Asymmetric Pictet–Spengler Reactions: Synthesis of 1,2,3,4-Tetrahydroisoquinoline Carboxylic Acid (Tic) Chimeras

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Dedicated to Professor Ivar Ugi on the occasion of his 70th birthday

Abstract: A preparatively simple diastereoselective synthesis of the amino acid chimera (1S,3S)-1,2,3,4-tetrahydroisoquinoline-1,3-dicarboxylic acid from hexafluoroacetone-protected phenylalanine and glyoxylic acid hydrate via Pictet–Spengler reaction is described. The potential of the reaction of hexafluoroacetone-protected phenylalanine with other aldehydes was scrutinized.

Key words: amino acids, cyclizations, diastereoselectivity, heterocycles, quinolines

1,2,3,4-Tetrahydroisoquinolines constitute a class of compounds attracting increasing interest, due to their various biological activities. The broad spectrum of pharma-cological properties includes *inter alia* antibacterial and antiplasmodial activity as well as cardiovascular and neuromodulating effects.¹

Much effort has been devoted to the asymmetric synthesis of optically active 1-substituted 1,2,3,4-tetrahydroisoquinolines, because they are substructures of many naturally occurring alkaloids and their non-natural derivatives.² The stereocenter at C-1 can be constructed, *inter alia* via Pictet–Spengler³ and Bischler–Napieralski⁴ reactions. The diastereoselective Pictet–Spengler reaction using tryptamines⁵ and tryptophanes⁶ and chiral aldehydes as optically active starting materials have been extensively studied, whereas enantioselective reactions are rarely found in literature.⁷

1,2,3,4-Tetrahydroisoquinoline-3-carboxylic acid (Tic-3, 1) is a phenylalanine analog in which the dihedral angle χ ($\chi^1 = N-C^{\alpha}-C^{\beta}-C^{\gamma}$, $\chi^2 = C^{\alpha}-C^{\beta}-C^{\gamma}-C^{\delta}$) is limited to a very small range because of its bicyclic nature.⁸ Tic-3 has been applied in many instances as a replacement of phenylalanine for the design of topographically constrained peptides to study effector/receptor interactions.⁹ Likewise, 1,2,3,4-tetrahydroisoquinoline-1-carboxylic acid (Tic-1, **2**) represents a conformationally rigid phenylglycine analog (Figure 1).





Figure 1 Structures of 1,2,3,4-tetrahydroisoquinoline carboxylic acids 1-3

Recently we described a new access to *N*-methylamino¹⁰ and *N*-ethylamino acids¹¹ using hexafluoroacetone as a protecting and activating reagent. When we applied the new strategy to hexafluoroacetone (HFA)-protected (*S*)-phenylalanine **5**, we found a preparatively simple synthesis of (1S,3S)-1,2,3,4-tetrahydroisoquinoline-1,3-dicarboxylic acid (**3**). The diastereoselective condensation reaction of **5** with glyoxylic acid hydrate in the presence of concentrated sulfuric acid at room temperature was successful and gave a single new product. The conversion of the starting material was complete within 3 days (monitored by ¹⁹F NMR). Based on the spectroscopic data we ascribe the structure of a HFA-protected 1,2,3,4-tetrahydroisoquinoline-1,3-dicarboxylic acid **6a** to the new compound (Scheme 1).



Scheme 1

The relative configuration of **6a** was determined by X-ray structure analysis: Both carboxylic groups are placed *trans* with respect to the bicyclic system. Since the configuration of C-3 is known to be (*S*) [sequence started from (*S*)-phenylalanine] we can assign the (*S*)-configuration to the new stereocenter at C-1. The compound crystallizes in the monoclinic space group $P2_1$ with two independent molecules in the unit cell. The configuration and bonding parameters of both molecules are very similar. Figure 2 shows one of the molecules. Both molecules establish along the b-axis a helical structure with hydrogen bonds between the carboxylic groups (Figure 3).



Figure 2 X-ray crystal structure of 6a

To determine the enantiomeric purity of **6a** we synthesized the enatiomeric reference compound **6b** obtainable from (*R*)-phenylalanine. HPLC experiments revealed that the reaction sequence $4 \rightarrow 5 \rightarrow 6$ occurs without racemization (ee >99.9%).



Figure 3 Helical structure along the b-axis in the unit cell of 6a

The unprotected amino acid **3**, which was unknown to the best of our knowledge, was obtained by hydrolysis of **6a** at room temperature in propan-2-ol-water. It possesses the Tic-3 as well as the Tic-1 substructure. Therefore, it belongs to the class of chimeric amino acids, ¹² which are interesting candidates for peptide modification. Furthermore, compound **3** should exhibit potential in the design of secondary structure mimetics and may be applied for inversion of the peptide sequence.

The 3-carboxy group of **6a** can be regioselectively functionalized on reaction with various nucleophiles. With amino acid amides for example dipeptide amides are formed ($6a \rightarrow 7$).

We tested the reaction of HFA-protected phenylalanine **5** with a series of aldehydes and ketones. However, only paraformaldehyde reacted with compound **5** in the presence of trifluoroacetic acid to give the Pictet–Spengler product **8** in good yields (Scheme 2). Compound **8** represents a carboxy group activated derivative of Tic-3. Therefore, it can be hydrolyzed under mild conditions (propan-2-ol–water, r.t.) to give Tic-3 (1). Comparison of the optical rotation measured for Tic-3 (1) ($[\alpha]_D - 134$) obtained via **5** \rightarrow **8** \rightarrow **1** with the value cited in literature ($[\alpha]_D - 139$)¹³ reveals that the product is optically pure. On reaction with amino acid esters dipeptide esters are formed (**8** \rightarrow **9**) (Scheme 2).



Scheme 2

When **5** was reacted with paraformaldehyde in the presence of thionyl chloride *N*-chloromethylation was observed. The *N*-chloromethyl compound **10** is stable at room temperature. On distillation in vacuo, at about 110 °C, HCl elimination occurs (Scheme 3). However, the Pictet–Spengler product could not be isolated. The formation of E/Z olefins **11a** and **11b**, which can be separated by distillation (Vigreux column) under reduced pressure, should be the result of a thermal induced 1,3-HCl elimination. The azomethine ylide¹⁴ formed first undergoes a sigmatropic 1,4-H shift to give a mixture of **11a** and **11b**.





Compounds of type 10 with a Cl-CH₂-N< moiety belong to the class of 1-halogenalkylamines which exist in an equilibrium with the resonance-stabilized alkylidene iminium salts.15 Equilibria of this type have been studied by ¹³C NMR spectroscopy.¹⁶ For example *N*,*N*-dimethyl(methylene)iminium chloride gives rise to four resonance signals at $\delta = 38.7$ (s), 49.4 (t), 79.0 (s) and 168.1 (t) in the ¹³C NMR spectrum (solvent: CH₂Cl₂/SO₂) demonstrating that there exists an equilibrium between 1-halogenalkylamine and the alkylidene iminium salt in solution. Introduction of electron-withdrawing substituents like trifluoromethyl groups results in a stabilization of the 1-halogenalkylamine form. The ¹³C NMR spectra of all *N*-chloromethyl¹⁰, *N*-bromomethyl¹⁷, *N*-iodomethyl¹⁸, and N-trifluoroacetoxy¹⁹ compounds of type **10** recorded so far give rise only to a single resonance signal for the Nmethylene carbon atom in the region of 30–73 ppm. No resonance absorption could be detected in the region of 160 ppm. From these findings we conclude that N-halogenmethyl-2,2-bis(trifluoromethyl)oxazolidin-5-ones exist mainly in the 1-halogenalkylamine form, with the halogen covalently bound. This could be the reason that olefins 11a and 11b are formed on heating compound 10, and not the Pictet-Spengler product, which should be generated via the methyleneiminium salt. In agreement with this assumption, compound 8 was obtained from 10 upon treatment with boron trifluoride etherate at room temperature.

On treatment with triethylsilane in the presence of trifluoroacetic acid compound **10** readily gave the HFA-protected *N*-methylphenylalanine **12**. Transformation into the unprotected *N*-methylphenylalanine **13** can be accomplished on heating (80 °C) in a dioxane-concd HCl mixture (Scheme 4). Compound **12** as activated species can be directly subjected to peptide synthesis ($12 \rightarrow 14$).





Solvents were purified and dried prior to use. Reagents were used as purchased. TLC was performed on alumina plates coated with Merck silica gel 60F₂₅₄. Compounds were visualized by spraying with a mixture of ceric(IV) nitrate, ammonium molybdate and H₂SO₄ followed by heating. Column chromatography was carried out on silica gel (32–63 μ m). For the determination of enantiomeric excess in 6a/6b, a Chiralpak AD column (Daicel Chemical Co. Ltd.) 25×0.46 cm, and as eluent system hexane-propan-2-ol-trifluoroacetic acid (90:10:0.05%, v/v), UV-detection (210 nm) and an injection volume of 20 µL were used. Under these conditions the enantiomers 6a and 6b eluted at 4.6 and 5.9 min, respectively. Melting points were determined with a Boëtius heating table. Optical rotations $([\alpha]_D)$ were measured using a Polartronic polarimeter (Schmidt & Haensch) in a 5 cm cell. For C, H, N analyses a CHNO-Rapid-Elemental-Analyser (Hereaus) was used. Mass spectra were recorded on a VG 12-250 (Masslab) electron ionization spectrometer (EI = 70 eV) or by VG ZAB-HSQ FAB spectrometer. ESI-FT-ICR-MS spectra were recorded on a Bruker Daltronics APEX II spectrometer (7 Tesla). IR spectra were obtained by using a FTIR spectrometer (Genesis ATI Mattson). ¹H (200.041 or 300.075 MHz), ¹³C (50.305 or 75.462 MHz) and ¹⁹F NMR (188.205 or 282.380 MHz) spectra were recorded on a Varian Gemini 200 or a Varian Gemini 300 spectrometer. Tetramethylsilane was used as reference standard for ¹H and ¹³C NMR spectra (internal) and trifluoroacetic acid for ¹⁹F NMR spectra (external). Abbreviations used are: Phe = phenylalanine, TFA = trifluoroacetic acid.

(4*S*)-4-Benzyl-2,2-bis(trifluoromethyl)-1,3-oxazolidin-5-one (5) Compound 5 was obtained²⁰ from 4 (16.5 g, 100 mmol) as a solid. Yield: 28.5 g (91%); mp 53–54 °C; $[\alpha]_D^{25}$ –44 (*c* = 1.6, CHCl₃).

 ^1H NMR (300 MHz, CDCl_3): δ = 2.98 (2 H, m, NH, CH_2), 3.25 (1 H, m, CH_2), 4.18 (1 H, m, CH), 7.23–7.41 (5 H, m, C_6H_5).

¹³C NMR (75 MHz, CDCl₃): δ = 38.5 (CH₂), 56.0 (CH), 88.4 [m, *C*(CF₃)₂], 120.1 (q, *J* = 284 Hz, CF₃), 121.1 (q, *J* = 287 Hz, CF₃), 127.8, 129.2, 129.3, 135.0 (C₆H₅), 170.6 (CO).

¹⁹F NMR (188 MHz, CDCl₃): $\delta = -2.77$ [m, C(CF₃)₂].

IR (KBr): v = 1827 (C=O) cm⁻¹.

MS (EI): m/z (%) = 313 (36) [M]⁺, 266 (16), 216 (12), 91 (100).

Anal. Calcd for $C_{12}H_9F_6NO_2$: C, 46.02; H, 2.90; N, 4.47. Found: C, 46.06; H, 2.87; N, 4.53.

(5S,10aS)-3,3-Bis(trifluoromethyl)-1-oxo-(1,2,3,4-tetra-

hydroisoquinolino)[2,3-*c*]oxazolidin-5-carboxylic Acid (6a) To a solution of oxazolidinone 5 (3.1 g, 10 mmol) and glyoxylic acid monohydrate (1.8 g, 20 mmol) in CH₂Cl₂ (30 mL) was added concd H₂SO₄ (4 mL) and stirred in a flask equipped with a bubbler at r.t. for several days. After consumption of the starting material (reaction control by ¹⁹F NMR), CH₂Cl₂ (60 mL) was added and then poured into a mixture of ice and aq NaOAc solution. After separation, the organic phase was washed with H₂O (2 × 50 mL), dried (MgSO₄) and evaporated. Triturating with pentane gave a crystalline powder, which was washed with pentane (2 × 30 mL). Filtration and drying in vacuo gave **6** as a white crystalline powder. Yield: 1.74 g (47%); mp 183–186 °C; $[\alpha]_D^{25}$ –101.8 (c = 1.0, CHCl₃); ee >99.9% (HPLC).

¹H NMR (200 MHz, acetone- d_6): $\delta = 3.05$ (1 H, m, CH₂), 3.24 (1 H, m, CH₂), 5.04 (1 H, m, CH), 5.11 (1 H, s, CHCO₂H), 7.29–7.58 (4 H, m, C₆H₄).

¹³C NMR (50 MHz, acetone-*d*₆): δ = 30.2 (CH₂), 49.7 (CH), 56.4 (CHCO₂H), 89.5 [m, *C*(CF₃)₂], 120.3 (q, *J* = 286 Hz, CF₃), 121.5 (q, *J* = 294 Hz, CF₃), 127.6, 127.9, 128.5, 129.4, 130.7, 131.5 (C₆H₄), 169.8 (CO), 177.1 (CO₂H).

¹⁹F NMR (188 MHz, acetone- d_6): $\delta = 0.43$ (q, J = 8.6 Hz, CF₃), 1.32 (q, J = 8.6 Hz, CF₃).

IR (KBr): v = 1838 (C=O), 1725 cm⁻¹.

MS (EI): m/z (%) = 369 (4) [M]⁺, 324 (33) [M - CO₂H]⁺, 302 (5), 226 (10), 130 (100).

Anal. Calcd for $C_{14}H_9F_6NO_4$: C, 45.54; H, 2.46; N, 3.79. Found: C, 45.14; H, 2.79; N, 3.78.

X-ray Crystallographic Data: Single crystals were grown from CHCl₃-hexane. Monoclinic, space group $P2_1/c$, T = 223 K; a = 13.391(1) Å, b = 7.009(1) Å, c = 15.627(1) Å, β = 93.77(1)°; V = 1464.5(2) Å³; Z = 4; D_c = 1.676 gcm⁻³; CCD-Diffractometer (BRUKER AXS), ω -scans (0.3°), 7982 data collected, 5260 independent reflections ($R_{int} = 0.037$), structure solution by direct methods,²¹ anisotropic refinement²² for all nonhydrogen atoms, hydrogen atoms refined isotropic, R1 = 0.0476; wR2 = 0.1232 [I >2 σ (I)]; and R1 = 0.0647; wR2 = 0.1398 for all data.²³

(5*R*,10a*R*)-3,3-Bis(trifluoromethyl)-1-oxo-(1,2,3,4tetrahydroisoquinolino)[2,3-*c*]oxazolidin-5-carboxylic Acid (6b)

Compound **6b** was prepared analogous to **6a** starting from (*R*)-phenylalanine. $[\alpha]_D^{25}$ +110 (*c* = 1, CHCl₃); ee >99.9% (HPLC). Other characteristic data are in agreement with compound **6a**.

(1*S*,3*S*)-1,2,3,4-Tetrahydroisoquinoline-1,3-dicarboxylic Acid (3)

Compound **6a** (0.74 g, 2 mmol) was dissolved in a mixture of propan-2-ol (2 mL)–H₂O (2 mL) and stirred for several days. After consumption of the starting material (reaction control by ¹⁹F NMR or TLC) the solvents were removed in vacuo. Recrystallization from EtOH–H₂O gave **3** as a white solid. Yield: 0.33 g (75%); mp 209– 211 °C (dec.); $[\alpha]_D^{25}$ –27 (*c* = 3, DMSO).

¹H NMR (200 MHz, D₂O): δ = 3.04 (1 H, m, CHCH₂), 3.22 (1 H, m, CHCH₂), 4.01 (1 H, m, CH), 5.07 (1 H, s, CHCO₂H), 7.11–7.39 (4 H, m, C₆H₄).

¹³C NMR (DMSO- d_6): $\delta = 29.3$ (CH CH_2), 51.9 (CH₂CH), 57.2 (CH), 126.1, 127.1, 128.1, 128.6, 130.0, 132.0 (C₆H₄), 170.1, 171.4 (2 CO₂H).

MS (EI): m/z (%) = 221 (5) [M]⁺, 176 (59) [M – CO₂H]⁺, 130 (100).

Anal. Calcd for C₁₁H₁₁NO₄·1.5H₂O: C, 57.38; H, 5.25; N, 6.08. Found: C, 57.20; H, 5.01; N, 5.77.

$(2S)-N^{\alpha}-[(1S,3S)-3-(1-Carboxy-1,2,3,4-tetrahydroisoquinolyl)-carbonyl]phenylalanine Amide (7)$

Compound **6a** (0.74 g (2 mmol) and (*S*)-phenylalanine amide (0.48 g, 3 mmol) were dissolved in propan-2-ol (4 mL) and stirred for several days. After consumption of the starting material (reaction control by ¹⁹F NMR or TLC) the precipitate was filtered and recrystallized (EtOH–H₂O) to give **7**. Yield: 0.34 g (47%); mp 218–221 °C; $[\alpha]_D^{25}$ –43 (*c* = 0.8, DMSO).

¹H NMR (200 MHz, DMSO-*d*₆): δ = 2.68 (1 H, m, CH_{2Phe}), 2.93 (3 H, m, CHC*H*₂, CH_{2Phe}), 4.14 (1 H, m, CH), 4.47 (1 H, m, CH_{Phe}), 4.55 (1 H, s, CHCO₂H), 7.00–7.43 (10 H, m, C₆H₄, C₆H₅ and CONH₂), 7.67 (1 H, s, CONH₂), 8.58 (1 H, m, CONH).

¹³C NMR (50 MHz, DMSO-*d*₆): δ = 30.2 (CHCH₂), 37.6 (CH_{2Phe}), 52.3 (CH), 54.0 (CH_{Phe}), 57.7 (CHCO₂H), 126.0, 126.5, 127.0, 128.0, 128.2, 128.4, 129.3, 131.8, 132.4, 137.9 (C₆H₄ and C₆H₅), 170.8, 171.3, 173.0 (CONH, CONH₂, CO₂H).

MS (ESI+): m/z = 368.1607 ([M + H]⁺ requires 368.1605).

$(10aS)\mbox{-}3,\mbox{3-Bis}(trifluoromethyl)\mbox{-}1\mbox{-}ox\mbox{-}o(1,2,3,\mbox{4-tetrahydroisoquinolino})[2,\mbox{3-}c]\mbox{0}ox\mbox{2}ox\mbox{2}ox\mbox{1}ox\mbox{1}ox\mbox{0}$

Compound **5** (1.56 g, 5 mmol) and paraformaldehyde (0.3 g, 10 mmol) were dissolved in CHCl₃ (2 mL)–TFA (2 mL). After 30 min (reaction control by ¹⁹F NMR or TLC) the solvents were removed in vacuo. Recrystallization from hexane gave **8** as a solid. Yield: 1.33 g (82%); mp 81–82 °C; $[\alpha]_D^{25}$ –114.3 (*c* = 1.0, CHCl₃).

¹H NMR (300 MHz, CDCl₃): δ = 3.07 (1 H, m, CHCH₂), 3.20 (1 H, m, CHCH₂), 3.89 (1 H, m, CH), 4.32 (1 H, m, CH₂), 4.43 (1 H, m, CH₂), 7.12–7.28 (4 H, m, C₆H₄).

¹³C NMR (75 MHz, CDCl₃): δ = 31.2 (CH₂CH), 47.3 (CH₂), 54.1 (CH), 89.7 [m, *C*(CF₃)₂], 120.5 (q, *J* = 285 Hz, CF₃), 121.7 (q, *J* = 293 Hz, CF₃), 126.6, 127.1, 127.4, 129.8, 131.2, 131.8 (C₆H₄), 168.6 (CO).

¹⁹F NMR (282 MHz, CDCl₃): δ = -0.26 (q, J = 8.7 Hz, CF₃), 3.76 (q, J = 8.7 Hz, CF₃).

IR (KBr): v = 1820 (C=O) cm⁻¹.

MS (EI): m/z (%) = 325 (46) [M]⁺, 278 (6), 256 (7), 228 (39), 104 (100).

Anal. Calcd for C₁₃H₉F₆NO₂: C, 48.01; H, 2.79; N, 4.31. Found: C, 47.94; H, 2.84; N, 4.23.

(3S)-1,2,3,4-Tetrahydroisoquinoline-3-carboxylic Acid (1)

Compound **8** (0.65 g, 2 mmol) was dissolved in a mixture of propan-2-ol (2 mL)–H₂O (2 mL) and stirred for several days. After consumption of the starting material (reaction control by ¹⁹F NMR or TLC) the solvents were removed in vacuo. Recrystallization from EtOH–H₂O gave **1** as a white solid. Yield: 0.25 g (72%); mp 304– 310 °C (Lit.¹ mp 325 °C); $[\alpha]_D^{25}$ –134 (*c* = 1.0, 0.1 N HCl) (Lit.¹³ $[\alpha]_D^{25}$ –139 (*c* = 1.0, 0.1 N HCl).

MS (EI): m/z (%) = 177 (7) [M]⁺, 132 (100) [M - CO₂H]⁺).

$(2S)-N^{u}-[(3S)-3-(1,2,3,4-tetrahydroisoquinolyl)carbonyl]-phenylalanine tert-Butyl Ester (9)$

Compound **8** (0.65 g, 2 mmol) and (*S*)-phenylalanine *tert*-butyl ester (0.67 g, 3 mmol) were dissolved in Et₂O (4 mL) and stirred for several days. After consumption of the starting material (reaction control by ¹⁹F NMR or TLC) the precipitate was filtered and stirred vigorously with Et₂O to give **9** as a white powder. Yield: 0.53 g (69%); mp 142–145 °C; $[\alpha]_D^{25}$ –5.3 (c = 1.0, CHCl₃).

¹H NMR (200 MHz, CDCl₃): $\delta = 1.45$ [9 H, s, C(CH₃)₃], 2.73 (1 H, m, CH_{2Phe}), 3.00–3.24 (3 H, m, CHCH₂, CH_{2Phe}), 3.56 (1 H, m, CH_{Phe}), 3.85 (1 H, m, CH₂), 3.96 (1 H, m, CH₂), 4.79 (1 H, m, CH), 7.04–7.27 (9 H, m, C₆H₄, C₆H₅), 7.70 (1 H, m, CONH).

¹³C NMR (50 MHz, CDCl₃): δ = 28.5 [C(CH₃)₃], 31.4 (CHCH₂), 38.6 (CH_{2Phe}), 47.6 (CH₂), 53.6 (CH_{Phe}), 56.7 (CH), 82.7 [C(CH₃)₃], 126.0, 126.7, 127.1, 127.4, 128.8, 129.5, 130.0, 134.8, 136.7, 136.8 (C₆H₄, C₆H₅), 171.2 (CONH), 173.2 (CO₂Bu-*t*).

MS (EI): m/z (%) = 380 (20) [M]⁺, 324 (71), 132 (100).

Anal. Calcd for $C_{23}H_{28}N_2O_3$: C, 72.60; H, 7.42; N, 7.36. Found: C, 72.84; H, 7.29; N, 7.72.

(4*S*)-4-Benzyl-2,2-bis(trifluoromethyl)-3-chloromethyl-1,3oxazolidin-5-one (10)

Compound **10** was obtained¹⁰ from **5** (6.27 g, 20 mmol) by Kugelrohr distillation as an oil. Yield: 4.70 g (65%); bp 90–100 °C (Kugelrohr)/0.7 Torr; $[a]_D^{25}$ +9.7 (c = 1.2, CHCl₃).

¹³C NMR (CDCl₃): δ = 35.9 (CH₂), 56.1 (CH), 59.1 (m, CH₂Cl), 90.1 [m, $C(CF_3)_2$], 120.4 (q, J = 287 Hz, CF₃), 121.0 (q, J = 290 Hz, CF₃), 128.4, 129.4, 129.8, 134.1 (C₆H₅), 168.2 (CO).

¹⁹F NMR (CDCl₃): $\delta = -0.43$ (q, J = 8.6 Hz, CF₃), 3.73 (q, J = 9.1 Hz, CF₃).

IR (film): v = 1845 (C=O) cm⁻¹.

MS (EI): m/z (%) = 361 (7) [M]⁺, 326 (30) [M – Cl]⁺, 132 (25), 91 (100).

Anal. Calcd for $C_{13}H_{10}ClF_6NO_2$: C, 43.18; H, 2.79; N, 3.87. Found: C, 43.20; H, 2.73; N, 4.08.

(Z)-4-Benzylidene-2,2-bis(trifluoromethyl)-3-methyloxazolidin-5-one (11a)

Compound **10** was stirred at 145 °C until the gas evolution (HCl) ceased (ca 1 h). The mixture of both isomers (**11a/11b**, 1:1) was separated by distillation over a Vigreux column (20 cm) in vacuo. Colorless oil; yield: 1.4 g (22%); bp 84 °C/1.0 Torr.

¹H NMR (200 MHz, CDCl₃): δ = 2.87 (3 H, m, CH₃), 6.64 (1 H, s, CH), 7.26–7.44 (5 H, m, C₆H₅). The olefinic proton gave no NOE with the *N*-methyl group.

¹³C NMR (50 MHz, CDCl₃): δ = 33.8 (CH₃), 84.0 [m, *C*(CF₃)₂], 110.0 (CH), 120.8 (q, *J* = 255 Hz, 2 CF₃), 126.2 (C), 128.7, 129.9, 130.1, 132.9 (C₆H₅), 162.7 (CO).

¹⁹F NMR (188 MHz, CDCl₃): $\delta = 1.10$ [s, C(CF₃)₂].

IR (film): v = 1823 (C=O), 1641 cm⁻¹.

MS (EI): m/z (%) = 325 (57) [M]⁺, 256 (36), 228 (64), 132 (11), 131 (100).

Anal. Calcd for $C_{13}H_9F_6NO_2$: C, 48.01; H, 2.79; N, 4.31. Found: C, 48.05; H, 2.76; N, 4.54.

(*E*)-4-Benzylidene-2,2-bis(trifluoromethyl)-3-methyloxazolidin-5-one (11b)

Distillation of the reaction mixture from **11a** gave as second fraction **11b**. Yield: 1.8 g (28%); slowly crystallizing colorless oil; mp 44 °C; bp ~100 °C/1.0 Torr.

¹H NMR (200 MHz, CDCl₃): δ = 3.09 (3 H, m, CH₃), 5.99 (1 H, s, CH), 7.26–7.36 (3 H, m, C₆H₅), 7.55–7.59 (2 H, m, C₆H₅). The olefinic proton gave a NOE with the *N*-methyl group.

¹³C NMR (50 MHz, CDCl₃): δ = 31.1 (CH₃), 88.2 [m, *C*(CF₃)₂], 112.6 (CH), 120.9 (q, *J* = 292 Hz, 2 CF₃), 126.2 (C), 128.6, 128.7, 130.2, 132.7 (C₆H₅), 160.0 (CO).

¹⁹F NMR (188 MHz, CDCl₃): $\delta = 0.76$ [s, C(CF₃)₂].

IR (film): v = 1823 (C=O), 1657 cm⁻¹.

MS (EI): m/z (%) = 325 (70) [M]⁺, 278 (9), 256 (51), 228 (7), 131 (100).

Anal. Calcd for $C_{13}H_9F_6NO_2$: C, 48.01; H, 2.79; N, 4.31. Found: C, 47.96; H, 2.81; N, 4.39.

(4S)-4-Benzyl-2,2-bis(trifluoromethyl)-3-methyl-1,3-oxazolidin-5-one (12)

Compound **12** was obtained¹⁰ from **5** (6.27 g, 20 mmol) via **10** (see above) as a colorless oil. Yield: 5.3 g (81%); bp 63–65 °C/0.7 Torr; $[\alpha]_D^{25}$ +14.6 (c = 3.6, CHCl₃).

 ^1H NMR (200 MHz, CDCl_3): δ = 2.64 (3 H, m, CH_3), 3.04 (1 H, m, CH_2), 3.18 (1 H, m, CH_2), 3.84 (1 H, m, CH), 7.19–7.38 (5 H, m, C_6H_5).

¹³C NMR (50 MHz, CDCl₃): δ = 33.8 (br, CH₃), 36.7 (CH₂), 62.3 (CH), 90.0 [m, *C*(CF₃)₂], 120.8 (q, *J* = 285 Hz, CF₃), 122.1 (q, *J* = 294 Hz, CF₃), 127.8, 129.0, 130.3, 135.2 (C₆H₅), 169.7 (CO).

¹⁹F NMR (188 MHz, CDCl₃): δ = -1.69 (q, J = 8.1 Hz, CF₃), 4.32 (q, J = 8.1 Hz, CF₃).

IR (film): v = 1842 (CO) cm⁻¹.

MS (EI): m/z (%) = 327 (60) [M]⁺, 280 (25), 258 (7), 236 (64), 91 (100).

Anal. Calcd for $C_{13}H_{11}F_6NO_2$: C, 47.70; H, 3.36; N, 4.28. Found: C, 47.44; H, 3.28; N, 4.47.

(2S)-N-Methylphenylalanine (13)

Compound **12** (0.65 g, 2 mmol) was dissolved in dioxane (3 mL)– concd HCl (3 mL) and stirred at 80 °C for 12 h. The solution was evaporated to dryness and the residue dissolved in EtOH (1 mL). Stirring with propylene oxide (3 mL) and drying the precipitate gave **13** as a white powder. Yield: 0.21 g (59%); mp 233–241 °C (Lit.²⁴ mp 243–246 °C); $[\alpha]_D^{25}$ +19.0 (*c* = 0.5, 5 N HCl) {Lit.²⁴ $[\alpha]_D^{25}$ +7.6 (*c* = 0.5, 5 N HCl}.

Analytical data are identical with the literature data.²⁴

(2*S*)-*N*^u-**[**(2*S*)-*N*-Methylphenylalanyl]phenylalanine Amide (14) Compound 12 (0.65 g, 2 mmol) and (*S*)-phenylalanine amide (0.48 g, 3 mmol) were dissolved in propan-2-ol (4 mL) and stirred for several days. After consumption of the starting material (reaction control by ¹⁹F NMR or TLC) the solvent was evaporated and the residue purified by flash chromatography (R_f 0.25) (eluent: light petroleum–CHCl₃–MeOH, 3:7:1) and recrystallization (EtOH–H₂O) to give 14 as needles. Yield: 0.49 g (72%); mp 79–81 °C; $[\alpha]_D^{25}$ +8.4 (*c* = 1.5, CHCl₃ + 5% TFA).

¹H NMR (300 MHz, CDCl₃ + 5% TFA): δ = 2.71 (3 H, s, CH₃), 2.74 (1 H, m, CH_{2Phe}), 2.92 (1 H, m, CH_{2Phe}), 3.15 (2 H, m, CH₂), 4.26 (1 H, m, CH), 4.85 (1 H, m, CH_{Phe}), 6.16 (1 H, s, CONH₂), 6.90 (1 H, s, CONH₂), 7.01–7.30 (10 H, m, 2 C₆H₅), 7.65 (1 H, m, CONH).

 ^{13}C NMR (75 MHz, CDCl_3 + 5% TFA): δ = 32.9 (CH_3), 37.6 (CH_2), 38.8 (CH_{2Phe}), 55.4 (CH_{Phe}), 63.3 (CH), 128.5, 129.5, 129.7, 129.8, 130.2, 132.6, 135.4 (2 C_6H_5), 168.1 (CONH), 175.4 (CONH_2).

MS (ESI+): m/z = 326.18607 ([M + H]⁺ requires 326.1863).

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