

Communication

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Chemoenzymatic synthesis of substituted azepanes by sequential biocatalytic reduction and organolithium-mediated rearrangement

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Supporting Information Placeholder

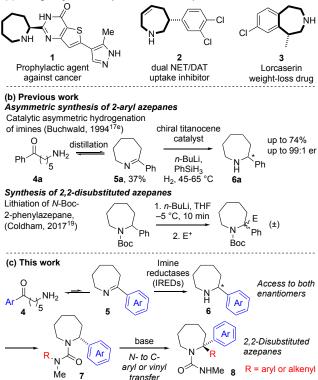
ABSTRACT: Enantioenriched 2-aryl azepanes and 2arylbenzazepines were generated biocatalytically by asymmetric reductive amination using imine reductases or by deracemisation using monoamine oxidases. The amines were converted to the corresponding N'-aryl ureas, which rearranged on treatment with base with stereospecific transfer of the aryl substituent to the 2position of the heterocycle via a configurationally stable benzyllithium intermediate. The products are previously inaccessible enantioenriched 2,2-disubstituted azepanes and benzazepines.

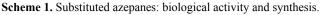
Biocatalysis provides a valuable enantioselective route to chiral amines. The discovery of enantiocomplementary imine reductases (IREDs) by Mitsukura *et al.*^{1a,b} enabled direct stereoselective reduction of imines,^{2a,b} and this approach has since been extended to intermolecular reductive amination.^{3a,b} However, these reductive methods are applicable only to amines in which the stereogenic center α to nitrogen carries a hydrogen atom. In this paper we report a way to overcome this limitation, by coupling a biocatalytic reduction with the stereospecific intramolecular arylation^{4a-e} of the resulting chiral amine.

The biocatalytic synthesis of seven-membered rings remains less explored^{2a,5a,b} than smaller ring sizes, and we chose to apply our strategy to the synthesis of 7-membered heterocyclic α -tertiary amine derivatives of potential utility in medicinal chemistry. Molecules incorporating 7-membered heterocyclic rings, particularly azepanes and benzazepines, find many applications in medicinal and pesticidal chemistry.⁶ Azepanes display activity as glycosidase inhibitors,^{7a-e} norepinephrine and dopamine transporters⁸ or protein kinase C inhibitors,⁹ and are promising candidates for the treatment of viral infections,^{7b} diabetes¹⁰ or cancer.^{7a,11a,b} Natural products and medicinal agents incorporating the azepane ring (Scheme 1a) include balanol,⁹ zilpaterol,⁹ azelastine,¹² capuramycin^{6,13} and *Stemona* alkaloids.¹⁴ Azepane 1 is a potential therapeutic agent against cancer,¹⁵ and **2** is an inhibitor of NET/DAT.8 The benzazepine core is also present in fenoldopam^{16a} and lorcaserin **3**.^{16b}

Strategies for the asymmetric synthesis of 2-aryl azepanes include C-H arylation of *N*-thioamide azepanes,^{17a} cyclisation of entities containing pre-formed stereogenic centres,^{17b-d} and enantioselective hydrogenation of the corresponding imines (Scheme 1b).^{17e,f} However, formation of seven-membered heterocyclic rings is generally not straightforward.^{18a-d} There are even fewer methods for the preparation of 2,2-disubstituted azepanes. Coldham *et al.* developed the lithiation-substitution of *N*- Boc-2-phenylazepane to yield racemic 2,2-disubstituted azepanes,¹⁹ and the synthesis of enantioenriched compounds with this core is limited to quaternary α -amino acid derivatives prepared by ring-closing methathesis^{20a} or structures obtained by the Schmidt rearrangement of enantioenriched α , α -disubstituted ketones.^{20b,c}







The 7-membered imine **5a** is a useful starting point for the asymmetric synthesis of 2-substituted azepanes, but its use is complicated by its equilibration to the corresponding aminoketone **4a**, even in organic solvents, and it must be purified by distillation.^{17e} A biocatalytic approach to the synthesis of 2-substituted azepanes **6** (Scheme 1c) could overcome this problem, allowing reductive amination to take place directly from the equilibrating mixture. As a preliminary to an exploration of biocatalytic reduction, equilibrium between **4a** and **5a** was

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quantified by acquiring ¹H NMR spectra of **5a** in aqueous 1 M phosphate solutions at pH values ranging from 1 to 8 (see SI for details). Across this whole pH range, the amount of 7-membered imine **5a** formed from aminoketone **4a** was undetectable by NMR. By comparison, the homologous 5-aminoketone **9** formed 80-90% of 6-membered cyclic imine **10** under the same conditions (Scheme 2).

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This instability of the 7-membered imine in the aqueous medium makes the biotransformation much more challenging than for the 6-membered ring equivalent. Indeed, neither *R*-IRED^{1a} nor *S*-IRED^{1b} (both of which displayed activity towards **10**) reduced imine **5a** to 2-phenyl azepane **6a**. We therefore tested two other enzymes, both of which catalyse intermolecular reductive amination: the reductive aminase AspRedAm from *Aspergillus oryzae*^{3a} and imine reductase IR-22 reported by Roiban *et al.* (Scheme 2 and Table 1).^{3b} Remarkably, IR-22 gave the *R* enantiomer of **6a** from **5a** in good conversion and excellent e.r.

In order to find an enantiocomplementary IRED, we screened a small panel of novel IREDs, both genomic and metagenomic, against the 6-membered ring imine **10** (Table 1; see SI for full screening data). Those displaying good conversions with **10** were also screened against the 7-membered ring imine **5a**. Three of the novel IREDs gave excellent enantioselectivities, forming the *S* enantiomer of **6a** with conversions exceeding 60%. CfIRED from *Cystobacter ferrugineus* gave the best conversion (92%). Metagenomic mIRED 10 was *R* selective, giving 64% conversion and 94:6 e.r. The reduction of **5a** with CfIRED was repeated preparatively on a 50 mg scale and gave **6a** in 85% yield.

Scheme 2. Biocatalytic routes to enantioenriched 2-phenyl azepane 6a.

IRED cell-free extract (2 mg/mL) Bacillus subtilis glucose 9(n = 1)NH₂ 4a (n = 2) dehydrogenase cell-free extract (1 mg/mL), NADP+ (1 mM), alucose (5 equiv.) Tris-HCl buffer (100 mM, pH 8.0) **11** (*n* = 1) 30 °C, 250 rpm, 20 h **10** (n = 1)6a (n = 2) 5a (n = 2) E. coli with MAO-N D11 (±)-6a KP_i buffer (1 M, pH 7.8), 37 °C, 250 rpm, 24 h 10-20 mM (R)-6a 41%; 99:1 er

Table 1. Enantioselective reduction of cyclic imines **10** and **5a** by imine reductases.^{*a*}

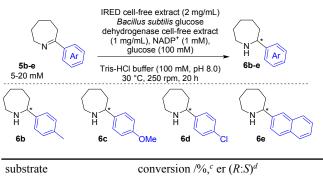
IRED	conversion	conversion
	to 11 $/\%$; (<i>R</i> : <i>S</i>) ^{<i>b</i>}	to 6a $/\%;^{c}(R:S)^{d}$
<i>R</i> -IRED ^{1a}	>99 (23:77)	0
S-IRED ^{1b}	>99 (96:4)	0
AspRedAm ^{3a}	0	0
IR-22 ^{3b}	86 (93:7)	70 ^e (98:2)
CfIRED	>99 (1:99)	92 (<1:99); 85 ^f (<1:99)
PcIRED	>99 (<1:99)	62 (<1:99)
mIRED 02	>99 (<1:99)	81 (<1:99)
mIRED 10	>99 (>99:1)	64 (94:6)

^{*a*}Experiments with **5a** performed with 2 mg/mL IRED cell-free extract, 20 mM substrate and 3 mL total reaction volume, while for **10** substrate concentration was 10 mM; ^{*b*} determined by GC; ^{*c*} determined by ¹H NMR; ^{*d*} determined by HPLC on chiral stationary phase; ^{*e*}5 mg/mL IRED cell-free extract concentration. ^{*f*} Isolated yield of preparative scale reaction per 50 mg batch. As an alternative to the asymmetric reduction of **5**, racemic **6** was kinetically resolved with enantioselective amine oxidases (MAO-N) (see SI for full scope).^{21a,b} On a 50 mg scale, *S*-selective oxidation of (\pm) -**6a** with MAO-N D11 gave (*R*)-**6a** in 41% yield and >99:1 er (Scheme 2). Attempts to incorporate this kinetic resultion into a deracemisation cycle by using ammonia-borane as a non-stereoselective reducing agent^{21a} failed as a result of the ring opening of **5a** to **4a**, which was reduced to the corresponding alcohol.

Differently substituted 7-membered cyclic imines **5b-e** were reduced using the *R*- and *S*-selective IREDs that gave best conversions with **5a** (CfIRED, IR-22, mIRED 02 and mIRED 10, Table 2). With the *p*-tolyl-substituted substrate **5b**, all of the enzymes gave good conversions to **6b**, and most also gave excellent enantiomeric ratios. Both enantiomers of **6c** were obtained in >99:1 er and at least 60% conversion for the less electrophilic *p*methoxyphenyl-substituted imine **5c**. The most electrophilic of the substrates tested, *p*-chlorophenyl-substituted **5d**, also gave excellent conversions and ers. Despite the insolubility of 2naphthyl-substituted **5e** in the aqueous buffer, **6e** was formed from two of the enzymes. In all of the enantioselective reductions, CfIRED and mIRED 02 consistently gave (*S*)-**6**, while IR-22 and mIRED 10 gave (*R*)-**6**.

 Table 2. Enantioselective reduction of differently substituted

 seven-membered ring imines 5b-e by imine reductases.



substrate	conversion / 70, er (K.S)				
	CfIRED	IR-22 ^{3b}	mIRED 02	mIRED 10	
$5b^a$	75, 1:99	67, 99:1	72, 1:99	67, 62:38	
5c ^b	69, 1:99	60, >99:1	65, <1:99	53, 50:50	
5d ^a	92, <1:99	81, 96:4	84, <1:99	63, 70:30	
5e	90, <1:99	0, -	64, <1:99	0, -	

^{*a*5} mg/mL IRED cell-free extract concentration; ^{*b*5} mM substrate and 25 mM glucose concentration; ^{*c*}determined by ¹H NMR spectroscopy; ^{*d*}determined by HPLC on chiral stationary phase.

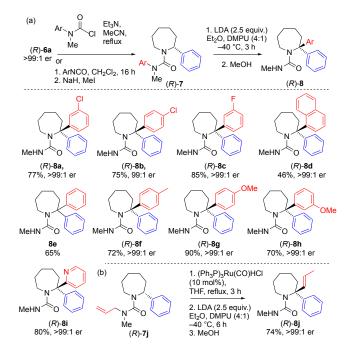
Although these 2-aryl azepanes are valuable scaffolds in their own right, they also offer a biocatalytic approach to the synthesis of synthetically challenging heterocyclic α -tertiary amines through stereospecific replacement of the α -CH bond by a carbon substituent. (Scheme 3). Such substitutions may be achieved by way of configurationally stable organolithium intermediates, with aryl migrations within lithiated ureas providing a method for stereospecific arylation.^{4a-e} An *N*'-substituted urea lithiated α to N undergoes a stereospecific intramolecular transfer of an aryl,^{4a} heteroaryl^{4b,22a} or vinyl^{22b} group to the α -position, giving α -tertiary amine derivatives that may be deprotected by solvolysis.

Ureas 7 were made from 6a using standard methods, and rearrangement to 8a was attempted by treatment of 7a with base (Scheme 3). After a series of unsuccessful rearrangements (see SI for full details) we found that treatment of (R)-7a with LDA in a

4:1 mixture of Et₂O and DMPU generated a benzylic organolithium that rearranged by *N*- to *C*- aryl migration, giving enantiopure **8a** (>99:1 er) in 77% yield. We assign the configuration of the product as *R* on the basis that related reactions^{4a,22a,23} proceed with retention of configuration.

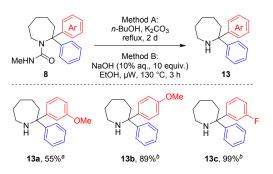
These optimised reaction conditions were applied to a range of 2-arylazepane-derived ureas 7 (Scheme 3a). These underwent organolithium-mediated arylation with a range of migrating groups to give products 8 with complete stereospecificity. Migrations of *meta*- (7a and 7c) and *para*- (7b) halogenated aromatic rings gave excellent yields and enantiomeric ratios. Even transfer of a sterically demanding 1-naphthyl ring gave enantiomerically pure product 8d, although in a lower yield. Despite the fact that this reaction is formally a nucleophilic aromatic substitution,^{4a} yields and stereospecificity were maintained in the migration of electron rich tolyl (8f) and methoxyphenyl rings (8g-h). Migration of a 2pyridyl ring gave enantiopure 8i incorporating both an azepane and pyridyl ring in 80% yield. These results are particularly remarkable as cyclic benzyllithiums are generally characterized by poor configurational stability^{24a-c} with racemization competing with rearrangement.4e,25

The method is not limited to aryl migrations.^{22b} An *N*-(*E*)propenylurea was made from the corresponding allyl urea **7j** (Scheme 3b; details of the synthesis in the SI) and treated under the same basic conditions. Stereospecific *N*- to *C*- alkenyl migration gave **8j** as a single geometrical isomer and in enantiomerically pure form, offering potential for further elaboration of the molecule at the double bond.



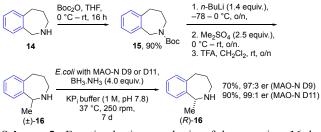
Scheme 3. Organolithium mediated α -functionalisation of azepinederived ureas. (a) Arylation by *ipso* attack on an N-aryl group; (b) Alkenylation by stereo-retentive substitution.

The rearrangement products **8** were cleanly deprotected either by solvolysis in *n*-butanol or by alkaline hydrolysis using microwave irradiation (Scheme 4). The products are previously inaccessible α -tertiary amines with the azepane scaffold **13**.²⁶



Scheme 4. Deprotection of ureas 8. aMethod A; bMethod B

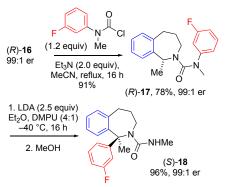
A similar chemo-enzymatic approach was applied to the biologically important 2-benzazepine scaffold, which is a component of a PNMT inhibitor.^{27a,b} Boc-protected 2,3,4,5-tetrahydro-1*H*-benzo[c]azepine **15** was methylated by way of its 2-lithio derivative^{24a,28} and deprotected to give **16** (Scheme 5).



Scheme 5. Enantioselective synthesis of benzazepine **16** by alkylation and deracemisation.

Benzazepine **16** was deracemised by treatment with *S*-selective oxidants MAO-N D9 and D11, using the unselective reductant BH₃.NH₃ to recycle the imine product (Scheme 5). Excellent enantioselectivities were obtained. The stability of the imine oxidation product of **16** appears to prevent hydrolysis to the corresponding aminoalcohol was evident in the crude ¹H NMR of the product. The methylation of **15** and deracemisation of **16** constitutes a novel approach to the asymmetric functionalisation of 2,3,4,5-tetrahydro-1*H*-benzo[*c*]azepines, which was previously achieved only with chiral auxiliaries.³⁰

To demonstrate the potential for further functionalization at the 2-position, the enantioenriched 2-benzazepine (*R*)-16 was converted into urea (*R*)-17 and treated with LDA in Et₂O/DMPU (Scheme 6). The *meta*-fluorophenyl ring migrated successfully to the 2-position of the urea, giving the heterocyclic α -tertiary amine derivative (*S*)-18 with a 2-benzazepine structure.



Scheme 6. Stereospecific arylation of a benzazepine-derived urea

In summary, we show that a chemo-enzymatic strategy for the preparation of heterocyclic α -tertiary amines with the azepane and

2-benzazepine scaffolds. The method makes use of enantioselective imine-reductase catalysed asymmetric imine reduction or deracemisation with amine oxidases, followed by stereospecific rearrangements of cyclic *N*-aryl benzyllithiums. Key aspects include the application of four novel imine reductases, and the stereospecific functionalisation of configurationally stable benzylic organolithium derivatives of nitrogen heterocycles. The products constitute a new class of functionalized azepanes and benzazepines of potential utility as drug molecules.

ASSOCIATED CONTENT

Supporting Information

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Experimental desriptions for chemistry and biocatalysis; spectroscopic data for all new compounds.

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The authors declare no competing financial interests.

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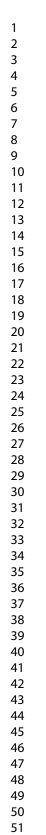
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