

Available online at www.sciencedirect.com

Bioorganic & Medicinal Chemistry Letters

Bioorganic & Medicinal Chemistry Letters 15 (2005) 297–302

Synthesis and antimalarial evaluation of new N^1 -(7-chloro-4-quinolyl)-1,4-bis(3-aminopropyl)piperazine derivatives

Adina Ryckebusch,^a Marie-Ange Debreu-Fontaine,^a Elisabeth Mouray,^b Philippe Grellier,^b Christian Sergheraert^a and Patricia Melnyk^{a,*}

^aInstitut de Biologie et Institut Pasteur de Lille—UMR CNRS 8525—Université de Lille II 1 rue du Professeur Calmette,

B.P. 447, 59021 Lille, France
¹USM 0504, "Biologie fonctionnelle des protozoaires", Département, "Régulations, Développement, Diversité Chimique", Muséum National d'Histoire Naturelle, 61 rue Buffon, 75005 Paris, France

> Received 19 April 2004; revised 21 October 2004; accepted 28 October 2004 Available online 18 November 2004

Abstract—Synthesis and evaluation of the activity of new N^1 -(7-chloro-4-quinolyl)-1,4-bis(3-aminopropyl)piperazine derivatives against a chloroquine-resistant strain of *Plasmodium falciparum* are described. Selectivity indices were improved for two compounds versus the lead 1, the bis-cyclopropylmethyl derivative, thus increasing the therapeutic interest of our family. As our previous studies conducted on the mode of action of our compounds made us hypothesize the existence of original mechanisms and/or original targets, terminal amino derivatives can be considered as promising tools further mechanistical studies, as probes for affinity chromatography.

2004 Elsevier Ltd. All rights reserved.

1. Introduction

Chloroquine (CQ) (Fig. 1) has been one of the two most widely used antimalarial drugs. The success of CQ was mainly due to its outstanding clinical efficacy, and the slow speed at which resistance developed to this drug. But the final arrival of resistance and the alarming spread of CQ-resistant Plasmodium falciparum on a global scale created an urgent need for the development of novel antimalarial drugs.^{[1](#page-4-0)}

Interest in 4-aminoquinolines is still prevailing^{[2,3](#page-4-0)} and several reports^{$2-6$} have shown that simple modifications on the side chain of CQ lead to analogues significantly more potent than CQ against CQ-resistant strains. These results highlighted the possibility to identify useful CQ analogues based on the same molecular mechanism of action: accumulation by weak-base effect in the acidic digestive vacuole (DV) of the parasite and inhibition of hemozoin formation, $7,8$ and circumventing the resistance mechanism.^{[9](#page-4-0)}

Figure 1. Chloroquine and compound 1.

Keywords: Antimalarial activity; Quinoline; Piperazine derivatives.

^{*} Corresponding author. Tel.: +33 3 20 87 12 19; fax: +33 3 20 87 12 33; e-mail: patricia.melnyk@ibl.fr

⁰⁹⁶⁰⁻⁸⁹⁴X/\$ - see front matter © 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2004.10.080

Figure 2. Compounds from Series 1 and 2.

Works in our laboratory focus on new N^1 -(7-chloro-4quinolyl)-1,4-bis(3-aminopropyl)piperazine derivatives. A library of sulfonamide derivatives^{[10](#page-4-0)} provided compounds up to 100-fold more potent than CQ on the CQ-resistant strain FcB1. More recently¹¹ we have reported a study concerning three series of N^1 -(7-chloro-4-quinolyl)-1,4-bis(3-aminopropyl)piperazine amides, secondary amines and tertiary amines. Among them, eleven compounds displayed a higher selectivity index (ratio CC_{50}/IC_{50} activity) than chloroquine (CQ) upon MRC-5 cells, and one of them, compound 1 [\(Fig. 1\)](#page-0-0), cured mice infected by Plasmodium berghei. Despite its curative properties, compound 1 displayed toxicity on treated mice above 20 mg/kg,^{[12](#page-4-0)} which hindered its further evaluation as a drug candidate.

Mechanistical studies conducted on our compounds suggested a CQ-like mode of action but also the involve-ment of additional mechanisms.^{[11](#page-4-0)} Fluorescence stud- $ies¹⁰$ $ies¹⁰$ $ies¹⁰$ also suggest the existence of additional targets outside the digestive vacuole, possibly involving an original mechanism of action.

Considering good pharmacological results obtained in this family, and taking into account some previously established structure–activity relationship, we decided to further enlarge chemical diversity in order to get more SAR information in our N^1 -(7-chloro-4-quinolyl)-1,4bis(3-aminopropyl)piperazine family and in the hope to improve activity/cytotoxicity ratio (using MRC-5 model). Firstly, considering compound 1 as a lead, we decided to introduce chemical diversity at the place of one of the two methylenecyclopropyl moieties (Series 1, Fig. 2). In addition, for the design of our compounds, we kept in mind their possible fixation on solid support for a ulterior use as pharmacological tools in affinity chromatography biological studies. Secondly, taking into account high antimalarial activities obtained for tertiary amines, in our previous series (among whom compound 1) we decided to replace both methylenecyclopropyl moieties by more constraint entities (Series 2, Fig. 2).

2. Chemistry

Compound 3 obtained by a procedure described previously, 11 11 11 was used as a precursor for synthesis of compounds 4–12 [\(Scheme 1](#page-2-0)). Amides 4 and 8 were prepared by reaction of amine 3 with respectively cyclopropyl carboxylic acid and Boc- β -Ala, using HBTU and HOBT as coupling agents and DIEA as a base. Amides 5 and 6 were obtained respectively by reaction with γ -butyrolactone and succinic anhydride. Synthesis of tertiary amine 11 was achieved by reductive amination from aldehyde 15 and amine 3 using sodium acetoxyborohydride. Compound 15 was previously synthesized by Boc-protection of commercially available 3-amino-propan-1-ol, followed by oxidation of alcohol to aldehyde with pyridinium chlorochromate. Primary amines 9 and 12 were obtained by deprotection of compounds 8 and 11. The succinamide 7 was prepared from acid 6, di-tert-butyl dicarbonate and ammonium carbonate.

Synthesis of cyclic amines 13–15 was achieved by nucleophilic substitution of primary amine 2, previously synthesized, with respectively dibromobutane, dibromo-pentane and dibromohexane [\(Scheme 2\)](#page-2-0).^{[13](#page-4-0)}

The compounds were characterized by MALDI-TOF and NMR and the data were consistent with the structure.[14](#page-4-0) The purity of final compounds was assumed by high-pressure liquid chromatography (HPLC).^{[15](#page-4-0)}

Scheme 1. Synthesis of compounds 4–12 (Series 1). Reagents and conditions: (a) appropriate carboxylic acid, DIEA, HBTU, HOBT, CH2Cl2, rt, 4h, 50–85%; (b) γ -butyrolactone, dry CH₃CN, 80°C, 48h, 53%; (c) succinic anhydride, dry CH₃CN, 80°C, 4h, 62%; (d) 5-chlorovaleronitrile, Nethylpiperidine, CH3CN, 40 °C, 6h, 42%; (e) compound 17, NaHB(OAc)3, CH2Cl2, rt, 4h then NaOH, 90%; (f) Boc2O, TEA, distilled THF, 0 °C, 1h then $(NH_4)_2CO_3$, rt, 16h, 50%; (g) TFA/CH₂Cl₂: 1/1, rt, 2h, 74–75%; (h) Boc₂O, dioxane/0.5N NaOH: 1/1, 16h, 80%; (i) pyridinium chlorochromate, dry CH_2Cl_2 , rt, 4h, 90%.

Scheme 2. Synthesis of compounds 13–15 (Series 2). Reagents and conditions: (a) appropriated dibromoalcane, DMF, K₂CO₃, rt, 48 h, 33–56%

3. Biological evaluation

The antimalarial activities of the two series of compounds were determined by their inhibition of parasite growth using the CQ-resistant strain FcB1 $(IC_{50}(CO) = 126 \text{ nM})$.^{[16,17](#page-5-0)} Results are given in [Table 1](#page-3-0).

In parallel, all compounds were tested for cytotoxicity upon a human diploid embryonic lung cell line (MRC-5 cells) using the colorimetric MTT assay.[18](#page-5-0)

4. Results and discussion

In Series 1 the introduction of a variety of fragments by means of amide and tertiary amine links provided compounds with a broad range of antimalarial activities.

The highest antimalarial activities were obtained for compounds 4, 10 and 11 which exhibited low IC_{50} values between 6.5 and 12.6 nM, up to 20-fold lower than CQ.

Replacement of the methylenecyclopropyl moiety in compound 1 by propylene–carbamic acid tert-butyl ester (compound 11) or butylcyanide (compound 10) provided compounds with activities 6-fold better than CQ and similar to that of compounds 1 and 3, suggesting that rather bulky and hydrophobic substituents can be introduced in this position without losing activity.

Boc-protection of the terminal primary amine in compounds 8 and 11 enhance considerably the antimalarial activity when compared to their free amine counterparts, compounds 9 and 12.

Introduction of R substituent by means of an amine link instead of amide led to more effective inhibition in the case of derivatives containing a terminal primary amine, free or protected by a Boc group. Indeed, activities of the amine compounds 11 and 12 were found to be more than 2-fold superior to those of their respective counterparts amides 8 and 9.

The average cytotoxicities of our compounds in Series 1 upon MRC-5 cells extended from 4μ M to more than 32μ M. This range provided two selectivity indices (compounds 4 and 10) superior to that of CQ, the selectivity of compound 4 being 5-fold superior to that of reference compound 1.

In Series 2, introduction of cyclic tertiary amines provided compounds between 6- and 11-fold more potent Table 1.

^a IC₅₀ values were obtained from triplicate experiments performed on the FcB1 strain. Standard error is given in brackets.
^b MRC-5 cells (human diploid embryonic lung cell line).
^c See Ref 11.

 d Nd: not determined.

than CQ. Shortening the heterocycle induced a slight decrease in activities but concomitantly lower cytotoxicities (compound 13 vs compounds 14, 15), leading for compound 13 to the best selectivity index in this series (2-fold better than compound 1).

Comparison of compound 13 with our previous results concerning its diethylamine counterpart 11 showed that rigidification in this region of the molecule yielded a slight loss in activity but also a 200-fold decrease in cytotoxicity, which finally provided for compound 13 a 22 fold increase in the selectivity index.

Further in vivo studies are going to be conducted on compounds 4 and 13 displaying selectivity indices superior to that of reference compound 1.

Our previous study^{[11](#page-4-0)} conducted on 69 derivatives displaying a common N^1 -(7-chloro-4-quinolyl)-1,4-bis(3aminopropyl)piperazine moiety showed that a selection of 19 representative compounds displayed high in vitro inhibition of β -hematin (equivalent of hemozoin) formation. Replacement of the quinoline moiety in one of these compounds by a series of heterocycles^{[19](#page-5-0)} led to a loss of the ability to inhibit β -hematin formation (with the exception of benzimidazole). Inhibition of β -hematin formation by our compounds appeared thus to be essentially due to the presence of the 7-chloro-4-aminoquinoline nucleus, consistent with studies conducted inside the DV on CQ and derivatives^{[20,21](#page-5-0)} that have demonstrated that the 7-chloro-4-amino-quinoline nucleus is responsible for hematin binding, inhibition of hemozoin formation and antimalarial activity, and that the side chain contributes only minimally to hematin binding. In order to assess the role of accumulation in the antimalarial activity we calculated the free intravacuolar accumulation ratio (VAR) ,^{[22](#page-5-0)} corresponding to equilibrium in the distribution of free compound. Even if our

compounds display generally good inhibition of hematin formation and high intravacuolar accumulation by weak-base character, no direct correlation could be established between these parameters and antimalarial activity. Exceptions such as aromatic carboxamide derivatives made us suppose the existence of additional mechanisms. Fluorescence studies conducted on sulfonamide derivatives¹⁰ made us assume that some of our compounds could have additional targets outside the digestive vacuole, possibly involving an original mechanism of action. The next step will consist in the search of these putative biological targets by the affinity chromatography technique, using a derivative of compound 1. Compounds 9 or 12 can be considered as good candidates for this technique, as their respective amino terminal function enable fixation on solid support while preserving significant antimalarial activities (respective IC_{50} 398 nM and 152 nM).

5. Conclusion

This work provided us with additional structure–activity and structure–cytotoxicity information in the N^1 -(7-chloro-4-quinolyl)-1,4-bis(3-aminopropyl)piperazine family. Indeed, this study proved that a number of substitutions lead to compounds with high activities and reduced cytotoxicities.

Synthesis and evaluation of the compounds in Series 1 showed that lipophilic substituents could be introduced in the place of one of the cyclopropyl methylene moieties in compound 1 while maintaining a high antimalarial activity. Introduction of cyclic tertiary amines on the N^1 -(7-chloro-4-quinolyl)-1,4-bis(3-aminopropyl)piperazine template provided three additional compounds with high antimalarial activities, among which one displayed improved selectivity index compared with reference compound 1.

Further investigation is required in order to enlarge diversity on the second methylenecyclopropyl member in compound 1. Parallel synthesis of two libraries of amides and tertiary amines compounds are being undertaken using compound 3 as a common precursor. Taking into account our first structure– activity relationship, these libraries will include on the terminal region of the molecule derivatives of primary amines and alcohols (i.e., etheroxides, esters, secondary and tertiary amines and amides, carbamic acid esters) and derivatives of carboxylic and hydroxamic acids.

Acknowledgements

These works are supported by CNRS (GDR 1077, FR CNRS 63, UMR CNRS 8525) and Universite´ de Lille II-Droit et Santé. A.R. was on scholarship granted by CNRS/Région Nord-Pas de Calais, France. The authors thank Gérard Montagne for NMR experiments and Hervé Drobecq for MS spectra.

References and notes

- 1. Ridley, R. G. Nature 2002, 415, 686–693.
- 2. Stocks, P. A.; Raynes, K. J.; Bray, P. G.; Park, B. K.; O'Neill, P. M.; Ward, S. A. J. Med. Chem. 2002, 45, 4975– 4983.
- 3. O'Neill, P. M.; Mukhtar, A.; Stocks, P. A.; Randle, L. E.; Hindley, S.; Ward, S. A.; Storr, R. C.; Bickley, J. F.; O'Neil, I. A.; Maggs, J. L.; Hughes, R. H.; Winstanley, P. A.; Bray, P. G.; Park, B. K. J. Med. Chem. 2003, 46, 4933– 4945.
- 4. De, D.; Krogstad, F. M.; Cogswell, F. B.; Krogstad, D. L. Am. J. Trop. Med. Hyg. 1995, 55, 579–583.
- 5. De, D.; Krogstad, F. M.; Byers, L. D.; Krogstad, D. L. J. Med. Chem. 1998, 41, 4926–4941.
- 6. Ridley, R. G.; Hofheinz, W.; Matile, H.; Jaquet, C.; Dorn, A.; Masciadri, R.; Jolidon, S.; Richter, W. F.; Guenzi, A.; Girometta, M. A.; Urwyler, H.; Huber, W.; Thaithong, S.; Peters, W. Antimicrob. Agents Chemother. 1996, 40, 1846– 1854.
- 7. Hawley, S. R.; Bray, P. G.; O'Neill, P. M.; Park, B. K.; Ward, S. A. Mol. Biochem. Parasitol. 1996, 80, 15-25.
- 8. Krogstad, D. J.; Gluzman, I. Y.; Kyle, D. E.; Oduola, A. M.; Martin, S. K.; Milhous, W. K.; Schlesinger, P. H. Science 1987, 238, 1283–1285.
- 9. Egan, T. J. Exp. Opin. Ther. Pat. 2001, 11, 185–209.
- 10. Ryckebusch, A.; Déprez-Poulain, R.; Debreu-Fontaine, M.-A.; Vandaele, R.; Mouray, E.; Grellier, P.; Sergheraert, C. Bioorg. Med. Chem. Lett. 2002, 12, 2595-2598.
- 11. Ryckebusch, A.; Déprez-Poulain, R.; Maes, L.; Debreu-Fontaine, M.-A.; Mouray, E.; Grellier, P.; Sergheraert, C. J. Med. Chem. 2003, 46, 542–557.
- 12. Two mice out of three died owing to toxicity at 40mg/kg of compound 1.
- 13. General procedure of the preparation of cyclic amines 13– 15: To a solution of amine 2 (150mg, 0.41mmol) in 5mL of DMF were added the appropriated dibromoalcane (1.2 equiv) and K_2CO_3 (287 mg, 5 equiv). After the mixture was stirred at room temperature for 48 h, dichloromethane was added and the mixture was washed with aqueous 1M $NaHCO₃$. The organic layer was separated and dried over $MgSO₄$, the solvent was evaporated and the residue was purified by thick layer chromatography to yield the desired product. 14. ¹ H and NMR spectra were obtained using a Bruker
- 300 MHz spectrometer. Chemical shifts (δ) were expressed in ppm relative to TMS used as an internal standard. Mass spectra were recorded on a time-of-flight (TOF) plasma desorption spectrometer using a californium source or on a MALDI-TOF Voyager-DE-STR spectrometer. Examples are given for compounds 4 and 13.

 $\mathbf{\hat{4}}$ ¹H NMR (CDCl₃), δ (ppm): 8.35 (d, J = 5.4 Hz, 1H, Ar-H), 8.27–8.20 (m, 1H, Ar–H), 8.01 (d, $J = 1.9$ Hz, 1H, Ar– H), 7.39–7.34 (m, 1H, Ar–H), 6.39 (d, $J = 5.5$ Hz, 1H, Ar– H), 3.49–3.45 (m, 4H, CH2), 3.30–3.27 (m, 2H, CH2), 2.80–2.75 (m, 10H, CH₂), 2.55–2.51 (m, 2H, CH₂), 2.07– 2.05 (m, 2H, CH₂), 1.98–1.62 (m, 3H, CH and CH₂), 0.96– 0.79 (m, 5H, CH and CH₂), 0.53–0.48 (m, 2H, CH₂), 0.27– 0.23 (m, 2H, CH₂); TOFMS $m/z = 484.4$. 13⁻¹H NMR (CDCl₃), δ (ppm): 8.42 (d, J = 5.4 Hz, 1H,

Ar–H), 7.86 (d, $J = 2.1$ Hz, 1H, Ar–H), 7.80 (d, $J = 8.9$ Hz, 1H, Ar–H), 6.24 (d, J = 5.4Hz, 1H, Ar–H), 3.33–3.27 (m, 2H, CH2), 2.58–2.40 (m, 18H, CH2), 1.93–1.83 (m, 2H, CH₂), 1.79–1.68 (m, 6H, CH₂); MALDI-MS $m/z = 416.2$.

15. Analytical HPLC was performed on a Shimadzu system equipped with a UV detector set at 254 nm. HPLC column: C18 nucleosil using the following eluent system: A (H2O/TFA:100/0.05) and B (CH3CN/H2O/TFA: 80/20/ 0.05). HPLC retention times (HPLC t_R) were obtained at flow rates of 1 mL/min using the following conditions: a gradient run from 100% eluent A for 1min, then to 100% eluent B over the next 30 min.4: P_{HPLC} < 99%, t_R 18.41 min; 5: P_{HPLC} < 99%, t_R 12.98 min, MALDI-MS $m/z = 502.3$; 6: P_{HPLC} < 99%, t_R 13.42min, MALDI-MS $m/z = 516.3;$ 7: P_{HPLC} 93%, t_{R} 20.74min, MALDI-MS $m/z = 515.3$; 8: P_{HPLC} < 99%, t_R 16.55min, MALDI-MS $m/z = 587.2$; 9: P_{HPLC} < 99%, t_R 11.83min, MALDI-MS $m/z = 487.6$; 10: $P_{\text{HPLC}} < 99\%$, t_R 12.72 min, MALDI-MS $m/z = 497.5$; 11: $P_{\text{HPLC}} < 99\%$, t_R 14.97 min, TOFMS $m/z = 573.3$; 12: $P_{\text{HPLC}} < 99\%$, t_{R} 11.31 min, TOFMS $m/z = 473.3;$ **13**: $P_{\text{HPLC}} < 99\%, t_{\text{R}}$ 11.41 min; **14**: P_{HPLC} < 99%, t_{R} 11.67 min, MALDI-MS $m/z = 430.2$; 15: P_{HPLC} 95%, t_{R} 12.26 min, MALDI-MS $m/z = 444.1$.

- 16. Trager, W.; Jensen, J. B. Science 1976, 193, 673–677.
- 17. Desjardins, R. E.; Canfield, C. J.; Haynes, J. D.; Chulay, J. D. Antimicrob. Agents Chemother. 1979, 16, 710–718.
- 18. Mossman, T. J. Immunol. Methods 1983, 65, 55–63.
- 19. Ryckebusch, A.; Déprez-Poulain, R.; Debreu-Fontaine, M.-A.; Vandaele, R.; Mouray, E.; Grellier, P.; Sergheraert, C. Bioorg. Med. Chem. Lett. 2003, 13, 2787–3783.
- 20. Egan, T. J.; Hunter, R.; Kaschula, C. H.; Marques, H. M.; Misplon, A.; Walden, J. J. Med. Chem. 2000, 43, 283–291.
- 21. Cheruku, S. R.; Maiti, S.; Dorn, A.; Scorneaux, B.; Bhattacharjee, A. K.; Ellis, W. Y.; Vennerstrom, J. L. J. Med. Chem. 2003, 46, 3166–3169.
- 22. Hawley, S.; Bray, P. C.; O'Neill, P. M.; Park, B. K.; Ward, S. A. Biochem. Pharmacol. 1996, 52, 723–733.