

A concise synthesis of β -sitosterol and other phytosterols

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ARTICLE INFO

Article history:

Received 25 February 2010
Received in revised form 6 May 2010
Accepted 22 May 2010
Available online 31 May 2010

Keywords:

Sitosterol
Campesterol
Phytosterol
Synthesis
Epoxide
Deoxygenation

ABSTRACT

A convenient synthesis of sidechain-modified phytosterols is achieved via a temporary masking of the stigmaterol 5,6-alkene as an epoxide. Following performance of the desired modification, the alkene is regenerated through a mild deoxygenation. The approach is applied to the syntheses of β -sitosterol and campesterol acetate, and suggests a facile route to the (*Z*)-isomers of Δ^{22-23} phytosterols.

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1. Introduction

Phytosterols and their derivatives are widely applied in the food and cosmetic industries, and have recently received a great deal of attention as nutraceutical additives [1–3]. Phytosterols have also attracted attention as inhibitors of sarcoplasmic reticulum calcium ATPase and potassium ion channels [4,5]. As part of a collaboration investigating the structural influences on uptake and processing of sterol esters [6], we required semipreparative amounts of β -sitosterol. However, β -sitosterol is commercially available in preparative amounts only as mixtures with other phytosterols, including stigmaterol, campesterol, and/or brassicasterol (Fig. 1); reported separations are relatively laborious [7,8].

Two routes have been reported for the synthesis of β -sitosterol from stigmaterol, which is available in pure form. The first, selective hydrogenation of the sidechain Δ^{22-23} alkene [9], was found to produce β -sitosterol contaminated with varying amounts of recovered stigmaterol as well as the fully saturated stigmastanol [10]. The second approach, which has been applied to the synthesis of sitosterol and related sterols (Fig. 2), circumvents the need for selective hydrogenation by protecting the Δ^{5-6} alkene as a cyclopropyl carbinyl ether [11,12]. Following hydrogenation of the Δ^{22-23} double bond, solvolysis of the cyclopropane reintroduces both the C_3 -alcohol and the Δ^{5-6} alkene. Although we found the latter approach very useful as a means of obtaining very pure sam-

ples of β -sitosterol, semipreparative applications were challenging in terms of removal of sterol methyl ether byproducts.

We now report a new strategy for the synthesis of sidechain-modified phytosterols based upon protection of the Δ^{5-6} alkene as an epoxide. The approach is illustrated with syntheses of β -sitosterol and campesterol acetate.

2. Experimental

2.1. General experimental procedures

All₃ and Cu(MnO₄)₂ were prepared by literature procedures [13,14]. All other reagents and solvents were used as supplied commercially, except CH₂Cl₂ (CaH₂) and THF (Na, Ph₂CO) which were distilled from the indicated reagent under an atmosphere of N₂. Melting points are uncorrected. Unless noted, NMR spectra were acquired at 400 MHz (¹H) or 100 MHz (¹³C) in CDCl₃; individual peaks are reported as: multiplicity, integration, coupling constant in Hz. IR spectra were recorded as neat films on a ZnSe crystal with selected absorbances reported in cm⁻¹. Mass spectroscopy was conducted at the Nebraska Center for Mass Spectrometry.

2.2. Stigmaterol acetate (2)

Stigmaterol acetate was prepared as a white solid (97%, mp 138–140 °C) by a variant of the procedure of Wang et al. [15]. Other physical and spectral data were identical to literature values.

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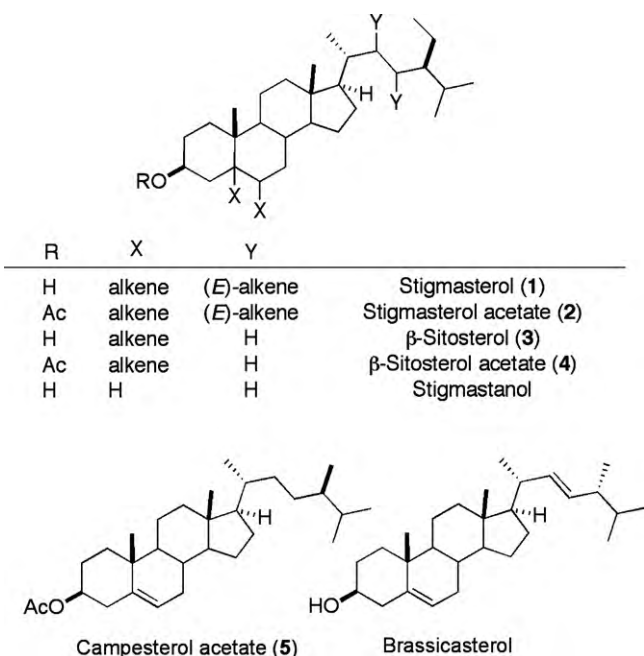


Fig. 1. Structural relationship of phytosterols.

2.3. $5\alpha,6\alpha$ - and $5\beta,6\beta$ -Epoxides of stigmasterol acetate (**6a**, **6b**)

A mixture of KMnO_4 (10 g, 60 mmol) and $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ (5.0 g, 20 mmol) was finely ground in a mortar and pestle [14]. Water (0.5 mL) was added, and the slightly wet mixture was transferred to the reaction flask. To the stirred suspension of this mixture in 25 mL CH_2Cl_2 was added stigmasterol acetate (**2**, 2.12 g, 4.51 mmol), followed by *t*-BuOH (2.5 mL). The reaction was heated to reflux for 1 h and cooled to room temperature. The reaction mixture was filtered through a silica pad, which was washed with ether. The residue obtained after concentration was recrystallized from CH_3OH to give a white solid (1.59 g, 75%) with mp 125–126 °C. NMR data indicated the product was a 1:6 mixture of the α - (**6a**) and β - (**6b**) epoxides of stigmasterol acetate [14]. Repeating this reaction with 2.23 g of stigmasterol acetate afforded 1.80 g (78%) of a 1:6 mixture of **6a** and **6b**.

Approach to $5\alpha,6\alpha$ - and $5\beta,6\beta$ -epoxides of stigmasterol acetate (**6a**, **6b**) via peracid epoxidation: to a 0 °C solution of **2** (0.308 g, 0.66 mmol) in CH_2Cl_2 (20 mL) was added mCPBA (0.170 g, 0.76 mmol). After 4 h at 0 °C, the reaction was diluted with sat. aq. K_2CO_3 (80 mL) and the aqueous layer was extracted with CH_2Cl_2 (3 \times 50 mL). The combined organic layers were dried over anhydrous Na_2SO_4 and the filtrate was concentrated in vacuo. The residue was purified by flash chromatography (hexane/EtOAc, 90:10) to afford 0.257 g (83%) of a white solid which was a 2.6:1 mixture of α - (**6a**) and β -isomers (**6b**) according to ^1H NMR.

2.4. $5\alpha,6\alpha$ - and $5\beta,6\beta$ -Epoxy sitosterol acetate (**7a**, **7b**)

To a solution of the 1:6 mixture of **6a** and **6b** (1.35 g, 3.0 mmol) in EtOAc (150 mL) was added 10% Pd/C (0.32 g), and the stirred reaction mixture was placed under an atmosphere of H_2 (balloon) for 12 h. The reaction mixture was filtered through a Celite pad, and the filtrate evaporated to furnish white solid (1.28 g, 96%, mp 113–114 °C) as a 1:6 mixture of epoxides **7a** and **7b** [8]. Repeating this reaction on 1.76 g of **6a/6b** afforded 1.75 g (99%) of a 1:6 mixture of **7a** and **7b**.

2.5. β -Sitosterol acetate (**4**)

The 1:6 mixture of epoxides **7a** and **7b** (470 mg, 1.0 mmol) was dissolved in 2:1 $\text{CH}_3\text{CN}/\text{CH}_2\text{Cl}_2$ (30 mL). Aluminum triiodide was added (610 mg, 1.5 mmol) and the resulting mixture was stirred at room temperature for 10 min. The reaction was quenched with aq. 10% $\text{Na}_2\text{S}_2\text{O}_3$ (100 mL) and the resulting mixture was extracted with CH_2Cl_2 (3 \times 100 mL). The combined organic layers were dried over Na_2SO_4 , and the residue from the concentrated filtrate was purified by flash chromatography (hexane/EtOAc, 95:5) to give 360 mg (80%) of **4** as a white solid: mp 111–112 °C, $[\alpha]_D = -34.5$ (CHCl_3 , $c = 1.0$). Other physical data were identical to values reported in the literature [11]. Repeating this reaction on 1.70 g of **7a/7b** afforded 1.40 g (85%) of **4**.

2.6. β -Sitosterol (**3**)

To a solution of β -sitosterol acetate (**4**, 240 mg, 0.47 mmol) in 1:1 $\text{CH}_3\text{OH}:\text{CH}_2\text{Cl}_2$ (30 mL) was added K_2CO_3 (140 mg, 1.01 mmol). The reaction mixture was stirred at room temperature for 12 h and then concentrated under vacuum. The residue was extracted with 30 mL CH_2Cl_2 . The organic layer was washed with 30 mL water and dried over Na_2SO_4 . The filtered organic layer was concentrated and the residue was purified through flash chromatography (hexane/EtOAc, 80:20) to give 220 mg (93%) of β -sitosterol **3** as a white solid. Mp 134–135 °C, $[\alpha]_D = -37.0$ (CHCl_3 , $c = 1.0$). Elemental analysis calculated for $\text{C}_{29}\text{H}_{50}\text{O}$: C 83.60, H 11.96; found: 83.99, 12.15. Other spectral properties were identical to values reported in the literature [11]. Repeating this reaction on 1.35 g of **4** afforded 1.20 g (98%) of beta sitosterol (**3**).

2.7. (*S*)-2,3-Dimethylbutan-1-ol (**8**)

(*S*)-2,3-Dimethylbutan-1-ol **8** was prepared as a colorless liquid (overall yield 60%, $[\alpha]_D = 4.4$, CHCl_3 , $c = 1.0$) by the procedure of Tietze, affording a product with spectral data identical to literature values [16].

2.8. (*S*)-2-(2,3-Dimethylbutylthio)benzothiazole (**9**)

To a mixture of dimethylbutanol **8** (102 mg, 1.00 mmol), 2-mercaptobenzothiazole (183 mg, 1.10 mmol) and PPh_3 (288 mg, 1.10 mmol) in freshly distilled THF (4 mL) was added diisopropyl

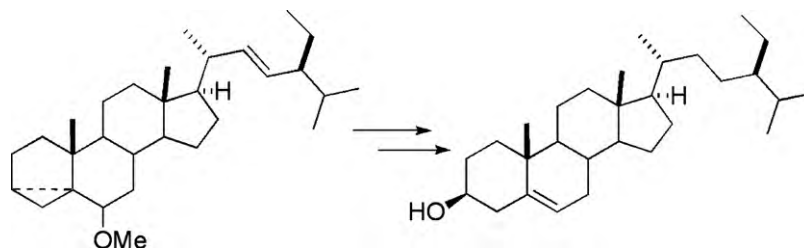


Fig. 2. Selective saturation of cyclopropyl carbonyl ether.

azodicarboxylate (DIAD, 0.21 mL, 1.10 mmol) dropwise at 0 °C under argon. The reaction was stirred for 3 h at 0 °C and then quenched with water. The aqueous layer was extracted with EtOAc (3 × 10 mL) and the combined organic layers were dried over anhydrous Na₂SO₄. The filtered organic layer was concentrated in vacuo and the residue purified by flash chromatography (hexane/EtOAc, 99:1) to afford thioether **9** (228 mg, 91%) as a light yellow oil. [α]_D = 42.3 (CHCl₃, c = 1.6); IR 2957, 1455, 1426, 1057, 991, 752 cm⁻¹; ¹H NMR: δ 7.89 (d, *J* = 8.1, 1H), 7.75 (d, *J* = 8.1, 1H), 7.42 (t, *J* = 7.2, 1H), 7.29 (t, *J* = 7.2, 1H), 3.50 (dd, *J* = 12.7, 4.8, 1H), 3.18 (dd, *J* = 12.7, 8.2, 1H), 1.88–1.77 (m, 2H), 1.04 (d, *J* = 6.7, 3H), 0.99 (d, *J* = 6.6, 3H), 0.94 (d, *J* = 6.6, 3H); ¹³C NMR: δ 167.75, 153.38, 135.15, 125.99, 124.08, 121.44, 120.91, 38.87, 38.78, 31.62, 20.37, 17.96, 15.29; HRFAB-MS (*m/z*) [M–H]⁺ calcd for [C₁₃H₁₈NS₂]⁺: 252.0881, found: 252.0875.

2.9. (S)-2-(2,3-Dimethylbutylsulfonyl)benzothiazole (**10**)

A 0 °C solution of **9** (183 mg, 0.73 mmol) in EtOH (10 mL) was oxidized with ammonium heptamolybdate tetrahydrate (1.8 g, 1.46 mmol) and 30% H₂O₂ (2.5 mL, 21.9 mmol) for 2 h. The mixture was extracted with EtOAc (3 × 10 mL) and the combined organic extracts were washed with brine (3 × 10 mL). The dried organic layers were filtered and the residue obtained upon concentration was purified by flash chromatography (hexane/EtOAc, 90:10) to afford sulfone **10** (177 mg, 86%) as a pale yellow oil. [α]_D = 15.5 (CHCl₃, c = 3.4); IR 2961, 1470, 1324, 1140, 1085, 758 cm⁻¹; ¹H NMR: δ 8.23 (d, *J* = 7.9, 1H), 8.03 (d, *J* = 7.9, 1H), 7.68–7.59 (m, 2H), 3.59 (dd, *J* = 14.4, 3.5, 1H), 3.31 (dd, *J* = 14.1, 8.9, 1H), 2.29–2.19 (m, 1H), 1.82–1.73 (m, 1H), 1.10 (d, *J* = 6.9, 3H), 0.89 (d, *J* = 6.8, 3H), 0.85 (d, *J* = 6.9, 3H); ¹³C NMR: δ 166.66, 152.70, 136.74, 128.00, 127.67, 125.43, 122.38, 58.83, 38.68, 32.47, 19.23, 17.89, 15.93; HRFAB-MS (*m/z*) [M–H]⁺ calcd for [C₁₃H₁₈NO₂S₂]⁺: 284.0779, found: 284.0778.

2.10. (3 β ,5 α ,6 α)- and (3 β ,5 β ,6 β)-Pregnane-20 α -carboxaldehyde-5,6-epoxy-3-yl acetate (**11a**, **11b**)

A –78 °C solution of **6a**, **6b** (~1:6 mixture, 100 mg, 0.21 mmol) in 10 mL of 50/50 CH₂Cl₂/MeOH was treated with a gaseous stream of ozone (2% O₃/O₂) for 5 min. The solution was purged with pure oxygen and then solvent was removed under vacuum. The residue was redissolved in 10 mL of 10/90 H₂O/AcOH and treated with zinc powder (55 mg, 0.84 mmol). The reaction mixture was stirred for 2 h at room temperature and then extracted with 50 mL CH₂Cl₂. The organic layer was washed with water (3 × 25 mL), then dried over anhydrous Na₂SO₄. The filtered organic layer was concentrated and the residue purified by flash chromatography (hexane/EtOAc, 90:10) to afford a 1:6 mixture of epoxides **11a** and **11b** as a white solid (81 mg, 99%), mp 87–8 °C. IR: 2950, 1727, 1367, 1262, 1238, 1042, 783 cm⁻¹; ¹H NMR: δ 9.57 (d, *J* = 3.3, 0.76H, β), 9.55 (d, *J* = 3.3, 0.16H, α), 4.99–4.91 (m, 0.14H, α), 4.81–4.73 (m, 0.87H, β), 3.09 (d, *J* = 2.2, 0.88H, β), 2.90 (d, *J* = 4.2, 0.13H, α), 2.38–2.31 (m, 1H), 2.13–1.82 (m, 9H), 1.54–0.89 (m, 20H), 0.7 (s, 3H); ¹³C NMR: δ 204.95, 170.52, 71.25, 63.41, 62.48, 55.39, 51.05, 50.93, 49.42, 42.89, 39.43, 37.95, 36.67, 35.06, 32.41, 29.74, 27.17, 26.97, 24.54, 21.84, 21.30, 17.03, 13.40, 12.11; HRFAB-MS (*m/z*) [M–Li]⁺ calcd for [C₂₄H₃₆LiO₄]⁺: 395.2774, found: 395.2778.

2.11. (3 β ,5 α ,6 α ,22Z)- and (3 β ,5 β ,6 β ,22Z)-Ergost-5,6-epoxy-22-en-3-yl acetate (**12a**, **12b**)

To a 78 °C solution of sulfone **10** (62 mg, 0.22 mmol) in THF (5 mL) was dropwise added LiHMDS (0.22 mL, nominally 1 M in THF, 0.22 mmol). The reaction was stirred for 1 h, whereupon the

mixture of aldehydes **11a** and **11b** (~1:6, 85 mg, 0.22 mmol) was added in 5 mL of THF. Stirring was continued for 1 h, and reaction was gradually warmed to room temperature. The reaction was quenched by 15 mL water and extracted with EtOAc (3 × 10 mL). The combined organic layers were dried over anhydrous Na₂SO₄ and filtered. The residue obtained upon concentration was purified by flash chromatography (hexane/EtOAc, 95:5) to afford a 1:16 mixture of epoxides **12a** and **12b** as a white solid (90 mg, 90%), mp 145–147 °C. IR: 2950, 2867, 1743, 1368, 1037, 764 cm⁻¹; ¹H NMR: δ 5.02 (dd, *J* = 10.9, 9.9, 2H), 4.83–4.73 (m, 1H), 3.09 (d, *J* = 2.0, 0.95H, β), 2.91 (d, *J* = 4.4, 0.06, α), 2.43–2.33 (m, 1H), 2.21–1.80 (m, 9H), 1.68–0.83 (m, 31H), 0.69 (s, 3H); ¹³C NMR: δ 170.56, 135.13, 131.22, 71.34, 63.58, 62.52, 56.27, 56.02, 51.02, 42.18, 39.72, 38.32, 38.01, 36.68, 35.04, 34.48, 33.35, 32.43, 29.74, 28.32, 27.20, 24.15, 21.92, 21.34, 20.55, 20.36, 19.94, 18.63, 17.06, 12.06; HRFAB-MS (*m/z*) [M–H]⁺ calcd for [C₃₀H₄₉O₃]⁺: 457.3682, found: 457.3668.

2.12. 5 α ,6 α - and 5 β ,6 β -Epoxides of campesterol acetate (**13a**, **13b**)

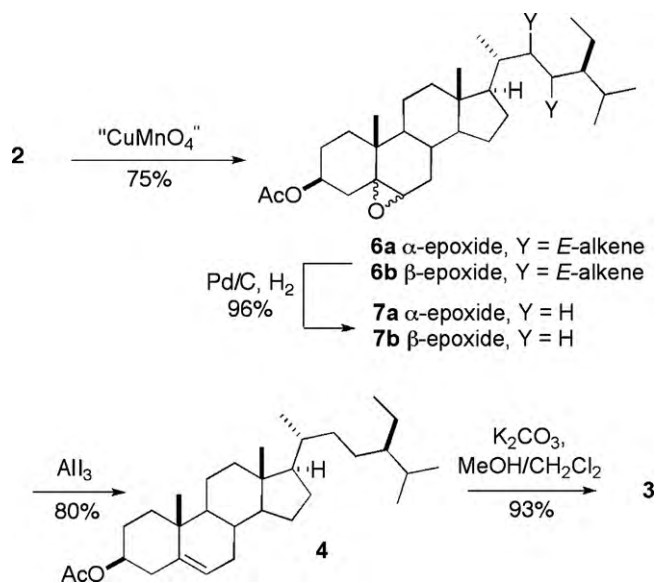
The mixture of epoxides **12a** and **12b** (30 mg, 0.065 mmol) was dissolved in 5 mL EtOAc. 10% Pd/C (7 mg) was added, and the reaction mixture was stirred at room temperature under an atmosphere of H₂ (balloon) for 12 h. The reaction mixture was filtered through a Celite pad, and the filtrate evaporated to a white solid (28 mg, 94%), mp 110–111 °C as 1:9 mixture of epoxides **13a** and **13b**. IR: 2953, 2867, 1729, 1367, 1263, 1043, 784 cm⁻¹; ¹H NMR: δ 5.01–4.93 (m, 0.15H, α), 4.83–4.73 (m, 0.96H, β), 3.09 (d, *J* = 2.1, 0.9H, β), 2.90 (d, *J* = 4.4, 0.1H, α), 2.12–1.8 (m, 8H), 1.58–0.77 (m, 37H), 0.65 (s, 3H); ¹³C NMR: δ 170.54, 71.34, 63.58, 62.51, 56.19, 56.14, 50.97, 42.28, 39.78, 38.81, 38.01, 36.66, 35.82, 35.03, 33.65, 32.47, 32.41, 30.26, 29.73, 28.14, 27.21, 24.18, 21.92, 21.31, 20.19, 18.66, 18.24, 17.03, 15.37, 11.76; HRFAB-MS (*m/z*) [M–H]⁺ calcd for [C₃₀H₅₁O₃]⁺: 459.3838, found: 459.3820.

2.13. Campesterol acetate (**5**)

The mixture of epoxides **13a** and **13b** (28 mg, 0.061 mmol) was dissolved in 2:1 CH₃CN/CH₂Cl₂ (3 mL). Aluminum triiodide (37 mg, 0.091 mmol) was added and the resulting mixture was stirred at room temperature for 40 min. The reaction was quenched with aq. 10% Na₂S₂O₃ (10 mL) and the resulting mixture was extracted with CH₂Cl₂ (3 × 10 mL). The combined organic layers were dried over anhydrous Na₂SO₄, and the residue from the concentrated filtrate was purified by flash chromatography (hexane/EtOAc, 95:5) to give 24 mg (91%) campesterol acetate (**5**) as a white solid. Mp 130–131 °C, [α]_D = –32 (CHCl₃, c = 0.7); IR 2954, 1730, 1367, 1247, 1037, 735 cm⁻¹; ¹H NMR: δ 5.39 (d, *J* = 4.8, 1H), 4.66–4.58 (m, 1H), 2.33 (d, *J* = 7.9, 2H), 2.05 (s, 3H), 1.90–1.84 (m, 2H), 1.59–0.78 (m, 38H), 0.68 (s, 3H); ¹³C NMR: δ 170.56, 139.66, 122.66, 73.99, 56.69, 56.08, 50.02, 42.31, 39.73, 38.84, 38.12, 36.99, 36.59, 35.90, 33.70, 32.43, 31.90, 31.86, 30.27, 28.24, 27.78, 24.29, 21.46, 21.03, 20.22, 19.32, 18.70, 18.26, 15.38, 11.87; HRFAB-MS (*m/z*) [M–Na]⁺ calcd for [C₃₀H₅₀O₂Na]⁺: 465.3709, found: 465.3703. Elemental analysis calculated for C₃₀H₅₀O₂: C 81.20, H 11.38; found: 81.39, 11.39. The ¹H NMR data matched that of a literature report [17].

3. Results and discussion

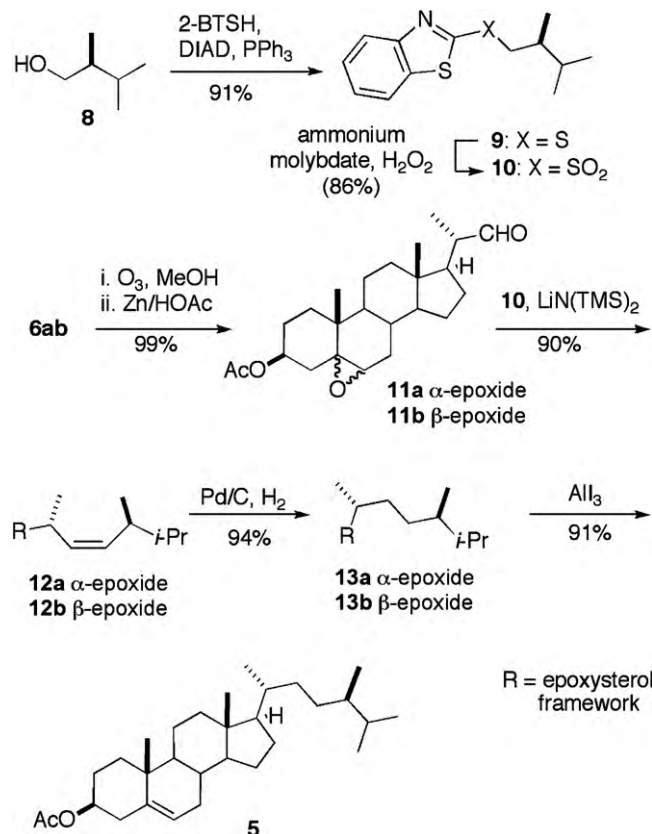
Our synthesis of β -sitosterol (**3**) is illustrated in Scheme 1. Selective epoxidation of the Δ^{5-6} alkene of stigmasterol acetate (**2**) with copper permananganate formed a 6:1 mixture of the 5 β ,6 β - and 5 α ,6 α -epoxides **6b** and **6a** [14,18]. Hydrogenation over Pd/C cleanly furnished a 1:6 mixture of sitosterol epoxides **7a** and **7b**. Deoxygenation of the saturated epoxides with AlI₃ [13] proceeded



Scheme 1. Synthesis of sitosterol.

rapidly to furnish a good yield of β -sitosteryl acetate **4**. Saponification afforded pure β -sitosterol (**3**), with mp 134–135 °C and $[\alpha]_D = -37.0$ [8,19].

Epoxidation with the commercially available peracid mCPBA also gave good selectivity for the Δ^{5-6} alkene, but now produced a 2.6:1 mixture of stigmaterol oxides favoring the α -isomer (**6a**). Hydrogenation proceeded uneventfully to furnish the corresponding mixture of sitosterol oxides **7a** and **7b**. However, attempted deoxygenation under the same conditions as employed



Scheme 2. Synthesis of campesterol acetate.

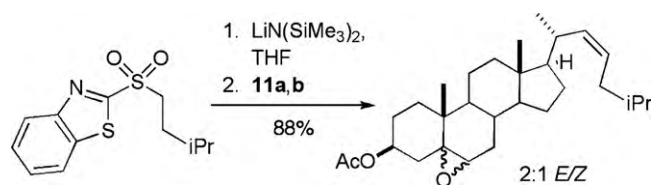


Fig. 3. Generality of Z-selective olefination.

earlier (AlI₃, 10 min, CH₃CN/CH₂Cl₂) now furnished only 33% of β -sitosterol acetate (**4**), accompanied by a significant amount (estimated >60% by mass) of a more polar product which yellowed immediately upon exposure to room light. The formation of the byproduct could be avoided almost completely by allowing the deoxygenation to proceed for 40 min. Alternatively, the byproduct could be converted to **4** by treatment with additional AlI₃. The results suggest that the deoxygenation of the α - and β -epoxides proceeds at very different rates, with the 5 α ,6 α diastereomer (**6a**) reacting via the intermediacy of a semistable iodohydrin.

As illustrated in Scheme 2, our strategy also provides a facile means of preparing other sidechain-modified phytosterols. For example, ozonolysis of the mixture of **6a/6b** furnished an approximately 1:6 mixture of aldehydes **11a** and **11b**. Julia-Kocienski olefination, using an enantiomerically pure sulfone (**10**) derived from (*S*)-2,3-dimethylbutanol (**8**) [16] furnished exclusively alkene **12** [20], corresponding to the monoepoxide of the *Z*-isomer of crinosterol [21]. Hydrogenation, followed by deoxygenation of the epoxide as before, furnished campesterol acetate (**5**).

The selective formation of *Z*-alkenes is unusual in Julia couplings [22] and we investigated the olefination of **11a** and **11b** with a known sulfone derived from isobutyl alcohol (Fig. 3) [23]. This reaction also selectively furnished the *Z*-alkene; the lower selectivity (2:1) compared with that observed for the synthesis of **12a** and **12b** may reflect the reduced degree of steric encumbrance in this model system. The ability to readily prepare *Z*-isomers of phytosterols opens the door to a number of steroid analogs not available from synthetic routes based upon Claisen rearrangements of C₂₂ allylic alcohols [12].

4. Conclusions

The formation of β -sitosterol has been achieved in 52% overall yield from commercially available stigmaterol using relatively simple chemistry and via easily purified intermediates. The core strategy, protection of the Δ^{5-6} alkene as an epoxide, holds potential for the synthesis of other phytosterols as well as unnatural analogs.

Acknowledgements

We thank Professor Tim Carr (University of Nebraska-Lincoln, Department of Nutrition and Health Sciences) for useful discussions. This research was supported by a USDA-NRI competitive grant (2007-35200-18298). Portions of this work were conducted in facilities remodeled with support from NIH (RR016544-01). NMR spectra were acquired, in part, on spectrometers purchased with NSF support (MRI 0079750, CHE 0091975).

Appendix A. Supplementary data

Supplementary data for this article, consisting of ¹H and ¹³C NMR spectra for compounds **3–13**, can be found in the online version, at doi:10.1016/j.steroids.2010.05.016.

References

- [1] Moghadasian MH. Pharmacological properties of plant sterols: *in vivo* and *in vitro* observations. *Life Sci* 2000;67:605–15.
- [2] Piironen V, Lindsay DG, Miettinen TA, Toivo J, Lampi AM. Plant sterols: biosynthesis, biological function and their importance to human nutrition. *J Sci Food Agric* 2000;80:939–66.
- [3] Ling WH, Jone PJH. Dietary phytosterols: a review of metabolism, benefits and side effects. *Life Sci* 1995;57:195–206.
- [4] Bao L, Li Y, Deng SX, Landry D, Tabas I. Sitosterol-containing lipoproteins trigger free sterol-induced caspase-independent death in ACAT-competent macrophages. *J Biol Chem* 2006;281:33635–49.
- [5] Promprom W, Kupittayanant P, Indrapichate K, Wray S, Kupittayanant S. The effects of pomegranate seed extract and β -sitosterol on rat uterine contractions. *Reprod Sci* 2010;17:288–96.
- [6] (a) Brown AW, Hang J, Dussault PH, Carr T. Plant sterol and stanol substrate specificity of pancreatic cholesterol ester lipase. *J Nutr Biochem* 2009. Available online 16 July 2009; (b) Rasmussen HE, Guderian Jr DM, Wray CA, Dussault PH, Schlegel VL, Carr T. Reduction in cholesterol absorption is enhanced by stearate-enriched plant sterol esters in hamsters. *J Nutr* 2006;136:2722–7.
- [7] Holman RT, Lundberg WO, Malkin T. Progress in the Chemistry of Fats and Other Lipids, vol. 1. London: Pergamon Press; 1952. Chapter 2.
- [8] Zhang X, Geoffroy P, Miesch M, Julien-David D, Raul F, Aoudiè-Werner D, et al. Gram-scale chromatographic purification of β -sitosterol: synthesis and characterization of β -sitosterol oxides. *Steroids* 2005;70:886–95.
- [9] Kircher HW, Rosenstein FU. Purification of sitosterol. *Lipids* 1973;8:97–100.
- [10] For an overview of stigmasterol hydrogenation, see: Geoffroy P, Julien-David D, Marchioni E, Raul F, Aoudiè-Werner D, Miesch M. Synthesis of highly pure oxyphytosterols and (oxy)phytosterol esters: Part I. Regioselective hydrogenation of stigmasterol: an easy access to oxyphytosterols. *Steroids* 2008;73:702–7.
- [11] (a) Steele JA, Mosettig E. The Solvolysis of Stigmasteryl tosylate. *J Org Chem* 1963;28:571–2; (b) McCarthy FO, Chopra J, Ford A, Hogan SA, Kerry JP, O'Brien NM, Ryan E, Maquire AR. Synthesis, isolation and characterization of β -sitosterol and β -sitosterol oxide derivatives. *Org Biomol Chem* 2005;3:3059–65.
- [12] For an application to sidechain-modified sterols, see: Khripach VA, Zhabinskii VN, Konstantinova OV, Khripach NB, Antonchick AV, Antonchick AP, Schneider B. Preparation of (25R)- and (25S)-26-functionalized sterols as tools for biosynthetic studies of cholic acids. *Steroids* 2005;70:551–62.
- [13] Sarmah P, Barua NC. Aluminum triiodide: a convenient reagent for deoxygenation of oxiranes. *Tetrahedron Lett* 1988;29:5815–6.
- [14] (a) Syamala MS, Das JJ, Baskaran S, Chandrasekaran S. A novel and highly β -selective epoxidation of Δ^5 -unsaturated sterols with permanganate ion. *J Org Chem* 1992;57:1928–30; (b) Baqi Y, Giroux S, Corey EJ. A study of the epoxidation of cycloolefins by the *t*-BuOH copper–permanganate system. *Org Lett* 2009;11:959–61.
- [15] Wang SM, Zhang YB, Liu HM, Yu GB, Wang KR. Mild and selective deprotection method of acetylated sterols and diterpenes by dibutyltin oxide. *Steroids* 2007;72:26–30.
- [16] Tietze LF, Raith C, Brazel CC, Hoelsken S, Magull J. Enantioselective synthesis of 2-substituted alcohols using (+)-(1S,2S)-pseudoephedrine as chiral auxiliary. *Synthesis* 2008:229–36.
- [17] Akihisa T, Tanaka N, Yokota T, Tanno N, Tamura T. 5α -Cholest-8(14)-en-3 β -ol and three 24-alkyl- $\Delta^{8(14)}$ -sterols from the bulbils of *Dioscorea batatas*. *Phytochemistry* 1991;30:2369–72.
- [18] Attempts to perform the $\text{Cu}(\text{MnO}_4)_2$ oxidation on unprotected stigmasterol resulted in formation of numerous byproducts.
- [19] Khripach VA, Zhabinskii VN, Konstantinova OV, Khripach NB, Antonchick AP, Schneider B. [3,3]-Claisen rearrangements in 24α -methyl steroid synthesis: application to campesterol, crinosterol, and Δ^{25} -crinosterol side chain construction. *Steroids* 2002;67:597–603.
- [20] Assigned from the 10.6 Hz coupling constant for H_{22} – H_{23} .
- [21] See, for example: Murakami K, Watanabe B, Nishida R, Mori N, Kuwahara Y. Identification of crinosterol from astigmatid mites. *Insect Biochem Mol Biol* 2007;37:506–11.
- [22] Blakemore PR. The modified Julia olefination: alkene synthesis via the condensation of metallated heteroarylalkylsulfones with carbonyl compounds. *Perkins* 2002;1:2563–85.
- [23] (a) Sutoris V, Foltinova P, Gaplovsky A. Benzothiazole compounds. XVII. 2-Alkyl- and 2-arylalkylsulfonylbenzothiazoles and their antimicrobial activity. *Chem Zvesti* 1980;34:404–12; (b) Bourdon B, Corbet M, Fontaine P, Goekjian PG, Gueyrard D. Synthesis of enol ethers from lactones using modified Julia olefination reagents: application to the preparation of tri- and tetrasubstituted exoglycals. *Tetrahedron Lett* 2008;49:747–9.