

Regular Article

Synthesis, Molecular Docking and Biological Evaluation of Quinolone Derivatives as Novel Anticancer Agents

Jie Li, Tu-cai Zheng, Yi Jin, Jian-guo Xu, Jian-gang Yu, and Yan-wen Lv*

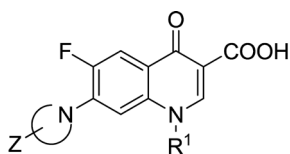
College of Chemistry and Materials Engineering, Quzhou University, Quzhou 324000, China.

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A series of novel quinolone derivatives (8a–j) were synthesized, and their anticancer activities were tested in human cancer cell lines, human lung carcinoma cell (A549), human promyelocytic leukemia cell (HL-60), and human cervical cancer cell (Hela). Compound 8i was found to be 5-times more potent in cell-killing activity for cell lines A549, HL-60, and Hela than the positive control irinotecan or cisplatin, with IC_{50} of 0.009, 0.008 and 0.010 μ M, respectively. The docking study revealed that compound 8i might have strong interactions with the active site of DNA-topoisomerase I.

Key words quinolone derivative; anticancer; synthesis; molecular docking

Quinolone as a privileged scaffold represents one of the most important structural unit prevalent in various naturally occurring and bioactive compounds.¹⁾ Quinolones consist of a bicyclic ring structure (Fig. 1) in which there is a substitution at position N-1, with various moieties. Most of the current agents have a carboxyl group at position 3, a keto group at position 4, a fluorine atom at position 6 and a nitrogen heterocycle at position 7.^{2,3)} In the late 1980s, reports emerged describing experimental antibacterial quinolones having sig-



R¹= Et, cyclopropyl, halo substituent aromatic ring, etc.

Z= attached group to nitrogen heterocyclic ring

Fig. 1. Structural Features of Quinolones

nificant potency against eukaryotic topoisomerases (Top) and showing cytotoxic activity against tumor cell lines.^{4–7)}

Li and colleagues designed and synthesized a series of quinolone derivatives as potential Top I inhibitors for cancer treatment.^{8–10)} Rajulu *et al.* designed a series of fluoroquinolones displaying good growth inhibition activities against human lung carcinoma cell (A549) and colon carcinoma (HCT-116).¹¹⁾ Recently, our group discovered a novel series of Top I inhibitors with quinolone scaffold. Quinolone derivative **1** was the most potent compound we synthesized.¹²⁾ Several important series of antimicrobial agents are associated with particular N-1 substituents. Tosufloxacin (**2**) and difloxacin (**3**) (Fig. 2) with the substituent of fluorine atom in the aromatic ring enhanced both potency against Gram-positives and pharmacokinetics.¹³⁾ The antibacterial tosufloxacin (**2**) can be viewed as intermediate agents on the evolutionary path toward both antibacterial and anticancer quinolone derivatives.⁷⁾ Using a scaffold modification strategy, our team changed the N-1 cyclopropyl group into N-1 aryl substituents with fluorine in the aromatic ring (Fig. 2).

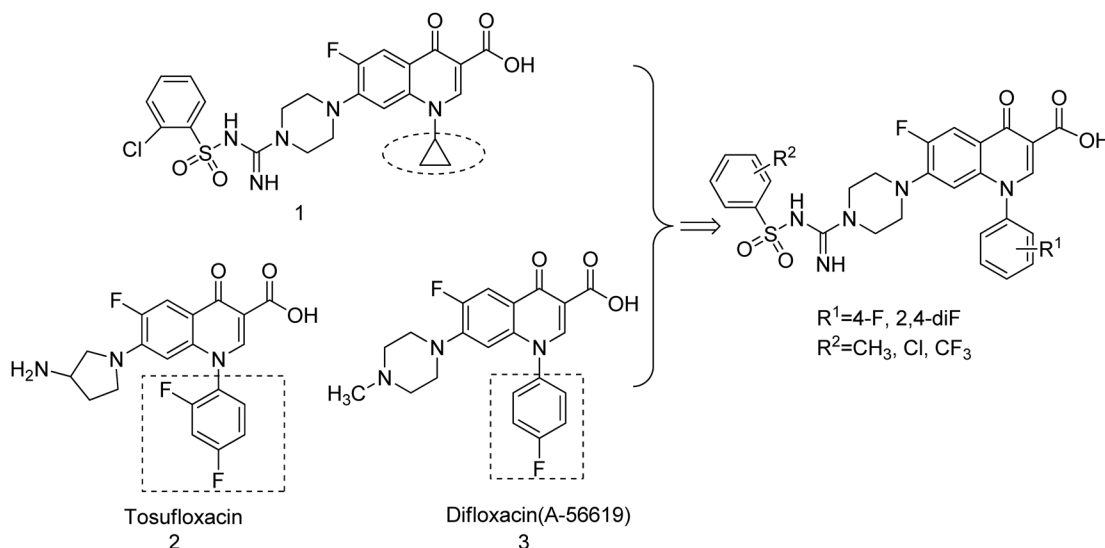


Fig. 2. Compound Generation by Scaffold Modification

* To whom correspondence should be addressed. e-mail: lyw@qzu.zj.cn

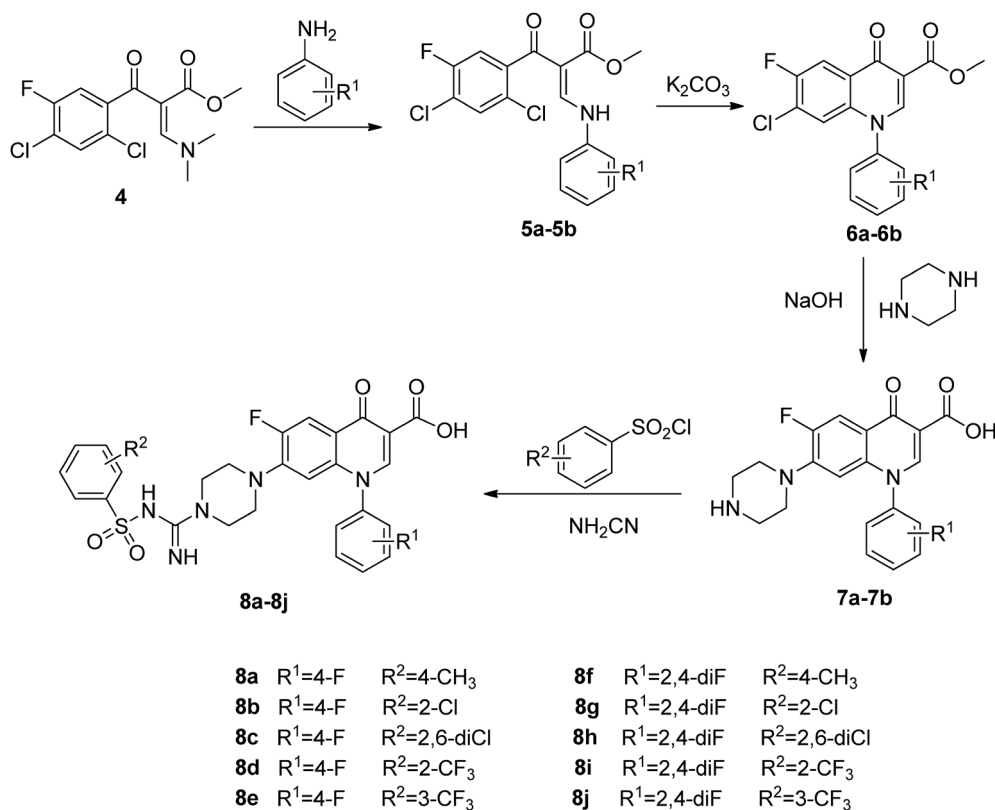


Chart 1. Synthesis of Compounds (8a–j)

Results and Discussion

Chemistry The method for the preparation of novel *N*-fluoroaromatic substituted piperazinylquinolone derivatives **8a–j** relies on the Grohe–Heitzer cycloacylation reaction.^{14,15} The synthetic route is outlined in Chart 1. The commercially available compound **4** was subjected to an addition–elimination reaction with a substituted primary amine, and the obtained product **5** was cyclised in a tandem addition–elimination reaction at the *ortho* position. Compound **7** was obtained by displacing the chlorine atom in compound **6** with anhydrous piperazine. Then the target compound **8** was prepared *via* a two-step one-pot tandem process.¹²

Biological Activity and Discussion All the target compounds (**8a–j**) were evaluated for their *in vitro* cytotoxic activity against three different human cancer cell lines, A549, human promyelocytic leukemia cell (HL-60) and human cervical cancer cell (Hela) by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay with irinotecan and cisplatin as positive control compounds. The assessments of anticancer activities were expressed as the concentration inhibiting 50% of cancer cell growth (IC₅₀). The Top I inhibitory activity assays was carried out using a topoisomerase I drug screening kit. The results are summarized in Table 1 and Fig. 3. As illustrated in Table 1, these compounds exhibited significant antitumor activity. Among them, compound **8i** displayed the most potent inhibitory activity (IC₅₀=0.009 μM for A549, IC₅₀=0.008 μM for HL-60, and IC₅₀=0.010 μM for Hela), which was better than the positive control irinotecan (IC₅₀=0.032 μM for A549, IC₅₀=0.044 μM for HL-60, and IC₅₀=0.038 μM for Hela) and cisplatin (IC₅₀=0.048 μM for A549, IC₅₀=0.057 μM for HL-60, and IC₅₀=0.047 μM for Hela). As expected, compound **8i** showed excellent Top I inhibitory

Table 1. IC₅₀ Cytotoxicity of Target Compounds

Compd.	IC ₅₀ (μM)		
	A549	HL-60	Hela
1	0.071	0.043	0.032
4	>0.157	>0.157	>0.157
8a	>0.086	0.081	>0.086
8b	>0.053	0.049	0.040
8c	0.041	0.039	0.043
8d	0.032	0.048	0.049
8e	0.036	0.016	0.037
8f	>0.082	0.050	0.069
8g	0.039	0.044	0.045
8h	0.038	0.033	0.029
8i	0.009	0.008	0.010
8j	0.028	0.018	0.010
Irinotecan	0.032	0.044	0.038
Cisplatin	0.048	0.057	0.047

activity comparable to irinotecan and cisplatin.

Subsequently, structure–activity relationships (SAR) studies were inferred from Table 1. In general, target compounds with *N*-1 substituent of 2,4-difluorophenyl group (**8f–j**) showed more potent activities than those with *N*-1 substituent of 4-fluorophenyl group (**8a–e**). In three different human cancer cell lines, using electron withdrawing group (Cl, CF₃) in the phenyl moiety as in compounds **8b–e** and **8g–j** increase the reactivity rather than using electron donating group (CH₃) in the phenyl moiety as in compounds **8a** and **8f**. And compounds **8i** and **8j** with trifluoromethyl in the phenyl moiety showed higher cytotoxic activity than in compounds **8g** and **8h** with

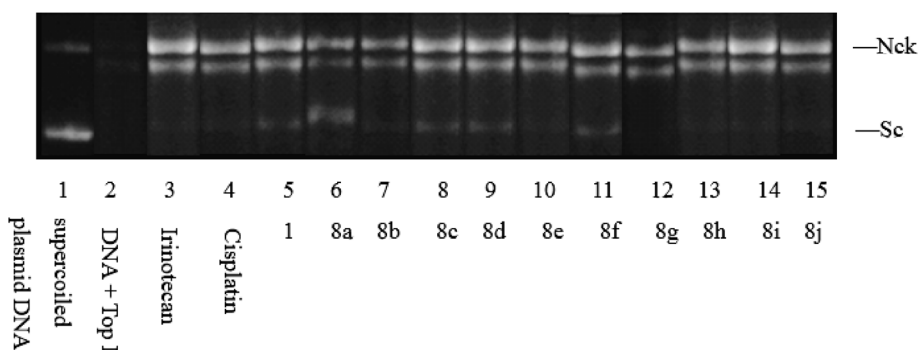


Fig. 3. Top I Inhibitory Activity

Inhibition of Top I at $1\mu\text{M}$. Lane 1: supercoiled plasmid DNA (pBR322); Lane 2: DNA+Top I; Lane 3: DNA+Top I+Irinotecan; Lane 4: DNA+Top I+Cisplatin; Lane 5–15: DNA+Top I+compounds (1, 8a, 8b, 8c, 8d, 8e, 8f, 8g, 8h, 8i and 8j). Sc—supercoiled DNA, Nck—nicked open circular DNA.

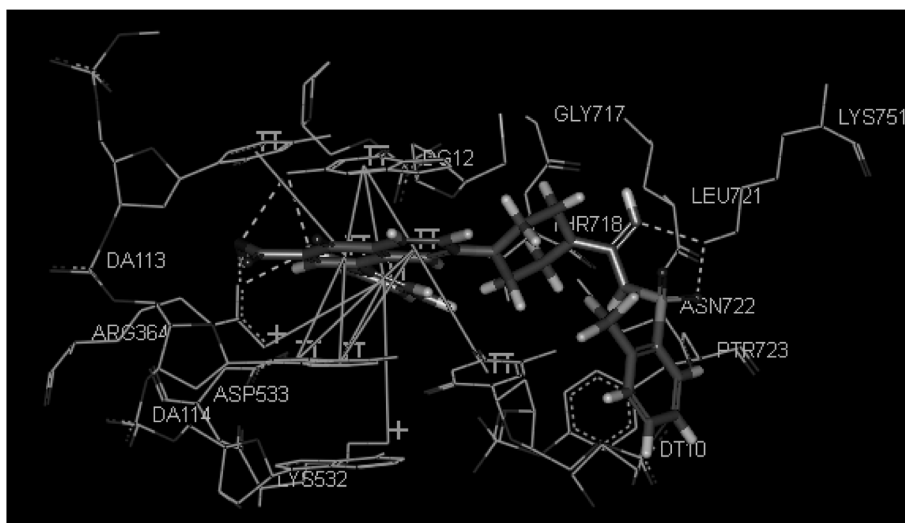


Fig. 4A. Compound 8i Bound to Top I-DNA Complex

The dotted lines show the hydrogen bonds and the solid lines show the π -cation and π - π interactions.

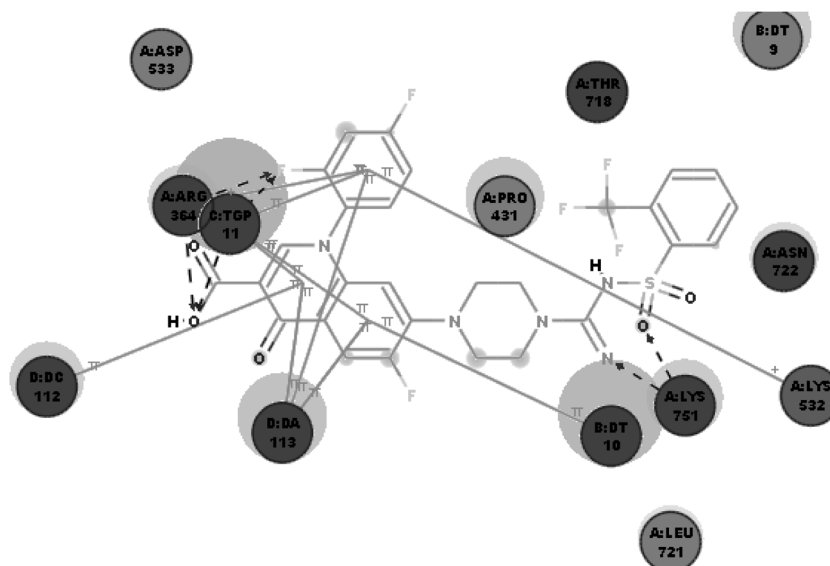


Fig. 4B. 2D Ligand Interaction Diagram of Compound 8i with Top I-DNA Complex Using Discovery Studio Program with the Essential Amino Acid Residues at the Binding Site Are Tagged in Circles

chlorine atom in the phenyl moiety. It is noted that the substitution R^2 at the 2-position of phenyl (8i) exhibiting better biological activities than the substitution R^2 at the 3-position

of phenyl (8j), which may be due to steric-hindrance effect.

Docking Study Docking was performed against DNA-Top I because it is reported as possible anticancer target of

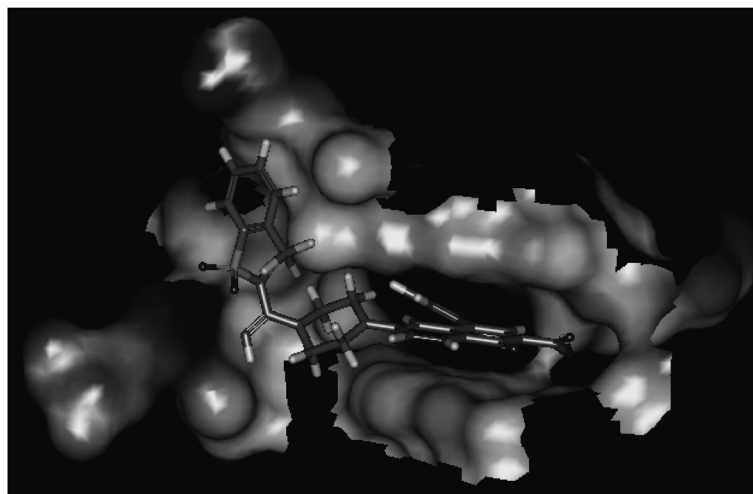


Fig. 4C. 3D Model of the Interaction between Compound **8i** and Top I-DNA Complex

The protein is represented by molecular surface. Compound **8i** is depicted by balls.

quinolone derivatives.^{16–19}) In order to understand the binding conformation of the most potent cytotoxic activity of the compound **8i**, its flexible molecular docking was carried out into the Top I (PDB code: 1K4T) active site using CDOCKER protocol of Discovery Studio 2.1.

The binding modes of compound **8i** and Top I-DNA complex were depicted in Fig. 4. The amino acid residue which had interaction with Top I-DNA complex were labeled in Figs. 4A and 4B. In the binding mode, compound **8i** was nicely bound to the Top I-DNA complex active site *via* six hydrogen bond, two cation– π interaction and ten π – π interaction. The nitrogen atom of guanidine group and one of the sulfonyl oxygen formed two hydrogen bond with amino of LYS 751. The fluorine atom at the 2-position of phenyl and the carboxyl group of **8i** formed two hydrogen bond with amino of ARG 364 and two hydrogen bond with base pair of TGP 11, respectively. And the benzene ring with two fluorine atom associated with LYS 532 and ARG 364 by two cation– π interactions. The quinolone skeleton and the benzene ring of compound **8i** stabilized by base-stacking interactions with both the upstream (–1) and downstream (+1) base pairs with ten π – π interactions.

The enzyme surface model was shown in Fig. 4C, which revealed that the molecule was well embedded in the active pocket. This molecular docking results and the biological assay data suggested that compound **8i** was a potential Top I inhibitors as anti-cancer agents.

Conclusion

We have designed and synthesized a novel series of quinolone derivatives (**8a–j**). These compounds exhibited *in vitro* cytotoxic activity against A549, HL-60, and Hela cells. Docking simulations were performed to position most active compound **8i** into the Top I-DNA complex active site to determine the probable binding conformation and the results confirmed that the compound was a potential Top I inhibitor.

Experimental

Chemistry Melting points (m.p.) were determined on Büchi B-540 melting point apparatus and are uncorrected. ¹H-NMR (300MHz) spectra were recorded on a Bruker AV

300MHz spectrometer. Mass spectra were obtained on a Thermo Finnigan LCQ-Advantage spectrometer (electrospray ionization (ESI), atmospheric pressure chemical ionization (APCI)), and high resolution (HR)-MS were carried out on an APEX (Bruker) mass III spectrometer. The compounds were dissolved in dimethyl sulfoxide (DMSO)-*d*₆. Chemical shifts were reported in ppm (δ) relative to tetramethylsilane (TMS) ($\delta=0$). Coupling constants, *J*, are reported in Hz, multiplicities being marked as: singlet (s), broad singlet (brs), doublet (d), triplet (t), quartet (q), multiplet (m).

General Procedure for Synthesis of Compounds (5a, b) Firstly, a solution of commercially available compound **4** (8g, 25mmol) in 25 mL ethanol, was added with the solution of fluoro-substituted aniline (26.25mmol) in 10 mL ethanol dropwise during 10 min. The reaction mixture was then stirred at 45°C for 6h. The cooled reaction was filtered to give the desired product **5** as light yellow solid.

General Procedure for Synthesis of Compounds (6a, b) A solution of **5** (10mmol), K₂CO₃ (2.2g, 16mmol) in 20 mL *N,N*-dimethylformamide (DMF) was stirred at 140°C for 2h. The hot mixture was filtered rapidly, then the solution was allowed to stir at room temperature for 10 min until a yellow precipitate is formed. The solid was filtered, washed with water twice, and dried *in vacuo*.

General Procedure for Synthesis of Compounds (7a, b) The compound **6** (3.5mmol), anhydrous piperazine (1.5g, 17.5mmol) and isopropanol (10mL) were mixed and stirred at 130°C for 3h, then the solvent was removed under reduced pressure. The residue was added with 10% NaOH (6mL), and stirred at 90°C for 1.5h, then added with activated carbon and stirred at reflux for 1h, respectively. The hot mixture was filtered and the aqueous phase was then acidified with hydrochloric acid to pH 7. The mixture was stirred at room temperature until a precipitated is formed. The solid was filtered on a Buchner funnel to give the desired product **7**.

General Procedure for Synthesis of Compounds (8a, j) Benzenesulfonyl chloride (1mmol) in butanone (5mL) was heated with stirring to 40°C, and cyanamide solution (50%) was added dropwise, then the temperature was raised to 60°C and stirring continued for 3h. The compound **7** (0.8mmol) was added and heated to 80°C for 3h. After cooling to 40°C,

the reaction mixture was poured into cold water while stirring, white crystals or powders were precipitated, filtered, washed with water, and dried. Analytically pure samples were obtained by recrystallization from aqueous ethanol.

(Z)-Methyl 2-(2,4-Dichloro-5-fluorobenzoyl)-3-((4-fluorophenyl)amino)acrylate (**5a**)

¹H-NMR (DMSO-*d*₆, 300 MHz) δ (ppm): 3.80 (s, 3H, -CH₃), 6.74–6.94 (m, 4H, -C₆H₄), 7.55 (d, 1H, *J*=4.0 Hz, 6-H), 7.81 (d, 1H, *J*=4.0 Hz, 3-H), 9.79 (s, 1H, -NH). MS (ESI): 385.0.

(Z)-Methyl 2-(2,4-Dichloro-5-fluorobenzoyl)-3-((2,4-difluorophenyl)amino)acrylate (**5b**)

¹H-NMR (DMSO-*d*₆, 300 MHz) δ (ppm): 3.83 (s, 3H, -CH₃), 6.60–6.78 (m, 3H, -C₆H₃), 7.61 (d, 1H, *J*=4.0 Hz, 6-H), 7.83 (d, 1H, *J*=4.0 Hz, 3-H), 9.56 (s, 1H, -NH). MS (ESI): 403.0.

Methyl 7-Chloro-6-fluoro-1-(4-fluorophenyl)-4-oxo-1,4-dihydroquinoline-3-carboxylate (**6a**)

¹H-NMR (DMSO-*d*₆, 300 MHz) δ (ppm): 3.82 (s, 3H, -CH₃), 7.09–7.18 (m, 4H, -C₆H₄), 7.25 (d, 1H, *J*=4.0 Hz, 8-H), 7.66 (d, 1H, *J*=4.0 Hz, 5-H), 8.29 (s, 1H, 2-H). MS (ESI): 349.0.

Methyl 7-Chloro-1-(2,4-difluorophenyl)-6-fluoro-4-oxo-1,4-dihydroquinoline-3-carboxylate (**6b**)

¹H-NMR (DMSO-*d*₆, 300 MHz) δ (ppm): 3.82 (s, 3H, -CH₃), 6.82–7.12 (m, 3H, -C₆H₃), 7.18 (d, 1H, *J*=4.0 Hz, 8-H), 7.66 (d, 1H, *J*=4.0 Hz, 5-H), 8.25 (s, 1H, 2-H). MS (ESI): 367.0.

6-Fluoro-1-(4-fluorophenyl)-4-oxo-7-(piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic Acid (**7a**)

¹H-NMR (DMSO-*d*₆, 300 MHz) δ (ppm): 1.20 (s, 1H, -NH), 2.82–2.85 (m, 4H, -CH₂, piperazine), 3.53–3.63 (m, 4H, -CH₂, piperazine), 6.48 (d, 1H, *J*=4.0 Hz, 8-H), 7.07–7.19 (m, 4H, -C₆H₄), 7.87 (d, 1H, *J*=4.0 Hz, 5-H), 8.32 (s, 1H, 2-H). MS (ESI): 385.1.

1-(2,4-Difluorophenyl)-6-fluoro-4-oxo-7-(piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic Acid (**7b**)

¹H-NMR (DMSO-*d*₆, 300 MHz) δ (ppm): 1.22 (s, 1H, -NH), 2.81–2.85 (m, 4H, -CH₂, piperazine), 3.34–3.70 (m, 4H, -CH₂, piperazine), 6.75 (d, 1H, *J*=4.0 Hz, 8-H), 6.85–7.13 (m, 4H, -C₆H₄), 8.02 (d, 1H, *J*=4.0 Hz, 5-H), 8.20 (s, 1H, 2-H). MS (ESI): 403.1.

6-Fluoro-1-(4-fluorophenyl)-4-oxo-7-(4-(*N*-tosylcarbamimidoyl)piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic Acid (**8a**)

M.p. 200–202°C, Yield 61%, ¹H-NMR (DMSO-*d*₆, 300 MHz) δ (ppm): 2.45 (s, 3H, -CH₃), 3.38–3.59 (m, 4H, -CH₂, piperazine), 3.78–3.88 (m, 4H, -CH₂, piperazine), 6.98 (d, 1H, *J*=4.0 Hz, 8-H), 7.08–7.10 (m, 4H, -C₆H₄), 7.30–7.65 (m, 4H, -C₆H₄), 8.01 (d, 1H, *J*=4.0 Hz, 5-H), 8.20 (s, 1H, 2-H); HR-MS Calcd for C₂₈H₂₅F₂N₅O₅S [M⁻]: 581.1544. Found: 581.1541.

7-(4-(*N*-((2-Chlorophenyl)sulfonyl)carbamimidoyl)piperazin-1-yl)-6-fluoro-1-(4-fluorophenyl)-4-oxo-1,4-dihydroquinoline-3-carboxylic Acid (**8b**)

M.p. 216–218°C, Yield 73%, ¹H-NMR (DMSO-*d*₆, 300 MHz) δ (ppm): 3.35–3.52 (m, 4H, -CH₂, piperazine), 3.75–3.81 (m, 4H, -CH₂, piperazine), 6.91 (d, 1H, *J*=4.0 Hz, 8-H), 7.06–7.708 (m, 4H, -C₆H₄), 7.40–7.78 (m, 4H, -C₆H₄), 7.92 (d, 1H, *J*=4.0 Hz, 5-H), 8.43 (s, 1H, 2-H); HR-MS Calcd for C₂₇H₂₂ClF₂N₅O₅S [M⁻]: 601.0998. Found: 601.0998.

7-(4-(*N*-((2,6-Dichlorophenyl)sulfonyl)carbamimidoyl)piperazin-1-yl)-6-fluoro-1-(4-fluorophenyl)-4-oxo-1,4-dihydroquinoline-3-carboxylic Acid (**8c**)

M.p. 211–213°C, Yield 71%, ¹H-NMR (DMSO-*d*₆, 300 MHz) δ (ppm): 3.40–3.54 (m, 4H, -CH₂, piperazine), 3.76–3.84 (m, 4H, -CH₂, piperazine), 6.83 (d, 1H, *J*=4.0 Hz, 8-H), 7.10–7.13 (m, 4H, -C₆H₄), 7.49–7.73 (m, 3H, -C₆H₃), 8.01 (d, 1H, *J*=4.0 Hz, 5-H), 8.26 (s, 1H, 2-H); HR-MS Calcd for C₂₇H₂₁Cl₂F₂N₅O₅S [M⁻]: 635.0609. Found: 635.0601.

6-Fluoro-1-(4-fluorophenyl)-4-oxo-7-(4-(*N*-((2-(trifluoromethyl)phenyl)sulfonyl)carbamimidoyl)piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic Acid (**8d**)

M.p. 207–209°C, Yield 60%, ¹H-NMR (DMSO-*d*₆, 300 MHz) δ (ppm): 3.35–3.57 (m, 4H, -CH₂, piperazine), 3.75–3.84 (m, 4H, -CH₂, piperazine), 7.01 (d, 1H, *J*=4.0 Hz, 8-H), 7.08–7.09 (m, 4H, -C₆H₄), 7.54–7.75 (m, 4H, -C₆H₄), 8.01 (d, 1H, *J*=4.0 Hz, 5-H), 8.21 (s, 1H, 2-H); HR-MS Calcd for C₂₈H₂₂F₅N₅O₅S [M⁻]: 635.1262. Found: 635.1260.

6-Fluoro-1-(4-fluorophenyl)-4-oxo-7-(4-(*N*-((3-(trifluoromethyl)phenyl)sulfonyl)carbamimidoyl)piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic Acid (**8e**)

M.p. 205–206°C, Yield 65%, ¹H-NMR (DMSO-*d*₆, 300 MHz) δ (ppm): 3.44–3.54 (m, 4H, -CH₂, piperazine), 3.76–3.84 (m, 4H, -CH₂, piperazine), 6.99 (d, 1H, *J*=4.0 Hz, 8-H), 7.10–7.14 (m, 4H, -C₆H₄), 7.53–7.92 (m, 3H, -C₆H₄), 7.93 (d, 1H, *J*=4.0 Hz, 5-H), 8.19 (s, 1H, -C₆H₄), 8.37 (s, 1H, 2-H); HR-MS Calcd for C₂₈H₂₂F₃N₅O₅S [M⁻]: 635.1262. Found: 635.1259.

1-(2,4-Difluorophenyl)-6-fluoro-4-oxo-7-(4-(*N*-tosylcarbamimidoyl)piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic Acid (**8f**)

M.p. 203–204°C, Yield 62%, ¹H-NMR (DMSO-*d*₆, 300 MHz) δ (ppm): 2.28 (s, 3H, -CH₃), 3.37–3.58 (m, 4H, -CH₂, piperazine), 3.78–3.87 (m, 4H, -CH₂, piperazine), 7.01 (d, 1H, *J*=4.0 Hz, 8-H), 7.09–7.42 (m, 3H, -C₆H₃), 7.44–7.92 (m, 4H, -C₆H₄), 8.27 (d, 1H, *J*=4.0 Hz, 5-H), 8.93 (s, 1H, 2-H); HR-MS Calcd for C₂₈H₂₄F₃N₅O₅S [M⁻]: 599.1450. Found: 599.1449.

7-(4-(*N*-((2-Chlorophenyl)sulfonyl)carbamimidoyl)piperazin-1-yl)-1-(2,4-difluorophenyl)-6-fluoro-4-oxo-1,4-dihydroquinoline-3-carboxylic Acid (**8g**)

M.p. 215–218°C, Yield 72%, ¹H-NMR (DMSO-*d*₆, 300 MHz) δ (ppm): 3.36–3.58 (m, 4H, -CH₂, piperazine), 3.79–3.86 (m, 4H, -CH₂, piperazine), 7.06 (d, 1H, *J*=4.0 Hz, 8-H), 7.18–7.44 (m, 3H, -C₆H₃), 7.52–8.03 (m, 4H, -C₆H₄), 8.26 (d, 1H, *J*=4.0 Hz, 5-H), 8.95 (s, 1H, 2-H); HR-MS Calcd for C₂₇H₂₁ClF₃N₅O₅S [M⁻]: 619.0904. Found: 619.0902.

7-(4-(*N*-((2,6-Dichlorophenyl)sulfonyl)carbamimidoyl)piperazin-1-yl)-1-(2,4-difluorophenyl)-6-fluoro-4-oxo-1,4-dihydroquinoline-3-carboxylic Acid (**8h**)

M.p. 210–212°C, Yield 79%, ¹H-NMR (DMSO-*d*₆, 300 MHz) δ (ppm): 3.47–3.52 (m, 4H, -CH₂, piperazine), 3.79–3.84 (m, 4H, -CH₂, piperazine), 7.06 (d, 1H, *J*=4.0 Hz, 8-H), 7.18–7.26 (m, 3H, -C₆H₃), 7.48–7.72 (m, 3H, -C₆H₃), 8.28 (d, 1H, *J*=4.0 Hz, 5-H), 8.95 (s, 1H, 2-H); HR-MS Calcd for C₂₇H₂₀Cl₂F₃N₅O₅S [M⁻]: 653.0514. Found: 653.0519.

1-(2,4-Difluorophenyl)-6-fluoro-4-oxo-7-(4-(*N*-((2-(trifluoromethyl)phenyl)sulfonyl)carbamimidoyl)piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic Acid (**8i**)

M.p. 205–207°C, Yield 68%, ¹H-NMR (DMSO-*d*₆, 300 MHz) δ (ppm): 3.36–3.58 (m, 4H, -CH₂, piperazine), 3.74–3.83 (m, 4H, -CH₂, piperazine), 6.84 (d, 1H, *J*=4.0 Hz, 8-H),

7.08–7.22 (m, 3H, $-C_6H_3$), 7.53–7.75 (m, 4H, $-C_6H_4$), 8.25 (d, 1H, $J=4.0$ Hz, 5-H), 8.67 (s, 1H, 2-H); HR-MS Calcd for $C_{28}H_{21}F_6N_5O_5S$ [M^-]: 653.1168. Found: 653.1163.

1-(2,4-Difluorophenyl)-6-fluoro-4-oxo-7-(4-(*N*-((3-(trifluoromethyl)phenyl)sulfonyl)carbamimidoyl)piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic Acid (**8j**)

M.p. 202–204°C, Yield 69%, 1H -NMR (DMSO- d_6 , 300MHz) δ (ppm): 3.40–3.55 (m, 4H, $-CH_2$, piperazine), 3.75–3.83 (m, 4H, $-CH_2$, piperazine), 6.82 (d, 1H, $J=4.0$ Hz, 8-H), 7.08–7.23 (m, 3H, $-C_6H_3$), 7.53–7.92 (m, 3H, $-C_6H_4$), 8.24 (d, 1H, $J=4.0$ Hz, 5-H), 8.28 (s, 1H, $-C_6H_4$), 8.67 (s, 1H, 2-H); HR-MS Calcd for $C_{28}H_{21}F_6N_5O_5S$ [M^-]: 653.1168. Found: 653.1161.

Anti-proliferation Assay The antiproliferative activities of the prepared compounds against A549, HL-60 and Hela cells were evaluated using a standard MTT-based colorimetric assay. Target tumor cell lines were grown to log phase in RPMI 1640 medium supplemented with 10% fetal bovine serum. After diluting to 1×10^6 cells mL^{-1} with the complete medium, 100 μL of the obtained cell suspension was added to each well of 96-well culture plates. The subsequent incubation was permitted at 37°C, 5% CO_2 atmosphere for 24h before the cytotoxicity assessments. Each concentration was in triplicate, and Irinotecan was used as the positive control. After 72h incubation at 37°C, 5% CO_2 atmosphere, 10 μL of MTT solution in Dulbecco's modified Eagle's medium (DMEM) (Invitrogen, U.S.A.) was added to each well. After three hours incubation at 37°C, 150 μL DMSO was added to each well. The plates were then vibrated for 10min for complete dissolution. The optical absorbance was measured at 570nm on an automated microplate spectrophotometer (Bio-Rad, U.S.A.). In all experiments three replicate wells were used for each drug concentration. The IC_{50} value was defined as the concentration at which 50% of the cells could survive. The results were summarized in Table 1.

Top I Inhibition Top I inhibition was assayed by determining relaxation of supercoiled DNA pBR322. The test compounds were dissolved in DMSO and diluted with the final concentration 1 or 10 μM respectively, the same method to irinotecan and cisplatin at the concentration of 1 or 10 μM . A mixture of 0.5 μg of plasmid pBR322 and 10U top I was incubated with the test compounds in final volume of 20 μL (in DMSO) at 37°C for 30min in relaxation buffer (20mmol/L Tris-HCl (pH 7.8), 50mmol/L KCl, 10mmol/L $MgCl_2$, 1mmol/L dithiothreitol (DTT), 0.2mmol/L ethylenediaminetetraacetic acid (EDTA)). The reactions were terminated by adding 2.5 μL of stop solution containing 10% sodium dodecyl sulfate (SDS), 0.2% bromophenol blue, 0.2% xylene cyanol and 30% glycerol. DNA samples were then electrophoresed on 1% agarose gel for 10h with Tris-borate-EDTA running buffer. Gels were stained for 30min in an aqueous solution of ethidium bromide and visualized by transillumination with UV light.

Molecular Docking The pdb file about the crystal structure of DNA-Top I bound to Topotecan (PDB code: 1T8I)²⁰ was obtained from the RCSB Protein Data Bank (<http://www.pdb.org>). The molecular docking procedure was performed by using CDOCKER protocol for receptor–ligand interactions of

Discovery Studio 2.1. For ligand preparation, the 3D structures of **8i** were generated and minimized using Discovery Studio 2.1. For protein preparation, the hydrogen atoms were added. The whole DNA-Top I domain defined as a receptor and the site sphere was selected based on the ligand binding location of Topotecan, then the Topotecan removed and the prepared ligand was placed during the molecular docking procedure. CHARMM was selected as the force field. The molecular docking was performed with a simulated annealing method.

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Conflict of Interest The authors declare no conflict of interest.

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