Communications

Molecular Recognition

Identifying Specific Conformations by Using a Carbohydrate Scaffold: Discovery of Subtype-Selective LPA-Receptor Agonists and an Antagonist**

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Lysophosphatidic acids (LPA) are a group of important extracellular signaling molecules that elicit a wide variety of fundamental biological responses, such as cell-growth stim-

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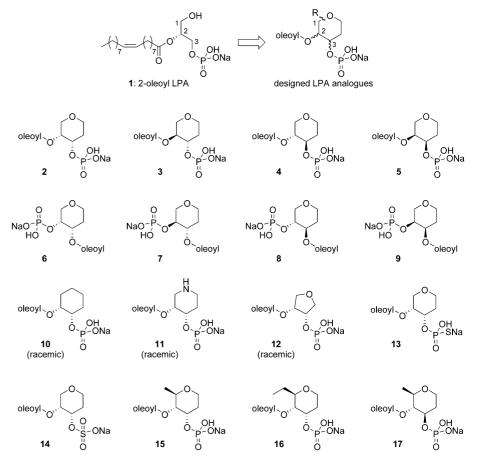
ulation, calcium mobilization, escape from apoptosis, tumor-cell invasion, and smoothmuscle contraction.^[1] The diverse properties are mediated through interactions of LPA with G-protein-coupled receptors named LPA₁, LPA₂, and LPA₃. Studies to explore the specific role of each subtype are in progress, and are essential for further advances, for example, for designing drugs to modulate a specific LPA-mediated signal-transduction pathway. Studies with subtype-selective agonists and antagonists are very powerful for this purpose. Herein, we describe a new strategy for the discovery of subtype-selective ligands for LPA receptors, based on the identification of specific active conformations of LPA by using carbohydrates as scaffolds.

2-Oleoyl LPA (1) is the most potent physiological molecule in the LPA family. Studies on this LPA-receptor ligand have focused mainly on the generation of stable analogues by preventing acyl migration between the 1- and 2-hydroxy groups.^[2-4] These studies led to the proposed LPA pharmacophore,^[5] as well as to several subtype-selective agonists and antagonists.^[6] The activity and selectivity of these analogues, however, are not yet satisfactory for their use as biological tools. This might be partly because of the conformational flexibility of these analogues. The population

and/or lifetime of analogues in the active three-dimensional conformation might not be sufficient for selective receptor activation as a result of a thermal disturbance. Therefore, we planned to restrict the conformational flexibility by introducing a ring structure.^[7]

Our molecular design for LPA analogues is summarized in Scheme 1. Based on the hypothesis that different LPAreceptor subtypes distinguish different three-dimensional arrangements of a negative charge (phosphate anion), a long hydrophobic tail (oleoyl group), and a hydrogen-bond acceptor (oxygen atom at the 1-position),^[8] we synthesized an array of molecules that display these recognition motifs at various relative positions defined by core carbohydrate scaffolds.^[9] By employing a variety of readily available enantiomerically pure carbohydrate configurational isomers, the arrangement can be finely tuned. The diversity of the arrangements can be further increased by introducing substituents on the ring to modulate the ring conformation. For each arrangement, a particular recognition motif can be modified independently to examine the effect of the motif. Furthermore, problematic acyl migration is prevented in these analogues, because the oxygen atom corresponding to the 1-hydroxy group of 2-oleoyl LPA is incorporated into the pyran ring. Based on this idea, we synthesized approximately 40 molecules; selected compounds are shown in Scheme 1.^[10]

First, the agonist activity of **2–9** was examined. Upon stimulation with the LPA analogues, the increase in Ca^{2+}



Scheme 1. Design of LPA analogues and selected synthesized compounds.

concentration in insect Sf9 cells that express LPA₃ receptors was assessed (Figure 1 a).^[11] Compounds **2–9** include all possible stereo- and regioisomers derived from the carbohydrate template. The isomer **2** had 5- to 10-fold higher agonist activity (EC₅₀ \approx 10 nM) relative to the commonly used agonist 1-oleoyl LPA (EC₅₀ \approx 50–100 nM). The other isomers **3–9** were 10- to 500-fold less potent than **2**. Specifically, the natural (2*R*)-oleoyl LPA analogue **2** was 50-fold more active than the non-natural 2*S* analogue **3** (EC₅₀ > 500 nM).^[12]

Next, we investigated the effect of individual recognition motifs on LPA₃ activation (compounds **10–14**) by fixing the stereochemistry to match that of the superior agonist **2**. Analogues **10**, **11**, and **12** did not show any agonist activity, which indicated the essential role of the oxygen atom in the six-membered ring as a hydrogen-bond acceptor.^[10] On the other hand, the activity of the thiophosphate analogue **13** was approximately 100- to 500-fold higher (EC₅₀ \approx 0.5 nM) than 1-oleoyl LPA. Thus, **13** is one of the most potent LPA₃ agonists reported to date.^[13] Moreover, **13** had only very weak agonist activity for LPA₁ (500- to 1000-fold weaker than 1-oleoyl LPA, Figure 1 b), and no agonist activity for LPA₂.^[10,14] Thus, **13** is a highly potent LPA₃-selective agonist that can be used as a biological tool.^[15]

We next targeted the discovery of LPA₁-selective agonists. An LPA₁-selective agonist would be highly desirable, because of the importance of this receptor. No agonist has been reported that can selectively activate the LPA₁ receptor in the

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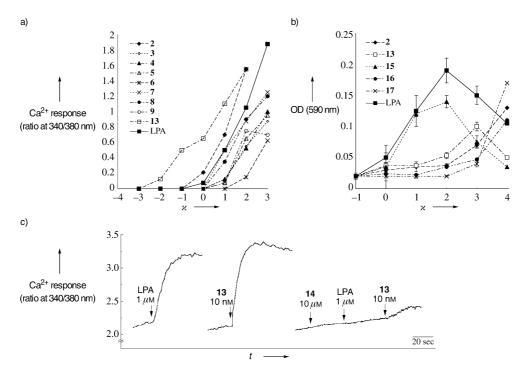


Figure 1. a) LPA₃-agonist activities. Assays were performed at least three times; representative data are shown. The change in the intracellular concentration of Ca^{2+} ions (Ca^{2+} response; y axis) upon activation by the agonists was determined by the emission ratio of Fura-2 AM fluorescence at an excitation wavelength of 340 nm (Ca^{2+} -bound form) and 380 nm (Ca^{2+} -free form); x = dose ($10^x nM$). b) LPA₁-agonist activities. The numbers of migrated cells upon activation by the agonists was determined by measuring the optical density (OD) at 590 nm (y axis). c) LPA₃-antagonist activity of **14**.

presence of the LPA₂ receptor. As compounds **2–14** had no significant agonist activity for LPA₁, we attempted to introduce a substituent on the six-membered ring to adjust the relative position of the recognition motifs for LPA₁ through fine-tuning of the ring conformation. Thus, we synthesized **15–17** and found that **15** had equivalent or stronger potency as an agonist for LPA₁ relative to 1-oleoyl LPA (Figure 1b).^[14,16] As **15** did not activate LPA₂,^[10] it is the first compound that can be used to distinguish between LPA₁-and LPA₂-agonist activity.^[15,17]

The subtype-selective agonist activity of 2, 13 (LPA₃selective), and 15 (LPA₁-selective) might be partly rationalized based on the hypothesis that a specific LPA receptor distinguishes a specific three-dimensional arrangement of the recognition motifs. To test this hypothesis, we determined the ring conformation in a solution state by using NMR techniques. All NOE data and coupling constant values suggested that 2 exists in a skewed-boat conformation with both the phosphate and oleoyl groups in pseudoequatorial positions (Figure 2a). On the other hand, 15 exists in a chair conformation with the methyl and oleoyl groups in equatorial positions and the phosphate group in an axial position (Figure 2b).^[18] The observed recognition-motif arrangements in 2 and 15 might correspond to the active binding structures of flexible 2-oleoyl LPA to LPA1 and LPA3, respectively, and the arrangements might be recognized selectively by each receptor.

Finally, we found that the analogue **14**, which contains a sulfate instead of a phosphate group, inhibited LPA₃ (Figure 1 c).^[10,19] No response was observed upon activation of

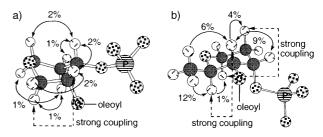


Figure 2. Observed conformations of 2 (a) and 15 (b) in solution in CD₃OD.

LPA₃ with 1-oleoyl LPA after pretreatment with **14** (10 μ M), and only weak activation was observed even with the highly potent agonist **13**. Moreover, **14** did not antagonize the 1oleoyl LPA induced migration of LPA₁-expressing cells (MDA, PC3, and 203g), nor the 1-oleoyl LPA-induced mobilization of Ca²⁺ ions in LPA₂-expressing cells (HT29). Thus, the antagonist activity of **14** is LPA₃-selective. Although the inhibitory activity was not very strong, **14** can be used as a lead LPA₃-selective antagonist for further structural optimization.

In conclusion, potent and subtype-selective agonists (2, 13, and 15) for LPA₁ and LPA₃ were developed by using carbohydrates as a core structure. The basic concept for ligand discovery was the selective extraction of active three-dimensional recognition-motif arrangements from conformationally flexible 2-oleoyl LPA. The concept allowed the discovery of a lead compound 14 for subtype-selective antagonists. To our knowledge, this is the first example of

receptor subtype-selective recognition by an array of small molecules by changing the relative three-dimensional arrangement of pharmacophores attached to a carbohydrate core. These compounds can be synthesized on a gram scale, and are stable for at least several months at -20 °C. Physiological studies on these subtype-selective agonists, as well as studies toward the development of potent and selective LPA antagonists are in progress.

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- For reviews, see: a) T. Hla, M. Lee, N. Ancellin, J. H. Paik, M. J. Kluk, *Science* 2001, 294, 1875; b) W. H. Moolenaar, O. Kranenburg, F. R. Postma, G. Zondag, *Curr. Opin. Cell Biol.* 1997, 9, 168.
- [2] For a recent review on LPA analogues, see: K. R. Lynch, T. L. Macdonald, *Biochim. Biophys. Acta* 2002, 1582, 289.
- [3] LPA exists as an equilibrium mixture of 1-acyl and 2-acyl LPA through intramolecular acyl migration: J. Chevallier, N. Sakai, F. Robert, T. Kobayashi, J. Gruenberg, S. Matile, *Org. Lett.* 2000, 2, 1859.
- [4] Several approaches were developed to generate stable LPA analogues by preventing acyl migration; the use of an amide:
 a) T. Sugiura, A. Tokumura, L. Gregory, T. Noguchi, S. T. Weintraub, D. J. Hanahan, Arch. Biochem. Biophys. 1994, 311, 358; the use of fluorinated analogues: b) Y. Xu, G. D. Prestwich, J. Org. Chem. 2002, 67, 7158; c) Y. Xu, L. Qian, G. D. Prestwich, J. Org. Chem. 2003, 68, 5320; d) Y. Xu, L. Qian, G. D. Prestwich, Org. Lett. 2003, 5, 2267; the use of an ether: e) Y. Hasegawa, J. R. Erickson, G. J. Goddard, S. Yu, S. Liu, K. W. Cheng, A. Eder, K. Bandoh, J. Aoki, R. Jarosz, A. D. Schrier, K. R. Lynch, G. B. Mills, X. Fang, J. Biol. Chem. 2003, 278, 11962; f) L. Qian, Y. Xu, H. Arai, J. Aoki, T. M. McIntyre, G. D. Prestwich, Org. Lett. 2003, 5, 4685.
- [5] D. W. Hopper, S. P. Ragan, S. B. Hooks, K. R. Lynch, T. L. Macdonald, J. Med. Chem. 1999, 42, 963.
- [6] a) C. E. Heise, W. L. Santos, A. M. Schreihofer, B. H. Heasley, Y. V. Mukhin, T. L. Macdonald, K. R. Lynch, *Mol. Pharmacol.* 2001, 60, 1173; b) D. J. Fisher, N. Nusser, T. Virag, K. Yokoyama, D. Wang, D. L. Baker, D. Baustista, A. L. Parrill, G. Tigyi, *Mol. Pharmacol.* 2001, 60, 776; and reference [4a].
- [7] Analogue design on the basis of a similar idea was attempted with aromatic compounds as scaffolds (reference [5]). Those analogues, however, had only weak (two orders of magnitude weaker) agonist activity compared to 1-oleoyl LPA. The lack of a three-dimensional concept in the molecular design might be a reason for the weak activity.
- [8] For computational modeling of the binding structure of LPA to LPA-receptor subtypes, see: a) D. Wang, Z. Lorincz, D. Bautista, K. Liliom, G. Tigyi, A. L. Parrill, *J. Biol. Chem.* 2001, 276, 49213;
 b) V. M. Sardar, D. Bautista, D. J. Fischer, K. Yokoyama, N. Nusser, T. Virag, D. Wang, D. L. Baker, G. Tigyi, A. L. Parrill,

Biochim. Biophys. Acta **2002**, *1582*, 309; results of these studies support our hypothesis.

- [9] For leading references on the use of carbohydrates as a scaffold for pharmacophores, see: a) R. Hirschmann, K. C. Nicolaou, S. Pietranico, J. Salvino, E. M. Leahy, P. A. Sprengeler, G. Furst, A. B. Smith III, C. D. Strader, M. A. Cascieri, M. R. Candelore, C. Donaldson, W. Vale, L. Maechler, J. Am. Chem. Soc. 1992, 114, 9217; b) R. Hirschmann, K. C. Nicolaou, S. Pietranico, E. M. Leahy, J. Salvino, B. Arison, M. A. Cichy, P. G. Spoors, W. C. Shakespeare, P. A. Sprengeler, P. Hamley, A. B. Smith III, T. Reisine, K. Raynor, L. Maechler, C. Donaldson, W. Vale, R. M. Freidinger, M. R. Cascieri, C. D. Strader, J. Am. Chem. Soc. 1993, 115, 12550; for recent examples, see: c) E. Locardi, M. Stöckle, S. Gruner, H. Kessler, J. Am. Chem. Soc. 2001, 123, 8189; d) M. Ghosh, R. G. Dulina, R. Kakarla, M. J. Sofia, J. Org. Chem. 2000, 65, 8387; e) T. Wunberg, C. Kallus, T. Opatz, S. Henke, W. Schmidt, H. Kunz, Angew. Chem. 1998, 110, 2620-2622; Angew. Chem. Int. Ed. 1998, 37, 2503; for a recent review, see: f) G. T. Le, G. Abbenante, B. Becker, M. Grathwohl, J. Halliday, G. Tometzki, J. Zuegg, W. Meutermans, Drug Discovery Today 2003, 8, 701.
- [10] For synthetic procedures and detailed assay results for these compounds, see Supporting Information.
- [11] K. Bandoh, J. Aoki, H. Hosono, S. Kobayashi, T. Kobayashi, K. Murakami-Murofushi, M. Tsujimoto, H. Arai, K. Inoue, *J. Biol. Chem.* **1999**, 274, 27776. In brief, Sf9 insect cells were transfected with cDNA of cloned human LPA receptors, and the increase in intracellular Ca²⁺ concentration upon activation by the agonists was quantified by using a Ca²⁺-specific fluorescent probe (Fura-2). No increase in Ca²⁺ concentration was observed in wild-type Sf9 cells in response to 1-oleoyl LPA or these agonists, thus eliminating the possibility that unknown receptors are affected.
- [12] For stereochemical properties of LPA-receptor activation, see: a) K. Yokoyama, D. L. Baker, T. Virag, K. Liliom, H.-S. Byun, G. Tigyi, R. Bittman, *Biochim. Biophys. Acta* 2002, *1582*, 295; b) L. Qian, Y. Xu, Y. Hasegawa, J. Aoki, G. B. Mills, G. D. Prestwich, J. Med. Chem. 2003, 46, 5575.
- [13] The analogue of Prestwich and co-workers is the only example with this level of activity (see reference [4c]). Compound 13 also had high potency (100- to 500-fold relative to 1-oleoyl LPA) in Ca²⁺-mobilization assays with several cells that express LPA₃.
- [14] The LPA₁-agonist activity was assessed by determining MDAcell migratory activity: M. Umezu-Goto, Y. Kishi, A. Taira, K. Hama, N. Dohmae, K. Takio, T. Yamori, G. B. Mills, K. Inoue, J. Aoki, H. Arai, *J. Cell Biol.* **2002**, *158*, 227. The LPA₂-agonist activity was assessed based on Ca²⁺-releasing activity from Sf9 cells transfected with LPA₂ (reference [11]).
- [15] Physiological studies with these analogues are ongoing.
- [16] A cell-migration assay with PC3 cells indicated that 15 is 10 times more potent than 1-oleoyl LPA as an LPA₁ agonist; see Supporting Information.
- [17] Compound **15** activates the LPA₃ receptor with comparable efficacy to 1-oleoyl LPA; see Supporting Information.
- [18] The ring in compound 16 should occupy the same conformation as that in 15; however, 16 might not fit into the binding pocket of LPA₁ as a result of the bulky substituent.
- [19] Only 14 showed inhibitory activity among compounds 2–17.