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Analogues of doxanthrine reveal differences between the dopamine D₁ receptor binding properties of chromanoisoquinolines and hexahydrobenzo[*a*] phenanthridines

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1. Introduction

ABSTRACT

Efforts to develop selective agonists for dopamine D_1 -like receptors led to the discovery of dihydrexidine and doxanthrine, two bioisosteric β -phenyldopamine-type full agonist ligands that display selectivity and potency at D_1 -like receptors. We report herein an improved methodology for the synthesis of substituted chromanoisoquinolines (doxanthrine derivatives) and the evaluation of several new compounds for their ability to bind to D_1 - and D_2 -like receptors. Identical pendant phenyl ring substitutions on the dihydrexidine and doxanthrine templates surprisingly led to different effects on D_1 -like receptor binding, suggesting important differences between the interactions of these ligands with the D_1 receptor. We propose, based on the biological results and molecular modeling studies, that slight conformational differences between the tetralin and chroman-based compounds lead to a shift in the location of the pendant ring substituents within the receptor.

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Central dopamine neurotransmission has evolved to mediate important sensory processing functions such as novelty detection, attention, memory formation, and coding of rewarding stimuli [1,2]. Executive and volitional functions such as reward seeking, behavioral reinforcement, and motor control are likewise mediated by dopaminergic pathways [3]. Furthermore, dopaminergic systems are involved in the manifestation of CNS pathologies including Parkinson's disease [4], schizophrenia [5], and substance abuse [6].

We have been interested for more than two decades in investigating the physiological role of the dopamine D_1 receptor by developing selective ligands for the activation of this receptor subtype. Structure-activity relationship studies of dopamine analogs have enabled the recognition of a "β-phenyldopamine pharmacophore" [7,8] as one template for the development of compounds displaying preference for the activation of D₁-like receptors over D₂-like receptors. This predictive model has enabled the discovery of several molecules that have shown significant potency and selectivity for dopamine D₁-like receptors [9–12]. The prototype of this type of compound is dihydrexidine 1 (DHX, Fig. 1.) [13], a hexahydrobenzo[a]phenanthridine developed as a conformationally-restricted molecule that incorporates the β-phenyldopamine pharmacophore, and is a potent dopamine D₁ full agonist with ten-fold higher affinity for D₁-like receptors over D₂like receptors [9]. This compound demonstrated significant therapeutic potential in an in vivo model of Parkinson's disease in African green monkeys [14] and helped to establish the importance of D₁ receptor activation in the control of motor function. Dihydrexidine also has been shown to increase blood flow in the prefrontal cortex [15], and improve working memory in aged monkeys [16].

Given the success that has been achieved with the development of bioisosteres of certain dopamine agonists, and in view of the

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Fig. 1. Structures of β -phenyldopamine-based dopamine D₁ receptor agonists.

results of Horn et al. [17], who reported a ten-fold loss of D₂ affinity by introduction of an oxygen atom into the structure of the D_2 agonist 6,7-ADTN, it was conjectured that a similar modification of the structure of DHX might yield a ligand with enhanced D₁selectivity. This line of reasoning led to the discovery of doxanthrine (+)–2, (DOX, Fig. 1.), a chromanoisoquinoline bioisostere of **1** that displays a greater than 300-fold selectivity for D_1 -like receptors over D₂-like receptors, with higher efficacy than dopamine for activation of the D_1 receptor [18].

To develop structure-activity relationships of compounds derived from the new chromanoisoquinoline template, we reviewed modifications of the hexahydrobenzo[*a*]phenanthridine ring system that had resulted in compounds with improved biological properties. In the DHX series, substitution on the β -phenyl ring at the 2-position with methyl and ethyl groups gave ligands with increased D_1 -selectivity as a result of decreased D_2 affinity [19] (Fig. 2). We hypothesized that compounds with analogous substitution at the 11-position of **2** would display D₁-selectivity greater than the parent compound. We also wished to investigate the effects of electronegative substituents on the β-phenyl ring of doxanthrine, which had not been chemically accessible in the DHX series. To pursue these directions, we prepared compounds bearing methyl, ethyl, fluoro, and trifluoromethyl substituents at the 11position of the chromanoisoquinoline template (compounds **5a**–**d**, Fig. 2) and assessed their affinities at D_1 and D_2 receptors. In addition, because the chroman oxygen of DOX appears to be instrumental in conferring D1-selectivity to the ligand, we prepared and tested the sulfur-containing thiochromanoisoquinoline 6. The differences in polarity and size of the sulfur atom in relation to the methylene group of DHX and the oxygen atom of DOX will allow a more complete survey of the nature of the receptor residue(s) that are situated in proximity to this important position when the ligand is bound to the D₁ receptor. The results of these studies are reported herein.

2. Results and discussion

2.1. Chemistry

The synthesis of compounds **5a**–**d** (Scheme 1) was based on a variation of the methodology employed to prepare doxanthrine, 2 [18]. The key step of this synthesis involves a Michael-type addition of an arvl-metal reagent to nitrochromene **11** (Scheme 1).

Starting from commercially available sesamol 7, aldehyde 10 was prepared in very modest yield by Friedel-Crafts-type formylation using dichloromethyl methyl ether and SnCl₄ (Scheme 2). Unfortunately, direct conversion of sesamol into the phenolic benzaldehyde 10 was accompanied by the formation of large amounts of insoluble, deep-blue-colored xanthylium ion 8, which typically precipitated out of the dichloromethane reaction medium.

Xanthylium ion 8 appears to form from a chloromethoxymethyl intermediate under the formylation conditions, which is sufficiently reactive, particularly in the presence of Lewis acids, to be attacked by the electron rich aromatic ring of another molecule of sesamol. This heterodimerization product is then poised to undergo intramolecular attack by the phenolic hydroxy to yield a hemiacetal, which can aromatize to yield a xanthydrol ether. Aided by the electron donating oxygen atoms that flank the aromatic rings, the xanthydrol ether species will readily eliminate methanol, forming a resonance-stabilized xanthylium ion, 8, which displays coloration and solubility properties consistent with similar compounds reported previously in the literature [20,21].

To avoid formation of this xanthylium side-product, sesamol was acetvlated prior to Friedel-Crafts formylation. This simple procedure allowed formation of aldehyde 10 in excellent yield and essentially free from the dimerization by-product. The desired nitrochromene 11 was then obtained in one step by treatment of 10 with nitroethylene generated in situ [22,23]. Extremely slow addition of 2-nitroethanol ensured minimization of base-catalyzed polymerization of nitroethylene, which was nevertheless a significant side reaction. Using this methodology, it was possible to prepare relatively large amounts of **11**.

Nitrochromene **11** is known to undergo Michael-type addition of aryl-metal reagents to provide exclusively trans adducts [18]. Thus, we anticipated that an appropriately substituted aryl-metal reagent would allow access to 11-substituted doxanthrine analogs 5a-d. In the original synthesis of DOX 2, a Grignard reagent was employed. That approach, however, suffered severe limitations because of the requirement for ortho/para-substituted aryl halides that are not readily available or easily synthesized. We therefore reasoned that *para*-substituted aryloxazolines **13a**-**d** (Scheme 3),



Fig. 2. Structures and dopamine receptor affinities of substituted hexahydrobenzo[a]phenanthridines and proposed analogues.



Scheme 1. Retrosynthetic analysis for the preparation of compounds 5a-d and 6.



Scheme 2. Synthesis of nitrochromene 11 and xanthylium side-product 8.

which are easily accessible from commercially available *para*substituted benzoic acids, could be *ortho*-lithiated, and after treatment with nitrochromene **11**, would allow exclusive production of the appropriately substituted *trans* adducts **14a**–**d**. The oxazoline moiety of these intermediates was anticipated to undergo facile acid hydrolysis to yield the carboxylic acids, which would allow eventual formation of the isoquinoline ring of the final products. Thus, we reasoned that *para*-substituted aryloxazoline



Scheme 3. Synthesis of chromanoisoquinolines 5a-d.



Scheme 4. Synthesis of thiochromanoisoquinoline 6.

reagents would function as versatile synthons in the preparation of a variety of substituted isoquinolines **17a**–**d**.

The completion of the synthesis of compounds **5a**–**d** is shown in Scheme 3. Treatment of nitrochromene **11** with organolithium reagents prepared by *ortho*-lithiation of aryloxazolines **13a**–**d**, gave, as anticipated, exclusively the *trans* adducts **14a**–**d**. Mild treatment of compounds **14a**–**d** with aqueous HCl gave the amine salts **15a**–**d** as products of partial hydrolysis of the oxazoline ring. Reduction of the nitro groups of these compounds with powdered zinc in acetic acid led to intermediate amine salts that upon basification and gentle heating readily cyclized to form insoluble lactams **16a**–**d**, which crystallized upon formation. These lactams were then reduced by treatment with borane in THF, and the resulting amines **17a**–**d** were demethylated with BCl₃ to yield the catecholamine hydrochloride target compounds **5a**–**d**.

To prepare the thiochromanoisoquinoline analog **6** we employed a strategy parallel to the synthesis of **2** [18], but employing the nitrothiochromene **20** as the Michael acceptor for the addition of aryl-Grignard reagent **21**. The transformations that led to **6** are shown in Scheme 4.

To prepare nitrothiochromene **20**, we relied on the Newman-Kwart rearrangement [24] of phenyl-*O*-thiocarbamate **18** into phenyl-*S*-thiocarbamate **19**. Thus, we prepared **18** from the previously synthesized sesamaldehyde **10**, and heated it in diphenyl

Table 1

Binding affinity of new compounds at D_1 - and D_2 -like receptors in porcine striatal tissue.

Ligand	Binding in porcine striatal homogenates ^a		
	D ₁ -like	D ₂ -like	Binding selectivity
	Ki (nM)	Ki (nM)	(D ₂ -like/D ₁ -like)
DOX (2)	26 ± 4.5	1700 ± 190	80 ± 19
11-Me-DOX (5a)	150 ± 25	6800 ± 860	55 ± 13
11-Et-DOX (5b)	180 ± 34	6400 ± 900	49 ± 14
11-F-DOX (5c)	110 ± 18	740 ± 110	8.5 ± 2.1
11-CF3-DOX (5d)	570 ± 87	6900 ± 980	14 ± 3.1
6	200 ± 41	2500 ± 410	15 ± 5.6
SCH23390	$\textbf{0.46} \pm \textbf{0.08}$	NA	NA
Chlorpromazine	NA	$\textbf{7.6} \pm \textbf{2}$	NA

 $^{a}\,$ All results shown are the mean \pm SEM for 4-11 independent experiments.

ether at 250 °C for 15 min to effect the rearrangement, obtaining modest yields of **19**. Basic hydrolysis of **19** gave a thiophenol that proved to be relatively unstable. By immediate treatment of the hydrolysate mixture with 2-nitroethanol, phthalic anhydride, and dibutylamine, we were able to prepare good quantities of nitro-thiochromene **20**, isolated as bright orange crystals. The methodology followed in the subsequent steps was basically identical to that employed in the synthesis of **2** [18].

2.2. Pharmacology

The affinities of compounds **5a**–**d** and **6** for dopamine D_1 -like and D_2 -like receptors were evaluated in a radioligand displacement assay using porcine striatal tissue preparations. Those results are shown in Table 1. Standard ligands for D_1 - and D_2 -like receptors, SCH-23390 and chlorpromazine, were included in the assays for comparison.

3. Conclusions

Competition binding assays using porcine striatal preparations revealed that all four of the 11-substituted analogs **5a**–**d** had significantly lower affinity at D₁-like receptors than their structural predecessor **2**, DOX. The methyl-substituted compound **5a** had a nearly six-fold decrease in D₁ affinity (D₁ K_i = 150 nM) compared to **2** (D₁ K_i = 26 nM). Given that introduction of a methyl group at the analogous 2-position of DHX (compound **3**) did not significantly reduce the affinity of the ligand in comparison to DHX, the loss of affinity for compound **5a** was unexpected. Extension of the hydrophobic bulk to an ethyl group (compound **5b**) incurred an even greater loss of D₁ affinity (D₁ K_i = 185 nM) compared to **2**. Although the ethyl substituted analog of DHX (compound **4**) also displayed affinity lower than the unsubstituted compound [19], its chroman isostere **5b** clearly shows a more dramatic negative effect of the ethyl substituent on affinity.

The disparity between the effects brought about by introduction of identical alkyl substitutions at the 11-position of doxanthrine (**2**) and the analogous 2-position of dihydrexidine, **1** must reflect critical differences between the interactions of these two isosteric ligands within the D_1 dopamine receptor. The D_1 binding affinities

of **1** and **2** reflect very similar ΔG° values for their interaction with the D₁ receptor, revealing that, from a thermodynamic point of view, the net effect of replacing the 8-methylene of 1 with an oxygen atom was insignificant. When one considers the potential for solvation of the chroman oxygen of 2 and its analogs in an aqueous environment, and thus, the desolvation energy that must be overcome upon binding of **2** to the D_1 receptor, the similar D_1 binding affinities of **1** and **2** can most easily be explained by invoking the formation of energetically favorable interactions between **2** and some element(s) of the D_1 binding site that can compensate for the energetically unfavorable desolvation process. Aside from inductive effects on the amine group, it appears likely that the chroman oxygen of DOX and its analogs may be engaged in a specific interaction when bound to the D₁ receptor. Replacement of the chroman oxygen of DOX with a sulfur atom resulted in a nearly eight-fold decrease in D₁ affinity.

After performing docking, molecular dynamics, and energy minimization of both methyl-substituted derivatives, **3** and **5a**, in our previously constructed "activated" model of the D₁ receptor [25], a slight conformational difference was observed between the DOX and DHX analogs, with the pendant phenyl ring deviating slightly more from the catechol plane in **5a** than in **3**. This change forces the DOX analog to lean more into the space occupied by TM6 (and TM7) than the DHX analog, toward residues at the extracellular end of TM6.

It is postulated that these relatively minor conformational differences can be tolerated by the D_1 receptor for the parent ligands, DHX (2) and DOX (5), but that they result in differing tolerance for substituents at the 2- or 11-positions of the pendant phenyl rings, respectively. This effect is most evident in the case of the methyl-substituted compounds, where the DHX analogue largely retains its affinity but the DOX-derived structure does not. Unfortunately, our molecular modeling studies have not allowed us to identify what can only be a subtle structural basis for the difference, but very recent mutagenesis experiments of the D₁ dopamine receptor in our laboratory have shown that residues in extracellular loop 2 (EL2), particularly L190, differentially affect the affinity of 3 and 5a (to be published elsewhere). Specifically, mutant L190A had significantly increased affinity for 5a, but not for 3, suggesting a reduced steric interaction and/or increased hydrophobic interaction between the 11-alkyl substituent of 5a and the smaller methyl side chain of alanine. Unfortunately, the state of the art in modeling extracellular loops in GPCRs is not very advanced, which may explain our inability to identify ligand-receptor interactions that can account for our results, and reinforcing our observation that the differences in the binding poses of the two series of ligands must be subtle.

Substitution with fluoro and trifluoromethyl groups gave sharp decreases in D_1 affinity compared with the unsubstituted ligand **2**. The effect was more prominent with the bulkier trifluoromethyl group (**5d**), which gave a 22-fold decrease in D_1 affinity compared to **2**, and an almost four-fold decrease compared to the methyl-substituted compound **5a**. These comparisons suggest that both steric and electronic factors may account for the decreased affinity of compound **5d**. In general, electron-withdrawing groups were especially detrimental to affinity, suggesting that aromatic stacking interactions may not be the driving force behind binding in this accessory region [26].

In summary, development of a novel methodology for the production of variously substituted *trans*-fused chromanoisoquinolines has allowed comparison of the receptor binding properties of bioisosteres and revealed important differences between the ligand/receptor complexes of DHX **1** and DOX **2** with the D₁ dopamine receptor. Explanations for the divergent SAR of the two series appear likely to be based on subtle conformational

differences caused by the presence of the heteroatom at the 4position of **2**, resulting in differential interactions with residues within the ligand binding site.

4. Experimental

4.1. Chemistry

All reagents were commercially available (Aldrich) and were used without further purification unless otherwise indicated. Dry THF and diethyl ether were obtained by distillation immediately before use from benzophenone-sodium under nitrogen. Column chromatography was carried out using silica gel 60 (230-400 mesh). J.T. Baker flexible thin layer chromatography sheets (silica gel IB2–F) were used to monitor reactions. Melting points were determined using a Thomas-Hoover apparatus and are uncorrected. H¹-NMR spectra were recorded using a 300 MHz Bruker ARX-300 NMR spectrometer. Chemical shifts are reported in δ values ppm relative to an internal reference (0.03%, v/v) of tetramethylsilane (TMS) in CDCl₃, except where noted. Chemical ionization mass spectra (CIMS) using isobutane as a carrier gas were obtained with a Finnigan 4000 spectrometer. Elemental analyses were performed by the Purdue University Microanalysis Laboratory. All the reactions were carried out under an inert atmosphere of argon.

4.2. Synthesis

4.2.1. 9H-Bis([1,3]dioxolo[4,5-b:4',5'-i])xanthylium chloride (8)

In a dry two-neck flask and under an inert atmosphere, sesamol (50.98 g, 369 mmol) was dissolved in 600 mL of CH₂Cl₂. Into this flask 52 mL (448 mmol) of SnCl₄ were added and the solution was cooled to 0 °C Cl₂CHOCH₃ (35 mL, 387.5 mmol) was then added dropwise and the mixture was warmed to room temperature and stirred for 3 h. The mixture was allowed to settle and the solvent was removed by decanting, keeping a positive pressure of nitrogen through one of the flask necks. Into the flask were then added 200 mL of CH₂Cl₂, and the mixture was vigorously stirred for 20 min. The solvent was again removed by decanting and 200 mL of CH₂Cl₂ were added. The mixture was filtered using a Buchner funnel and the solids rinsed with CH₂Cl₂. The collected solids were placed under vacuum to yield 51.8 g (46%) of a bluegreen solid. ¹H NMR (acetone-d₆): δ 5.85 (4H, OCH₂O), 6.11 (1H, CH), 6.32 (2H, ArH), 6.45 (2H, ArH); EIMS: *m*/*z* 269 (M⁺, 100); HRMS Calculated for $C_{15}H_{11}O_6$ (xanthylium-H₂O solvate) 287.0556, found 287.0166.

4.2.2. 5-Hydroxybenzo[1,3]dioxole acetate (9)

In a two-neck flask fitted with a mechanical stirrer, a suspension of 26 g (1.086 mol) of NaH in 300 mL of dry THF was cooled and stirred on an ice bath. A solution of 75 g (0.543 mol) of sesamol in 300 mL of dry THF was added dropwise over 1 h. After the addition, 205 mL (2.174 mol) of acetic anhydride were added, resulting in a thick suspension. This mixture was stirred for 3 h at RT. The reaction mixture was then cooled to 0 °C and quenched by slow careful addition of 2N HCl. Water (300 mL) was then added and the solution was extracted with CH_2Cl_2 (3 × 50 mL). These extracts were washed once with 50 mL of water, 50 mL of brine, and were then dried over MgSO₄. The mixture was filtered and the solvents removed by rotary evaporation. Kugelrohr distillation at 80 °C and 0.1 torr gave 94.89 g (97%) of the product as a clear oil. This material had properties identical to those reported previously for this compound [27].

4.2.3. 6-Hydroxybenzo[1,3]dioxole-5-carboxaldehyde (10)

In a two-neck flask, 40 g of **9** (0.222 mol) were dissolved in 500 mL CH₂Cl₂ and the solution cooled to 0 °C. Through a dropping funnel, 52 mL (0.444 mol) of SnCl₄ were added dropwise, followed by slow addition of 22 mL (0.244 mol) of Cl₂CHOCH₃, causing the formation of a precipitate. The reaction was stirred for 2 h and was then poured over ice. The mixture was partitioned and the organic layer was washed with of 2M HCl (3×20 mL), and then with water (2×50 mL). The organic solution was dried over Na₂SO₄, filtered, and the solvent removed by rotary evaporation to afford a solid, which was triturated under cold MeOH, filtered, and dried to yield 34.38 g (93.2%) of **10** as a tan solid: mp 120–121 °C (Lit [28]. mp 125–126). ¹H NMR (CDCl₃): δ 9.63 (s, 1H, CHO), 6.87 (s, 1H, ArH), 6.47 (s, 1, ArH), 6.02 (s, 2H, ArOCH2O), 1.54 (s, 1H, ArOH). CIMS: *m*/*z* 167 (M + H⁺, 100). Anal. (C₈H₆O₄) C, H.

4.2.4. 3-Nitro-2H-6,7-methylenedioxychromene (11)

In a three-necked 1 L flask equipped with a Dean Stark trap, condenser, and mechanical stirring, 34.38 g (0.207 mol) of sesamaldehyde 10 were dissolved in 500 mL of toluene containing 17.5 mL of di-n-butylamine (0.104 mmol) and 61.3 g (0.414 mol) of phthalic anhydride. This mixture was vigorously stirred while heating at reflux. Through a cannula, 32.65 mL (0.455 mol) of 2nitroethanol were added into the reaction flask over 4.5 days using a syringe pump to maintain a rate of 0.3 mL/h. The reaction was stirred at reflux for 12 more hours, and then cooled to RT. The contents were filtered through Celite, and the mixture was washed with 2 N NaOH (4×300 mL), brine (100 mL), and then dried over MgSO₄. After filtration, the organic solution was concentrated under reduced pressure and the crude concentrate was passed through a short column of silica to remove dark polar impurities. The solvent was removed under reduced pressure, and the residual red powder was triturated under 2:1 hexanes/EtOAc. The solid was collected by filtration and dried under vacuum to yield 35.56 g (77.7%) of pure nitrochromene **11** as a bright red powder: mp 139 °C; ¹H NMR (CDCl₃): δ 5.20 (s, 2H, ArOCH₂); 6.02 (s, 2H, OCH₂O); 6.49 (s, 1H, ArH); 6.69 (s, 1H, ArH); 7.75 (s, 1H, ArCH). CIMS: *m*/*z* 222 (MH⁺, 100). Anal. (C₁₀H₇NO₅) C, H, N.

4.2.5. (\pm) -Trans-4-[2-(4,4-dimethyloxazolin-2-yl)-5methylphenyl]-6,7-methylenedioxy-3-nitrochroman (**14a**)

Toluyloxazoline 13a [29] (3.85 g, 20.34 mmol) was dissolved in 40 mL of dry THF and stirred at -45 °C in a dry ice/chlorobenzene bath. Using a syringe, 8.14 mL (20.34 mmol) of a 2.5 M solution of *n*BuLi in hexanes was slowly added, and the solution turned bright orange. The reaction was stirred for 1 h and was then cannulated into a flask containing 3.0 g (13.56 mmol) of nitrochromene 11 dissolved in 200 mL of dry THF previously cooled to -78 °C. This mixture was allowed to warm to room temperature over 1 h, and was then quenched with an aqueous solution of saturated NH₄Cl. The mixture was extracted with CH_2Cl_2 (4 \times 30 mL), the pooled organic extracts were rinsed with water (50 mL), and then brine (20 mL). The extracts were dried over MgSO₄, filtered, and the solvent removed under reduced pressure to yield a dark oil. This oil was dissolved in 40 mL of MeOH, whereupon immediate crystallization occurred. The crystallizing mixture was then kept at 0 °C overnight, filtered, and the collected solids rinsed with cold methanol to yield 3.27 g of pure product as cream-colored crystals. A second crop was obtained from MeOH to yield a total of 3.52 g (65.5%) of the product mp 172–175 °C. ¹H NMR (CDCl₃): δ 7.85 (d, 1H, ArH); 7.14 (d, 1H, ArH); 6.79 (s, 1H, ArH); 6.45 (s, 1H, ArH); 6.32 (s, 1H, ArH); 5.92-5.86 (m, 2H, OCH₂); 4.21 (br, 1H, ArCHAr); 4.93 (br, 1H, CHNO₂); 4.65-4.59 (m, 1H, OCH₂); 4.15-4.11 (dd, 1H, $J_{\text{gem}} = 11.7 \text{ Hz}, J_{\text{vic}} = 2.4 \text{ Hz}$; 4.06 (s, 2H, oxazoline CH₂); 2.28 (s, 3H, CH₃); 1.32–1.3 (2s, 6H, 2CH₃). CIMS: m/z 411 (M + H⁺, 100). Anal. (C₂₂H₂₂N₂O₆) C, H, N.

4.2.6. (±)-Trans-2,3-methylenedioxy-11-methyl-6a,12b-dihydro-6H-chromeno[3,4-c]isoquinolin-8-one (**16a**)

One gram of nitro-oxazoline 14a was dissolved in 60 mL THF and 20 mL of a 2N aqueous HCl solution were added. This reaction was stirred for 48 h. and then the solution was concentrated under reduced pressure to one-half of its initial volume. The solution was then extracted with EtOAc (3 \times 30 mL), the extracts were washed once with water (10 mL), dried over MgSO₄, filtered, and the solvent removed under reduced pressure to yield 940 mg of the hydrochloride salt 15a as a tan solid that was used without further purification. Hydrochloride 15a (2.9 g, 6.237 mmol) was dissolved in 50 mL CH₃COOH, 5 g of Zn powder were added, and the mixture was stirred under an inert atmosphere for 3 h. The reaction was filtered through Celite, and the zinc salts on the filter were rinsed with THF. The filtrate was concentrated under reduced pressure and 10 mL of MeOH were added to the residue. This solution was then basified to pH 9 by slow addition of concentrated aqueous ammonia. This mixture was warmed to 50 °C and stirred for 1 h. After cooling, the solid was collected by filtration, washed on the filter with cold MeOH, and dried to yield 1.45 g (67%) of fine, white, cotton-like needles: mp >300 °C (dec). ¹H NMR (CDCl₃): δ 7.98 (d, 1H, ArH); 7.46 (s, 1H, ArH); 6.99 (s, 1H, ArH); 6.51 (s, 1H, NH); 6.00–5.96 (2d, 2H, OCH₂O); 4.30–4.26 (dd, 1H, OCH₂, *J*_{gem} = 9.1 Hz, $J_{\text{vic}} = 3.6 \text{ Hz}$; 4.21 (d, 1H, ArCHAr, $J_{trans} = 11.1 \text{ Hz}$); 3.95 (t, 1H, OCH₂, $J_{\text{gem}} = 9.1 \text{ Hz}$; 3.89–3.86 (dd, 1H, CHN, $J_{trans} = 11.1 \text{ Hz}$, $J_{\text{vic}} = 3.6 \text{ Hz}$); 2.41 (s, 3H, CH₃). EIMS: *m*/*z* 309 (M⁺, 100). Anal. (C₁₈H₁₅NO₄) C, H, N.

4.2.7. (±)-Trans-2,3-methylenedioxy-11-methyl-6a,7,8,12btetrahydro-6H-chromeno[3,4-c]-isoquinoline (**17a**)

Lactam 16a (1.3 g, 4.28 mmol) was suspended in 200 mL of dry THF and stirred at reflux. Into this flask, 21.4 mL (21.4 mmol) of a 1M BH₃ solution in THF were added and the reaction was stirred at reflux for 30 h. The mixture was then cooled to 0 °C and water was added carefully to quench the reaction. The mixture was concentrated under reduced pressure to about one-third of its original volume and 100 mL of water were added. The mixture was then extracted with EtOAc (2 \times 15 mL) and the extracts dried over MgSO₄. The drying agent was then removed by filtration and the filtrate concentrated under reduced pressure to afford a solid that was dissolved in 30 mL of a 2 M solution of HCl in ethanol. This solution was stirred at 70 $^\circ C$ for 40 min and then cooled to 0 $^\circ C$ overnight to induce crystallization. The crystals were collected by filtration to obtain 1.06 g of the HCl salt of 17a. A second batch was obtained by crystallization from EtOH to obtain an additional 0.13 g. The combined crops of this salt were suspended in MeOH, and ammonia was added to pH 9. Water was added, the suspension was extracted with CH₂Cl₂, the extracts dried over MgSO₄, filtered, and concentrated under reduced pressure to yield 1.05 g (85%) of the amine. An analytical sample was obtained by recrystallization from MeOH: mp: 97–99 °C. ¹H NMR (MeOD): δ 7.26–7.23 (m, 2H, 2ArH); 7.16-7.13 (d, 1H, ArH); 6.86 (s, 1H, ArH); 6.48 (s, 1H, ArH); 5.91-5.88 (2d, 2H, OCH₂O); 4.46-4.39 (m, 3H, OCH₂, CH₂N); 4.22 (d, 1H, ArHAr, $J_{trans} = 11.4 \text{ Hz}$; 4.03 (t 1H, OCH₂, $J_{gem} = 10.5 \text{ Hz}$); 2.30 (s, 3H, CH₃). EIMS: *m*/*z* 295 (M⁺, 100). Anal. (C₁₈H₁₇NO₃) C, H, N.

4.2.8. (±)-Trans-11-methyl-2,3-dihydroxy-6a,7,8,12b-tetrahydro-6H-chromeno[3,4-c]isoquinoline hydrobromide (**5a**)

Amine **17a** (300 mg, 1.02 mmol) was dissolved in 15 mL of CH_2Cl_2 under an inert atmosphere and the solution was cooled to -78 °C. Into this flask, 8.1 mL (8.1 mmol) of 1M BBr₃ in CH_2CL_2 were added through a syringe. The solution was allowed to warm to

0 °C and was stirred for 4 h. To quench the reaction, 10 mL of dry MeOH were added and the solution was stirred for another hour. The solvents were removed under reduced pressure and 10 mL of MeOH were again added. The solvents were then removed to yield a yellow film, which was dissolved in 0.5 mL of isopropanol and stored at -15 °C until crystals appeared (60 days). The solvents were then removed under reduced pressure to yield a yellow solid that was triturated under cold EtOH and filtered to provide 221 mg (60%) of the product, pure by NMR: mp: 195–200 °C dec. ¹H NMR (D₂O): δ 7.30 (d, 1H, ArH); 7.28 (s, 1H, ArH); 7.22 (d, 1H, ArH); 6.99 (s, 1H, ArH); 6.52 (s, 1H, ArH); 4.56–4.51 (dd, 1H, OCH₂, *J*_{gem} = 10.2 Hz, *J*_{vic} = 4.5 Hz); 4.51–4.42 (2d, 2H, CH₂N); 4.22 (d, 1H, ArCHAr, *J*_{trans} = 11.7 Hz); 4.13 (t, 1H, OCH₂); 3.24 (dt, 1H, CHN, *J*_{trans} = 11.7 Hz, *J*_{vic} = 4.5 Hz). ESIMS: *m*/*z* 283 (M⁺, 100). High res. ESIMS for C₁₇H₁₉NO₃ (M + H⁺): calc. 284.1287, found 284.1289.

4.2.9. 2-(4-Ethylphenyl)-4,4-dimethyloxazoline (13b)

A solution of 2-amino-2-methylpropanol (12.76 mL, 133.1 mmol) in 100 mL CH₂Cl₂ was cooled to 0 °C. Into this flask, 4ethylbenzoyl chloride (11.22 g, 66.59 mmol) was introduced dropwise. This mixture was stirred for 4 h, then 10.2 mL (139.8 mmol) of SOCl₂ were introduced dropwise. The mixture was allowed to warm to room temperature overnight. The solvents were removed by rotary evaporation, and water (50 mL) was added. The aqueous mixture was washed twice with 15 mL CH₂Cl₂, and then basified with aqueous ammonia. The cloudy mixture was extracted with CH₂Cl₂ (3 × 30 mL), and the pooled organic extracts were washed once with 20 mL of water and dried over MgSO₄. The mixture was then filtered and the solvents removed under reduced pressure to yield 7.5 g (55%) of oxazoline **13b** as a colorless oil. ¹H NMR (CDCl₃): δ 7.93 (d, 2H, ArH); 7.25 (d, 2H, ArH); 4.18 (s, 2H, OCH₂); 2.68 (q, 2H, CH₂); 1.43 (s, 6H, 2CH₃); 1.22 (t, 3H, CH₃).

4.2.10. (±)-Trans-4-[2-(4,4-dimethyl-oxazolin-2-yl)-5ethylphenyl]-6,7-methylenedioxy-3-nitrochroman (**14b**)

Under dry nitrogen, 7.50 g (36.89 mmol) of oxazoline 13b were dissolved in 15 mL of dry THF, and cooled to -78 °C in a dry ice/ acetone bath. Into this flask, 15 mL (37.5 mmol) of a 2.5M solution of *n*BuLi in hexanes were introduced through a syringe. The mixture was stirred for 4 h, then a solution of 5.44 g (34.59 mmol) of nitrochromene 11 in 40 mL of dry THF, previously to cooled to -78 °C, was introduced through a cannula. The reaction was removed from the dry ice bath and stirred for 30 min, and then was quenched with a saturated solution of NH₄Cl. The mixture was then extracted with 3 \times 30 mL CH₂Cl₂, washed once with water, once with brine, and dried over MgSO4. The mixture was then filtered and the filtrate was concentrated by rotary evaporation to yield a dark oil. This oil was then dissolved in hot EtOH and left to cool overnight in a freezer. The resulting crystals were collected by filtration and air-dried to yield 5.29 g of the product as yellowish crystals. A second crop provided an additional 0.35 g (50.6% total yield): mp 138 °C. ¹H NMR (CDCl₃): δ 7.85 (dd, 1H, ArH); 7.17 (d, 1H, ArH); 6.80 (s, 1H, ArH); 6.45 (s, 1H, ArH); 6.33 (s, 1H, ArH); 5.92-5.86 (2d, 2H, OCH₂O); 5.66 (br, 1H, ArCHAr); 4.95 (br, 1H, CHNO₂); 4.66-4.60 (m, 1H, OCH₂); 4.14-4.05 (m, 3H, OCH₂, oxazoline CH₂); 2.60 (q, 2H, ArCH₂); 1.29 (2s, 6H, 2CH₃); 1.12 (t, 3H, CH₃). ESIMS: *m*/*z* (rel. intensity) 424 (M⁺, 100). Anal. (C₂₃H₂₄NO₂) C, H, N.

4.2.11. (±)-Trans-2,3-methylenedioxy-11-ethyl-6a,12b-dihydro-6Hchromeno[3,4-c]isoquinolin-8-one (**16b**)

In a 100 mL flask, 500 mg (1.18 mmol) of nitro-oxazoline **14b** were dissolved in 20 mL of THF and 20 mL 2 M HCl and stirred overnight at room temperature. The solution was concentrated under reduced pressure to one-half of its original volume and

extracted with EtOAc (4×15 mL). The extracts were washed once with brine, dried over MgSO₄, filtered, and the solvents removed by rotary evaporation to yield 516 mg of the crude HCl salt 15b as a tan foam. This salt was dissolved in a mixture of 50 mL CH₃COOH and 10 mL of THF. Powdered zinc metal (2 g) was added and the suspension was stirred overnight. The mixture was then filtered and the zinc salts on the filter were rinsed with CH₃COOH. The filtrate was concentrated by rotary evaporation, the residue was dissolved in 20 mL EtOH, and then basified with ammonia. This mixture was stirred at 50 °C for 4 h and was then cooled overnight. The resulting crystals were collected by filtration, rinsed on the filter with cold EtOH, and then dried under vacuum to yield 0.318 g (83.5%) of lactam 16b as a white fluffy product: mp >240 °C ¹H NMR (CDCl₃): δ 8.01 (d, 1H, ArH); 7.47 (s, 1H, ArH); 7.26 (m, 1H, ArH); 6.99 (s, 1H, ArH); 6.87 (s, 1H, NH); 6.51 (s, 1H, ArH); 6.00-5.96 (2d, 2H, OCH₂O); 4.35-4.31 (dd, 1H, OCH₂, J_{gem} = 9.1 Hz, J_{vic} = 3.1 Hz); 4.25–4.21 (d, 1H, ArCHAr, $J_{trans} = 11.4$ Hz); 3.99–3.93 (dd, 1H, OCH₂, $J_{gem} = 9.1$ Hz, $J_{\text{vic}} = 11.1 \text{ Hz}$; 3.91–3.82 (dt, 1H, CHN, $J_{trans} = 11.1 \text{ Hz}$, $J_{\text{vic}} = 3.1 \text{ Hz}$); 2.74-2.67 (q, 2H, ArCH₂); 1.29-1.22 (t, 3H, CH₃). EIMS: m/z 323 (M⁺, 100). Anal. (C₁₉H₁₇NO₄) C, H, N.

4.2.12. (±)-Trans-2,3-methylenedioxy-11-ethyl-6a,12b-tetrahydro-6H-chromeno[3,4-c]isoquinoline hydrochloride (**17b**)

In a 500 mL flask, 1.97 g (6.09 mmol) of lactam 16b was suspended in 200 mL of dry THF. Into this suspension, 61 mL (60.87 mmol) of a 1 M solution of BH₃ in THF were introduced, and the reaction was stirred and heated at reflux for 48 h. The clear solution was then reduced to about one-third volume by rotary evaporation, cooled to 0 °C, and quenched with water. This mixture was extracted with 3×30 mL CH₂Cl₂, and the organic extracts were washed once with 10 mL water and once with brine. The pooled extracts were then dried over MgSO₄, filtered, and the solvents removed by rotary evaporation to yield a viscous oil. This oil was stirred for 4 h at 40° C with 25 mL of 2 M HCl in EtOH, whereupon the product crystallized. The crystals were collected by filtration and dried to yield 1.97 g (94%) of HCl salt **17b** as a white powder: mp >240 °C ¹H NMR (DMSO-d₆): δ 10.07 (br, 2H, ⁺NH₂); 7.36 (d, 1H, ArH); 7.22 (br, 2H, 2ArH); 6.98 (s, 1H, ArH); 6.67 (s, 1H, ArH); 6.05 (2d, 2H, OCH₂O); 4.49-4.31 (m, 4H, OCH₂, CH₂N, ArCHAr); 4.16 (dd, 1H, OCH₂, $J_{gem} = J_{vic} = 10.5$ Hz); 3.16 (dt, 1H, CHN, *J*_{trans} = *J*_{vic} = 10.8 Hz, *J*_{vic2} = 4.5 Hz); 2.61 (q, 2H, ArCH₂); 1.16 (t, 3H, CH₃). EIMS: *m*/*z* 309 (M⁺, 100). Anal. (C₁₉H₁₉NO₃) C, H, N.

4.2.13. (±)-Trans-2,3-dihydroxy-11-ethyl-6a,7,8,12b-tetrahydro-6H-chromeno[3,4-c]isoquinoline hydrobromide (**5b**)

In a 50 mL flask, 150 mg (0.485 mmol) of amine 17b (free base) was dissolved in 10 mL CH₂Cl₂. Under an inert, dry atmosphere, this solution was cooled to -78 °C and 2 mL (2 mmol) of a 1M solution of BBr₃ in CH₂Cl₂ were added. The flask was then placed in an icewater bath, stirred overnight at 0 °C, and then 5 mL of dry MeOH were added. The reaction flask was allowed to warm to RT and the solvents were removed by rotary evaporation. Dry MeOH (5 mL) was added to the residue and the solvents were removed under reduced pressure. Again, 5 mL of MeOH were added and the solvents were removed to yield a yellow foam that was pure by NMR, and was crystallized from EtOAc/EtOH to yield 19 mg (12%) of **5b** as light yellow powder: mp 188–203 °C dec. ¹H NMR (D₂O): δ 7.37 (d, 1H, ArH); 7.36 (s, 1H, ArH); 7.31 (d, 1H, ArH); 7.05 (s, 1H, ArH); 6.59 (s, 1H, ArH); 4.61–4.56 (dd, 1H, OCH₂, *J*_{vic} = 4.2 Hz); 4.55-4.43 (2d, 2H, CH2N); 4.32-4.28 (d, 1H, ArCHAr, $J_{trans} = 11.4 \text{ Hz}$; 4.22–4.15 (dd, 1H, OCH₂, $J_{gem} = J_{vic} = 10.8 \text{ Hz}$); 3.36-3.26 (dt, 1H, CHN, $J_{trans} = 11.4$ Hz, 4.72-4.52, $J_{vic} = 4.2$ Hz); 2.66-2.63 (q, 2H, ArCH₂); 1.18 (t, 3H, CH₃) ESIMS: m/z 297 (M⁺,

100). High res. ESIMS for $C_{18}H_{21}NO_3$ (M + H⁺): calc. 297.1365, found 297.1369.

4.2.14. (±)-Trans-4-[2-(4,4-dimethyloxazolin-2-yl)-5fluorophenyl]-6,7-methylenedioxy-3-nitrochroman (**14c**)

Under a dry N₂ atmosphere, 1.747 g (9.04 mmol) of oxazoline **13c** [30] were dissolved in 15 mL of dry THF. and cooled to -45 °C in a dry ice/chlorobenzene bath. Into this flask. 3.6 mL (9.04 mmol) of a 2M solution of *n*BuLi in hexanes were introduced through a syringe. The mixture was stirred for 1.25 h and then a solution of 1.0 g (4.52 mmol) of nitrochromene 11 in 40 mL of dry THF, previously cooled to -78 °C, was introduced through a cannula. The reaction was removed from the dry ice bath and stirred for 30 min, and then 20 mL of a saturated solution of NH₄Cl were added. The mixture was extracted with CH_2Cl_2 (3 × 30 mL), washed once with water, and once with brine. The organic extracts were dried over MgSO₄, filtered, and the solvents removed by rotary evaporation to yield a crude oil. This oil was then dissolved in hot MeOH, whereupon immediate crystallization occurred. The mixture was cooled, and the crystals were collected and dried to afford 1.22 g of the product as yellowish crystals. A second crop yielded 0.06 g of crystals (68% total yield): mp 183 °C ¹H NMR (CDCl₃): δ 7.85 (dd, 1H, ArH); 7.08–7.01 m, 1H, ArH); 6.75 (dd, 1H, ArH); 6.48 (s, 1H, ArH); 6.35 (s, 1H, ArH); 5.95-5.91 (2d, 2H, OCH₂O); 5.75 (s, 1H, ArCHAr); 4.98 (br, 1H, CHNO₂); 4.72-4.66 (m, 1H, OCH₂); 4.10 (dd, 1H, OCH₂, $J_{\text{gem}} = 12.6 \text{ Hz}$; 4.06 (s, 2H, oxazoline CH₂); 1.31 (d, 6H, 2CH₃). ESIMS: *m*/*z* 415 (M⁺, 100). Anal. (C₂₁H₁₉FN₂O₆) C, H, N.

4.2.15. (\pm)-Trans-2,3-methylenedioxy-11-fluoro-6a,12b-dihydro-6H-chromeno[3,4-c]isoquinolin-8-one (**16c**)

A solution of 500 mg (1.21 mmol) of oxazoline 14c were dissolved in 10 mL of THF and 10 mL 2M HCl. This solution was stirred for 4 h at 60 °C, allowed to cool, and extracted four times with 15 mL EtOAc. The extracts were washed once with brine. Upon standing, the organic layer produced crystals, which were filtered and rinsed with EtOAc to yield 549 mg (97%) of HCl salt 15c. These crystals were then dissolved in 15 mL CH₃COOH, and 0.5 g of powdered zinc metal were added. This suspension was stirred overnight and then filtered, rinsing the zinc salts on the filter with CH₃COOH. The filtrate was then concentrated by rotary evaporation. The residue was dissolved in 5 mL EtOH and then basified with ammonia to produce crystallization. This suspension was heated at 50 °C and stirred for 4 h, then cooled and stirred overnight. The crystals were collected by filtration, and rinsed on the filter with EtOAc/EtOH. Drying under vacuum afforded 265 mg (72.3%) of lactam **16c** as a white fluffy product: mp >240 °C 1 H NMR (DMSO-d₆): δ 7.95 (dd, 1H, ArH); 7.29 (m, 2H, 2ArH); 7.17 (s, 1H, ArH); 6.60 (s, 1H, ArH); 6.02 (d, 2H, OCH₂O); 4.31 (dd, 1H, OCH₂, $J_{\text{gem}} = 9.9 \text{ Hz}, J_{\text{vic}} = 3.6 \text{ Hz}$; 4.25 (d, 1H, ArCHAr, $J_{trans} = 11.4 \text{ Hz}$); 3.87 (dd, 1H, J_{gem} = J_{trans} = 10.5 Hz); 3.67 (dt, 1H, CHN, $J_{trans} = 11.4$ Hz, $J_{vic} = 3.9$ Hz). EIMS: m/z 313 (M⁺, 100). Anal. (C₁₇H₁₂FNO₄) C, H, N.

4.2.16. (\pm)-Trans-2,3-methylenedioxy-11-fluoro-6a,7,8,12b-tetrahydro-6H-chromeno[3,4-c]-isoquinoline (**17c**)

In a 250 mL flask, 1.3 g (4.86 mmol) of lactam **16c** were suspended in 40 mL of dry THF. Into this suspension, 20 mL (20 mmol) of a 1M solution of BH₃ in THF were introduced, and the solution was stirred at reflux overnight. The clear solution was then cooled to 0 °C, quenched with water, and reduced to one-third of its volume by rotary evaporation. This mixture was extracted with CH_2Cl_2 (3 × 15 mL), and the extracts were washed once with 10 mL water and once with brine. This solution was then dried over MgSO₄, filtered, and the filtrate was concentrated by rotary evaporation to yield a viscous oil. This oil was stirred for

45 min with 10 mL of a 2 M solution of HCl in EtOH, to produce a white precipitate. The mixture was filtered and the collected solids dried to yield 1.05 g of the HCl salt as a white powder. Water (30 mL) was added to the filtrates and the mixture was washed with 20 mL of ether, basified with NH₄OH and extracted with CH₂Cl₂ (10 mL x 3). The extracts were dried over MgSO₄, filtered and the solvent removed under reduced pressure. The residue was then crystallized from cold EtOH to yield an additional 170 mg of the free base as white crystals (88.7% total yield): mp 132–135 °C. ¹H NMR (CDCl₃): δ 7.21 (dd, 1H, ArH); 7.14 (dd, 1H, ArH); 6.95 (dt, 1H, ArH); 6.89 (s, 1H, ArH); 6.52 (s, 1H, ArH); 5.95 (s, 2H, OCH₂O); 4.31 (dd, 1H, OCH₂, $J_{gem} = 10.2$ Hz, $J_{vic} = 5.1$ Hz); 4.05 (s, 2H, ArCH₂N); 3.87 (d, 1H, ArCHAr, J_{trans} = 10.8 Hz); 3.88 (t, 1H, OCH₂, $J_{\text{vic}} = 10.2 \text{ Hz}$; 3.03–2.94 (dt, 1H, CHN, $J_{trans} = 10.8 \text{ Hz}$). EIMS: m/z299 (M⁺, 100). High res. EIMS for: C₁₇H₁₅FNO₃ calc. 299.2964, found 299.2966.

4.2.17. (\pm) -Trans-2,3-dihydroxy-11-fluoro-6a,7,8,12b-tetrahydro-6H-chromeno[3,4-c]isoquinoline hydrochloride (**5c**)

In a 50 mL flask, 100 mg (0.334 mmol) of amine 17c was dissolved in 10 mL CH₂Cl₂. The solution was cooled to -78 °C and 1.33 mL (1.33 mmol) of a 1 M solution of BCl₃ in CH₂Cl₂ were added. The reaction flask was then placed into an ice-water bath, stirred for 6 h, and then 5 mL of dry MeOH were added. The reaction flask was allowed to warm to RT and the solvents were removed by rotary evaporation. Another 5 mL of MeOH were added and the solution was concentrated again by rotary evaporation, leaving a vellow foam that was crystallized from EtOAc/EtOH to yield 31 mg (28%) of **5c** as a light yellow powder: mp 181–200 °C dec. ¹H NMR (DMSO-d₆): δ 10.18 (br, 2, ⁺NH₂); 9.43 (s, 1H, OH); 9.10 (s, 1H, OH); 7.77 (t, 1H, ArH); 7.46 (m, 2H, 2ArH); 7.07 (s, 1H, ArH); 6.63 (s, 1H, ArH); 4.72-4.52 (m, 4H, ArCHAr, ArCH₂N, OCH₂,); 4.32 (t, 1H, OCH₂); 3.36 (dt, 1H, CHN, *J*_{trans} = 11.1 Hz). ESIMS: *m*/*z* 288 (M⁺, 100). High res. ESIMS for (M⁺): C₁₆H₁₆FNO₃ calc. 288.1036, found 288.1036.

4.2.18. 2-(4-trifluoromethylphenyl)-4,4-dimethyloxazoline (13d)

This compound was prepared using the methodology of Meyers et al. [31] and had properties identical to those reported previously [32]. Its NMR spectrum has not been previously reported. ¹H NMR (CDCl₃): δ 8.03 (d, 2H, ArH); 7.63 (d, 2H, ArH); 4.12 (s, 2H, OCH₂); 1.47 (s, 6H, 2CH₃); 1.22 (t, 3H, CH₃).

4.2.19. (±)-Trans-4-[2-(4,4-dimethyl-oxazolin-2-yl)-5-

trifluoromethylphenyl]-6,7-methylenedioxy-3-nitrochroman (14d)

Under a dry atmosphere, 2.20 g (9.04 mmol) of oxazoline 13d were dissolved in 35 mL of dry THF and cooled to -78 °C. Into this flask, 3.8 mL (9.5 mmol) of a 2M solution of *n*BuLi in hexanes were introduced through a syringe. The mixture was stirred for 1 h, at which point a solution of 1 g (4.52 mmol) of nitrochromene 11 in 150 mL of dry THF, previously cooled to -78 °C, was introduced through a cannula. The reaction was removed from the dry ice bath and stirred for 30 min, and was then quenched by adding 10 mL of a saturated solution of NH₄Cl. The solution was then extracted with 3×50 mL CH₂Cl₂, the extracts washed once with water, once with brine, dried over MgSO₄, filtered, and the filtrate was then concentrated by rotary evaporation to yield a crude oil. This oil was dissolved in MeOH and cooled to RT overnight. The resulting crystals were collected by filtration and dried to yield 1.45 g (69%) of the product as yellow crystals: mp 176 °C. ¹H NMR (CDCl₃): δ 8.07 (d, 1H, ArH); 7.63 (d, 1H, ArH); 7.30 (s, 1H, ArH); 6.50 (s, 1H, ArH); 6.30 (s, 1H, ArH); 5.96-5.91 (2d, 2H, OCH₂O); 5.76 (s, 1H, ArCHAr); 4.96 (br, 1H, CHNO₂); 4.72-4.62 (m, 1H, OCH₂); 4.12-4.08 (s, m, 3H, oxazoline CH₂, OCH₂); 1.39–1.33 (m, 6H, 2CH₃). ESIMS: m/z 464 (M⁺, 100). Anal. (C₂₂H₁₉F₃N₂O₆) C, H, N.

4.2.20. (\pm) -Trans-2,3-methylenedioxy-11-trifluoromethyl-6a,12b-dihydro-6H-chromeno[3,4-c]-isoquinolin-8-one (**16d**)

In a 250 mL flask, 2.57 g (5.56 mmol) of oxazoline 14d were dissolved in a mixture of 40 mL of THF and 40 mL 2M HCl. This solution was stirred overnight, 100 mL of water were added, and the mixture was then extracted with 4×30 mL CH₂Cl₂. The pooled organic extracts were washed twice with 10 mL of water and dried over MgSO₄. The mixture was filtered, and the filtrate was concentrated by rotary evaporation to afford 2.57 g (89.5%) of a yellow foam that could be crystallized from iPrOH, but was used without further purification. In a 250 mL flask, 2.79 g of HCl salt 15d were dissolved in 150 mL of CH₃COOH. Powdered Zn (1 g) was added and the suspension was stirred overnight. The reaction was filtered, the solids on the filter were rinsed with CH₃COOH, and the filtrate was reduced to dryness under vacuum. The residue was dissolved in 15 mL EtOH, and ammonia was added to basify (pH 10) the mixture. This solution was cooled to 0 °C overnight. The resulting solid was collected by filtration, washed on the filter with cold EtOH, and then dried under vacuum to yield 1.77 g (90.6%) of lactam **16d** as a white, fluffy product: mp >240 °C. ¹H NMR (DMSOd₆): δ 8.85 (s, 1, NH); 8.10 (d, 1H, ArH); 7.82 (d, 1H, ArH); 7.79 (s, 1H, ArH); 7.19 (s, 1H, ArH); 6.63 (s, 1H, ArH); 6.04 (2d, 2H, OCH₂O, $J_{\text{gem}} = 14.1 \text{ Hz}$); 4.33 (d, 1H, ArCHAr, $J_{trans} = 11.7 \text{ Hz}$); 4.32 (dd, 1H, OCH₂, $J_{gem} = 10.2$ Hz, $J_{vic} = 3.6$ Hz); 3.89 (t, 1H, OCH₂, $J_{\text{gem}} = 10.2$ Hz), 3.73 (dt, 1H, NCH, $J_{trans} = 11.7$ Hz, $J_{\text{vic}} = 3.6$ Hz). ESIMS: *m*/*z* 363 (M⁺, 100). Anal. (C₁₈H₁₂F₃NO₄) C, H, N.

4.2.21. (\pm) -Trans-2,3-methylenedioxy-11-trifluoromethyl-6a,7,8,12b-tetrahydro-6H-chromeno[3,4-c]isoquinoline hydrochloride (**17d**)

In a 250 mL flask, 1.765 g (4.86 mmol) of lactam 16d were suspended in 50 mL of dry THF. Into this suspension, 24.3 mL (24.3 mmol) of a 1 M solution of BH₃ in THF were introduced, and the reaction was heated at reflux and stirred overnight. The clear solution was then reduced to about one-third of its volume by rotary evaporation, cooled to 0 °C, and diluted with H₂O. This mixture was extracted with 3×25 mL CH₂Cl₂, and the extracts were washed once with 10 mL water and once with brine. This solution was then dried over MgSO₄, filtered, and the solvents removed by rotary evaporation to yield a viscous oil. A 2M solution of HCl in EtOH (10 mL) was added to this oil, and the mixture was stirred for 45 min to produce, after 10 min, a white precipitate. The suspension was cooled to 0 °C for 2 h and filtered. The solid on the filter was rinsed with cold EtOH and dried to afford 1.57 g (84%) of the HCl salt as a white powder: mp >240 °C. ¹H NMR (CDCl₃): δ 7.78 (d, 1, ArH); 7.71 (d, 2H, ArH); 7.63 (s, 1H, ArH); 7.02 (s, 1H, ArH); 6.69 (s, 1H, ArH); 6.05 (2d, 2H, OCH₂O); 4.61-4.44 (m, 4H, OCH₂, NCH₂, ArCHAr); 4.18 (t, 1H, OCH₂); 3.26 (dt, 1H, *J*₁ = 10.8 Hz, *J*₂ = 3.9 Hz). ESIMS: *m*/*z* 349 (M⁺, 100). Anal. (C₁₈H₁₄F₃NO₃) C, H, N.

4.2.22. (\pm) -Trans-2,3-dihydroxy-11-trifluoromethyl-6a,7,8,12b-tetrahydro-6H-chromeno[3,4-c]-isoquinoline hydrochloride (**5d**)

In a 50 mL flask, 702 mg (2.01 mmol) of amine **17d** was dissolved in 20 mL CH₂Cl₂ under a dry N₂ atmosphere. The flask was cooled to -78 °C and 8 mL (8 mmol) of a 1 M solution of BCl₃ in CH₂Cl₂ were added. The reaction flask was then placed in an icewater bath, and stirred for 4.5 h, at which time 4 mL of dry MeOH were added. The reaction was allowed to warm to RT and the solvents were removed by rotary evaporation. Addition of 5 mL of dry MeOH and concentration by rotary evaporation was repeated twice more, to leave a tan-orange foam, which was crystallized from CH₃CN/MeOH to afford a light orange powder: mp >187 °C dec. ¹H NMR (DMSO-d₆): δ 10.03 (br, 2, ⁺NH₂); 9.25 (s, 1H, OH); 8.92 (s, 1H, OH); 7.77 (d, 1H, ArH); 7.71 (s, 1H, ArH); 7.69 (d, 1H, ArH); 6.81 (s, 1H, ArH); 6.41 (s, 1H, ArH); 4.59–4.45 (2d, 2H,

ArCH₂N, $J_{gem} = 15.1$ Hz); 4.18 (t, 1H, OCH₂); 3.26 (dt, 1, CHN, $J_{trans} = 10.8$ Hz, $J_{vic} = 3.6$ Hz). ESIMS: m/z (rel. intensity) 337 (M⁺, 100). High res. ESIMS for C₁₇H₁₆F₃NO₃ (M + H⁺): calc. 338.1004, found 338.1000.

4.2.23. 6-(O-Dimethylthiocarbamoyl)-1,3-benzodioxole (18)

Under N₂, NaH (0.32 g, 13.25 mmol) was slurried with stirring in 20 mL of dry THF at -78 °C. A solution of sesamaldehyde **10** (2 g, 12.05 mmol) dissolved in 50 mL of dry THF was then added dropwise to the suspension, followed by a solution of 1.638 g (13.25 mmol) of dimethylthiocarbamoyl chloride dissolved in 15 mL of dry THF. The reaction was allowed to warm to RT, was stirred for 12 h, and was then carefully quenched with water and extracted with CH₂Cl₂ (3 × 15 mL). The pooled extracts were washed with brine, dried over MgSO₄, filtered, and the solvent removed under reduced pressure to afford an orange solid, which was recrystallized from EtOH to yield 2.30 g (75%) of pure thiocarbamate **18** as lightbrown flakes: mp 137–140 °C. ¹H NMR (CDCl₃): δ 9.89 (s, 1H, CHO); 7.31 (s, 1H, ArH); 6.60 (s, 1H, ArH); 6.10 (s, 2H, OCH₂O); 3.48 (s, 3H, CH₃); 3.41 (s, 3H, CH₃). CIMS: 254 (M + H⁺, 100). Anal. (C₁₁H₁₁NO₄S) C, H, N.

4.2.24. 6-(S-Dimethylthiocarbamoyl)-1,3-benzodioxole (19)

In a single-necked flask attached to a condenser, and under an inert atmosphere, 2.27 g (8.96 mmol) of thiocarbamate **18** were dissolved in 35 mL of diphenyl ether. This solution was heated at 250 °C for 15 min and then immediately cooled to RT with an ice bath. Hexanes (100 mL) was added to the solution, which was stored overnight at 0 °C to cause precipitation. The solid was collected by filtration, washed repeatedly on the filter with hexanes, and purified by column chromatography to yield 1.27 g (56%) of the product as a brown solid. This compound was used without further characterization.

4.2.25. 3-Nitro-2H-6,7-methylenedioxythiochromene (20)

In a single-neck flask, 2.96 g of thiocarbamate **19** (11.70 mmol) were dissolved in 400 of mL of MeOH and heated to 60 °C. Next, 35 mL of a 2M solution of NaOH in MeOH were added, and the solution was heated at reflux for 2.5 h until TLC indicated complete hydrolysis. The mixture was acidified to pH 7 with 2 M HCl in EtOH. The solvents were then removed under reduced pressure, avoiding exposure to air. Toluene (400 mL) was added to the mixture, followed by 1 mL (5.85 mmol) of dibutylamine, 3.46 g (23.39 mmol) of phthalic anhydride, and 0.3 mL (4.19 mmol) of 2nitroethanol. This mixture was heated at reflux for 3 h, at which time addition of 2-nitroethanol was begun at a rate of about 1 drop every 15 min, for a total of 1.6 mL (22.33 mmol) of the reagent added over 8 h. The reaction was stirred at reflux overnight and was then cooled and filtered through Celite. The filtrate was extracted several times with 2 M NaOH, washed once with water, dried over Na₂SO₄, filtered, and reduced to dryness by rotary evaporation. The crude nitrochromene was purified by column chromatography to yield 1.78 g (64%) of a bright red powder: mp: 128–131 °C. ¹H NMR (CDCl₃): δ 7.82 (s, 1H, ArCH); 6.82 (s, 1H, ArH); 6.81 (s, 1H, ArH); 6.03 (s, 2H, OCH₂O); 4.07 (s, 2H, OCH₂). EIMS: 237 (M⁺, 100). High res. EIMS for $C_{10}H_8NO_4S$ (M⁺): calc. 237.0096, found 237.0099.

4.2.26. (\pm) -Trans-2,3-methylenedioxy-6a,12b-dihydro-6H-

thiochromeno[3,4-c]isoquinoline (**22**)

In a dry flask, 2.99 g (12.30 mmol) of 2-(2-bromophenyl)-1,3dioxane were dissolved in 25 mL of dry THF and 597 mg (26.61 mmol) of magnesium powder were added, along with 1 drop of 1,2-dibromoethane. This solution was heated at reflux and stirred for 1 h until formation of the Grignard reagent was complete. The mixture was then cooled to 0 °C and, through a dropping funnel, a solution of 972 mg (4.10 mmol) of nitrothiochromene 20 in 35 mL of dry THF were added dropwise over 0.5 h. The reaction was stirred for 10 min and was then guenched with a satd solution of NH₄Cl. The mixture was then filtered and extracted several times with EtOAc. The combined organic extracts were washed once with water, once with brine, dried over MgSO₄. and filtered. The filtrate was reduced to drvness to vield an oil that was purified using silica gel flash column chromatography, eluting with a 4:1 mixture of hexanes:EtOAc to provide 867 mg (52.7%) of pure **22** as a white powder: mp 154 °C ¹H NMR (CDCl₃): δ 7.22 (m, 1H, ArH); 7.27-7.24 (m, 2H, ArH); 6.60 (s, 1H, ArH); 6.48 (s, 1H, ArH); 5.85 (s, 2H, OCH₂O); 5.80 (d, 1H, ArCHAr, J = 3.3 Hz); 5.62 (s, 1H, OCHO); 5.28–5.25 (m, 1H, CHNO₂); 4.27–4.18 (m, 2H, OCH₂C); 3.98–3.90 (m, 2H, OCH₂C); 3.43 (m, 2H, SCH₂); 2.26–2.20 (m, 1H, CCH₂C); 1.40 (bd, 1H, CCH₂C). EIMS: 401 (M⁺, 100). Anal. (C₂₀H₁₉NO₆S) C, H, N.

4.2.27. (\pm) -Trans-2,3-methylenedioxy-6a,7,8,12b-tetrahydro-6H-thiochromeno[3,4-c]isoquinoline (**24**)

Into a solution of 550 mg (1.37 mmol) of compound **22** in 10 mL of THF and 30 mL CH₃COOH were added 2 g of powdered zinc. This suspension was vigorously stirred for 6 h at 70 °C and was then cooled to RT, filtered, and the filtrate was concentrated under reduced pressure. To the residue was added 30 mL of 2 M ethanolic HCl and this solution was stirred for 30 min. The solvents were removed once again and 30 mL of a 2 M solution of NaOH and 30 mL of CH₂Cl₂ were added. This biphasic mixture was stirred overnight. The organic layer was then separated, washed with water, dried over Na₂SO₄, filtered, and the solvent removed to leave 418 mg of a light brown solid (**23**), which was used without further purification.

To a solution of 400 mg of the crude imine 23 (1.356 mmol) in 10 mL of dry THF and 30 mL of EtOH was added 85 mg of NaCNBH₃ (1.36 mmol) followed by dropwise addition of 4 mL of a 2M solution of HCl in EtOH over 10 min. The suspension was stirred for 4 h and was then reduced under vacuum to about one-third of its original volume. Into this suspension, 10 mL of a 2M solution of NaOH were added and the mixture was extracted with CH₂Cl₂ $(3 \times 10 \text{ mL})$. The pooled organic extracts were washed once with water, dried over Na₂SO₄, filtered, and the solvent was removed. The resulting residue was purified by column chromatography, eluting with EtOAc to yield 316 mg (78%) of amine 24 as a tan solid. An analytical sample was prepared by crystallizing the HCl salt from EtOH/Et₂O: mp >260 °C dec. ¹H NMR (DMSO- d_6): δ 10.11 (bs, 1H, ⁺NH); 9.69 (bs, 1H, ⁺NH); 7.49–7.40 (m, 4H, ArH); 7.07 (s, 1H, ArH); 6.48 (s, 1H, ArH); 6.00 (d, 2H, OCH₂O); 4.41 (bs, 2H, ArCH₂N); 4.27 (d, 1H, ArCHAr); 3.45-3.42 (m, 2H, SCH2, CHN); 3.11 (t, 1H, SCH₂). High res. ESIMS for $C_{17}H_{16}NO_2S (M + H^+)$: calc. 298.0902, found 298.0904.

4.2.28. (\pm) -2,3-Dihydroxy-6a,7,8,12b-tetrahydro-6H-thiochromeno [3,4-c]isoquinoline hydro-chloride (**6**)

A solution of 300 mg (1.01 mmol) of amine **24** in 8 mL of CH₂Cl₂ was placed into a flask, cooled to -78 °C, and 4 mL (4.0 mmol) of a 1M solution of BBr₃ in CH₂Cl₂ were added through a syringe. This solution was allowed to warm to 0 °C in an ice bath and was stirred for 3 h. Dry EtOH (5 mL) was then added to quench the reaction, and the solution was stirred for 30 min at RT. The solvents were removed by rotary evaporation, and 5 mL of dry EtOH were again added. Removal of the solvent yielded a white powder that was collected by filtration to afford a total of 104 mg (34%) of pure catecholamine **6** as a tan powder, in two crops: mp 288–294 °C dec. ¹H NMR (DMSO-*d*₆): δ 10.03 (bs, 2H, ⁺NH₂); 9.18 (s, 1H, ArOH); 9.04 (s, 1H, ArOH); 7.49–7.38 (m, 4H, 4ArH); 6.79 (s, 1H, ArH); 6.49 (s,

1H, ArH); 4.43 (t, 2H, ArCH₂N); 4.26–4.22 (d, 1H, ArCHAr, $J_{trans} = 10.5$ Hz); 3.45–3.35 (m, 1H, NCH₂); 3.28–3.19 (m, 1H, SCH₂); 3.10–3.03 (t, 1H, SCH₂, J = 10.5 Hz). ESIMS: 286 (M + H⁺, 100). Anal. (C₁₆H₁₆CINO₂S) C, H, N.

4.3. Pharmacology methods

4.3.1. Materials

[³H]spiperone (95 Ci/mmol) and [³H]SCH-23390 (81 Ci/mmol) were purchased from Amersham Biosciences (Piscataway, NJ, USA). [³H]Cyclic AMP (30 Ci/mmol) was purchased from PerkinElmer (Boston, MA, USA). Chlorpromazine, SCH-23390, butaclamol, and most other reagents were purchased from Sigma–Aldrich Chemical Company (St. Louis, MO, USA).

4.3.2. Competition binding experiments

Porcine striatal tissue was obtained fresh from the Purdue Butcher Block and prepared as previously described [18]. Briefly, striatal tissue was homogenized using a potter-type homogenizer, suspended in homogenization buffer (20 mM Hepes, 0.32 M sucrose, pH 7.4), and centrifuged at $1000 \times g$ for 10 min at 4 °C. The pellet (P1) was discarded and the supernatant was centrifuged at $30,000 \times g$ for 10 min at 4 °C. The resulting pellet (P2) was resuspended in 50 mM Tris buffer (pH 7.4) by briefly using a Kinematica homogenizer, and then centrifuged at $30,000 \times g$ for 30 min at 4 °C. This pellet was resuspended again in 50 mM Tris buffer, dispensed into 1 mL aliquots, and centrifuged for 10 min at 13,000 $\times g$ and 4 °C. A BCA protein assay was used to determine the final protein concentration of the pellets. Supernatant was aspirated and pellets were frozen at -80° C until use.

Radioligand binding assays were performed as previously described [18], with only minor modifications. Pellets were resuspended (1 mL/mg) in receptor binding buffer (50 mM Hepes, 4 mM MgCl₂, pH 7.4) and 75 µg protein was used per assay tube. Receptor isotherms were performed with [³H]SCH-23390 and [³H]spiperone to determine B_{max} and K_d for D₁-like and D₂-like receptor sites, respectively (760 fmol/mg and 0.44 nM for [³H]SCH-23390, and 250 fmol/mg and 0.075 nM for [³H]spiperone). All D₂-like receptor binding assays were performed in the presence of 50 nM ketanserin to mask 5-HT_{2A} sites. Nonspecific binding was defined with 5 μ M butaclamol. Drug dilutions for competitive binding experiments were made in receptor binding buffer and added to assay tubes containing 75 µg protein and either 1 nM [³H]SCH-23390 or 0.15 nM [³H]spiperone. All binding experiments were incubated for 30 min at 37 °C, and were terminated by filtration with ice cold wash buffer using a 96-well Packard Filtermate cell harvester. After the samples were dried, 30 µL of Packard Microscint O was added to each well. Radioactivity was counted using a Packard Topcount scintillation counter.

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