* ATCC 29213; ^c*K. p.*, *Klebsiella pneumonia*; *E. c.*, *Escherichia coli*; *P. a.*, *Pseudomonas aeruginosa*; *A. b.*, *Acinetobacter baumannii*; *P. a.* 27853, *Pseudomonas aeruginosa* ATCC 27853; *E. c.* 25922, *Escherichia coli* ATCC 25922. ^dA = Norfloxacin

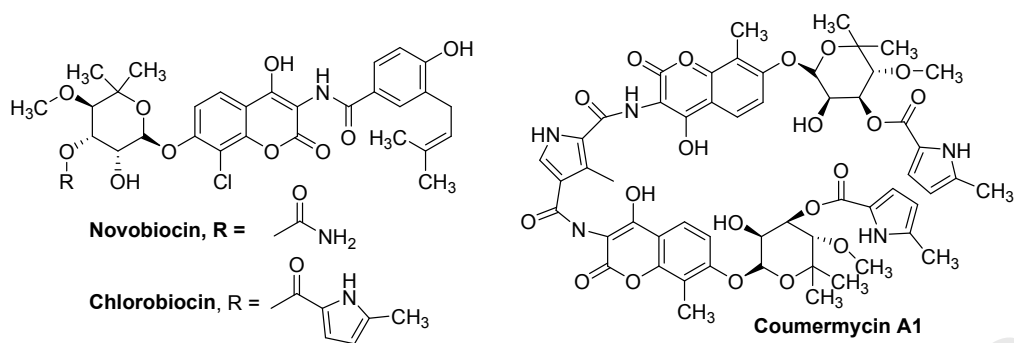
Fig. 1. Drug molecules containing coumarin scaffold.

Fig. 2. Design of novel coumarin thiazolidinediones.

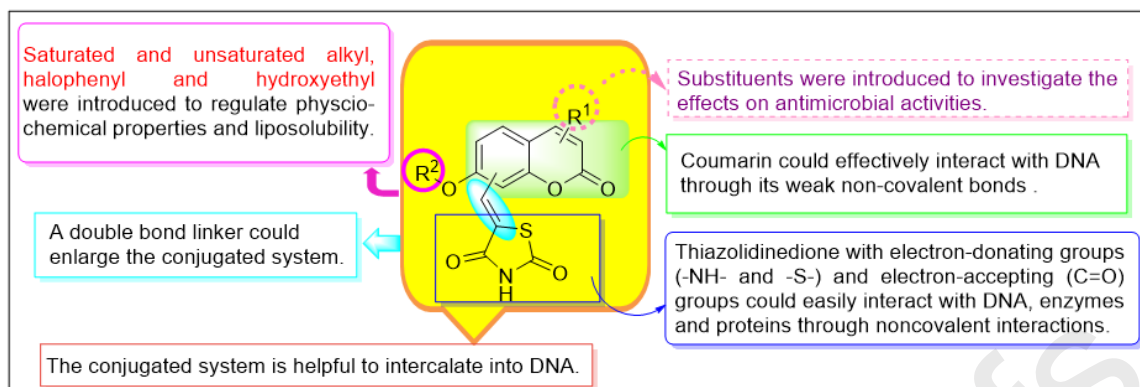


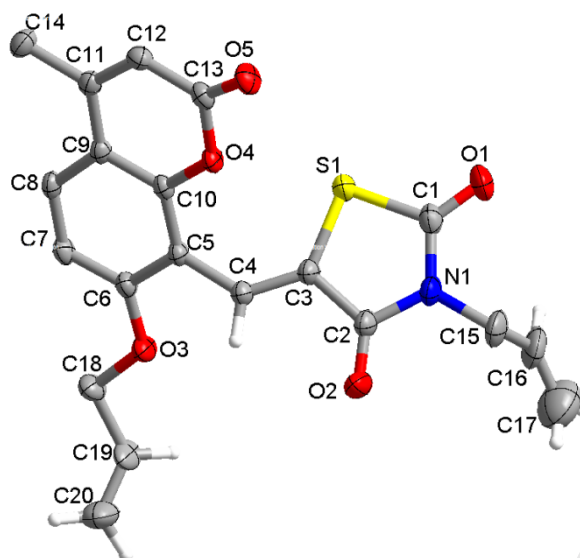
Fig. 3. The crystal structure of compound **13a**.

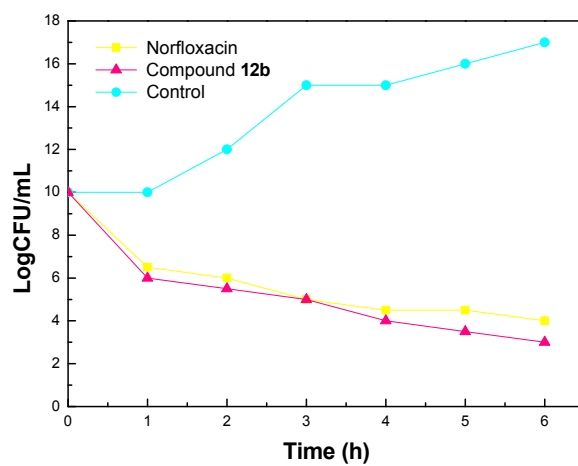
Fig. 4. Time-kill kinetics of compound **12b** ($4 \times \text{MIC}$) against MRSA.

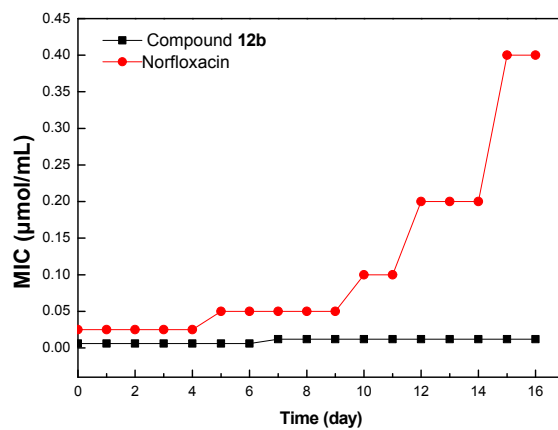
Fig. 5. Drug resistance development of compound **12b** against MRSA.

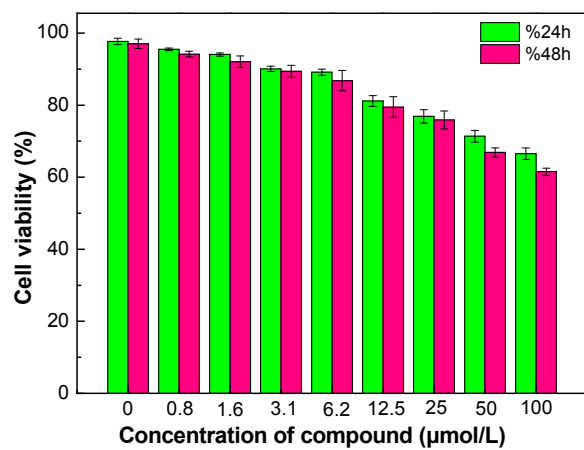
Fig. 6. Cytotoxic assay of target compound **12b** on human hepatocyte LO2 cells tested by MTT methodology.

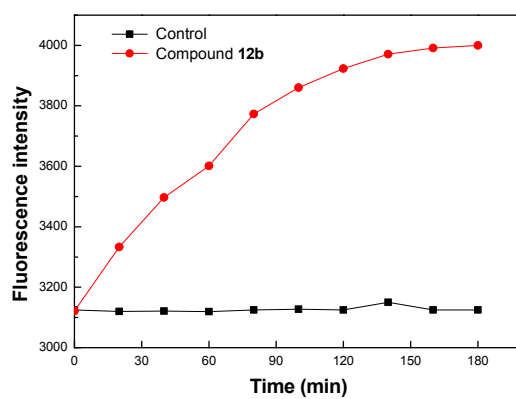
Fig. 7. Membrane permeabilization of resistant bacterial MRSA toward compound **12b** ($12 \times \text{MIC}$).

Fig. 8. UV absorption spectra of MRSA DNA with different concentrations of compound **12b** (pH = 7.4, T = 290 K). Inset: comparison of absorption at 260 nm between the value of compound **12b**–DNA complex and the sum values of free DNA and free compound **12b**. $c(\text{DNA}) = 7.4 \times 10^{-5} \text{ mol/L}$, and $c(\text{compound } \mathbf{12b}) = 0\text{--}2.0 \times 10^{-5} \text{ mol/L}$ for curves *a*–*i* respectively at an increment 0.25×10^{-5} .

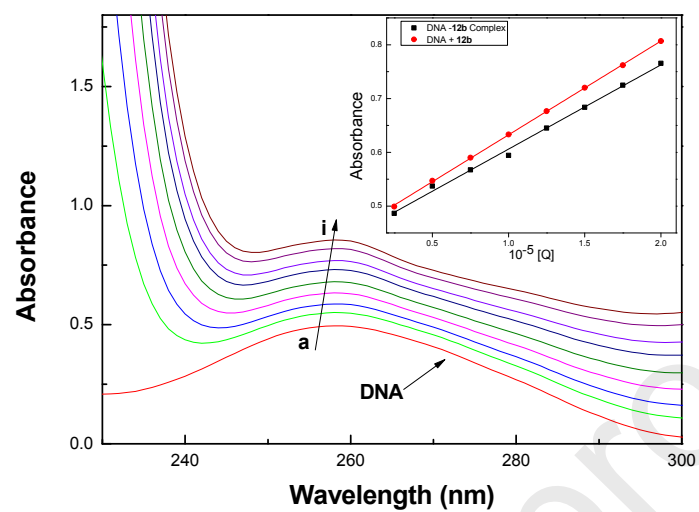


Fig. 9. UV absorption spectra of the competitive reaction between **12b** and NR with MRSA DNA. $c(\text{DNA}) = 7.4 \times 10^{-5} \text{ mol/L}$, $c(\text{NR}) = 2 \times 10^{-5} \text{ mol/L}$, and $c(\text{compound } \mathbf{12b}) = 0-4.5 \times 10^{-5} \text{ mol/L}$ for curves a-i respectively at increment 0.5×10^{-5} . (Inset) Absorption spectra of the system with the increasing concentration of **12b** in the wavelength range of 350–600 nm absorption spectra of competitive reaction between compound **12b** and NR with DNA.

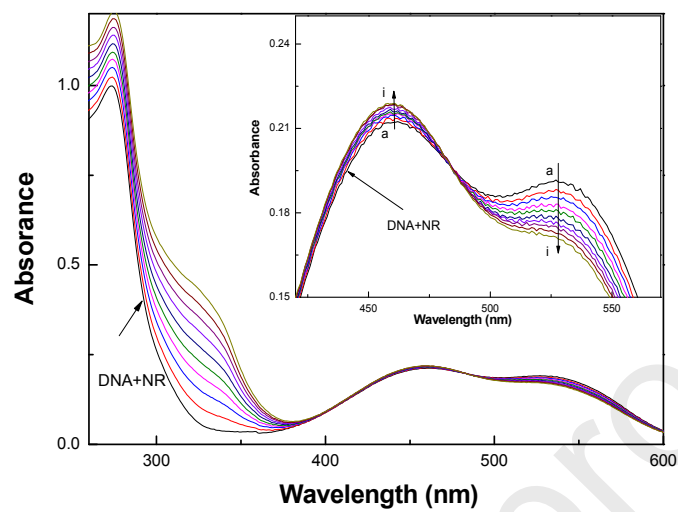
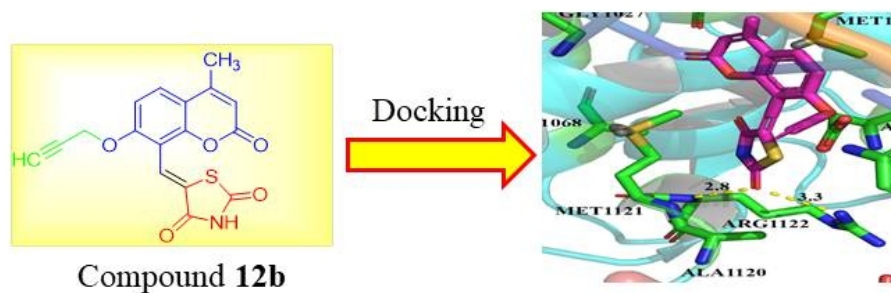
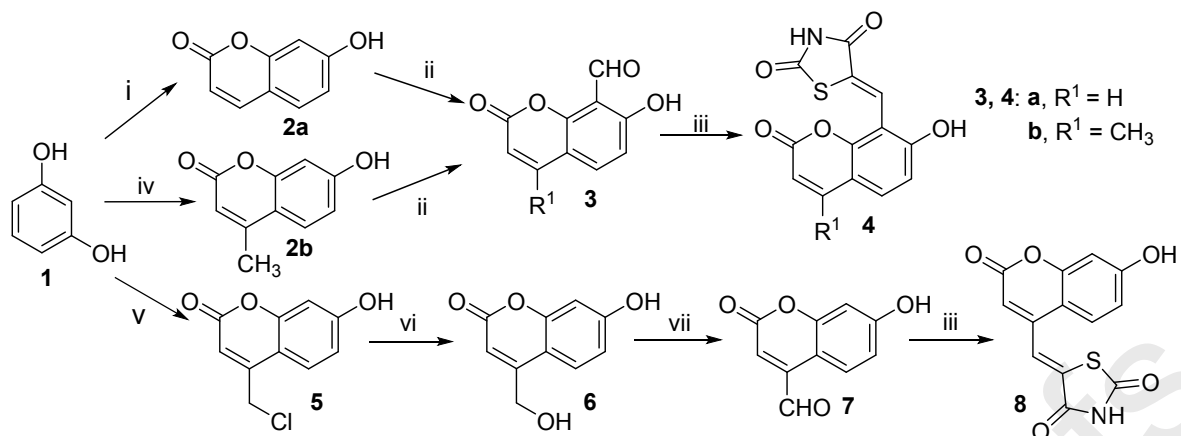


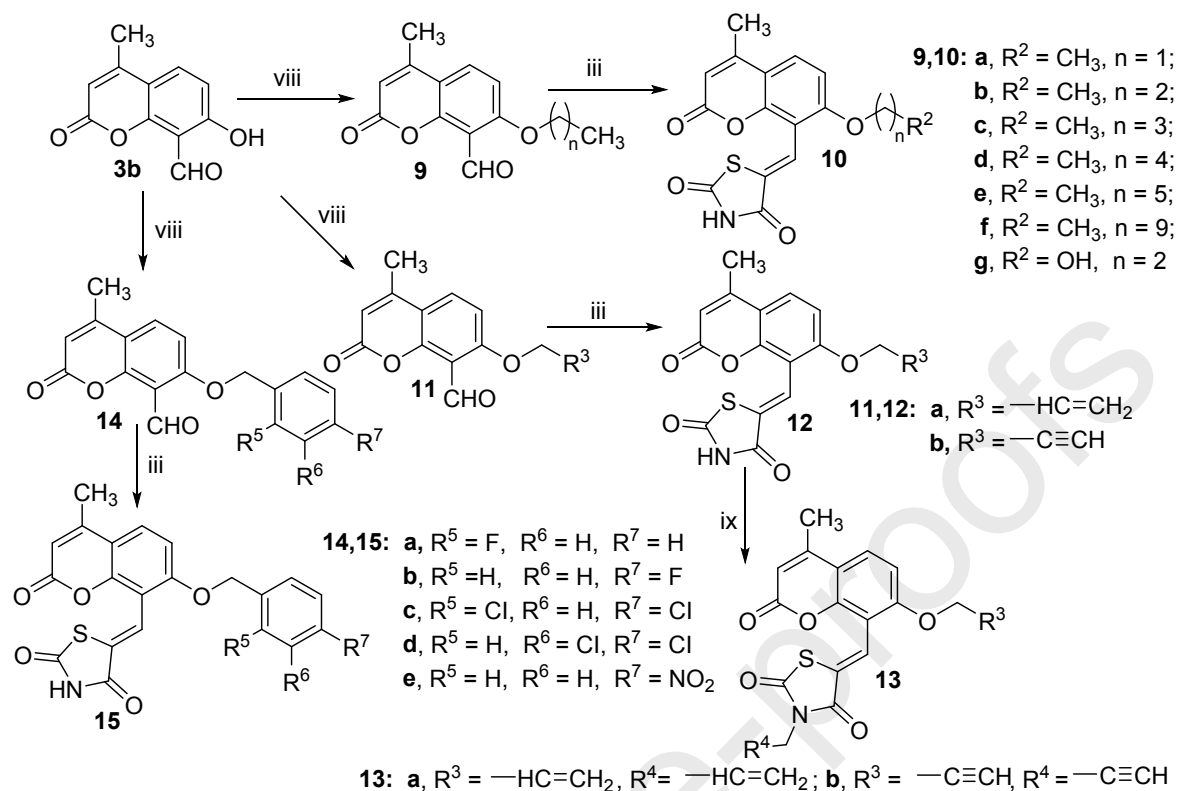
Fig. 10. Molecular modeling of compound **12b** docked in gyrase-DNA (gyrase-DNA complex code: 2XCS).

Scheme 1 Synthetic route of coumarin thiazolidinediones 1–8.



Reagents and conditions: (i) 2-hydroxysuccinic acid; (ii) (a) HMTA/AcOH, 75 °C, 8 h; (b) 10% HCl, 75 °C, 1 h; (iii) thiazolidinedione, EtOH, piperidine, N₂, 80 °C; (iv) H₂SO₄, ethylacetoacetate, 0 °C, 24 h; (v) ethyl 4-chloroacetoacetate, H₂SO₄, 0 °C, 24 h; (vi) H₂O, 100 °C, 3 days; (vii) MnO₂, ethyl acetate, N₂, 80 °C, 7 h.

Scheme 2 Synthetic route of coumarin thiazolidinediones 9–15.



Reagents and conditions: (iii) thiazolidinedione, EtOH, piperidine, N_2 , 80 °C, 6 h; (viii) aliphatic bromides, benzyl halides, 2-bromoethanol, potassium carbonate, DMF, 80 °C, 4–8 h; (ix) aliphatic bromides, potassium carbonate, DMF, 80 °C, 4 h.

Graphical Abstract

Ethylenic conjugated coumarin thiazolidinediones as efficient antimicrobial modulators against clinical methicillin-resistant *Staphylococcus aureus*

Author Names and Affiliations:

Chun-Fang Hu^a, Peng-Li Zhang^a, Yan-Fei Sui^a, Jing-Song Lv^a, Mohammad Fawad Ansari^{a,†}, Narsaiah Battini^{a,‡}, Shuo Li^{b,*}, Cheng-He Zhou^{a,*} and Rong-Xia Geng^{a,*}

^a Institute of Bioorganic & Medicinal Chemistry, Key Laboratory of Luminescent and Real-Time Analytical Chemistry (Southwest University), Ministry of Education, School of Chemistry and Chemical Engineering, Southwest University, Tiansheng street 2, Beibei district, Chongqing 400715, PR China.

^b School of Chemical Engineering, Chongqing University of Technology, Chongqing 400054, PR China.

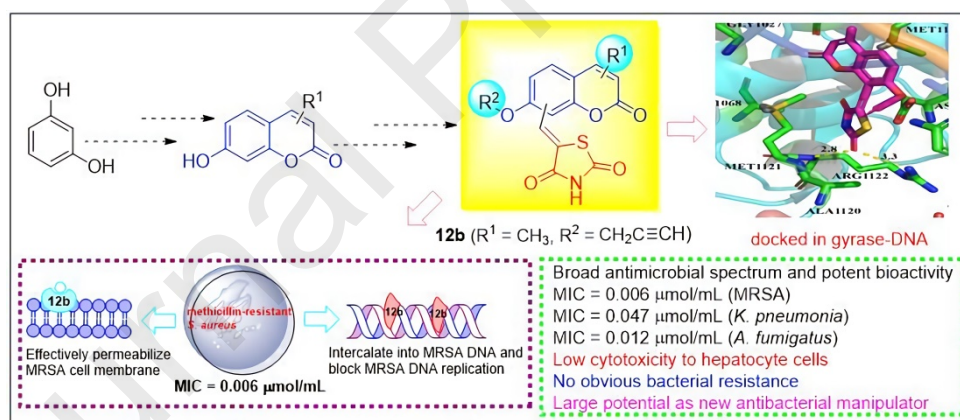
Corresponding Address:

* Tel.: +86-23-68254967; Fax: +86-23-68254967; E-mail: zhouch@swu.edu.cn (Cheng-He Zhou); geng0712@swu.edu.cn (Rong-Xia Geng); Lishuo@cqut.edu.cn (Shuo Li).

[†] Postdoctoral fellow from Jamia Millia Islamia, Delhi, India

[‡] Postdoctoral fellow from CSIR-Indian Institute of Integrative Medicine, Jammu, India

A series of novel ethylenic conjugated coumarin thiazolidinediones were synthesized and screened for their antimicrobial activities.



- ▶ Novel ethylenic conjugated coumarin thiazolidinediones with good antimicrobial potency were developed.
- ▶ Compound **12b** showed excellent inhibitory efficacy against MRSA and *A. fumigatus* with MIC values of 0.006 $\mu\text{mol/mL}$ and 0.012 $\mu\text{mol/mL}$, respectively.
- ▶ Compound **12b** with low toxicity did not trigger the resistance development in bacteria.
- ▶ Molecular docking rationalized the antimicrobial activity.
- ▶ Compound **12b** could effectively permeabilize MRSA cell membrane and intercalate into DNA to block MRSA DNA replication.

Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

Journal Pre-proofs