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Tandem IBX-Promoted Primary Alcohol Oxidation/Opening of Intermediate β , γ -Diolcarbonate Aldehydes to (*E*)- γ -Hydroxy- α , β -enals

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Dedication ((optional))

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Abstract: A tandem IBX-promoted oxidation of primary alcohol to aldehyde and opening of intermediate β , γ -diolcarbonate aldehyde to (*E*)- γ -hydroxy- α , β -enal has been developed. Remarkably, the carbonate opening delivered exclusively (*E*)-olefin and no overoxidation of γ -hydroxy was observed. The method developed has been extended to complete the stereoselective total synthesis of both (*S*)- and (*R*)-coriolides and D-*xylo*- and D-*arabino*-C-20 guggultetrols.

Introduction

 γ -Hydroxy- α , β -enals are a group of reactive aldehydes that have dose-dependent regulatory roles, as well as detrimental effects in various cell types and organs. Their origin and cellular functions are well documented.^[1] These can act as Michael acceptors and are also involved in Schiff base formation with biological nucleophiles, which very well explains their bioactivity.^[1] While γ hydroxy- α , β -enals are the degradation products of lipid peroxidation, the major product is 4-hydroxy-2E-nonenal (HNE), which is involved in several biological processes, oxidative stress, regulation of mitochondrial uncoupling, etc.^[2,3] Apart from this, the γ -hydroxy- α , β -enal moiety is formally present as skeletal unit in many natural products, for e.g. as vicinal diene-hydroxy unit in coriolide 1a. [4a] and 4-hydroxy enoic ester moiety in patulolides 1b and 1c,^[4b,c] aspicilin 1d,^[4d] cineromycin B 1e,^[4e] albocycline 1f,^[4f] and macrosphelides A 1g and E 1h,^[4g,h] etc (Figure 1). Thus, this renders the γ -hydroxy- α , β -enal as a compelling moiety on both bioactivity and synthesis fronts.

The early synthesis of γ -hydroxy- α , β -enals has been described by Esterbauer *et al.*^[5] and then by Erickson^[6] using conventional routes. This was followed by the work of Grée *et al.*^[7a] based on alkyl Grignard addition to fumaraldehyde monodimethyl acetal. A similar approach was used by Iriye and co-workers^[7b] based on orthogonal functionalization of 2-butene-1,4-diol. Wittig reaction based approaches^[8a,b] and opening of β , γ -epoxy aldehyde with NaOH or Et₃N with limited examples^[9a-d] are also explored. Chakraborty *et al.* employed Swen oxidation, PDC, IBX or TPAP/NMO conditions for oxidative opening of epoxide **2** in the synthesis of γ -hydroxy- α , β -enals 3 (only four examples, Scheme 1A).^[9d] The recent interesting report by Sasaki and Takeda^[10] involved γ -p-toluenesulfonyl- α , β -epoxysilane **2**' as acrolein



Supporting information for this article is given via a link at the end of the document.

X-ray data and NMR spectra (PDF)

Accession Codes

CCDC 1882370 contain the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif.



β-anion equivalent for a tandem base-promoted Brook-type epoxide opening, allylsilane-based aldehyde allylation and subsequent γ -*p*-toluenesulfonyl elimination, giving the γ -hydroxy- α,β -enals **3**, in one-pot manner (Scheme 1B). Our work reported herein, on preparing γ -hydroxy- α , β -enals was a serendipitous discovery. In the 2-iodoxybenzoic acid (IBX) oxidation of compound 5i (R = C_5H_{11}) to get aldehyde 6i, we observed exclusive formation of γ -hydroxy- α , β -enal **3i**, stereoselectively with (E)-olefin bond (Scheme 1C). The reaction involved IBXpromoted primary alcohol oxidation/opening of intermediate $\beta_{,\gamma}$ diolcarbonate aldehyde providing stereoselectively (E)-y-hydroxy- α,β -enal. IBX-mediated oxidation of saturated carbonyl compounds to α,β -unsaturated carbonyls has been developed by Nicolaou et al.[11] This method may not be applicable for yhydroxy- α , β -enals synthesis (γ -hydroxy being susceptible to oxidation). Our work compliments Takeda's approach and is simpler in terms of starting materials required and reagents involved. No over-oxidation of the γ -hydroxy group was observed. Hence we took up this reaction for further optimization and to study its scope and applications in natural product synthesis.



[B] Sasaki and Takeda





Scheme 1. Methods for the Synthesis of γ -Hydroxy- α , β -enals

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Results and Discussion

We chose compound 5i to investigate various oxidants and to optimize the reaction conditions. In EtOAc, of the various oxidants investigated (Table 1, entries 1-6), IBX and Des-Martin periodinane (DMP) provided reasonable yields of 3i (entries 1 and 2). PhI(OAc)₂ with TEMPO or oxone was not successful for this reaction (entries 5 and 6) and the substare 5i was mostly recovered, in the latter case even after reflux. IBX (1.5 equiv) in different solvents (entries 7-13) was next examined and EtOAc (entry 1) was revealed as the best solvent. In 1,2-dichloroethane (DCE) solvent under reflux, trace of furan product was isolated, which may get formed through the hydroxy-enal cyclization. Increase in reaction temperature from room temperature to 60 °C increased the yield marginally (entry 1 v/s 14). Further use of additives such as NaOAc, pyridine, p-TsOH or DMSO at 60 °C did not improve the yield (entries 15-18). A reaction at reflux conditions improved the yield of 3i to 81% (entry 19). An increase in amount of IBX (entries 20-22) showed best results with 2.0 equiv IBX providing 3i in 94% yield in a 3 h reaction (entry 20). The higher loading of IBX over 2.0 equiv was not beneficial.

Table 1. Screening of Reaction Conditions.^a

7	D OH	oxidant solvent, T ^c	÷ ∀	3 OH	∕∼₀
(syr	o-diol carbonate)	31			
Entry	Oxidant (equiv)	Solvent	T (° C)	Time (h)	Yield ^b 3i (%)
1	IBX (1.5)	EtOAc	rt rt	12	53
2	PDC(15)	EtOAc	rt	4	16
2	PCC(15)	EtOAc	rt	8	24
5	TEMPO (1.5)/	EtOAc	rt	8	complex
)	$PhI(OAc)_{2}(1.5)$				mixture
6	PhI(OAc) ₂ (1.5)/	EtOAc	rt-reflux	50	NR
	Oxone(1.5)				
7	IBX (1.5)	CH_2Cl_2	rt	12	36
8	IBX (1.5)	DCE	rt-reflux	30	15 ^c
9	IBX (1.5)	THF	rt	13	40
10	IBX (1.5)	CH₃CN	rt	13	6
11	IBX (1.5)	DMSO	rt	15	5
12	IBX (1.5)	Benzene	rt	30	21
13	IBX (1.5)	Acetone	rt	20	NR
14	IBX (1.5)	EtOAc	60	14	59
15	IBX (1.5)/	EtOAc	60	8	19
	NaOAc (1.0)				
16	IBX (1.5)/	EtOAc	60	8	49
	Pyr (1.0)				
17	IBX (1.5)/	EtOAc	60	9	37
	p-TsOH (1.0)			V	
18	IBX (1.5)/	EtOAc	60	7	48
	DMSO (1.0)				
19	IBX (1.5)	EtOAc	reflux	8	81
20	IBX (2.0)	EtOAc	reflux	3	94
21	IBX (2.5)	EtOAc	reflux	3	94
22	IBX (3.0)	EtOAc	reflux	2.5	93
^a Reaction conditions: 5i (1.0 mmol), oxidant (1.5–3.0 equiv),					
solvent, rt–reflux, 2.5–30 h. ^b Isolated yield. ^c Traces of furan					
product observed. NR = No reaction, rt = room temperature					

With the optimized conditions, the scope of this tandem oxidation of primary alcohol/subsequent opening of intermediate diol carbonate aldehyde was investigated. As shown in Scheme 2, the IBX-promoted reaction of various diolcarbonates **5a-n** with aliphatic or aryl groups delivered the γ -hydroxy- α , β -enals **3a-n** in good to excellent yields in 3-4 h reactions. A few chiral vicinal diolcarbonates **5i'** and **5m'** prepared through Sharpless asymmetric dihydroxylation, yielded the chiral γ -hydroxy- α , β -enals **3i'** and **3m'** in 93 and 78% yields, respectively with no loss in optical purity at the γ -hydroxy centre.



Scheme 2. Substrate Scope for γ-Hydroxy- α ,β-unsaturated Aldehyde Synthesis. Reaction Conditions: 5a-n (1.0 mmol), IBX (2.0 equiv), EtOAc, reflux, 3-4 h.

A selectivity reaction was investigated with acetonide group v/s the carbonate (Scheme 3). The reaction of 1,2-diol-acetonide compound (prepared from 7i) with IBX (2.0 equiv) delivered the aldehyde 8i with no opening of the acetonide moiety. In the carbonate case 5m, a reaction quenched after 1 h showed the presence of starting material 5m and the carbonate aldehyde 6m in 1:1.5 ratio. The reaction for 3 h, with the complete consumption of 5m showed the presence of product 3m and the intermediate 6m in 5:1 ratio. Further continuation of this reaction for another 1 h resulted in complete conversion to 3m. Thus, the reaction involves first oxidation of primary hydroxy to the aldehyde followed by opening of the carbonate to give the enal 3m. The reaction also requires the presence of the labile carbonate rather than the acetonide. The 1,2-diol carbonates used in this study were prepared by cis-dihydroxylation of (E)-olefins (see Experimental Section). The IBX-mediated reaction resulted exclusively in (*E*)- $\alpha\beta$ -unsaturated aldehydes. To determine the stereospecificity of the reaction, we prepared the anti-diol carbonate 5i" (through cis-dihydroxylation of cis-olefin) and subjected to IBX oxidation (Scheme 3). This resulted in the same product, (*E*)- $\alpha\beta$ -unsaturated aldehyde **3i** in 80% yield. Thus, the reaction is not stereospecific with regards to relative stereochemistry of the diol carbonate and the geometry of olefin formed in the enal.

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We further considered the synthetic modifications of the γ hydroxy- α , β -enal compounds (Scheme 4). Pinnick oxidation of aldehyde group of **3m** delivered the corresponding γ -hydroxy- α , β unsaturated acid 9 in 76% yield. Further esterification of 9 gave γ -hydroxy- β , γ -unsaturated ester **10** in 83% yield. the Homologation of enals 31, 3m and 3n with the ylide from triethylphosphonoacetate gave the dienoates 11a, 11b and 11c, in 63-76% yields. respectively Addition of lithiated phenylacetylene to enal 3m furnished the skipped ene-yne alcohol 12 in 58% yield, with no conjugate addition being observed.



Scheme 3. Selectivity Experiments and Stepwise Formation of 3m



Scheme 4. Synthetic Modifications of γ -Hydroxy- α , β -enals

The application of a synthetic methodology toward the synthesis of natural products is an important validation of its synthetic potential and usefulness. Thus, the developed reaction

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was also utilized in the total synthesis of both enantiomers of coriolide. It is a major component in the scent gland extracts of the male long wing tropical butterflies, *Heliconius pachinus*.^[4a,12] It was also isolated from the seed oil of *Monninae marginata* (polygalaceae), a plant native to Uruguay.^[13] A few syntheses of coriolide are reported in the literature.^[12,14] Our synthesis started from 1,9-nonanediol **13**, which on monobromination (90%) and further oxidation of the remaining hydroxy group delivered the carboxylic acid **14** in 98% yield (Scheme 5). The phosphonium salt **15** was prepared from bromo acid **14** in 72% yield. This was converted into its ylide using KHMDS and reacted with the (*S*)- γ -hydroxy- α , β -enal **3i**' at -78 °C to provide stereoselectively the *Z*,*E*-diene acid **16** in 64% yield. The latter on macrolactonization by the Yamaguchi method^[15] afforded (*S*)-coriolide **1a** in 58% yield.^[14b] On the other hand, inversion-cyclization of **16** under Mitsunobu's conditions^[16] delivered (*R*)-coriolide **1a**.



Scheme 5. Total Synthesis of (S)-and (R)-Coriolides

Further, the total synthesis of D-xylo- and D-arabino-C20 guggultetrols was undertaken. These are long chain aliphatic 1.2.3.4-tetrols, with the former as major naturally occurring lipid isolated from the gum resin of the tree Commiphora mukul, known as guggul.^[17] It has been used in Ayurveda, the ancient Indian medicinal system for the treatment of inflammation, rheumatoid arthritis, obesity, and disorders of lipid metabolism besides several other ailments.^[18] The C18 and C20 guggultetrols have been synthesized before by various methods.[19] Our synthesis of C20 guggultetrol is shown in Scheme 6. The reduction of the aldehyde group in compound 3m' with DIBAL-H gave the ene-diol 17 in 80% yield. This was converted into its diacetate 18 (85% vield) and then subjected to asymmetric dihydroxylation using (DHQ)₂-PHAL ligand to afford the diol diacetate 19, which without further purification was converted into its tetraacetate as a diastereomer mixture (dr = 9:1). The mixture was separated by column chromatography to furnish the D-xylo-C20 guggultetrol tetraacetate 20a^[20] in 70% overall yield. The minor diastereomer D-arabino-C20 guggultetrol 20b was obtained in 8% yield. While compound 17 could be subjected directly for dihydroxylation, the tetrol product purification was hampered due to its high polarity and resulted in lower yields of the tetraacetate in the subsequent reaction. The stepwise conversion, first into the diacetate and then dihydroxylation resulted in better overall yields and diastereomeric ratio.

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Scheme 6. Total Synthesis of D-xylo- and D-arabino-C20 Guggultetrol Tetraacetates

Conclusions

In summary, in this paper we have executed an interesting chemistry of IBX promoted primary alcohol oxidation to aldehyde and the subsequent opening of the intermediate β , γ -diolcarbonate aldehyde to give stereoselectively the (*E*)- γ -hydroxy- α , β -enals. These are important compounds for their inherent bioactivity and also as useful synthons in natural products synthesis. This has been demonstrated by completing the total synthesis of both enantiomers of coriolide and the C20 D-*xylo*- and D-*arabino*-guggultetrol tetraacetates.

Experimental Section

General information. Solvents were dried by using standard procedures. Thin-layer chromatography was performed on EM 250 Kieselgel 60 F254 silica gel plates. The spots were visualized by staining with KMnO₄ or by using a UV lamp. ¹H-NMR and ¹³C-NMR were recorded with a spectrometer operating at 400 or 500 and 100 or 125 MHz for proton and carbon nuclei, respectively. The chemical shifts are based on the TMS peak at $\delta = 0.00$ pm for proton NMR and the CDCl₃ peak at $\delta = 77.00$ ppm (t) for carbon NMR. For other deuterated solvents the standard chemical shifts were considered. IR spectra were obtained on an FT-IR spectrometer. Optical rotations were measured using sodium D-line (589 nm). HRMS (ESI-TOF) spectra were recorded using positive electrospray ionization by the TOF method. The enantiomeric excess was determined by the HPLC method using Chiralpak OD-H or AD-H columns at wavelength 230 nm.

General Procedure for the Synthesis of Alcohols (22a-e). To a stirred solution of malonic acid (2.71 g, 26.0 mmol, 1.3 equiv) in pyridine (4 mL) was added piperidine (0.2 mL, 2.0 mmol, 0.1 equiv) and then aryl aldehyde 21a-e (20 mmol) drop wise under N₂ atmosphere. The resulting mixture was stirred for 6 h at 85 °C, when TLC indicated the disappearance of the starting material. The mixture was cooled to room temperature and neutralized with 10% aq. HCl (5 mL) to give a white solid that was filtered

and washed with cold water. Recrystallization from aqueous EtOH (1:1) afforded (E)-3-(aryl)-acrylic acids, which were used directly for the next reaction.

To a suspension of LiAlH₄ (1.52 g, 40.0 mmol, 2.0 equiv) in THF (60 mL) under N₂ atmosphere at 0 °C was added the solution of above (*E*)-3-(aryl)-acrylic acids in THF (15 mL) dropwise. The resulting mixture was stirred at reflux for 6-8 h. The mixture was then cooled to room temperature and quenched with saturated aq. Na₂SO₄ solution (15 mL) at 0 °C. Then EtOAc (100 mL) was added and the mixture stirred for 1 h. It was then filtered through a sintered glass funnel and the filtrate was concentrated. The residue was purified by a silica gel column chromatography using petroleum ether/EtOAc (3:1) as eluent to give the corresponding alcohols **22a-e** in 44-58% overall yields over two steps.



3-Phenylpropan-1-ol (**22a**).^[21] Colourless oil (1.362 g, 50%). ¹H NMR (500 MHz, CDCl₃): δ 1.65 (br s, 1H, *OH*), 1.88–1.93 (m, 2H), 2.72 (t, *J* = 7.9 Hz, 2H), 3.68 (t, *J* = 6.4 Hz, 2H), 7.18–7.22 (m, 3H), 7.28–7.31 (m, 2H) ppm. ¹³C{¹H} NMR (125 MHz, CDCl₃): δ 32.0, 34.2, 62.2, 125.8, 128.36, 128.4, 141.8 ppm.IR (CHCl₃): ω_{max} = 3361, 3084, 3062, 3026, 2937, 2863, 1603, 1583, 1496, 1454, 1266, 1218, 1154, 1059, 1032, 918, 805, 753, 699, 668, 574 cm⁻¹.

3-(*p*-Tolyl)propan-1-ol (**22b**).^[21] Colourless oil (1.683 g, 56%). ¹H NMR (500 MHz, CDCl₃): δ 1.61 (br s, 1H, *OH*), 1.87–1.90 (m, 2H), 2.33 (s, 3H), 2.68 (t, *J* = 7.7 Hz, 2H), 3.68 (t, *J* = 6.4 Hz, 2H), 7.10 (s, 4H) ppm. ¹³C{¹H} NMR (125 MHz, CDCl₃): δ 21.0, 31.6, 34.3, 62.3, 128.3, 129.1, 135.3, 138.7 ppm. IR (CHCl₃): υ_{max} = 3351, 3018, 2927, 2862, 1515, 1452, 1266, 1160, 1112, 1046, 911, 837, 805, 781, 756, 669, 561 cm⁻¹. HRMS (ESI-TOF) *m/z*: [M + Na]⁺ Calcd for C₁₀H₁₄ONa 173.0937; Found 173.0935.

3-(4-Methoxyphenyl)propan-1-ol (22c).^[21] Colourless oil (1.928 g, 58%). ¹H NMR (400 MHz, CDCl₃): δ1.82–1.89 (m, 2H), 2.58 (br s, 1H, OH), 2.65 (t, J = 7.6 Hz, 2H), 3.65 (t, J = 6.5 Hz, 2H), 3.79 (s, 3H), 6.84 (d, J = 8.6 Hz, 2H), 7.12 (d, J = 8.6 Hz, 2H) ppm. ¹³C(¹H) NMR (100 MHz, CDCl₃): δ 31.0, 34.3, 55.1, 61.9, 113.6, 129.2, 133.8, 157.6 ppm. IR (CHCl₃): υ_{max} = 3389, 2994, 2939, 1612, 1584, 1513, 1465, 1372, 1299, 1247, 1178, 1111, 1038, 912, 832, 814, 748, 701, 638, 563, 522 cm⁻¹. HRMS (ESI-TOF) *m*/*z*. [M + Na]⁺ Calcd for C₁₀H₁O₂Na 189.0886; Found 189.0888.

3-(4-Fluorophenyl)propan-1-ol (**22d**).^[22] Colourless oil (1.357 g, 44%). ¹H NMR (400 MHz, CDCl₃): δ 1.61 (br s, 1H, OH), 1.83–1.89 (m, 2H), 2.68 (t, J = 7.6 Hz, 2H), 3.66 (t, J = 6.4 Hz, 2H), 6.94–6.99 (m, 2H), 7.12–7.17 (m, 2H). ¹³C(¹H} NMR (100 MHz, CDCl₃): δ 31.2, 34.3, 62.0, 115.08 (d, J = 20.9 Hz), 129.70 (d, J = 7.9 Hz), 137.34 (d, J = 3.1 Hz), 161.30 (d, J = 242.8 Hz) ppm. ¹⁹F NMR (376 MHz, CDCl₃): δ –117.73 ppm. IR (CHCl₃): $\upsilon_{max} = 3343, 2932, 2868, 1664, 1511, 1450, 1273, 1222, 1158, 1094, 1041, 912, 822, 760, 552 cm⁻¹.$

3-(3-Bromophenyl)propan-1-ol (**22e**).^[23] Colourless oil (2.151 g, 50%). ¹H NMR (500 MHz, CDCl₃): δ 1.75 (br s, 1H, *OH*), 1.87–1.93 (m, 2H), 2.71 (t, *J* = 7.7 Hz, 2H), 3.68 (t, *J* = 6.5 Hz, 2H), 7.20–7.22 (m, 2H), 7.28–7.32 (m, 2H) ppm. ¹³C{¹H} NMR (125 MHz, CDCl₃): δ 32.0, 34.2, 62.2, 125.8, 126.9, 128.4, 129.9, 131.4, 141.8 ppm. IR (CHCl₃): υ_{max} = 3344, 2928, 2854, 1664, 1535, 1454, 1278, 1149, 1085, 946, 758, 700 cm⁻¹.

General Procedure for the Synthesis of Homoallyl Alcohols (23a-n). Oxalyl chloride (1.523 g, 12.0 mmol, 1.5 equiv) was gradually added to a solution of DMSO (1.875 g, 24.0 mmol, 3.0 equiv) in CH₂Cl₂ (25 mL) at – 78 °C over a period of 2 min. After stirring for 15 min, a solution of alcohol **22a-n** (8.0 mmol) in CH₂Cl₂ (5 mL) was added and the reaction mixture stirred for 45 min. A solution of NEt₃ (4.5 mL, 32.0 mmol, 4.0 equiv) in CH₂Cl₂ (3 mL) was added and the reaction mixture stirred for 30 min and then gradually warmed to room temperature over 1 h. After addition of water (20 mL), the organic layer was separated and the aqueous layer extracted with CH₂Cl₂ (3 × 30 mL). The combined organic extracts were washed with water, brine, dried (Na₂SO₄) and concentrated to give the crude aldehyde, which was used directly in the next reaction.

To the solution of above aldehyde in DMSO (30 mL) was added sequentially malonic acid (1.665 g, 16.0 mmol, 2.0 equiv) and piperidinium acetate (34.8 mg, 0.24 mmol, 0.03 equiv). The mixture was stirred vigorously at 100 °C for 6-12 h and then cooled to room temperature. EtOAc (30 mL) was added and the organic layer was separated and the aqueous layer was extracted with EtOAc (2 \times 50 mL). The combined organic layers were washed with water, brine, dried (Na₂SO₄) and filtered through a plug of silica gel. The filtrate was concentrated and the residue was used in the next reaction.

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To a slurry of LiAlH₄ (0.607 g, 16.0 mmol, 2.0 equiv) in dry THF (50 mL) at 0 °C was added dropwise a solution of the above acid in THF (20 mL). The reaction mixture was refluxed for 14 h and then quenched with saturated aq. Na₂SQ₄ solution (15 mL) at 0 °C. Then EtOAc (80 mL) was added and the mixture stirred for 1 h. The mixture was filtered through a sintered glass funnel and the filtrate was concentrated. The residue was purified by silica gel column chromatography using petroleum ether/EtOAc (3:1) as eluent to give the corresponding alcohols **23a-n**.

R _____OH

23a-n

(*E*)-5-*Phenylpent*-3-*en*-1-*ol* (**23a**).^[24] Pale yellow oil (0.856 g, 66%). ¹H NMR (400 MHz, CDCl₃): δ 2.31 (q, *J* = 6.0 Hz, 2H), 3.37 (d, *J* = 6.7 Hz, 2H), 3.66 (t, *J* = 6.3 Hz, 2H), 5.49 (tdt, *J* = 14.5, 6.9, 1.4 Hz, 1H), 5.73 (dtt, *J* = 14.5, 6.7, 1.3 Hz, 1H), 7.18–7.22 (m, 3H), 7.28–7.32 (m, 2H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 358, 39.0, 61.9, 125.9, 127.4, 128.35, 128.4, 132.3, 140.4 ppm. IR(CHCl₃): υ_{max} = 3408, 2952, 1654, 1636, 1219, 1104, 1042, 855, 822, 775 cm⁻¹. LRMS (ESI-TOF) *m/z*: [M + Na]* Calcd for C₁₁H₁₄ONa 185.0936; Found 185.1049.

(*E*)-5-(4-Methoxyphenyl)pent-3-en-1-ol (**23**c). Pale yellow oil (0.969 g, 63%). ¹H NMR (400 MHz, CDCl₃): δ 1.58 (br s, 1H, *OH*), 2.30 (q, *J* = 6.5 Hz, 2H), 3.30 (d, *J* = 6.7, 2H), 3.65 (t, *J* = 6.3 Hz, 2H), 3.79 (s, 3H), 5.46 (dtt, *J* = 14.7, 6.9, 1.4 Hz, 1H), 5.70 (dtt, *J* = 14.4, 6.8, 1.3 Hz, 1H), 6.83 (d, *J* = 8.7 Hz, 2H), 7.09 (d, *J* = 8.7 Hz, 2H) ppm. ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 35.9, 38.2, 55.3, 62.0, 113.8, 127.1, 129.4, 132.5, 132.9, 157.9 ppm. IR (CHCl₃) υ_{max} = 3392, 3030, 2996, 2932, 2835, 1611, 1511, 1440, 1300, 1245, 1176,1108, 1036, 969, 817, 708, 638, 563, 513 cm⁻¹. LRMS (ESI-TOF) *m*/z: [M + Na]⁺ Calcd for C₁₂H₁₆O₂Na 215.1042; Found 215.0425.

(*E*)-5-(4-*Fluorophenyl)pent*-3-*en*-1-*ol* (**23***d*). Colourless oil (1.081 g, 75%). ¹H NMR (500 MHz, CDCl₃): δ 2.31 (q, *J* = 7.0 Hz, 2H), 3.35 (dd, *J* = 11.7, 6.7 Hz, 2H), 3.66 (t, *J* = 6.4 Hz, 2H), 5.48 (dtt, *J* = 14.5, 6.8, 1.3 Hz, 1H), 5.66–5.76 (m 1H), 6.95–6.99 (m, 1H), 7.11–7.21 (m, 2H), 7.28–7.31 (m, 1H) ppm. ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 35.9, 38.2, 62.0, 115.15 (d, *J* = 3.1 Hz), 161.4 (d, *J* = 243.4 Hz) ppm. ¹⁹F NMR (470 MHz, CDCl₃): δ –117.5 ppm. IR (CHCl₃): υ_{max} = 3343, 2931, 1660, 1510, 1453, 1222, 1158, 968, 909, 826, 759, 669 cm⁻¹. LRMS (ESI-TOF) *m*/*z* [M + Na]* Calcd for C1₁H₁₃FONa 203.0842; Found 203.0536.

(*E*)-5-(3-*Bromophenyl*)*pent*-3-*en*-1-*ol* (**23e**). Colourless oil (1.543 g, 80%). ¹H NMR (400 MHz, CDCl₃): δ 2.31 (q, *J* = 6.1 Hz, 2H), 3.37 (d, *J* = 6.7 Hz, 2H), 3.66 (t, *J* = 6.3 Hz, 2H), 5.49 (dtt, *J* = 14.9, 6.7, 1.4 Hz, 1H), 5.73 (dtt, *J* = 14.4, 6.8, 1.4 Hz, 1H), 7.18–7.22 (m, 2H), 7.28–7.32 (m, 2H) ppm.¹³C{¹H} NMR (100 MHz, CDCl₃): δ 35.9, 39.1, 62.0, 126.0, 127.4, 128.4, 128.44, 132.5, 140.5 ppm. IR (CHCl₃): ω max = 3352, 2931, 2878, 1665, 1535, 1453, 1240, 1159, 1046, 769 cm⁻¹. LRMS (ESI-TOF) *m*/*z* [M + Na]⁺ Calcd for C₁₁H₁₃BrONa 265.0022; Found 265.0888.

(*E*)-4-Phenylbut-3-en-1-ol (**23***f*). Pale yellow oil (0.901 g, 76%). ¹H NMR (500 MHz, CDCl₃): δ 1.79 (br s, 1H, *OH*), 2.49 (q, *J* = 6.2 Hz, 2H), 3.76 (t, *J* = 6.3 Hz, 2H), 6.21 (dt, *J* = 15.9, 7.2 Hz, 1H), 6.50 (d, *J* = 15.9 Hz, 1H), 7.21–7.24 (m, 1H), 7.29–7.32 (m, 2H), 7.36–7.38 (m, 2H) ppm. ¹³C{¹H} NMR (125 MHz, CDCl₃): δ 36.4, 62.0, 126.0, 126.3, 127.2, 128.5, 132.8, 137.2 ppm. IR (CHCl₃) ω_{max} = 3399, 3025, 2922, 2850, 1656, 1493, 1449, 1377, 1046, 966, 693, 668 cm⁻¹. LRMS (ESI-TOF) *m/z* [M + Na]⁺ Calcd for C₁₀H₁₂ONa 171.0780; Found 171.0729.

(*E*)-Hex-3-en-1-ol (**23g**).^[26] Colourless oil (0.553 g, 69%). ¹H NMR (400 MHz, CDCl₃): δ 0.97 (t, *J* = 7.4 Hz, 3H), 1.99–2.07 (m, 2H), 2.23–2.28 (m, 2H), 3.61 (t, *J* = 6.3 Hz, 2H), 5.36 (dtt, *J* = 14.6, 6.9, 1.4 Hz, 1H), 5.59 (dtt, *J* = 14.1, 6.3, 1.2 Hz, 1H) ppm. ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 13.7, 25.6, 35.9, 62.0, 124.7, 135.8 ppm. IR (CHCl₃): υ_{max} = 3393, 2952, 2925, 2854, 1654, 1464, 1374, 1266, 1048, 911, 802, 764 cm⁻¹.

(*E*)-Oct-3-en-1-ol (**23h**).^[26] Pale yellow oil (0.667 g, 65%). ¹H NMR (400 MHz, CDCl₃): δ 0.88 (t, *J* = 7.1 Hz, 3H), 1.23–1.36 (m, 4H), 1.99–2.03 (m, 2H), 2.25 (q, *J* = 6.1 Hz, 2H), 3.61 (t, *J* = 6.3 Hz, 2H), 5.36 (dtt, *J* = 14.8,

6.9, 1.3 Hz, 1H), 5.54 (dtt, *J* = 14.5, 6.8, 1.2 Hz, 1H) ppm. $^{13}C\{^{1}H\}$ NMR (100 MHz, CDCI₃): δ 13.9, 22.2, 31.6, 32.3, 36.0, 62.0, 125.7, 134.3 ppm. IR (CHCI₃): υ_{max} = 3368, 2957, 2927, 2873, 2857, 1653, 1466, 1378, 1266, 1049, 969, 911, 738, 706, 541, 472 cm^{-1}.

(*E*)-*Non*-3-*en*-1-*ol* (**23***i*).^[26] Pale yellow oil (0.774 g, 68%).¹H NMR (400 MHz, CDCl₃): δ 0.88 (t, *J* = 6.8 Hz, 3H), 1.24–1.38 (m, 6H), 2.00 (q, *J* = 7.0 Hz, 2H), 2.25 (q, *J* = 6.1 Hz, 2H), 3.62 (t, *J* = 6.3 Hz, 2H), 5.37 (dtt, *J* = 14.6, 6.9, 1.3 Hz, 1H), 5.55 (dtt, *J* = 14.5, 6.7, 1.2 Hz, 1H) ppm. ¹³C(¹H) NMR (100 MHz, CDCl₃): δ 14.0, 22.5, 29.1, 31.4, 32.6, 36.0, 62.0, 125.6, 134.5 ppm. IR (CHCl₃): ω_{max} = 3365, 2956, 2927, 2857, 1668, 1542, 1467, 1376, 1171, 1127, 1049, 969 cm⁻¹. LRMS (ESI-TOF) *m*/z [M + K]* Calcd for C₉H₁₈OK 181.0989; Found 181.1087.

(*E*)-*Dec*-3-*en*-1-*ol* (**23***j*).^[26] Pale yellow oil (0.788 g, 63%). ¹H NMR (400 MHz, CDCl₃): δ 0.87 (t, *J* = 6.8 Hz, 3H), 1.26–1.36 (m, 8H), 2.06 (q, *J* = 6.9 Hz, 2H), 2.26 (q, *J* = 6.2 Hz, 2H), 3.62 (t, *J* = 6.2 Hz, 2H), 5.36 (dtt, *J* = 14.6, 6.9, 1.2 Hz, 1H), 5.54 (dtt, *J* = 14.5, 6.6, 1.1 Hz, 1H) ppm. ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 14.0, 22.6, 28.8, 29.4, 31.7, 32.7, 36.0, 62.0, 125.6, 134.4 ppm. IR (CHCl₃): υ_{max} = 3367, 2952, 2925, 2854, 1465, 1378, 1176, 1049, 967 cm⁻¹.

(*E*)-*Undec-3-en-1-ol* (**23***k*).^[26] Pale yellow oil (0.844 g, 62%). ¹H NMR (500 MHz, CDCl₃): δ 0.87 (t, *J* = 6.9 Hz, 3H), 1.26–1.36 (m, 10H), 2.00 (q, *J* = 6.9 Hz, 2H), 2.25 (q, *J* = 6.2 Hz, 2H), 3.61 (t, *J* = 6.3 Hz, 2H), 5.36 (dtt, *J* = 14.7, 7.0, 1.2 Hz, 1H), 5.54 (dtt, *J* = 15.2 6.7, 1.5 Hz, 1H) ppm. ¹³C{¹H} NMR (125 MHz, CDCl₃): δ 14.1, 22.6, 27.3, 29.1, 29.4, 31.8, 32.6, 36.0, 62.0, 125.6, 134.4 ppm. IR (CHCl₃): υ_{max} = 3363, 2952, 2929, 2854, 1629, 1465, 1377, 1259, 1177, 1047, 968, 887, 758, 722, 668 cm⁻¹.

(*E*)-*Tetradec-3-en-1-ol* (**23***I*). Pale yellow oil (1.0 g, 59%). ¹H NMR (500 MHz, CDCl₃): δ 0.87 (t, *J* = 6.9 Hz, 3H), 1.25–1.36 (m, 16H), 2.00 (q, *J* = 7.1 Hz, 2H), 2.25 (q, *J* = 6.4 Hz, 2H), 3.61 (t, *J* = 6.3 Hz, 2H), 5.36 (dt, *J* = 14.7, 6.9 Hz, 1H), 5.54 (dt, *J* = 14.7, 6.7 Hz, 1H) ppm. ¹³C(¹H) NMR (125 MHz, CDCl₃): δ 14.1, 22.7, 27.4, 29.2, 29.3, 29.4, 29.5, 29.6, 31.9, 32.7, 36.0, 62.0, 125.6, 134.4 ppm. IR (CHCl₃): $\upsilon_{max} = 3411$, 2924, 2853, 1466, 1378, 1264, 1178, 1047, 970, 909, 743, 705 cm⁻¹. HRMS (ESI-TOF) *m/z* [M + Na]⁺ Calcd for C1₄H₂₈ONa 235.2032; Found 235.2027.

(*E*)-*I*cos-3-*en*-1-*ol* (**23***m*). White semisolid (1.73 g, 73%). ¹H NMR (400 MHz, CDCl₃): δ 0.87 (t, *J* = 6.8 Hz, 3H), 1.25–1.36 (m, 28H), 2.00 (q, *J* = 7.0 Hz, 2H), 2.26 (q, *J* = 6.3 Hz, 2H), 3.62 (q, *J* = 6.3 Hz, 2H), 5.36 (dtt, *J* = 14.6, 6.9, 1.3 Hz, 1H), 5.54 (dtt, *J* = 14.6, 6.7, 1.2 Hz, 1H) ppm. ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 14.1, 22.7, 29.2, 29.4, 29.48, 29.5, 29.6, 29.7, 31.9, 32.7, 36.0, 62.0, 125.6, 134.4 ppm. IR (CHCl₃) υ_{max} = 3429, 3052, 2925, 2854, 1466, 1378, 1264, 1183, 1047, 970, 909, 742, 706 cm⁻¹. HRMS (ESI-TOF) *m*/*z*: [M + Na]* Calcd for C₂₀H₄₀ONa 319.2971; Found 319.2971.

(*E*)-6-(*Benzyloxy*)*hex*-3-*en*-1-*ol* (**23***n*). Pale yellow oil (1.106 g, 67%). ¹H NMR (400 MHz, CDCl₃): δ 2.26 (q, J = 6.3 Hz, 2H), 2.35 (q, J = 6.7 Hz, 2H), 3.49 (t, J = 6.8 Hz, 2H), 3.61 (t, J = 6.3 Hz, 2H), 4.51 (s, 2H), 5.47 (dt, J = 14.1, 6.6Hz, 1H), 5.56 (dt, J = 14.4, 6.3 Hz, 1H), 7.28–7.36 (m, 5H) ppm. ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 33.1, 36.0, 61.8, 69.7, 72.8, 127.5, 127.6, 128.1, 128.3, 130.0, 138.3 ppm. IR (CHCl₃): $\upsilon_{max} = 3398$, 2929, 2859, 1605, 1448, 1356, 1097, 1048, 973 cm⁻¹. HRMS (ESI-TOF) *m*/z. [M + Na]⁺ Calcd for C₁₃H₁₈O₂Na 229.1199; Found 229.1172.

(Z)-Non-3-en-1-ol (23i"). Was prepared following literature procedure.[27]



General Procedure for the Synthesis of Diol Carbonates (5a-n). To a mixture of K₃[Fe(CN)₆] (3.951 g, 12.0 mmol, 3.0 equiv), K₂CO₃ (1.658 g, 12.0 mmol, 3.0 equiv) and few drops of pyridine or (DHQ)₂PHAL or (DHQ)₂PHAL (31.2 mg, 0.04 mmol, 1.0 mol%) in *t*-BuOH–H₂O (11, 20 mL) cooled at 0 °C was added K₂OsO₄·2H₂O (5.9 mg, 0.016 mmol, 0.4 mol%) followed by methanesulfonamide (380.5 mg, 4.0 mmol, 1.0 equiv). After stirring for 5 min at 0 °C, the olefin **23a-n** (4.0 mmol, 1.0 equiv). After stirring for 5 min at 0 °C, the olefin **23a-n** (4.0 mmol) was added in one portion. The reaction mixture was stirred at 0 °C for 24 h and then quenched with solid Na₂SO₃ (0.4 g). The stirring was continued for an additional 45 min and the solution extracted with EtOAc (3 × 40 mL). The combined organic layers were washed with water, brine, dried (Na₂SO₄) and concentrated. Silica gel column chromatography of the crude product using petroleum ether/EtOAc (2:3) as eluent gave the triol as a colourless oil, which was used for the next reaction.

To the solution of above triol in CH_2Cl_2 (40 mL) was added pyridine (1.3 mL, 16.0 mmol, 4.0 equiv) under N₂ atmosphere. The reaction mixture was stirred for 5-10 min and then triphosgene (1.19 g, 4.0 mmol, 1.0 equiv) was

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added under N₂ atmosphere and stirring was maintained at room temperature for 40 min. The reaction was quenched with saturated aq. NH₄Cl (2 mL). The phases were separated and the organic layer was diluted with EtOAc (30 mL), washed with brine, dried (Na₂SO₄) and concentrated. The residue was purified by silica gel column chromatography using petroleum ether/EtOAc (4:1) as eluent to afford **5a**-**n**.

4-Benzyl-5-(2-hydroxyethyl)-1,3-dioxolan-2-one (5a). Colourless semisolid (0.435 g, 49%). ¹H NMR (400 MHz, CDCl₃): δ1.68–1.76 (m, 1H), 1.83–1.91 (m, 1H), 2.00 (br s, 1H, OH), 3.00 (dd, J = 14.2, 6.0 Hz, 1H), 3.11 (dd, J = 14.1, 6.2 Hz, 1H), 3.70 (t, J = 6.6 Hz, 2H), 4.55–4.63 (m, 2H), 7.22–7.24 (m, 2H), 7.28–7.35 (m, 3H) ppm. ¹³C{1H} NMR (100 MHz, CDCl₃): δ 35.9, 39.2, 57.9, 78.8, 82.1, 127.4, 128.9, 129.4, 134.1, 154.4 ppm. IR (CHCl₃): $_{0max} = 3432$, 2919, 2850, 1788, 1631, 1453, 1381, 1266, 1169, 1055, 755, 701, 465 cm⁻¹. HRMS (ESI-TOF) *m*/*z* [M + Na]⁺ Calcd for C1₂H₁₄O₄Na 245.0784; Found 245.0786.

 $\begin{array}{ll} \mbox{4-}(4\mbox{-}Fluorobenzyl)\mbox{-}5\mbox{-}2\mbox{-}hyperbox{-}2\mbox{-}nyperbox{-}1\mbox{-}nyperbox{-}nyp$

4-(3-Bromobenzyl)-5-(2-hydroxyethyl)-1,3-dioxoan-2-one (**5e**). Colourless semisolid (0.650 g, 54%). ¹H NMR (400 MHz, CDCl₃): δ 1.68–1.77 (m, 1H), 1.83–1.91 (m, 1H), 2.16 (br s, 1H, OH), 3.00 (dd, J = 14.4, 5.4 Hz, 1H), 3.10 (dd, J = 14.4, 6.0 Hz, 1H), 3.69 (t, J = 5.8 Hz, 2H), 4.54–4.63 (m, 2H), 7.22–7.35 (m, 4H) ppm. ¹³C(¹H) NMR (100 MHz, CDCl₃): δ 35.9, 39.2, 57.9, 78.8, 82.1, 127.4, 128.6, 128.8, 129.3, 129.4, 134.2, 154.4 ppm. IR (CHCl₃): υ_{max} = 3414, 2927, 2888, 1795, 1660, 1546, 1497, 1455, 1384, 1242, 1215, 1166, 1058, 756, 701 cm⁻¹. LRMS (ESI-TOF) *m/z*: [M + Na]⁺ Calcd for C₁₂H₁₃BrO₄Na 303.0050; Found 303.1249.

 $\begin{array}{l} \label{eq:2-Hydroxyethyl} \text{J}\text{-}5\text{-}phenyl-1,3\text{-}dioxolan-2\text{-}one} \quad \textbf{(5f)}. \quad \text{Pale yellow oil} \\ (0.566 \text{ g}, 68\%). \ ^1\text{H} \ \text{NMR} \ (400 \ \text{MHz}, \ \text{CDCl}_3): \\ \delta 2.05 \ (\text{q}, \textit{J}=6.0 \ \text{Hz}, 2\text{H}), 2.80 \\ (\text{br s}, 1\text{H}, \textit{OH}), 3.76-3.86 \ (\text{m}, 2\text{H}), 4.72 \ (\text{q}, \textit{J}=6.5 \ \text{Hz}, 1\text{H}), 5.33 \ (\text{d}, \textit{J}=7.6 \\ \text{Hz} \ 1\text{H}), 7.34-7.42 \ (\text{m}, 5\text{H}) \ \text{ppm}. \ ^{13}\text{C}(^{11}\text{H}) \ \text{NMR} \ (100 \ \text{MHz}, \ \text{CDCl}_3): \\ \delta 35.3, \\ 57.9, 81.7, 83.3, 126.1, 129.1, 129.7, 135.2, 154.5 \ \text{ppm}. \ \text{IR} \ (\text{CHCl}_3): \\ \textit{umax} = 3433, \ 3020, 2918, 2849, 1805, 1498, 1458, 1376, 1353, 1279, 1186, \\ 1159, 1069, 1050, 1027, 973, 699, 667, 626, 588, 496 \ \text{cm}^{-1}. \ \text{LRMS} \ (\text{ESI-TOF}) \ \textit{m/z}. \ [\text{M} + \text{Na}]^+ \ \text{Calcd for } C_{11}\text{H}_{12}\text{O4}\text{Na} \ 231.0627; \ \text{Found } 231.0729. \end{array}$

5-*Ethyl-4-(2-hydroxyethyl)-1,3-dioxolan-2-one* (**5g**). Colourless oil (0.333 g, 52%). ¹H NMR (400 MHz, CDCl₃): δ 1.02 (t, J = 7.4 Hz, 3H), 1.72–1.79 (m, 2H), 1.89–1.96 (m, 2H), 2.57 (br s, 1H, *OH*), 3.79 (t, J = 6.5 Hz, 2H), 4.31 (q, J = 6.3 Hz, 1H), 4.47–4.52 (m, 1H) ppm. ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 8.7, 26.5, 36.2, 57.9, 79.1, 83.3, 154.8 ppm. IR (CHCl₃): ω_{max} = 3432, 2969, 2925, 2854, 1794, 1551, 1464, 1377, 1264, 1185, 1055, 910, 775, 734 cm⁻¹. LRMS (ESI-TOF) *m/z*. [M + K]* Calcd for C₇H₁₂O₄K 199.0367; Found 199.0581.

4-(2-Hydroxyethyl)-5-butyl-1,3-dioxolan-2-one (**5**h). Pale yellow oil (0.496 g, 66%). ¹H NMR (400 MHz, CDCl₃): δ 0.90 (t, J = 6.7 Hz, 3H), 1.34–1.45 (m, 4H), 1.66–1.77 (m, 2H), 1.90–1.96 (m, 2H), 3.81 (t, J = 5.3 Hz, 2H), 4.32–4.37 (m, 1H), 4.46–4.51 (m, 1H) ppm. ¹³C{¹H} NMR (100 MHz, 100 MHz,

CDCl₃): δ 13.7, 22.2, 26.6, 33.1, 36.1, 58.0, 79.4, 82.3, 154.8 ppm. IR (CHCl₃): υ_{max} = 3434, 2959, 2931, 2871, 1797, 1556, 1466, 1381, 1264, 1215, 1181, 1057, 914, 776, 754 cm⁻¹. HRMS (ESI-TOF) *m/z*: [M + Na]⁺ Calcd for C₉H₁₆O₄Na 211.0941; Found 211.0959.

(4\$,5\$)-4-(2-Hydroxyethyl)-5-pentyl-1,3-dioxolan-2-one (5i'). Prepared using (DHQ)₂-PHAL ligand in asymmetric dihydroxylation. Pale yellow oil (0.501 g, 62%). [a]₂²⁵ = -32.7 (c = 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 0.87 (t, J = 6.9 Hz, 3H), 1.29–1.49 (m, 6H), 1.63–1.75 (m, 2H), 1.90–1.96 (m, 2H), 2.53 (br s, 1H, *OH*), 3.81 (t, J = 5.5 Hz, 2H), 4.32–4.37 (m, 1H), 4.46–4.51 (m, 1H) ppm. ¹³C(¹H) NMR (100 MHz, CDCl₃): δ 13.8, 22.3, 24.2, 31.2, 33.4, 36.1, 58.0, 79.5, 82.3, 154.8 ppm. IR (CHCl₃): ν_{max} = 3458, 2955, 2932, 2864, 1801, 1654, 1552, 1464, 1381, 1175, 1061, 773, 668 cm⁻¹. HRMS (ESI-TOF) *m/z*. [M + Na]⁺ Calcd for C₁₀H₁₈O₄Na 225.1097; Found 225.1099. Following a similar procedure, racemic **5**i was prepared from **24**i using catalytic pyridine instead of (DHQ)₂-PHAL ligand, followed by preparation of the carbonate. Analytical data is same as **5**i'. The enantiomeric excess was determined by converting the triol **7**i' obtained after asymmetric dihydroxylation of **23i** into its tribenzoate **24i**' as described below.

(3S,4S)-Nonane-1,3,4-triyl tribenzoate (**24i**'). To a stirred solution of (3S,4S)-nonane-1,3,4-triol **7i**' (25 mg, 0.142 mmol) in pyridine (1.5 mL) was added benzoyl chloride (0.17 mL, 1.43 mmol, 10.0 equiv) at 0 °C. The reaction mixture was stirred for 12 h and then quenched with ice water. The solution was extracted with CHCl₃(2 × 10 mL). The combined organic layers were concentrated and the residue purified by silica gel coloumn chromatography using petroleum ether/EtOAc (19:1) to give the tribenzoate **24i**' as colourless semisolid (59 mg, 85%).



$$\begin{split} & [\alpha]_{D}^{25} = -0.8 \ (c = 0.25, \ CHCl_3). \ ^{1}\text{H} \ \text{NMR} \ (400 \ \text{MHz}, \ CDCl_3): \ \delta \ 0.81-0.85 \\ & (m, 3H), \ 1.26-1.38 \ (m, 4H), \ 1.39-1.43 \ (m, 2H), \ 1.78-1.79 \ (m, 2H), \ 2.24-2.27 \ (m, 2H), \ 4.36-4.42 \ (m, 1H), \ 4.46-4.54 \ (m, 1H), \ 5.46-5.50 \ (m, 1H), \ 5.65-5.69 \ (m, 1H), \ 7.36-7.45 \ (m, 6H), \ 7.50-7.57 \ (m, 3H), \ 7.98-8.07 \ (m, 6H) \ ppm. \ ^{13}\text{C}(^{1}\text{H}) \ \text{NMR} \ (100 \ \text{MHz}, \ CDCl_3): \ \delta \ 1.39, \ 2.2.4, \ 24.9, \ 30.4, \ 30.7, \ 31.5, \ 61.2, \ 71.6, \ 74.6, \ 128.3, \ 128.4, \ 129.6, \ 129.6, \ 129.7, \ 129.8, \ 129.9, \ 132.9, \ 133.06, \ 133.1, \ 165.9, \ 166.1, \ 166.4 \ ppm. \ \text{IR} \ (CHCl_3): \ \ \text{vmax} = 3063, \ 3032, \ 2955, \ 2928, \ 2857, \ 1720, \ 1602, \ 1584, \ 1491, \ 1451, \ 1378, \ 1314, \ 1268, \ 1176, \ 1108, \ 1070, \ 1026, \ 1001, \ 937, \ 913, \ 850, \ 803, \ 757, \ 711, \ 687, \ 617 \ \text{cm}^{-1} \ \text{HMS} \ (ESI-TOF) \ m/x \ [M+Na]^* \ Calcd \ for \ C_{30}H_{32}O_6 \ Na \ 511.2091; \ \text{Found} \ 511.2091; \ \text{Found}$$

4-(2-Hydroxyethyl)-5-hexyl-1,3-dioxolan-2-one (5j). Pale yellow oil (0.450 g, 52%). ¹H NMR (500 MHz, CDCl₃): δ 0.89 (t, *J* = 6.4 Hz, 3H), 1.26–1.50 (m, 8H), 1.69–1.73 (m, 2H), 1.95–2.01 (m, 2H), 3.85 (t, *J* = 4.9 Hz, 2H), 4.33–4.37 (m, 1H), 4.47–4.51 (m, 1H) ppm. ¹³C{¹H} NMR (125 MHz, CDCl₃): δ 14.0, 22.4, 24.5, 28.8, 31.5, 33.5, 36.1, 58.2, 79.4, 82.2, 154.6 ppm. IR (CHCl₃): ω_{max} = 3433, 2952, 2928, 2857, 1796, 1464,1380, 1268, 1213, 1174, 1057, 903, 692, 666 cm⁻¹. HRMS (ESI-TOF) *m/z*. [M + Na]⁺ Calcd for C11H₂₀O4Na 239.1254; Found 239.1257.

4-(2-Hydroxyethyl)-5-heptyl-1,3-dioxolan-2-one (**5**k). Pale yellow oil (0.442 g, 48%). ¹H NMR (500 MHz, CDCl₃): δ 0.88 (t, *J* = 6.7 Hz, 3H), 1.24–1.49 (m, 8H), 1.66–1.78 (m, 4H), 1.92–1.99 (m, 2H), 3.83–3.85 (m, 2H), 4.33–4.37 (m, 1H), 4.46–4.50 (m, 1H) ppm. ¹³C{¹H} NMR (125 MHz, CDCl₃): δ 14.0, 22.6, 24.6, 29.0, 29.1, 31.6, 33.5, 36.1, 58.2, 79.4, 82.2, 154.6 ppm. IR (CHCl₃): ω_{max} = 3485, 3018, 2926, 2855, 1801, 1655, 1465, 1379, 1349, 1180, 1056, 893, 695, 667, 620, 520, 488 cm⁻¹. HRMS (ESI-TOF) *m/z*: [M + Na]⁺ Calcd for C₁₂H₂₂O₄Na 253.1410; Found 253.1412.

4-(2-Hydroxyethyl)-5-decyl-1,3-dioxolan-2-one (**5**I). Pale yellow oil (0.534 g, 49%). ¹H NMR (400 MHz, CDCl₃): δ 0.87 (t, J = 7.1 Hz, 3H), 1.25–1.48 (m, 16H), 1.67–1.73 (m, 2H), 1.91–1.97 (m, 2H), 3.83 (t, J = 5.9 Hz, 2H), 4.34 (q, J = 6.6 Hz, 1H), 4.48 (q, J = 6.6 Hz, 1H) ppm. ¹³C(¹H) NMR (125 MHz, CDCl₃): δ 14.1, 22.6, 24.6, 29.1, 29.25, 29.3, 29.4, 29.5, 31.8, 33.5, 36.1, 58.1, 79.4, 82.2, 154.6 ppm. IR (CHCl₃): υ_{max} = 3482, 3018, 2925, 2855, 1794, 1561, 1465, 1379, 1344, 1181, 1058, 889, 760, 694, 667, 620, 536 cm⁻¹. HRMS (ESI-TOF) *m*/z. [M + Na]⁺ Calcd for C₁₅H₂₈O₄Na 295.1880, Found 295.1876.

(3*R*,4*R*)-4-(2-Hydroxyethyl)-5-hexadecyl-1,3-dioxolan-2-one (5*m*). Prepared using (DHQD)₂-PHAL ligand in asymmetric dihydroxylation. White solid (0.813 g, 57%), m.p. = 48-50 °C. [α] $_{D}^{25}$ = +15.4 (*c* = 0.3, CHCl₃). ¹H NMR (500 MHz, CDCl₃): δ 0.87 (t, *J* = 6.6 Hz, 3H), 1.24–1.48 (m, 26H),

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1.65–1.77 (m, 4H), 1.92–1.98 (m, 2H), 3.83–3.85 (m, 2H), 4.31–4.37 (m, 1H), 4.46–4.50 (m, 1H) ppm. $^{13}C\{^{1}H\}$ NMR (125 MHz, CDCl₃): δ 14.1, 22.7, 24.6, 29.2, 29.3, 29.5, 29.56, 29.6, 29.63, 29.7, 31.9, 33.5, 36.1, 58.2, 79.4, 82.2, 154.7 ppm. IR (CHCl₃): υ_{max} = 3396, 2918, 2852, 1778, 1760, 1471, 1378, 1051, 771 cm⁻¹. HRMS (ESI-TOF) *m*/*z* [M + Na]⁺ Calcd for C₂₁H₄₀O₄Na 379.2819; Found 379.2814. Following a similar procedure, racemic 5m was prepared from 23m using catalytic pyridine instead of (DHQD)₂-PHAL, followed by preparation of the carbonate. Analytical data is same as 5m'. The enantiomeric excess was determined by converting the triol 7m' obtained after asymmetric dihydroxylation of 23m into its tribenzoate 24m' as described below.

(*3R*,*4R*)-*lcosane-1,3,4-triyl tribenzoate* (*24m'*). The tribenzoate *24m'* was prepared from *7m'* by a similar procedure as described for *24i'* to afford *24m'* as white semisolid (85%).



 $\label{eq:approx_star} \begin{array}{l} [\alpha]_{D}{}^{25} = +16.3 \ (c=2.0, \mbox{ CHC}_{l_3}). \ ^{1}\mbox{H} NMR \ (400 \ \text{MHz}, \mbox{ CDC}_{l_3}): \ ^{0}\ ^{0$

(±)-anti-4-(2-Hydroxyethyl)-5-pentyl-1,3-dioxolan-2-one (5i").



cis-Dihydroxylation of (*Z*)-non-3-en-1-ol (**23i**", 142 mg, 1.0 mmol) and subsequent carbonate preparation by following similar procedure as described for **5i** produced **5i**" (103 mg, 51%) as pale yellow oil. ¹H NMR (500 MHz, CDCl₃): ∂ 0.88 (t, *J* = 6.6 Hz, 3H), 1.26–1.44 (m, 6H), 1.53–1.57 (m, 2H), 1.84–1.93 (m, 2H), 3.79–3.88 (m, 2H), 4.67–4.70 (m, 1H), 4.88–4.93 (m, 1H) ppm. ¹³C{¹H} NMR (125 MHz, CDCl₃): ∂ 13.9, 22.4, 25.2, 29.1, 31.3, 31.4, 58.5, 76.8, 79.9, 154.6 ppm. IR (CHCl₃): υ_{max} = 3471, 302, 917, 2850, 1778, 1757, 1469, 1378, 1310, 1178, 1051, 997, 959, 924, 912, 876, 848, 715, 666, 624 cm⁻¹. HRMS (ESI-TOF) *m/z*: [M + Na]* Calcd for C₁₀H₁₈O₄Na 225.1097; Found 225.1096.

General Procedure for the Synthesis of Enals (3a-n). To a stirred solution of 5a-n (1.0 mmol) in EtOAc (5 mL) was added IBX (0.56 g, 2.0 mmol, 2.0 equiv) under N₂ atmosphere. The resulting suspension was refluxed for 3-4 h (TLC monitoring). The reaction was cooled to room temperature and filtered through a sintered glass funnel. The filter cake was washed with EtOAc (3 \times 2 mL) and the combined filtrates were concentrated. The residue was purified by silica gel column chromatography using petroleum ether/EtOAc (7:2) as eluent to afford 3a-n (54–94%).

(*E*)-4-Hydroxy-5-phenylpent-2-enal (**3a**). Pale yellow oil (155 mg, 88%). ¹H NMR (500 MHz, CDCl₃): δ 2.35 (br s, 1H, *OH*), 2.87 (dd, *J* = 13.6, 8.3 Hz, 1H), 2.96 (dd, *J* = 13.7, 5.3 Hz, 1H), 4.62–4.66 (m, 1H), 6.31 (ddd, *J* = 15.7, 7.9, 1.6 Hz, 1H), 6.85 (dd, *J* = 15.7, 4.3 Hz, 1H), 7.22–7.24 (m, 2H), 7.27–7.28 (m, 1H), 7.32–7.35 (m, 2H), 9.54 (d, *J* = 7.9 Hz, 1H) ppm. ¹³C{¹H} NMR (125 MHz, CDCl₃): δ 43.0, 71.7, 127.1, 128.7, 129.4, 130.9, 136.4, 157.8, 193.6 ppm. IR(CHCl₃): υ_{max} = 3426, 2925, 2851, 2361, 1686, 1636,

1450, 1138, 1095, 1020, 976, 773, 701, 464 cm⁻¹. HRMS (ESI-TOF) m/z: [M + H]⁺ Calcd for C₁₁H₁₃O₂ 177.0910; Found 177.0913.

(*E*)-4-Hydroxy-5-(*p*-tolyl)*pent*-2-*enal* (**3b**). Pale yellow oil (152 mg, 80%). ¹H NMR (400 MHz, CDCl₃): δ 2.34 (s, 3H), 2.82 (dd, *J* = 13.7, 8.2 Hz, 1H), 2.94 (dd, *J* = 13.6, 5.2 Hz, 1H), 4.59–4.65 (m, 1H), 6.32 (ddd, *J* = 15.7, 7.9, 1.7 Hz, 1H), 6.85 (dd, *J* = 15.7, 4.3 Hz, 1H), 7.10–7.16 (m, 4H), 9.56 (d, *J* = 7.9 Hz, 1H) ppm. ¹³C(¹H) NMR (100 MHz, CDCl₃): δ 21.0, 42.6, 71.8, 129.3, 129.5, 130.9, 133.1, 136.8, 157.6, 193.5 ppm. IR (CHCl₃): υ_{max} = 3434, 2921, 2857, 1686, 1515, 1438, 1263, 1136, 1090, 1022, 978, 911, 866, 803, 752, 735, 672, 529, 507 cm⁻¹. HRMS (ESI-TOF) *m*/*z* [M + K]⁺ Calcd for C₁₂H₁₄O₂K 229.0625; Found 229.0623.

(*E*)-4-Hydroxy-5-(4-methoxyphenyl)pent-2-enal (**3***c*). Pale yellow oil (155 mg, 75%). ¹H NMR (400 MHz, CDCl₃): δ 2.80 (dd, *J* = 13.8, 8.2 Hz, 1H), 2.94 (dd, *J* = 13.8, 5.2 Hz, 1H), 3.80 (s, 3H), 4.59–4.64 (m, 1H), 6.33 (ddd, *J* = 15.6, 7.9, 1.6 Hz, 1H), 6.84 (dd, *J* = 15.7, 4.4 Hz, 1H), 6.87–6.89 (m, 2H), 7.14 (d, *J* = 8.6 Hz, 2H), 9.57 (d, *J* = 7.8, 1H) ppm. ¹³C(¹H) NMR (100 MHz, CDCl₃): δ 42.2, 55.3, 71.9, 114.3, 128.1, 130.4, 131.0, 157.5, 158.8, 193.4 ppm. IR (CHCl₃): υ_{max} = 3436, 2916, 2836, 1687, 1612, 1513, 1464, 1441, 1399, 1301, 1248, 1178, 1136, 1110, 1090, 1033, 981, 827, 735, 672, 600, 527 cm⁻¹. HRMS (ESI-TOF) *m/z*: [M + Na]⁺ Calcd for C1₂H₁₄O₃Na 229.0835; Found 229.0839.

(*E*)-5-(4-*Fluorophenyl*)-4-*hydroxypent-2-enal* (**3d**). Pale yellow oil (120 mg, 62%). ¹H NMR (500 MHz, CDCl₃): δ 2.84 (dd, J = 13.8, 8.2 Hz, 1H), 2.96 (dd, J = 13.8, 5.1 Hz, 1H), 4.61–4.65 (m, 1H), 6.32 (ddd, J = 15.7, 7.7, 1.7 Hz, 1H), 6.83 (dd, J = 15.7, 4.4 Hz, 1H), 7.01–7.05 (m, 2H), 7.18–7.21 (m, 2H), 9.58 (d, J = 7.8 Hz, 1H) ppm. ¹³C{¹H} NMR (125 MHz, CDCl₃): δ 42.2, 71.7, 115.65 (d, J = 21.2 Hz), 130.9 (d, J = 7.7 Hz), 131.2, 132.0 (d, J = 2.9 Hz), 156.9, 162.0 (d, J = 245.5 Hz), 193.2 ppm. ¹⁹F NMR (470 MHz, CDCl₃): δ –115.5 ppm.IR (CHCl₃) _{Umax} = 3425, 2924, 2847, 1685, 1601, 1510, 1222, 1159, 1138, 1101, 1016, 979, 913, 830, 758, 736, 534 cm⁻¹. HRMS (ESI-TOF) *m*/z [M + K]⁺ Calcd for C₁₁H₁₁FO₂K 233.0375; Found 233.0372.

 $\begin{array}{l} (E)-5-(3\text{-}Bromophenyl)-4\text{-}hydroxypent-2\text{-}enal~(3e). Pale yellow oil (0.145 g, 57\%). 1H NMR (500 MHz, CDCl_3): <math display="inline">\delta2.09~(\text{br s}, 1H, OH), 2.86~(\text{dd}, J=13.6, 8.3 Hz, 1H), 2.98~(\text{dd}, J=13.6, 5.3 Hz, 1H), 4.63-4.67~(m, 1H), 6.32~(\text{dd}, J=15.7, 7.9, 1.7 Hz, 1H), 6.85~(\text{dd}, J=15.7, 4.4 Hz, 1H), 7.22-7.36~(m, 4H), 9.56~(\text{d}, J=7.8 Hz, 1H)~\text{ppm}. ^{13}\text{C}^{1}\text{H}\}$ NMR (125 MHz, CDCl_3): $\delta43.1, 71.7, 127.1, 128.3, 128.7, 128.8, 129.4, 131.0, 136.3, 157.4, 193.4~\text{ppm}. IR (CHCl_3): \upsilon_{max} = 3435, 3029, 2926, 2849, 1686, 1639, 1605, 1495, 1454, 1138, 1094, 1027, 981, 912, 751, 702, 521~\text{cm}^{-1}. HRMS~(ESI-TOF)~m/z [M + K]^+ Calcd for C1_{11}H_{11}BrO_2K 292.9574; Found 292.9586. \end{array}$

 $\begin{array}{l} (E)\mbox{-}4\mbox{-}phenylbut\mbox{-}2\mbox{-}enal\mbox{ (3f)}. Colourless oil (87.6 mg, 54%). \mbox{^1H} \\ NMR (400 MHz, CDCl_3): $\ensuremath{\mathcal{S}} 5.48 (dd, J = 4.5, 1.6 Hz, 1H), 6.43 (ddd, J = 15.6, 7.8, 1.7 Hz, 1H), 6.91 (dd, J = 15.6, 4.6 Hz, 1H), 7.32\mbox{-}7.42 (m, 5H), 9.57 (d, J = 7.9 Hz, 1H) ppm. \mbox{^{13}C} \mbox{^{1}H} NMR (100 MHz, CDCl_3): $\ensuremath{\mathcal{S}} 73.6, 126.6, 128.7, 129.0, 130.3, 140.4, 157.0, 193.6 ppm. IR(CHCl_3): $\ensuremath{\mathcal{U}} max\mbox{=} 3430, 2921, 2852, 1686, 1136, 1093, 1010, 978, 912, 738, 702 cm^{-1}. \\ HRMS (ESI-TOF) m/z [M + K]^+ Calcd for $C_{10}H_{10}O_2K$ 201.0312; Found 201.0312. \\ \end{array}$

(*E*)-4-Hydroxyhex-2-enal (**3g**). Colourless oil (82.2 mg, 72%). ¹H NMR (400 MHz, CDCl₃): δ 1.0 (t, J = 7.5 Hz, 3H), 1.61–1.73 (m, 2H), 1.89 (br s, 1H, OH), 4.36–4.41 (m, 1H), 6.31 (ddd, J = 15.7, 7.8, 1.5 Hz, 1H), 6.82 (dd, J = 15.7, 4.7 Hz, 1H), 9.59 (d, J = 7.8 Hz, 1H) ppm. ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 9.4, 29.5, 72.3, 130.9, 158.5, 193.5 ppm. IR (CHCl₃): υ_{max} = 3433, 2955, 2924, 2853, 1685, 1559, 1463, 1377, 1261, 1086, 1041, 1024, 980, 859, 801, 762, 671, 540 cm⁻¹. HRMS (ESI-TOF) *m/z* [M + Na]⁺ Calcd for C₆H₁₀O₂Na 137.0573; Found 137.0568.

(*E*)-4-Hydroxyoct-2-enal (**3h**). Colourless oil (122.3 mg, 86%). ¹H NMR (400 MHz, CDCl₃): δ 0.91 (t, *J* = 7.2 Hz, 3H), 1.32–1.46 (m, 4H), 1.57–1.66 (m, 2H), 2.05 (br s, 1H, *OH*), 4.42 (q, *J* = 5.3 Hz, 1H), 6.30 (ddd, *J* = 15.7, 7.8, 1.3 Hz, 2H), 6.82 (dd, *J* = 15.7, 4.7 Hz, 1H), 9.57 (d, *J* = 7.8 Hz, 1H) ppm. ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 139, 22.5, 27.3, 36.2, 71.1, 130.6, 159.1, 193.7 ppm. IR (CHCl₃): υ_{max} = 3429, 2955, 2927, 2856, 1689, 1505, 1465, 1378, 1261, 1124, 1095, 1022, 978, 801, 761 cm⁻¹. HRMS (ESI-TOF) *m/z*: [M + H]⁺ Calcd for C₈H₁₅O₂ 143.1067; Found 143.1061.

 $\begin{array}{l} (S,E)\mbox{-}4\mbox{-}Hydroxynon\mbox{-}2\mbox{-}enal\ (3i\mbox{)}.^{[28]}\ Colourless\ oil\ (145.4\ mg,\ 93\%).\ [\alpha]\mbox{D}^{25} \\ =\ +43.8\ (c\ =\ 0.6,\ CHCl_3),\ lit.^{[28]}\ [\alpha]\mbox{D}^{25} \\ =\ +48.0\ (c\ =\ 0.69,\ CHCl_3).^1\ H\ NMR \\ (500\ MHz,\ CDCl_3):\ \partial\ 0.89\ (t,\ J\ =\ 6.9\ Hz,\ 3H),\ 1.28\mbox{-}1.47\ (m,\ 6H),\ 1.59\mbox{-}1.66 \\ (m,\ 2H),\ 1.89\ (br\ s,\ 1H,\ OH),\ 4.43\ (q,\ J\ =\ 5.6\ Hz,\ 1H),\ 6.30\ (ddd,\ J\ =\ 15.7,\ 7.9,\ 1.3\ Hz,\ 1H),\ 6.82\ (dd,\ J\ =\ 15.7,\ 4.7\ Hz,\ 1H),\ 9.58\ (d,\ J\ =\ 7.9\ Hz,\ 1H) \\ ppm.\ ^{13}C\{^1H\}\ NMR\ (125\ MHz,\ CDCl_3):\ \partial\ 14.0,\ 22.5,\ 24.8,\ 31.6,\ 36.4,\ 71.1,\ 103.6,\ 159.0,\ 193.6\ ppm.\ IR\ (CHCl_3):\ \upsilon_{max}\ =\ 3429,\ 2955,\ 2930,\ 2858,\ 1692,\ 1635,\ 1466,\ 1380,\ 1312,\ 1260,\ 1219,\ 1185,\ 1125,\ 1023,\ 973,\ 909,\ 797, \end{array}$

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758, 699, 595 cm $^{1}.$ HRMS (ESI-TOF) m/z [M + K]* Calcd for C_9H_{16}O_2K 193.0782; Found 193.0778.

Following a similar procedure, racemic **3i** (94%) was prepared from **5i**. Analytical data is same as **3i**'. Aslo *anti*-diol carbonate **5i''** on oxidation with IBX (2.0 equiv) produced **3i** in 80% yield.

(*E*)-4-Hydroxydec-2-enal (**3**): Pale yellow oil (153 mg, 90%). ¹H NMR (400 MHz, CDCl₃): δ 0.88 (t, *J* = 6.9 Hz, 3H), 1.29–1.45 (m, 8H), 1.60–1.66 (m, 2H), 1.75 (br s, 1H, OH), 4.43 (q, *J* = 5.6 Hz, 1H), 6.31 (ddd, *J* = 15.7, 7.9, 0.9 Hz, 1H), 6.82 (dd, *J* = 15.7, 4.7 Hz, 1H), 9.58 (d, *J* = 7.8 Hz, 1H) ppm. ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 14.0, 22.5, 25.1, 29.1, 31.6, 36.5, 71.1, 130.6, 159.2, 193.7 ppm. IR (CHCl₃): υ_{max} = 3436, 2952, 2930, 2856, 2733, 1691, 1682, 1639, 1467, 1288, 1146, 1126, 1076, 976, 757, 724 cm⁻¹. HRMS (ESI-TOF) *m/z*: [M + K]⁺ Calcd for C₁₀H₁₈O₂K 209.0938; Found 209.0943.

(*E*)-4-Hydroxytetradec-2-enal (*3*). Pale yellow oil (172 mg, 76%). ¹H NMR (500 MHz, CDCl₃): δ 0.87 (t, *J* = 7.0 Hz, 3H), 1.25–1.46 (m, 16H), 1.59–1.65 (m, 2H), 1.85 (br s, 1H, OH), 4.42 (q, *J* = 5.6 Hz, 1H), 6.30 (ddd, *J* = 15.7, 7.9, 1.2 Hz, 1H), 0.82 (dd, *J* = 15.6, 4.7 Hz, 1H), 9.57 (d, *J* = 7.9 Hz, 1H) ppm. ¹³C(¹H) NMR (125 MHz, CDCl₃): δ 14.1, 22.7, 25.2, 29.3, 29.4, 29.46, 29.5, 29.6, 31.9, 36.5, 71.1, 130.6, 159.9, 193.6 ppm. IR (CHCl₃): υ_{max} = 3433, 2927, 2855, 1693, 1467, 1378. 1272, 1035, 978, 925, 823, 763, 617, 556, 469 cm⁻¹. HRMS (ESI-TOF) *m/z*: [M + Na]⁺ Calcd for C₁₄H₂₆O₂Na 249.1825; Found 249.1823.

(*R*,*E*)-4-Hydroxyicos-2-enal (**3***m*⁷). White solid (242 mg, 78%), m.p. = 55 °C. [a]p²⁵ = -9.5 (*c* = 1.85, CHCl₃). ¹H NMR (500 MHz, CDCl₃): δ 0.87 (t, *J* = 7.1 Hz, 3H), 1.25–1.48 (m, 28H), 1.60–1.66 (m, 2H), 1.71 (br s, 1H, *OH*), 4.41–4.45 (m, 1H), 6.30 (ddd, *J* = 15.7, 7.9, 1.5 Hz, 1H), 6.82 (dd, *J* = 15.6, 4.7 Hz, 1H), 9.58 (d, *J* = 7.8 Hz, 1H) ppm. ¹³C(¹H) NMR (125 MHz, CDCl₃): δ 14.1, 22.7, 25.2, 29.4, 29.44, 29.5, 29.6, 29.63, 29.66, 29.7, 31.9, 36.5, 71.2, 130.7, 158.9, 193.6 ppm. IR (CHCl₃): _{Umax} = 3454, 2923, 2852, 1742, 1685, 1600, 1527, 1488, 1463, 1437, 1378, 1347, 1268, 1247, 1225, 1164, 1082, 1016, 852, 712, 669, 534 cm⁻¹. HRMS (ESI-TOF) *m*/z [M + Na]⁺ Calcd for C₂₀H₃₈O₂Na 333.2764; Found 333.2765.

Following a similar procedure, racemic 3m (76%) was prepared from 5m. Analytical data is same as 3m'. The enantiomeric excess of 3m' was determined by converting it into its benzoate derivative 26m' as described below.

(*R*,*E*)-1-oxoicos-2-en-4-yl benzoate (**26***m*): To a stirred solution of (*R*,*E*)-4-Hydroxyicos-2-enal **3m'** (20 mg, 0.064 mmol) in pyridine (1.5 mL) was added benzoyl chloride (0.074 mL, 0.64 mmol, 10.0 equiv) at 0 °C. The reaction mixture was stirred for 12 h and then quenched with ice water. The solution was extracted with CHCl₃(2 × 10 mL). The combined organic layers were concentrated and the residue purified by silica gel coloumn chromatography using petroleum ether/EtOAc (19:1) as eluent to give the tribenzoate **26m'** as colourless semisolid (20 mg, 80%).



[α]_D²⁵ = -20.0 (*c* = 0.10, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 0.88 (t, *J* = 6.7 Hz, 3H), 1.20–1.35 (m, 26H), 1.40–1.47 (m, 2H), 1.83–1.90 (m, 2H), 5.74–5.80 (m, 1H), 6.29 (ddd, *J* = 15.8, 7.7, 1.6 Hz, 1H), 6.85 (dd, *J* = 15.8, 4.6 Hz, 1H), 7.47 (t, *J* = 7.8 Hz, 2H), 7.60 (tt, *J* = 7.5, 1.2 Hz, 1H), 8.07 (d, *J* = 7.1 Hz, 2H), 9.59 (d, *J* = 7.7 Hz, 1H) ppm. ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 14.1, 22.7, 25.0, 29.3, 29.34, 29.4, 29.5, 29.6, 29.7, 31.9, 33.8, 72.8, 128.5, 129.6, 129.7, 131.5, 133.4, 154.1, 165.6, 193.0 ppm. IR (CHCl₃): υ_{max} = 3792, 3701, 3434, 2928, 2857, 2413, 2304, 1720, 1586, 1461, 1272, 1108, 986, 837, 793, 713, 639 cm⁻¹. HRMS (ESI-TOF) *m*/z [M + K]⁺ Calcd for C₂₇H₄₂O₃K 453.2766; Found 437.2758. The racemic benzoate was prepared following similar procedure. HPLC analysis: Chiralpak AD-H coloumn, hexane-*i*PrOH = 93:7, flow rate = 1 mL/min, t_{major} = 6.39 min, 100% ee.

(*E*)-6-(*Benzyloxy*)-4-hydroxyhex-2-enal (**3***n*). Pale yellow oil (194 mg, 88%). ¹H NMR (500 MHz, CDCl₃): δ1.82–1.89 (m, 1H), 1.95–2.01 (m, 1H), 3.65–3.74 (m, 2H), 4.50 (d, *J* = 1.2 Hz, 2H), 4.61–4.65 (m, 1H), 6.34 (ddd,

 $\begin{array}{l} J=15.6,\ 7.9,\ 1.7\ Hz,\ 1H),\ 6.79\ (dd,\ J=15.6,\ 4.2\ Hz,\ 1H),\ 7.27-7.36\ (m,\\ 5H),\ 9.54\ (d,\ J=7.9,\ 1H)\ ppm.\ ^{13}C\{^{1}H\}\ NMR\ (125\ MHz,\ CDCI_3):\ \delta\ 35.3,\\ 67.9,\ 70.1,\ 73.3,\ 127.7,\ 127.9,\ 128.4,\ 130.6,\ 137.3,\ 158.7,\ 193.6\ ppm.\ IR\ (CHCI_3):\ \upsilon_{max}=3451,\ 2921,\ 2863,\ 1685,\ 1636,\ 1367,\ 1249,\ 1114,\ 977,\ 910,\\ 738,\ 700,\ 609,\ 464\ cm^{-1}.\ HRMS\ (ESI-TOF)\ {\it m/z}.\ [M\ +\ Na]^+\ Calcd\ for\ C_{13}H_{16}O_3Na\ 243.0992;\ Found\ 243.0986. \end{array}$

3,4-Isopropylidinedioxy-nonanol (25i). To a stirred solution of triol 7i (30 mg, 0.170 mmol) in acetone (3 mL) was added p-TsOH-H₂O (1 mg, catalytic) followed by 2,2-dimethoxypropane (19.5 mg, 0.187 mmol, 1.1 equiv) at 0 °C. The reaction mixture was stirred at room temperature for 1 h under N₂ atmosphere and then quenched with 2 drops of saturated aq. NaHCO₃ and stirred for additional 5 min. The solvents were removed and the residue purified by silica gel column chromatography using petroleum ether/EtOAc (4:1) as eluent to afford 25i (28.5 mg, 77%) as colorless oil.



¹H NMR (500 MHz, CDCl₃): δ 0.89 (t, J = 6.3 Hz, 3H), 1.28–1.45 (m, 6H), 1.38 (s, 6H), 1.51–1.52 (m, 2H), 1.72–1.75 (m, 1H), 1.82–1.84 (m, 1H), 2.16 (br s, 1H, *OH*), 3.66–3.67 (m, 1H), 3.74–3.77 (m, 1H), 3.81 (t, J = 4.9 Hz, 2H) ppm. ¹³C(¹H) NMR (100 MHz, CDCl₃): δ 14.0, 22.5, 25.8, 27.2, 27.3, 31.9, 32.5, 34.8, 60.9, 80.2, 81.0, 108.4 ppm. IR (CHCl₃): υ_{max} = 3441, 2955, 2926, 2855, 1632, 1463, 1378, 1242, 1166, 1054, 876, 761, 722 cm⁻¹. HRMS (ESI-TOF) *m*/z. [M + Na]⁺ Calcd for C₁₂H₂₄O₃Na 239.1618; Found 239.1614.

3,4-Isopropylidinedioxy-nonanal (8i). Reaction of compound 25i (25 mg, 0.115 mmol) with IBX (64.4 mg, 0.230 mmol, 2.0 equiv) following similar procedure as described for **3a-n** for 10 h gave **8i** (14.6 mg, 59%) as pale yellow oil. ¹H NMR (500 MHz, CDCl₃): δ 0.89 (t, J = 6.6 Hz, 3H), 1.28–1.39 (m, 4H), 1.38, 1.40 (2s, 6H), 1.48–1.55 (m, 4H), 2.62–2.63 (m, 2H), 3.68 (q, J = 7.4 Hz, 1H), 4.07 (q, J = 7.5 Hz, 1H), 9.82 (s, 1H) ppm. ¹³C{¹H} NMR (125 MHz, CDCl₃): δ 14.0, 22.5, 25.7, 27.1, 27.2, 31.9, 32.3, 46.5, 75.5, 80.7, 108.8, 200.2 ppm. IR (CHCl₃): υ_{max} = 3440, 2956, 2928, 2856, 2728, 1729, 1629, 1460, 1241, 1167, 1009, 1031, 988, 911, 874, 735, 511 cm⁻¹. HRMS (ESI-TOF) *m/z*: [M + Na]⁺ Calcd for C₁₂H₂₂O₃Na 237.1461; Found 237.1464.

The IBX oxidation of compound **5m** after 1 h showed the presence of **5m** and **6m** in 1:1.5 ratio. The ¹H NMR of the crude reaction mixture (after concentration) showed characteristic peaks for **6m** at $\delta = 2.90$ (dd, J = 18.4, 6.8 Hz, 1H) and 3.10 (dd, J = 18.5, 6.0 Hz, 1H) ppm. These correspond to the α -hydrogens of compound **6m** (See SI for the spectra). In a 3 h reaction of **5m**, we found the presence of **6m** in minor amount and the corresponding peaks (as above) completely disappeared after another 1 h of reaction giving the final product **3m**.

(E)-4-Hydroxyicos-2-enoic acid (9). To a solution of aldehyde 3m (0.05 g, 0.16 mmol) and cyclohexene (39.4 mg, 0.48 mmol, 3.0 equiv) in t-BuOH (0.5 mL) was added sodium chlorite (14.5 mg, 0.16 mmol, 1.0 equiv) in NaH₂PO₄ (1 mL, pH 3.5 buffer) dropwise at -5 °C. The resulting colourless solution was stirred for 10 h at room temperature. Then the mixture was basified with 6 N NaOH (pH 10), and t-BuOH was removed at reduced pressure. The residue was dissolved in water and extracted with hexane $(2 \times 10 \text{ mL})$. The hexane layer was discarded and the aqueous layer was acidified (6 N HCl, pH 3) and extracted with Et₂O (2 × 5 mL). The combined organic layers were washed with water, brine, dried (Na₂SO₄) and concentrated. The residue was purified by silica gel column chromatography using petroleum ether/EtOAc (1:1) to give 9 (40 mg, 76%) as white solid, m.p. = 55–60 °C. ¹H NMR (400 MHz, CDCl₃): δ 0.87 (t, J = 7.0 Hz, 3H), 1.15–1.45 (m, 28H), 1.52–1.64 (m, 2H), 4.34 (q, J = 5.3 Hz, 1H), 6.05 (dd, J = 15.7, 1.3 Hz, 1H), 7.05 (dd, J = 15.6, 4.7 Hz, 1H) ppm. ¹³C{¹H} NMR (100 MHz, CDCl₃): δ14.1, 22.7, 25.2, 29.3, 29.4, 29.5, 29.55, 29.6, 29.64, 29.7, 31.9, 36.5, 71.1, 119.2, 152.8, 170.9 ppm. IR (CHCl₃): υmax = 3475, 2914, 2848, 1676, 1644, 1471, 1286, 1202, 1141, 1078, 976, 893, 741, 710 cm⁻¹. HRMS (ESI-TOF) *m/z*: [M + Na]⁺ Calcd for C₂₀H₃₈O₃Na 349.2713; Found 349.2717.

Methyl (E)-4-hydroxyicos-2-enoate (10). The α , β -unsaturated acid 9 (35 mg, 0.107 mmol) was treated with a solution of concentrated sulphuric acid (1 drop) in methanol (2 mL) and refluxed for one hour. The solution was cooled to room temperature and concentrated in vacuo. Et₂O (1 mL) was added and the organic layer was washed with 5% aqueous solution of NaHCO₃ followed by brine, dried (Na₂SO₄) and concentrated. The crude compound was purified by silica gel coloumn chromatography using petroleum ether/EtOAc (7:3) as eluent to give the ester 10 (30.2 mg, 83%) as light brown solid, m.p. = 48–52 °C. ¹H NMR (400 MHz, CDCl₃): δ 0.87

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(t, *J* = 7.0 Hz, 3H), 1.24–1.46 (m, 28H), 1.54–1.60 (m, 2H), 1.89 (br s, 1H, *OH*), 3.73 (s, 3H), 4.29 (q, *J* = 5.6 Hz, 1H), 6.03 (dd, *J* = 15.7, 1.2 Hz, 1H), 6.95 (dd, *J* = 15.7, 4.9 Hz, 1H) ppm. $^{13}C\{^{1}H\}$ NMR (100 MHz, CDCl₃): δ 14.1, 22.7, 25.2, 29.3, 29.4, 29.5, 29.54, 29.6, 29.7, 31.9, 36.6, 51.6, 71.1, 119.6, 150.6, 167.0 ppm. IR (CHCl₃): υ_{max} = 3361, 3244, 2918, 2847, 1726, 1662, 1464, 1430, 1325, 1274, 1191, 1178, 1137, 980, 909, 734 cm⁻¹. HRMS (ESI-TOF) m/z [M + Na]+ Calcd for C₂₁H₄₀O₃Na 363.2870; Found 363.2869.

General Procedure for the Synthesis of Diene Esters (11a-c). To a stirred solution of triethylphosphonoacetate (33.6 mg, 0.15 mmol, 1.5 equiv) in dry THF (3 mL) at 0 °C was added NaH (3.6 mg, 0.15 mmol, 1.5 equiv). The reaction mixture was stirred for 0.5 h at room temperature and then cooled to 0 °C and the aldehyde **3** (0.1 mmol) in THF (1 mL) was added dropwise. The reaction mixture was stirred for 4 h from 0 °C to room temperature (monitored through TLC). The reaction mixture was the extracted with saturated aqueous NH₄Cl solution (1 mL). The solution was extracted with EtOAc (3 × 5 mL) and the combined organic layers were washed with H₂O, brine, dried (Na₂SO₄) and concentrated. The residue was purified by silica gel coloumn chromatography using petroleum ether/EtOAc (4:1) as eluent to afford the diene esters **11a-c** (63-76%).

Ethyl (2*E*,4*E*)-6-*hydroxyhexadeca-2*,4-*dienoate* (**11a**). Pale yellow oil (22.5 mg, 76%). ¹H NMR (500 MHz, CDCl₃): δ 0.87 (t, *J* = 6.7 Hz, 3H), 1.25–1.30 (m, 19H), 1.54–1.58 (m, 2H), 4.18–4.26 (m, 3H), 5.29 (s, 1H, *OH*), 5.88 (d, *J* = 15.3 Hz, 1H), 6.11 (dd, *J* = 15.3, 5.9 Hz, 1H), 6.33–6.38 (m, 1H), 7.24–7.29 (m, 1H) ppm. ¹³C{¹H} NMR (125 MHz, CDCl₃): δ 14.1, 14.3, 22.7, 25.2, 29.3, 29.48, 29.5, 29.54, 29.6, 31.9, 37.1, 60.3, 72.0, 121.5, 127.4, 143.8, 144.9, 167.0 ppm. IR (CHCl₃): ω_{max} = 3372, 3237, 2953, 2919, 2849, 1714, 1641, 1617, 1463, 1366, 1306, 1261, 1233, 1179, 1139, 1093, 1068, 1039, 1000, 955, 921, 868, 825, 721, 656 cm⁻¹. HRMS (ESI-TOF) *m/z*: [M + Na]* Calcd for C₁₈H₃₂O₃Na 319.2244; Found 319.2247.

Ethyl (2*E*,4*E*)-6-*hydroxydocosa-2*,4-*dienoate* (**11b**). White semisolid (26.3 mg, 69%). ¹H NMR (500 MHz, CDCl₃): δ 0.88 (t, *J* = 6.8 Hz, 3H), 1.25–1.33 (m, 32H), 1.54–1.58 (m, 2H), 4.17–4.26 (m, 3H), 5.89 (d, *J* = 15.4 Hz, 1H), 6.11 (dd, *J* = 15.3, 5.9 Hz, 1H), 6.33–6.39 (m, 1H), 7.25–7.28 (m, 1H) ppm. ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 14.1, 22.7, 24.7, 25.3, 29.1, 29.2, 29.3, 29.4, 29.5, 29.53, 29.6, 29.64, 29.7, 31.9, 33.7, 37.1, 60.4, 72.0, 121.5, 127.4, 143.8, 144.9, 167.0 ppm. IR (CHCl₃): ω_{max} = 2922, 2851, 1714, 1644, 1608, 1466, 1374, 1302, 1259, 1181, 1139, 1042, 995, 910, 735 cm⁻¹. HRMS (ESI-TOF) *m/z* [M + Na]⁺ Calcd for C₂₄H₄₄O₃Na 403.3183; Found 403.3179.

Ethyl (2E,4E)-8-(benzyloxy)-6-hydroxyocta-2,4-dienoate (11c). Pale yellow oil (18.3 mg, 63%). ¹H NMR (400 MHz, CDCl₃): δ 1.29 (t, J = 7.1 Hz, 3H), 1.78–1.94 (m, 2H), 3.62–3.73 (m, 2H), 4.20 (q, J = 7.1 Hz, 2H), 4.46–4.49 (m, 1H), 4.51 (s, 2H), 5.87 (d, J = 15.3 Hz, 1H), 6.09 (dd, J = 15.2, 5.2 Hz, 1H), 6.37–6.44 (m, 1H), 7.23–7.26 (m, 1H), 7.29–7.37 (m, 5H) ppm. ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 14.3, 36.0, 60.3, 68.3, 70.9, 73.4, 121.3, 127.2, 127.7, 127.9, 128.5, 137.6, 143.9, 144.3, 167.0 ppm. IR (CHCl₃): υ_{max} = 3306, 2933, 2877, 1667, 1532, 1452, 1412, 1278, 1239, 1150, 1097, 947, 762, 665 cm⁻¹. HRMS (ESI-TOF) *m*/*z*. [M + Na]⁺ Calcd for C₁₇H₂₂O4Na 313.1410; Found 313.1413.

(*E*)-1-Phenyldocos-4-en-1-yne-3,6-diol (**12**). To a stirred solution of phenylacetylene (36.2 mg, 0.354 mmol, 2.2 equiv) in dry THF (1 mL) was added *n*-BuLi (0.22 mL, 1.6 M solution in THF, 2.2 equiv) at –78 °C. The reaction mixture was stirred for 15 min and then aldehyde **3m** (50 mg, 0.161 mmol) in THF (1 mL) was added dropwise and stirred for additional 1.5 h. After completion of reaction, it was quenched with saturated aqueous NH₄Cl (few drops) and the solution was concentrated. The residue was purified by silica gel coloumn chromatography using petroleum ether/EtOAc (4:1) as eluent to afford the diol **12** (38.5 mg, 58%) as white semisolid. ¹H NMR (500 MHz, CDCl₃): δ 0.88 (t, *J* = 7.0 Hz, 3H), 1.25–1.34 (m, 28H), 1.51–1.59 (m, 2H), 4.19 (q, *J* = 6.4 Hz, 1H), 5.12 (d, *J* = 5.4 Hz, 1H), 5.89 (ddd, *J* = 15.4, 5.5, 0.9 Hz, 1H), 5.98–6.03 (m, 1H), 7.29–7.33 (m, 3H), 7.44–7.46 (m, 2H) ppm. ¹³Cl¹H} NMR (125 MHz, CDCl₃): δ 14.1, 22.7, 25.4, 29.4, 29.5, 29.58, 29.6, 29.65, 29.7, 31.9, 37.1, 62.8, 71.9, 86.3, 87.9, 122.3, 128.3, 128.6, 129.2, 131.7, 135.7 ppm. IR (CHCl₃): $\upsilon_{max} = 3286$, 5920, 2849, 1790, 1652, 1466, 1266, 1137, 1078, 970, 917, 818, 723, 686, 564, 529 cm⁻¹. HRMS (ESI-TOF) *m*/z [M + Na]⁺ Calcd for C₂₈H₄Q₂Na 435.3234; Found 435.3233.

9-Bromononanol (**27**)^[29] To a solution of nonane-1,9-diol **13** (0.5 g, 3.12 mmol) in toluene (15 mL) was added aqueous HBr (48%, 0.360 mL, 3.12 mmol, 1.0 equiv) dropwise with stirring. The mixture was refluxed for 10 h and then cooled to room temperature and washed with dilute aqueous NaOH and brine. The organic layer was dried over Na₂SO₄ and concentrated. The residue was purified by silica gel column

chromatography using petroleum ether/EtOAc (4:1) as eluent to afford 9bromononanol **27** (0.627 g, 90%) as a colourless oil.



¹H NMR (400 MHz, CDCl₃): δ 1.31 (m, 8H), 1.40–1.43 (m, 2H), 1.53–1.59 (m, 2H), 1.85 (quintet, *J* = 7.2 Hz, 2H), 3.40 (t, *J* = 6.9 Hz, 2H), 3.64 (t, *J* = 6.6 Hz, 2H) ppm. ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 25.7, 28.1, 28.7, 29.3, 29.4, 32.7, 32.8, 34.0, 63.0 ppm. IR (CHCl₃): ω_{max} = 3390, 2930, 2855, 1659, 1464, 1437, 1264, 1247, 1220, 1057, 758 cm⁻¹. HRMS (ESI-TOF) *m/z*. [M + K]⁺ Calcd for C₉H₁₉BrOK 261.0251; Found 261.0248.

9-Bromononanoic acid (14).^[29] A solution of CrO₃ (238 mg, 2.386 mmol, 1.5 equiv) in water (1.0 mL) was cooled to 0 °C and conc H₂SO₄ (0.17 mL, 3.18 mmol, 2.0 equiv), was added cautiously followed by water (0.5 mL). After 5 min, a solution of 9-bromononanol 27 (355 mg, 1.591 mmol) in acetone (2 mL) was added dropwise. The reaction mixture was stirred for 2 h at 0 °C before warming to room temperature and then stirred for 12 h. Et₂O (5.0 mL) and H₂O (5.0 mL) were added and the aq. layer was extracted with Et₂O (3 × 2 mL). The combined organic layers were washed with brine, dried (Na₂SO₄), filtered and concentrated. The residue was purified by silica gel column chromatography using petroleum ether/EtOAc (4:1) as eluent to afford the acid 14 (370 mg, 98%) as colourless oil. ¹H NMR (500 MHz, CDCl₃): δ 1.32–1.37 (m, 6H), 1.39–1.45 (m, 2H), 1.62 (quint, J = 7.1 Hz, 2H), 1.84 (quint, J = 7.0 Hz, 2H), 2.34 (t, J = 7.5 Hz, 2H), 3.39 (t, J = 6.8 Hz, 2H) ppm. ¹³C{¹H} NMR (125 MHz, CDCl₃): δ 24.6, 28.0, 28.5, 28.9, 29.0, 32.7, 33.9, 34.0, 180.2 ppm. IR (CHCl₃): v_{max} = 3420, 2932, 2852, 1702, 1464, 1434, 1410, 1302, 1204, 1098, 937, 760, 721, 648, 542. cm⁻¹. HRMS (ESI-TOF) m/z. [M + Na]⁺ Calcd for C₉H₁₇BrO₂Na 259.0304; Found 259.0304.

Preparation of Wittig Salt (15).^[29] 9-Bromononanoic acid **14** (230 mg, 0.970 mmol) and triphenylphosphine (254 mg, 0.970 mmol, 1.0 equiv) were dissolved in toluene (10 mL). This mixture was heated to reflux for 24 h. On cooling, the upper layer was decanted and the lower layer was dissolved in CH₂Cl₂ and concentrated in vacuo. The resulting brown viscous solid was washed with boiling EtOAc, and a white viscous solid **15** (349 mg, 72%) was collected by filtration and dried in vacuo.

(S,9Z,11E)-13-Hydroxyoctadeca-9,11-dienoic acid (16). To a slurry of Wittig salt 15 (0.831 g, 1.664 mmol, 2.0 equiv) in THF (5 mL), stirred at -78 °C was added KHMDS (3.33 mL, 1 M solution in toluene, 3.33 mmol, 4.0 equiv) dropwise. The mixture was warmed to room temperature over 2 h before re-cooling to -78 °C. To this was added the aldehyde 3i' (130 mg, 0.832 mmol) in THF (3 mL) dropwise and the mixture stirred for 12 h at room temperature. EtOAc (5 mL) and 1 M HCl (5 mL) were added and the aq. layer was separated. This was further extracted with EtOAc (2×10 mL). The combined organic layers were dried (Na₂SO₄) and the solvent evaporated under reduced pressure. The residue was purified by silica gel column chromatography using petroleum ether/EtOAc (7:3) as eluent to give **16** (157.8 mg, 64%) as colourless oil. $[\alpha]_D^{25} = +6.3$ (c = 0.6, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 0.88 (t, J = 6.7 Hz, 3H), 1.25–1.39 (m, 14H), 1.46–1.64 (m, 4H), 2.14–2.19 (m, 2H), 2.33 (t, J = 7.4 Hz, 2H), 4.16 (q, J = 6.7 Hz, 1H), 5.39–5.46 (m, 1H), 5.65 (dd, J = 15.2, 6.8 Hz, 1H) 5.96 (t, J = 10.9 Hz, 1H), 6.48 (dd, J = 15.2, 11.2 Hz, 1H) ppm. ¹³C{¹H} NMR (125 MHz, CDCl₃): *δ* 14.0, 22.6, 24.6, 25.1, 25.8, 27.6, 28.8, 28.82, 29.3, 31.7, 33.9, 37.2, 72.9, 125.8, 127.8, 132.8, 135.7, 179.0 ppm. IR (CHCl₃): Umax = 3429, 2928, 2855, 1724, 1713, 1590, 1449, 1402, 1380, 1242, 1220, 1047, 1024, 948, 770. cm⁻¹. HRMS (ESI-TOF) *m/z*: [M + Na]⁺ Calcd for C₁₈H₃₂O₃Na 319.2244; Found 319.2243.

(S, 10Z, 12E)-14-Pentyloxacyclotetradeca-10, 12-dien-2-one, (S)-coriolide (1a).[12,14b] A mixture of acid 16 (50 mg, 0.169 mmol) and Et₃N (18.8 mg, 0.186 mmol, 1.1 equiv) in dry THF (3 mL) was stirred for 10 min at room temperature and then 2,4,6-tri-chlorobenzoyl chloride (41.2 mg, 0.169 mmol, 1.0 equiv) was added under N2 atmosphere. After stirring for 2 h at room temperature, the resulting precipitate was filtered and washed with a small amount of THF (1 mL). The filtrate was diluted with benzene (90 mL) and slowly added to a refluxing solution of 4-dimethylaminopyridine (DMAP) (124 mg, 1.014 mmol, 6.0 equiv) in benzene (22 mL) over a period of 8 h with dropping condenser. The reaction mixture was washed successively with a saturated aq. citric acid solution, water, and aq. NaHCO₃, dried (Na₂SO₄) and concentrated. The residue was purified by silica gel column chromatography using petroleum ether/EtOAc (9.5:0.5) as eluent to afford **1a** (27.3 mg, 58%) as colourless oil. $[\alpha]_D^{25} = +31.9$ (c = 0.5, hexane), lit.^[14b] $[\alpha]_D^{25} = +33.0$ (c = 2.82, hexane). ¹H NMR (400 MHz, CDCl₃): δ 0.88 (t, J = 6.7 Hz, 3H), 1.25–1.36 (m, 14H), 1.56–1.63 (m, 4H), 1.83-1.92 (m, 2H), 2.35-2.41 (m, 1H), 2.53-2.61 (m, 1H), 5.41-5.45 (m,

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1H), 5.48–5.55 (m, 1H), 5.72 (dd, J = 15.4, 3.8 Hz, 1H), 6.03 (t, J = 10.8 Hz, 1H), 6.46–6.54 (m, 1H) ppm. ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 14.0, 22.5, 24.8, 24.9, 25.1, 25.4, 26.3, 26.7, 27.0, 31.6, 33.0, 35.1, 72.3, 123.8, 128.3, 131.2, 132.0, 172.9 ppm. IR (CHCl₃): v_{max} = 2929, 2856, 1739, 1464, 1373, 1261, 1180, 1156, 1094, 1024, 949, 804, 545. cm⁻¹. HRMS (ESI-TOF) m/z [M + Na]⁺ Calcd for C₁₈H₃₀O₂Na 301.2138; Found 301.2134.

(R, 10Z, 12E)-14-Pentyloxacyclotetradeca-10, 12-dien-2-one, (R)-coriolide (ent-1a).[14b] To a stirred solution of diethyl azodicarboxylate (60 mg, 0.202 mmol, 2.0 equiv) and acid 16 (30 mg, 0.101 mmol) in dry CH₂Cl₂ (3 mL) at -15 °C was added triphenylphosphine (53 mg, 0.202 mmol, 2.0 equiv) under vigorous stirring. The reaction mixture was stirred for 2 h and then at room temperature for 12 h and the precipitate formed was removed by filtration. The filtrate was concentrated and the residue was purified by silica gel column chromatography using petroleum ether/EtOAc (9.5:0.5) as eluent to give **ent-1a** (13.8 mg, 49%) as colorless oil. $[a]_D^{25} = -30.7$ (c = 0.5, hexane), lit.^[14b] $[a]_D^{25} = -31.9$ (c = 1.73, hexane). Spectral data is same as 1a.

(R,E)-lcos-2-ene-1,4-diol (17). To stirred solution of 3m' (125 mg, 0.402 mmol) in dry CH2Cl2 (10 mL) was added DIBAL-H (0.58 mL, 1.75 M solution in toluene, 1.006 mmol, 2.5 equiv) dropwise under N2 atmosphere at 0 °C. The mixture was stirred for 1 h and then quenched with saturated aq. solution of sodium-potassium tartrate (5 mL) and further stirred for 3 h until two separate layers were formed. The aqueous layer was extracted with CH_2CI_2 (3 × 4 mL) and the combined organic layers were dried (Na₂SO₄)and concentrated. The residue was purified by silica gel column chromatography using petroleum ether/EtOAc (3:2) as eluent to afford 17 (100 mg, 80%) as white solid, m.p. = 76 °C. $[\alpha]_D^{25} = -4.6$ (*c* = 0.2, CHCl₃). ¹H NMR (500 MHz, CDCl₃): δ 0.88 (t, J = 6.8 Hz, 3H), 1.25–1.41 (m, 28H), 1.47–1.57 (m, 2H), 4.11–4.13 (m, 1H), 4.16 (d, *J* = 5.2 Hz, 2H), 5.72–5.76 (m, 1H), 5.81–5.86 (m, 1H) ppm. ¹³C{¹H} NMR (125 MHz, CDCl₃): *δ* 14.1, 22.7, 25.4, 29.4, 29.5, 29.58, 29.6, 29.65, 29.66, 29.7, 31.9, 37.3, 63.0, 72.3, 129.7, 134.6 ppm. IR (CHCl₃): υ_{max} = 3285, 2916, 2849, 1685, 1463, 1315, 1273, 1071, 1025, 912, 770 cm 1 . HRMS (ESI-TOF) m/z [M + Na]* Calcd for C₂₀H₄₀O₂Na 335.2921; Found 335.2921.

(R,E)-lcos-2-ene-1,4-diyl diacetate (18). To a stirred solution of 17 (50 mg, 0.160 mmol) in CH₂Cl₂ (3 mL) was sequentially added DMAP (2 mg, catalytic), anhydrous Et₃N (65 mg, 0.64 mmol, 4.0 equiv) and Ac₂O (49 mg, 0.48 mmol, 3.0 equiv) under N2 atmosphere at room temperature. The mixture was stirred for 12 h and concentrated. The residue was purified by silica gel coloumn chromatography using petroleum ether/EtOAc (5:1) as eluent to give **18** (53.9 mg, 85%) as white semisolid. [a] $_{\rm D}^{25}$ = -5.00 (c = 0.10, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 0.87 (t, J = 6.6 Hz, 3H), 1.24-1.39 (m, 28H), 1.52–1.62 (m, 2H), 2.05 (s, 3H), 2.07 (s, 3H), 4.55 (d, J = 5.3 Hz, 2H), 5.24 (q, J = 6.5 Hz, 1H), 5.66–5.80 (m, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 14.1, 20.9, 21.2, 22.7, 25.1, 29.3, 29.47, 29.5, 29.6, 29.7, 31.9, 34.2, 64.1, 73.7, 126.3, 132.6, 170.3, 170.7 ppm. IR (CHCl₃): υ_{max} = 2959, 2927, 2858, 1713, 1466, 1417, 1380, 1290, 1219, 1171, 1080, 1028, 969, 931, 760, 705, 671, 627 cm⁻¹. HRMS (ESI-TOF) m/z. [M + K]⁺ Calcd for C24H44O4K 435.2871; Found 435.2871.

(2S,3S,4R)-Icosane-1,2,3,4-tetrayl tetraacetate, D-xylo-C20-guggultetrol (20a)[19f] and (2R,3R,4R)-Icosane-1,2,3,4-tetrayl tetraacetate, D-arabino-C20-guggultetrol (20b). To a mixture of K3[Fe(CN)6] (183.7 mg, 0.558 mmol, 3.0 equiv), K₂CO₃ (77.1 mg, 0.558 mmol, 3.0 equiv), (DHQ)₂PHAL (2.2 mg, 0.0028 mmol, 1.5 mol%) in t-BuOH-H2O (1:1, 1 mL) cooled at 0 C was added K2OsO4.2H2O (0.8 mg, 0.00188 mmol, 1.0 mol%) followed by methanesulfonamide (18.6 mg, 0.186 mmol, 1.0 equiv). After stirring for 5 min at 0 °C, the olefin 18 (74 mg, 0.186 mmol) was added in one portion. The reaction mixture was stirred at 0 °C for 24 h and then quenched with solid Na₂SO₃ (60 mg). The stirring was continued for an additional 45 min and the solution extracted with EtOAc (5 \times 2 mL). The combined organic layers were washed with water, brine, dried (Na₂SO₄) and concentrated. The colourless semisolid 19 (72 mg) was used for the next reaction.

To a stirred solution of $19\ (72\ mg)$ in dry $CH_2Cl_2\ (3\ mL)$ was added sequentially DMAP (2 mg, catalytic), anhydrous Et_3N (75.3 mg, 0.744 mmol, 4.0 equiv) and Ac_2O (57 mg, 0.558 mmol, 3.0 equiv) under N_2 atmosphere at room temperature. The mixture was stirred for 12 h and then concentrated. The residue was purified by silica gel column chromatography using petroleum ether/EtOAc (4:1) to give the mixture of 20a and 20b (79 mg, dr = 9:1)). The mixture was separated by flash column chromatograpgy using petroleum ether/EtOAc (5:1) to give 20a (67 mg, 70%) and then 20b (7.6 mg, 8%) as white solids.

Data for 20a. M.p. = 44–48 °C. [α]_D²⁵ = +1.1 (c = 1.15, CHCl₃), lit.^[19f] [α]_D²⁵ = +0.99 (c = 1.0, CHCl₃). ¹H NMR (500 MHz, CDCl₃): δ 0.87 (t, J = 7.1 Hz, 3H), 1.23–1.32 (m, 28H), 1.50–1.53 (m, 2H), 2.04, 2.07, 2.07, 2.09 (4s, 12H), 3.97 (dd, J = 12.0, 5.4 Hz, 1H), 4.36 (dd, J = 11.8, 3.7 Hz, 1H), 5.07

(q, J = 6.4 Hz, 1H), 5.23–5.27 (m, 2H) ppm. ¹³C{¹H} NMR (125 MHz, CDCl₃): δ 14.1, 20.56, 20.6, 20.7, 20.9, 22.7, 24.9, 29.2, 29.3, 29.4, 29.5, 29.6, 29.63, 30.6, 31.9, 62.0, 69.6, 71.3, 71.4, 169.96, 170.0, 170.3, 170.4 ppm. IR (CHCl₃): v_{max} = 3021, 2926, 2855, 1746, 1456, 1435, 1373, 1047, 1030, 959, 669 cm⁻¹. HRMS (ESI-TOF) m/z [M + Na]⁺ Calcd for C₂₈H₅₀O₈Na 537.3398; Found 537.3396. For crystal structure see supporting information.

Data for 20b. M.p. 50–54 °C. [a]p²⁵ = -0.87 (c = 0.35, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 0.87 (t, J = 7.0 Hz, 3H), 1.24–1.39 (m, 28H), 1.51– 1.53 (m, 2H), 2.03, 2.05, 2.07, 2.11 (4s, 12H), 3.96 (dd, J = 11.7, 6.9 Hz, 1H), 4.23 (dd, J = 11.6, 5.0 Hz, 1H), 5.03 (q, J = 6.7 Hz, 1H), 5.22 (dd, J = 7.1, 3.3 Hz, 1H), 5.32–5.36 (m, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 14.1, 20.67, 20.7, 20.8, 20.9, 22.7, 25.0, 29.4, 29.42, 29.44, 29.5, 29.6, 29.66, 29.7, 30.4, 31.9, 62.1, 68.3, 70.3, 71.3, 170.0, 170.2, 170.24, 170.5 ppm. IR (CHCl₃): Umax = 2924, 2853, 1751, 1465, 1437, 1371, 1219, 1049, 958, 890, 855, 819, 756, 721, 665, 651, 629, 602, 570 cm⁻¹. HRMS (ESI-TOF) m/z. [M + Na]⁺ Calcd for C₂₈H₅₀O₈Na 537.3398; Found 537.3396.

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Conflict of Interest

The authors declare no conflict of interest.

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A tandem IBX-promoted oxidation of primary alcohol to aldehyde, opening of intermediate β , γ -diolcarbonate aldehyde to (*E*)- γ -hydroxy- α , β -enal has been developed with application to the stereoselective total synthesis of both (S)- and (R)-coriolides and D-xylo- and D-arabino-C-20-guggultetrols

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Tandem IBX-Promoted Primary Alcohol Oxidation/Opening of Intermediate β , γ -Diolcarbonate Aldehydes to (*E*)- γ -Hydroxy- α , β -enals