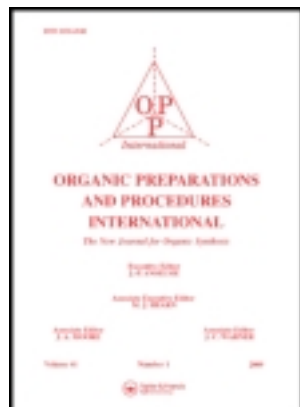


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### An Improved Procedure for the Preparation of Ribavirin

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## An Improved Procedure for the Preparation of Ribavirin

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In recent decades, there has been considerable interest in *ribavirin* (1- $\beta$ -D-ribofuranosyl-1,2,4-triazole-3-carboxamide, **4**), as a broad-spectrum antiviral agent.<sup>1–3</sup> Because of its activity against both RNA and DNA viruses, it is extensively used for the treatment of influenzas, hepatitis B and C virus,<sup>4</sup> SARS,<sup>5</sup> syncytial virus,<sup>6</sup> and its value has been enhanced since oral ribavirin was approved by the FDA in 1998, and became part of a combination therapy for the treatment of hepatitis.<sup>7</sup>

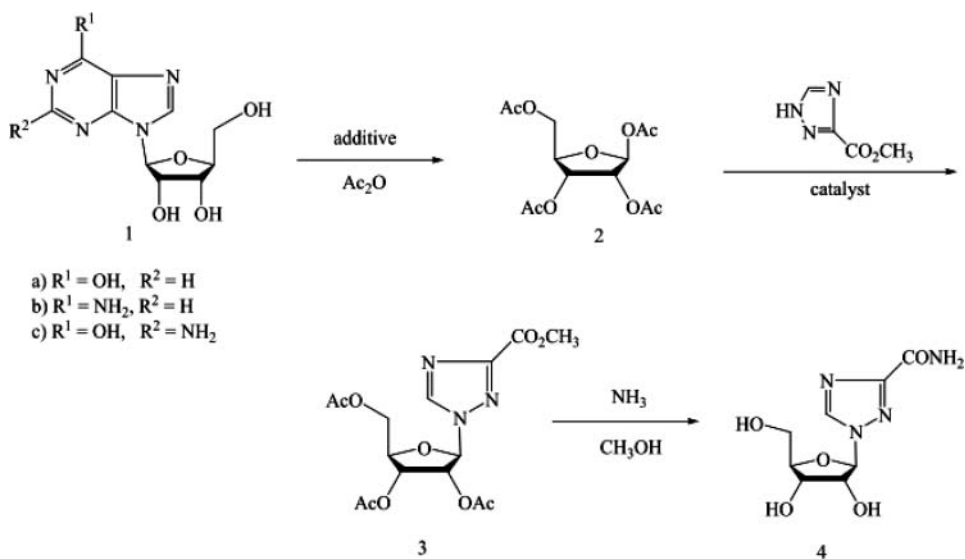
There are two types of methods for the preparation of *ribavirin*, a chemical method<sup>8–11</sup> and an enzymatic synthesis.<sup>12–15</sup> A typical chemical protocol involves a three-step manufacturing process.<sup>11</sup> Condensation of 1,2,3,5-tetra-*O*-acetyl- $\beta$ -D-ribofuranose (**2**), prepared by the *p*-toluenesulfonic acid catalyzed acetylation of inosine (**1a**), with methyl 1,2,4-triazole-3-carboxylate gave methyl 1-(2,3,5-tri-*O*-acetyl- $\beta$ -D-ribofuranosyl)-1H-1,2,4-triazole-3-carboxylate (**3**) which is subsequently converted to *ribavirin* (**4**) by deacetylation and ammonolysis (*Scheme 1*).

The process for the preparation of **2** from **1** suffer from drawbacks such as high cost, low yields, need for large amounts of catalyst and work-ups unsuitable for industrial production.<sup>10</sup> A major factor affecting this reaction is the acidity of the catalyst; if the catalyst is too acidic, the product decomposes while a catalyst of low acidity leads to long reaction times or incomplete cleavage of the nucleoside bond. Herein we report a practical, convenient and efficient process for the synthesis of **4** from inosine (**1a**, R<sup>1</sup> = OH, R<sup>2</sup> = H), adenosine (**1b**, R<sup>1</sup> = NH<sub>2</sub>, R<sup>2</sup> = H), or guanosine (**1c**, R<sup>1</sup> = OH, R<sup>2</sup> = NH<sub>2</sub>) in three steps.

*Table 1* shows the results of acetylation and cleavage of purine nucleosides promoted by various catalysts. According to published results, inosine, adenosine and guanosine can be converted into **2** and to the corresponding acetyl purines with excess of acetic acid used both as a catalyst and a solvent (*Table 1, Entries 2–4*). Better results were obtained by the

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Scheme 1

use of *p*-Toluenesulfonic acid (Table 1, Entry 1). The acetylation and cleavage of purine nucleosides required less catalyst and gave better yields. Our results show that 0.45% of TFA was sufficient to accomplish this reaction (Table 1, Entries 5–7). Compared to other reported catalysts, the amount of catalyst used was greatly reduced, the yields were improved and reaction temperatures were milder.

Similarly, previous methods<sup>8,11,17,18</sup> for the conversion of **2** to **3** suffers from the need to use higher temperatures (–), longer reaction times (Table 2, Entries

**Table 1**  
Comparison of Present Method with Previous Procedures for the Synthesis of **2**<sup>a</sup>

Entry	Nucleoside	Catalyst	Amount of catalyst% (mol)	Temp. (°C)	Yield (%) <sup>b</sup>
1	inosine	TsOH	3.10	90	84 <sup>10</sup>
2	guanosine	acetic acid <sup>c</sup>	307	138	81 <sup>16</sup>
3	adenosine	acetic acid <sup>c</sup>	296	138	73 <sup>16</sup>
4	inosine	acetic acid <sup>c</sup>	296	138	83 <sup>16</sup>
5	inosine	TFA	0.45	100	90
6	adenosine	TFA	0.45	100	84
7	guanosine	TFA	0.45	100	86

<sup>a</sup>Reaction mixtures containing 0.15 mol inosine or adenosine or guanosine and 1.5 mol acetic anhydride.

<sup>b</sup>Yield of **2** based on starting nucleosides.

<sup>c</sup>Used as the catalyst and the solvent.

**Table 2**  
Comparison of Present Catalysis with Other Procedures for the Synthesis of **3**

Entry	Ribofuranose (mol)	Methyl 1,2,4-triazole-3- 3-carboxylate/(mol)	Catalyst	Time (hrs)	Temp. (°C)	Yield (%) <sup>a</sup>
1	0.10	0.1	( <i>p</i> -O <sub>2</sub> NPh) <sub>2</sub> PO <sub>2</sub> H	0.25–0.30	160–165	78 <sup>8</sup>
2	1.26	1.46	SnCl <sub>4</sub>	2	5–40	80 <sup>17</sup>
3	0.19	0.21	<i>p</i> -TosOH	6	80–90	79 <sup>18</sup>
4	0.01	0.01	PhSO <sub>3</sub> H	5	90–100	80 <sup>18</sup>
5	0.01	0.01	MeSO <sub>3</sub> H	5	90–100	79 <sup>18</sup>
6	0.15	0.17	CF <sub>3</sub> SO <sub>3</sub> H	2	130–135	88

<sup>a</sup>Isolated yield based on 1,2,3,5-tetra-*O*-acetyl- $\beta$ -D-ribofuranose.

3–5) and give lower yields (Table 2, Entries 1–5). Of several agents studied [*bis*-(*p*-nitrophenyl)phosphinic acid, stannic chloride, *p*-toluenesulfonic, benzenesulfonic and methanesulfonic acids), trifluoromethanesulfonic acid (TfOH)] (Table 2, Entry 6) was found to be the best catalyst for the conversion of **2** to **3** (88%) in high purity at 135° in 2 hrs.

In conclusion, we have developed an improved method for the preparation of ribavirin by using TFA as the catalyst for the conversion **1** to **2** and TfOH for the conversion **2** to **3**. The advantages of our procedure include short reaction times, easy work-up, low amount of catalyst, good yields and purity of the products. A further advantage in using **1b** and **1c** is that the by-products, adenine and guanine obtained in the first step maybe useful in the synthesis of *adefovir* and *acyclovir* respectively. The overall yields of **4** from **1a**, **1b** and **1c** were 75%, 70%, 71% respectively compared to 48%, 47%, 52% by the precious method.<sup>11,15,18</sup>

## Experimental Section

All the starting materials were purchased from commercial sources and used without further purification. The IR spectra were obtained as KBr pellets on an AVATAR 370 FI-IR Thermo Nicolet spectrometer. <sup>1</sup>H NMR spectra were recorded on an AVANCE III (500 MHz) with TMS as an internal standard. Mps were determined on a Yanagimoto micromelting point apparatus and are uncorrected.

**Typical Procedure for the Preparation of 1,2,3,5-Tetra-*O*-Acetyl- $\beta$ -D-ribofuranose (2) from 1c.** A suspension of guanosine 1c (42.5 g, 0.15 mol) and acetic anhydride (153.0 g, 1.5 mol) was stirred in a three-neck 250 mL flask at 136°C for 1 h. The temperature was reduced to 60°C and TFA (0.077 g, 0.68 mmol) was then added and heating at 100°C was continued for 20–30 hrs. The progress of the reaction was monitored by TLC (1:1 ethyl acetate-acetone). Upon cooling, the precipitate formed was collected and dried to give 33.75 g (96%) of *N*<sup>2</sup>,*N*<sup>9</sup>-diacetylguanine. The filtrate was evaporated under reduced pressure and the residue was recrystallized from methanol to give 41.1 g (86%) of the 1,2,3,5-tetra-*O*-acetyl- $\beta$ -D-ribofuranose (**2**) as a white solid, mp. 82–83°C, lit.<sup>8</sup> mp. 80–82°C.

**Typical Procedure for the Preparation of 3.** A mixture of 1,2,3,5-tetra-*O*-acetyl- $\beta$ -D-ribofuranose (47.1 g, 0.15 mol) and methyl 1,2,4-triazole-3-carboxylate (21.6 g, 0.17 mol) and TfOH (0.11 g, 0.75 mmol) was heated with vigorous stirring at 135°C for 2 h under vacuum (about 55 mmHg). The solid obtained was then recrystallized from methanol, collected and dried to give 53.1 g (92%) of **3** as a white solid, mp. 106–108°C, lit.<sup>8</sup> mp. 107–109°.

**Typical Procedure for the Preparation of Ribavirin (4).** A mixture of 1,2,3,5-tetra-*O*-acetyl- $\beta$ -D-ribofuranose (47.1 g, 0.15 mol) and methyl 1,2,4-triazole-3-carboxylate (21.6 g, 0.17 mol), TfOH (0.11 g, 0.75 mmol) was heated under vigorous stirring at 135° for 2 h under vacuum (about 55 mmHg). Methanol (100 mL) was added and NH<sub>3</sub> gas was introduced into the reaction mixture for 40 hrs at 20°. The solvent was removed by distillation, and the residual solid was recrystallized from EtOH and dried under vacuum at 60° for 10 h to give 31.1 g (83%) of **4** as a white solid, mp. 169–170°C, lit.<sup>8</sup> mp. 166–168°C. The overall yield from the former procedure was 71% based on guanosine.

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