



## Using (+)-carvone to access novel derivatives of (+)-*ent*-cannabidiol: The first asymmetric syntheses of (+)-*ent*-CBDP and (+)-*ent*-CBDV

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### ABSTRACT

(–)-Cannabidiol [(–)-CBD] has recently gained prominence as a treatment for neuro-inflammation and other neurodegenerative disorders; interest is also developing in its synthetic enantiomer, (+)-CBD, which has a higher affinity to CB1/CB2 receptors than the natural stereoisomer. We have developed an inexpensive, stereoselective route to access *ent*-CBD derivatives using (+)-carvone as a starting material. In addition to (+)-CBD, we report the first syntheses of (+)-cannabidivarin, (+)-cannabidiphorol as well as C-6/C-8 homologues.

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### Introduction

Nature remains inspiring in its ability to manufacture a diverse array of chiral secondary metabolites from relatively simple starting compounds. Perhaps even more remarkable is the fact that many of these building blocks exist as achiral, sparsely functionalized materials that are transformed *in vivo* into highly decorated molecules that exist as single stereoisomers. Importantly, while epimeric/diastereomeric metabolites are oftentimes isolated [1], with very few exceptions [eg., (+)- and (–)-carvone], natural product enantiomers are rarely found in Nature [2], but rather are almost exclusively manufactured in the laboratory. More often than not, this occurs serendipitously, *en route* to the total synthesis of a compound with unknown, undefined, or otherwise ambiguous absolute stereochemical assignments [3]. If the molecule is sufficiently small in size, a stereoselective synthesis may also be performed to probe the potentially unique activity of the non-natural *ent*-derivative, as there exists a prodigious amount of data that demonstrates the difference of one enantiomer *versus* the other in a biological context [4]. Additionally, there has been at several studies that have documented the increased activity of a natural product diastereomer relative to the natural stereoisomer itself [5]. Therefore, the targeted study of *ent*-natural products,

and related stereoisomers, is a viable and valuable approach to the discovery of potential new leads for drug discovery.

Recently, terpene derived (–)-Cannabidiol [(–)-2, (–)-CBD, Fig. 1], the major non-psychoactive constituent found in hemp, has gained popularity amongst the synthetic community [6], as cannabinoids, in general, have been increasingly shown to possess potent anti-inflammatory activity [7], especially against a number of neurological ailments including, but not limited to Alzheimer's [8a] and Parkinson's disease [8b]. Additionally, many naturally occurring cannabinoids have been studied in animal/clinical trials for a number of other uses, exploiting their antiepileptic, [8b] anxiolytic [8c], antiarthritic [8d], and antiemetic [8e] properties. There is also emerging evidence that (–)-CBD can interact with endocannabinoid receptors in the brain and protect against oxidative stress in neural cells [8f]. This in turn helps to reduce inflammation, the effects of which can cause the buildup of neurotoxic substances over time and lead to neuro-degeneration [8b]. In recent years, neuroinflammation has been identified as contributing more to the pathogenesis of Alzheimer's than even senile plaques and neurofibrillary tangles [9].

Both natural and synthetic cannabinoids have been involved in numerous clinical trials with several approved in multiple countries for their beneficial and quantifiable medicinal applications. While most of these treatments are CBD/THC mixtures, for example, Epidiolex [8b], Cannador [8b], and Sativex [10] (Nabiximol), some are pure THC-derived drugs, such as Nabilone [8b]

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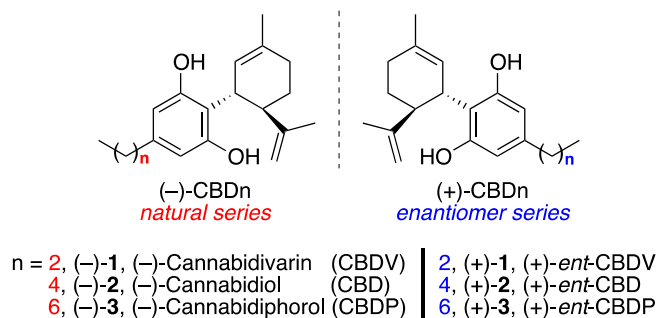


Fig. 1. Cannabidiol and related analogs.

[(±)-Cesamet] and Dronabinol [8b]. Also of significance, the cannabinoid drug Dexanabinol (**HU-211**, Fig. 1), based on the (+)-*ent*-cannabinoid skeletal structure, surprisingly has no affinity for CB<sub>1</sub> or CB<sub>2</sub> receptors, yet has significant non-competitive antagonist effects on *N*-methyl-D-aspartic acid [11]. This is notable since it is based on **HU-219**, which is a synthetic and more potent derivative of (-)-CBD [11].

While data suggests that (-)-CBD exhibits a low affinity for CB<sub>1</sub> (found mainly in the brain) and CB<sub>2</sub> (in peripheral cells), its non-natural synthetic enantiomer *ent*-CBD [(+)-2] and related derivatives are known to have a higher affinity for these same membrane receptors [12]. We believe *ent*-CBD derivatives will continue to prove valuable as novel derivatives of (-)-CBD continue to be explored as potential new therapeutics. To help support this statement, Table 1 shows the nM binding affinities of select cannabinoids towards the CB<sub>1</sub> and CB<sub>2</sub> receptors, demonstrating that (+)-*ent*-2 has increased binding when compared to its natural stereoisomer [8c]. Interestingly, another trend that warrants attention is the increased binding affinity of Δ<sup>9</sup>(-)-THC derivatives as their alkyl tails increase in length [13]; (-)-THCP, which has a seven carbon tail, binds an order of magnitude tighter to CB<sub>1</sub> and CB<sub>2</sub> than Δ<sup>9</sup>(-)-THC (Table 1).

In 2018, the Maio laboratory reported a new synthetic method that allowed for the expedient construction of non-natural CBD derivatives via the Lewis Acid mediated union of (-)-carvone, a readily available and inexpensive starting material, with resorcinol derivatives [14]. Importantly, by using (+)-carvone, this protocol also allowed access to enantiomers of the CBD scaffold in only three synthetic operations, two of which are general and can be carried out on gram scale, yielding a relatively stable epoxy-carvone silyl ether. However, difficulty in Δ<sup>8</sup> to Δ<sup>9</sup>-alkene transposition forced us to explore an alternative route for converting our scaffold into (+)-*ent*-CBD itself, as well as its C-3 and C-7 alkyl chain isomers, (+)-*ent*-cannabidivarin [(+)-1, *ent*-CBDV] and (+)-*ent*-cannabidiphorol [(+)-3, *ent*-CBDP], respectively, neither of which have been previously prepared in their non-natural, enantiomeric form. Our interest in these latter two derivatives stems from structure activity relationship data that demonstrate the importance of the alkyl chain length and how these derivatives may bind to CB<sub>1</sub> and CB<sub>2</sub> receptors (Table 1) [15]. Also of note, nat-

**Table 1**  
Previously reported binding affinities of select cannabinoids [8c-e,12].

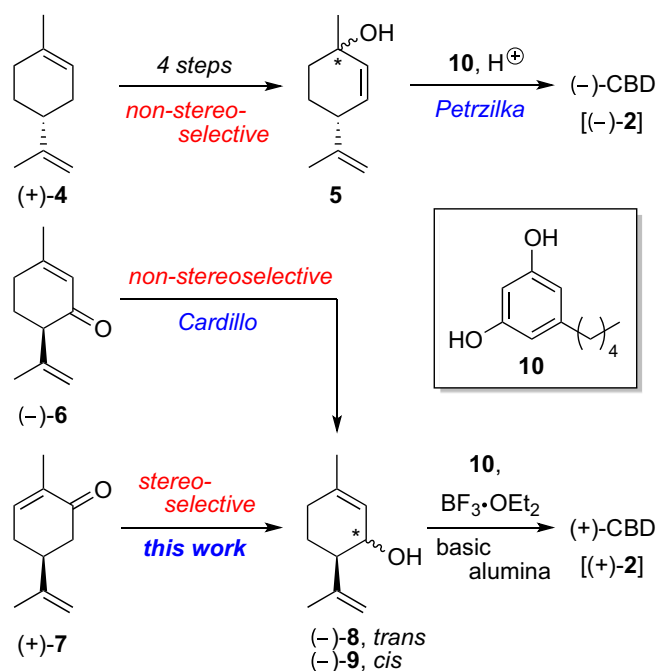
Cannabinoid	CB <sub>1</sub> K <sub>i</sub> (nM)	CB <sub>2</sub> K <sub>i</sub> (nM)
(-)-CBDV	>10,000	>10,000
(-)-CBD	>10,000	>10,000
(+)-CBD	842	203
(-)-THCV	22–75	62–105
(-)-THC	18–40	36–42
(-)-THCP	1.2	6.2

ural (-)-CBDV is in early clinical development for the treatment of autism spectrum disorders [16] and recently, (-)-CBDP has emerged as a more potent cannabinoid than (-)-CBD itself, making it an alternative to THC therapy without the signature psychoactivity of the latter [17].

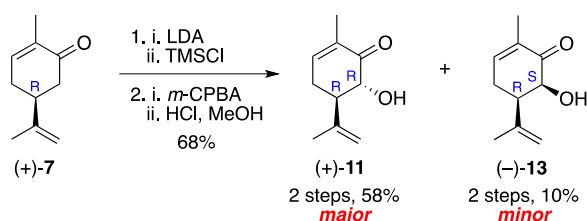
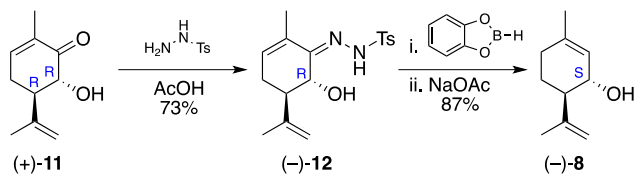
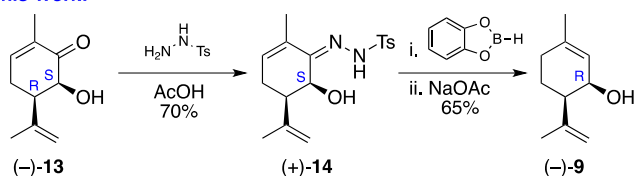
At the onset of our synthetic campaign, we evaluated the currently known syntheses of (-)- and (+)-CBD, many of which involve the acid-catalyzed union of a terpene derivative with olivetol, several of which are noteworthy here. The report by Petržilka utilized limonene-derived **5** as one of the coupling partners (Scheme 1), uniting this compound with olivetol (**10**) under mildly acidic conditions [18]. While this process does permit access to (-)-CBD, its key step suffers from a long reaction time (days), modest yield, and the overall number of steps in which **5** was derived from (+)-**4** [19]. A separate approach, first pioneered by Cardillo [8g] and later employed by Mechoulam [8c], utilized isopiperitenone [(+)-**6**] as a starting material. From this terpene, (+)-CBD could be accessed in two steps involving (1) LiAlH<sub>4</sub> reduction, and (2) treatment of the resultant alcohol mixture (**8** and **9**) with **10** in the presence of BF<sub>3</sub>•OEt<sub>2</sub>. Unfortunately, the relatively high cost of isopiperitenone (in either *enantiomer* form, ~\$1000/g) challenged us to think of potential ways to synthesize enantiopure **8** from more readily available starting compounds (Scheme 1) [20]. Recognizing the structural similarity between the southern hemisphere of **8** and (+)-carvone, we began to envision strategies to convert this inexpensive (\$0.15/g), caraway-derived terpene into the requisite chiral, non-racemic isopiperitenol. Scheme 2

## Results and discussion

In terms of retrosynthesis, based on literature precedent, we believed it would be possible to access **8** from tosylhydrazone **12** by exploiting the McIntosh reduction/rearrangement chemistry, which would effectively transpose the alkene from the Δ<sup>8</sup> to the Δ<sup>9</sup> location (*note*: cannabinoid notation) [21]. Hydrazone **12**, in turn, could be easily derived from hydroxycarvone **11**, which is



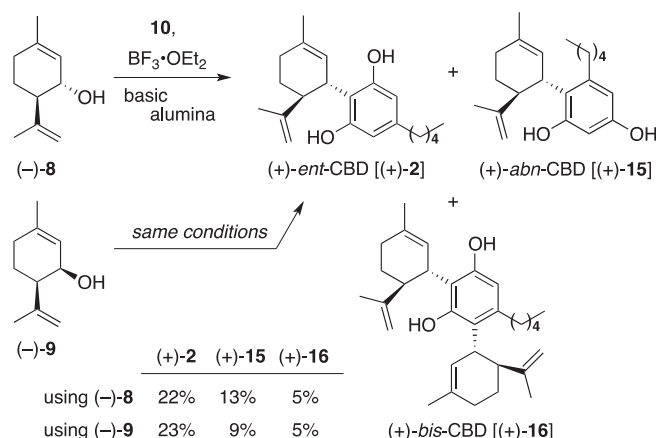
Scheme 1. Previous syntheses in the context of this work.

**Known literature protocol & yields:** <sup>22b</sup>**This Work:****This Work:****Scheme 2.** Synthesis of (1*S*, 6*R*)-isopiperitenol and (1*R*, 6*R*)-isopiperitenol.

already known to be the major product formed upon the Rubottom oxidation of (+)-carvone [22].

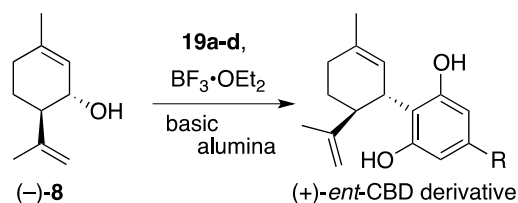
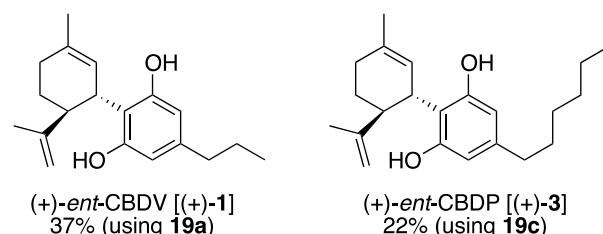
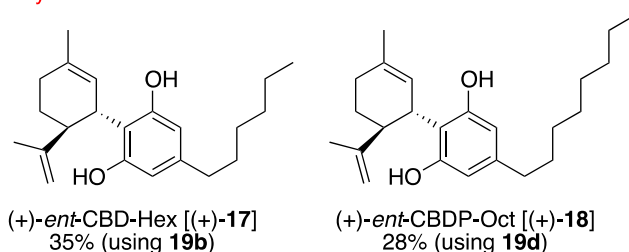
In the forward direction, treatment of (+)-carvone (7) with LDA, followed by the addition of TMSCl to the *in situ*-generated enolate allowed access to the corresponding silyl enol ether, which was directly treated with *m*-CPBA to afford a mixture of  $\alpha$ -hydroxycarvone isomers *trans*-(+)-11 (major) and *cis*-(+)-13 (minor), respectively. Although the diastereomer ratio and yield oftentimes varied, it consistently provided *trans*-hydroxy-carvone (+)-11 as the major product. Pleasingly, this result was in good agreement with literature precedent for this reaction [22b] and these C(6)-epimers could be easily separated by flash column chromatography. Next, each of these compounds was separately treated with tosylhydrazide and the corresponding hydrazones [(-)-12 and (+)-14] were successfully subjected to a one-pot reduction/rearrangement [21] sequence to afford the desired products, (1*S*, 6*R*)-isopiperitenol (-)-8 and (1*R*, 6*R*)-isopiperitenol (-)-9 in excellent overall yield (87% and 65%). Notably, the catecholborane used for this step can be formed *in situ* for a fraction of the cost [23]. Also of note, while previously demonstrated on related systems [21b], this alkene transposition reaction has yet to be reported for  $\alpha$ -hydroxycarvone. Importantly, this operationally simple and robust 4-step sequence can be carried out on gram scale, representing the first asymmetric total syntheses of (-)-8 and (-)-9 from (+)-carvone, circumventing the need to source these same alcohol products from costly (-)-isoperitenone (6).

Once synthetic (+)-isopiperitenol was in hand, we chose to repeat the Mechoulam buffered Lewis Acid protocol [8c] for the synthesis of (+)-*ent*-CBD [(+)-2] before exploiting this same method for the synthesis of novel cannabinoids (+)-*ent*-CBDV [(+)-1] and (+)-*ent*-CBDP [(+)-3] (Scheme 3). Pleasingly, when a solution of (-)-8 and olivetol (10) or, separately, (-)-9 and 10 was added to a solution of  $\text{BF}_3 \cdot \text{OEt}_2$  and basic alumina at reflux, (+)-*ent*-CBD [(+)-2] was produced as the major product, along with its abnormal regioisomer (+)-*abn*-CBD [(+)-15] in only 10 s and in yields consistent with literature values [8c]. Also observed, as documented by Crombie [24], was the formation of *bis*-(+)-16 as a minor by-product. Importantly, these three reaction

**Scheme 3.** Confirmation of the Mechoulam (+)-*ent*-CBD synthesis.

products have substantially different  $R_f$  values, making their separation by flash column chromatography an efficient way in which to separate them (see ESI for a photo of a representative TLC plate). Also, as an interesting side note, when Baek [25] repeated this reaction protocol in the absence of basic alumina, union of (-)-8 and 10 was followed by rapid cyclization to form (+)-*ent*-THC. We found similar results were obtained when the basic alumina was flame-dried prior to use.

Encouraged by the successful repetition of the Mechoulam (+)-*ent*-CBD synthesis, our attention turned to the construction of the C-3 and C-7 alkyl chain isomers, (+)-*ent*-cannabidivarin [(+)-1] and (+)-*ent*-cannabidiphorol [(+)-3], the natural stereoisomers of which are both known compounds [24]. It was during this time that we also began exploring the literature and discovered that the analogous C-6 isomer [(+)-17, CBD-Hex] was only reported in the patent

**a. Synthesized Natural Product Enantiomers****b. Synthesized Non-Natural *ent*-CBD Derivatives****Scheme 4.** First asymmetric synthesis of (+)-*ent*-CBDV, CBDP, and related C-6 and C-8 alkyl chain derivatives.

literature [26], with no synthesis shown, and the C-8 isomer [(+)-**18**, CBD-Oct] had yet to be proposed. We believed this latter CBD derivative would be of value since a  $\Delta^8$ -(-)-THC-Oct derivative has been previously reported and showed optimal binding to the CB<sub>1</sub> and CB<sub>2</sub> receptor when compared to its heptyl, pentyl, butyl, and propyl derivatives [27]. Clearly, the targeted synthesis of this congener in enantiomeric form, should prove valuable for future study.

In order to target these four derivatives, it was first necessary to synthesize their corresponding resorcinol fragments. In each case, this was easily accomplished in three steps involving, (1) olefination using 3,5-dimethoxybenzaldehyde and the appropriately sized ylide partner (see ESI for details), (2) hydrogenation of the resultant *E/Z*-alkene mixture, and (3) acid-catalyzed ether cleavage. It should be noted that all three of these operations are relatively high yielding and can be performed without intermediate purification, in a single 8 h period.

Once in hand, each of these C(6)-substituted resorcinol derivatives (**19a-d**) was separately united with (+)-isopiperitenol [(−)-**8**] using alumina buffered BF<sub>3</sub>·OEt<sub>2</sub> to afford the corresponding *ent*-CBD derivative, along with the concomitant formation of their *ent-abn*-CBD and *ent-bis*-CBD congeners (Scheme 4, see ESI for full details). Importantly, this represents the first asymmetric total syntheses of (+)-CBDV [(+)-**1**] and (+)-CBDP [(+)-**3**], and the first targeted syntheses of the related congeners (+)-CBD-Hex [(+)-**17**] and CBD-Oct [(+)-**18**].

## Conclusion

In summary, we report here the first asymmetric synthesis of both (1*S*, 6*R*)-isopiperitenol (37% overall) and (1*R*, 6*R*)-isopiperitenol (5% overall) in four synthetic steps from (+)-carvone as a starting material. Of note, this was made possible by exploiting the McIntosh alkene *trans*-position reaction as a key step. We then demonstrated the utility of this protocol by synthesizing (in one additional step for each) the enantiomer of cannabidiol, (+)-CBD (22%), and the related congeners (+)-CBDV (37%), (+)-CBDP (22%), (+)-CBD-Hex (35%), and (+)-CBD-Oct (28%). Also of note, this manuscript reports the first documentation and characterization of nearly all of their associated abnormal and *bis*-addition byproducts. We believe these enantiomer CBD derivatives will be of great interest and may lead to the discovery of even more active CBD-analogs. We are currently investigating the biological potency of these new *ent*-CBD derivatives and our findings will be reported in due course.

## Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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## Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.tetlet.2021.152891>.

## References

- [1] T. Ito, H. Ito, M. Oyama, T. Tanaka, J. Murata, D. Darnaedi, M. Iinuma, *Phytochem. Lett.* 5 (2012) 325–328.
- [2] Interesting examples include: limonene, pinene, and notoamide B, for a full review, see: J.F. Finefield, D.H. Sherman, M. Kreitman, R.M. Williams *Angew. Chem. Int. Ed.* 51 (2012) 4802–4836.
- [3] R. Tello-Aburto, T.D. Newar, W.A. Maio, *J. Org. Chem.* 77 (2012) 6271–6289.
- [4] K. Mori, *Chirality* 23 (2011) 449–462.
- [5] A. de Fatima, L.K. Kohn, J.E. de Carvalho, R.A. Pilli, *Bioorg. Med. Chem.* 14 (2006) 622–631.
- [6] (a) M.C. Pirrung, *J. Med. Chem.* 63 (2020) 12131–12136; (b) B. Jung, J.K. Lee, J. Kim, E.K. Kang, S.Y. Han, H.-Y. Lee, I.S. Choi, For recent syntheses of (−)-CBD and related analogs, see, *Chem. Asian. J.* 14 (2019) 3749–3762; (c) Z.P. Shultz, G.A. Lawrence, J.M. Jacobson, E.J. Cruz, J.W. Leahy, *Org. Lett.* 20 (2018) 381–384; (d) W.A. Kinney, M.E. McDonnell, H.M. Zhong, C. Liu, L. Yang, W. Ling, T. Qian, L. Chen, Z. Cai, D. Petkanas, D.E. Brenneman, *ACS Med. Chem. Lett.* 7 (2016) 424–428; (e) M.R. Gotz, J.A. Collado, J. Fernandez-Ruiz, B.L. Fiebich, L. Garcia-Toscano, M. Gomez-Canas, O. Koch, A. Leha, E. Munoz, C. Navarrete, M.R. Pazos, U. Holzgrabe, *Front. Pharmacol.* 10 (2019) 1284; (f) X. Gong, C. Sun, M.A. Abama, W. Shi, Y. Xie, W. Xu, F. Zhu, Y. Zhang, J. Shen, H.A. Aisa, *J. Org. Chem.* 85 (2020) 2704–2715.
- [7] S. Ben-Shabat, L.O. Hanus, G. Katzavian, R. Gallily, *J. Med. Chem.* 49 (2006) 1113–1117.
- [8] (a) G. Mazzocanti, O.H. Ismail, I. D'Acquarica, C. Villani, C. Manzo, M. Wilcox, A. Cavazzini, F. Gasparrini, *Chem. Commun.* 53 (2017) 12262–12265; (b) E.C. Rosenberg, R.W. Tsien, B.J. Whalley, O. Devinsky, *Neurotherapeutics* 12 (2015) 747–768; (c) L.O. Hanus, S. Tchilibon, D.E. Ponde, A. Breuer, E. Fride, R. Mechoulam, *Org. Biomol. Chem.* 3 (2005) 1116–1123; (d) D.P. Papahatjis, V.R. Nahmias, S.P. Nikas, T. Andreou, S.O. Alapafuja, A. Tsoinias, J. Guo, P. Fan, A. Makriyannis, *J. Med. Chem.* 50 (2007) 4048–4060; (e) E. Fride, D. Ponde, A. Breuer, L. Hanus, *Neuropharmacology* 48 (2005) 1117–1129; (f) R. Mechoulam, *J. Clin. Pharmacol.* 42 (2002) 115–195; (g) B. Cardillo, L. Merlini, S. Servi, *Tetrahedron Lett.* 13 (1972) 945–948; (h) E. Fride, C. Feigin, D.E. Ponde, A. Breuer, L. Hanus, N. Arshavsky, R. Mechoulam, *Eur. J. Pharmacol.* 506 (2004) 179–188.
- [9] M.T. Heneka, M.J. Carson, G.E. Landreth, F. Brosseron, D.L. Feinstein, A.H. Jacobs, T. Wyss-Coray, J. Victorica, R.M. Ransohoff, K. Herrup, S.A. Frautschy, B. Finsen, G.C. Brown, A. Verkhratsky, K. Yamanaoka, J. Koistinaho, E. Latz, A. Halle, G.C. Petzold, T. Town, D. Morgan, M.L. Shinohara, V.H. Perry, C. Holmes, N.G. Bazan, D.J. Brooks, S. Hunot, B. Joseph, N. Deigendesch, O. Garaschuk, E. Boddeke, C.A. Dinarello, J.C. Breitner, G.M. Cole, D.T. Goldenbock, M.P. Kummer, *Lancet. Neurol.* 14 (2015) 388–405.
- [10] M. Maccarrone, I. Bab, T. Biro, G.A. Cabral, S.K. Dey, V. Di Marzo, J.C. Konje, G. Kunos, R. Mechoulam, P. Pacher, K.A. Sharkey, A. Zimmer, *Trends Pharm. Sci.* 36 (2015) 277–296.
- [11] E. Pop, *Curr. Pharm. Design* 6 (2000) 1347–1359.
- [12] T. Bisogno, L. Hanus, L. De Petrocellis, S. Tchilibon, D.E. Ponde, I. Brandi, A.S. Moriello, J.B. Davis, R. Mechoulam, V. Di Marzo, *Brit. J. Pharm.* 134 (2001) 845–852.
- [13] D. An, S. Peigneur, L.A. Hendrickx, J. Tytgat, *Int. J. Mol. Sci.* 21 (5046) (2020) 1–32.
- [14] S.J. Bailey, R.S. Sapkota, A.E. Gollither, B. Dungan, M. Talipov, F.O. Holguin, W.A. Maio, *Org. Lett.* 20 (2018) 4618–4621.
- [15] H. Chung, A. Fierro, C.D. Pessoa-Mahana, *PLoS One* 14 (2019) 1–18.
- [16] X. Gong, C. Sun, M.A. Abama, W. Shi, Y. Xie, W. Xu, F. Zhu, Y. Zhang, J. Shen, H.A. Aisa, *J. Org. Chem.* 85 (4) (2020) 2704–2715.
- [17] C. Citti, P. Linciano, F. Russo, L. Luongo, M. Iannotta, S. Maione, A. Lagana, A.L. Capriotti, F. Forni, M.A. Vandelli, G. Gigli, G. Cannazza, *Sci. Rep.* 9 (20335) (2019) 1–13.
- [18] T. Petrzilka, W. Haefliger, C. Sikemeier, G. Ohloff, A. Eschenmoser, *Helv. Chim. Acta* 50 (1967) 719–723.
- [19] S.M. Wilkinsons, J. Price, M. Kassiou, *Tetrahedron Lett.* 54 (2013) 52–54.
- [20] Z.G. Brill, M.L. Condakes, C.P. Ting, T.J. Maimone, *Chem. Rev.* 117 (2017) 11753–11795.
- [21] (a) Y. Chai, D.A. Vicic, M.C. McIntosh, *Org. Lett.* 5 (2003) 1039–1042; (b) D.T. Bateman, A.L. Joshi, K. Moon, E.N. Galitovskaya, M. Upreti, T.C. Chambers, M.C. McIntosh, *Bioorg. Med. Chem. Lett.* 19 (2009) 6898–6901.
- [22] (a) O.V. Ardashov, A.V. Pavlova, I.V. Il'ina, E.A. Morozova, D.V. Korchagina, E.V. Karpova, K.P. Volcho, T.G. Tolstikova, N.F. Salakhutdinov, *J. Med. Chem.* 54 (2011) 3866–3874; (b) R.B. dos Santos, T.J. Brocksom, P.R. Zanotto, U. Brocksom, *Molecules* 7 (2002) 129–134.
- [23] K.M. Waltz, J.F. Hartwig, *J. Am. Chem. Soc.* 122 (2000) 11358–11369.
- [24] L. Crombie, W.M.L. Crombie, *Phytochemistry* 14 (1975) 213–220.
- [25] S. Baek, M. Srebnik, R. Mechoulam, *Tetrahedron Lett.* 26 (1985) 1083–1086.
- [26] Horwitz, A.; D'Espaux, L.; Wong, J.; Bector, R.; Hjelmeland, A. K.; Platt, D.; Ubersax, J. U.S. Patent 2020069214, 2019, (patent application).
- [27] B.R. Martin, R. Jefferson, R. Winckler, J.L. Wiley, J.W. Huffman, P.J. Crocker, B. Saha, R.K. Razdan, *J. Pharmacol. Exp. Ther.* 290 (1999) 1065–1079.