



Pergamon

Bioorganic & Medicinal Chemistry Letters 12 (2002) 1327–1330

BIOORGANIC &
MEDICINAL
CHEMISTRY
LETTERS

Design and Synthesis of a Piperazinylalkylisoxazole Library for Subtype Selective Dopamine Receptor Ligands

Mi Young Cha,^a Byung Chul Choi,^a Kyung Ho Kang,^b Ae Nim Pae,^a
Kung Il Choi,^a Yong Seo Cho,^a Hun Yeong Koh,^{a,*} Hee-Yoon Lee,^{b,*}
Daeyoung Jung^c and Jae Yang Kong^c

^aBiochemicals Research Center, Korea Institute of Science and Technology, Cheongryang, Seoul 130-650, South Korea

^bCMDS, Department of Chemistry (BK21), Korea Advanced Institute of Science and Technology, Daejeon 305-701, South Korea

^cPharmaceutical Screening Research Team, Korea Research Institute of Chemical Technology, Daejeon 305-600, South Korea

Received 22 February 2002; accepted 23 March 2002

Abstract—A piperazinylbutylisoxazole library was designed, synthesized and screened for the binding affinities to dopamine D₂, D₃, and D₄ receptors. Several ligands were identified to possess high binding affinity and selectivity for the D₃ and D₄ receptors over the D₂ receptor. Compounds **6s** and **6t** showed K_i values of 2.6 nM and 3.9 nM for the D₃ receptor with 46- and 50-fold selectivity over the D₂ receptor, respectively. © 2002 Elsevier Science Ltd. All rights reserved.

Introduction

Recently we have reported¹ a simple and efficient way of constructing libraries of heterocycles focused on the potential GPCR (G-Protein Coupled Receptor) binding capability.² In a classical pharmacology two types of dopamine receptors acting through G-proteins had been defined: the D₁ receptor which mediates the activation of adenylyl cyclase³ and the D₂ receptor which mediates its inhibition.⁴ Today at least five dopamine receptors were identified through molecular pharmacology and they were grouped into two subfamilies, the D₁-like (D₁⁵ and D₅⁶) and the D₂-like group (D₂,⁷ D₃⁸ and D₄⁹), based on amino acid homology and similar pharmacologic properties. The D₂-like receptors show high affinities for the most of the drugs used to treat schizophrenia and Parkinson's disease. Classical dopamine D₂ antagonists are effective in treating schizophrenia. However, they cause severe extrapyramidal side-effects¹⁰ that are thought to result from the blockade of D₂ receptors in the striatum of brain. The distribution of the D₃ and D₄ receptors in the limbic areas of brain suggests that these receptors may be particularly an attractive target for the design of potent and selective antipsychotic drugs without causing extrapyramidal side effects.¹¹

*Corresponding author. Tel.: +82-42-869-2835; fax: +82-42-869-8370; e-mail: leehy@kaist.ac.kr

Design of the Library

Based on SAR (structure–activity relationships) analysis of ligands for various GPCR's, especially dopamine D₃ and D₄ receptors antagonists and privileged structures for GPCRs,¹² the major aspects of the structural design to generate potent and selective ligands were divided into three parts: part **A** containing a biogenic amine, part **B** with alkyl chain as a linker and part **C** as electrostatically similar groups to carbonyl group or aromatic group at Q. Part **A** generally includes heterocyclic rings such as piperazine, piperidine or pyrrolidine. Part **B** serves as a linker between the biogenic amine and the hydrophobic aromatic group will be alkyl chain with varying length or conformationally rigid carbocyclic group. The chain length presumably reflects relative position of aromatic groups and the biogenic amine in spatial arrangement for binding to receptors, and could impose selectivity among receptors. The Q position of part **C** will both rigidify the part of the linker chain and be used as alternatives to bioisosteres such as amide,¹³ carbonyl,¹⁴ oxygen,¹⁵ secondary alcohol,¹⁶ and heterocycles.¹⁷ The substituents, S₁ and S₂ occupy the hydrophobic binding sites for aromatic residues. In practice, we selected a scaffold with an arylpiperazine and an isoxazole connected by a linker for the library. We designed a synthetic scheme of piperazinylalkylisoxazole library by combination of diverse arylpiperazines with

the substituted phenylisoxazole aldehydes where $n = 3$ or $n = 4$. (Fig. 1).

Chemistry

The synthetic strategy for obtaining a series of piperazinylalkylisoxazoles was adopted from the solution phase combinatorial synthesis,^{1,18} where the amine functionality in the scaffold provides particularly rapid and easy purification of products. In practice, purification was achieved by forming HCl salt of target molecules without using chromatographic purification in the last step.

The preparation of starting materials was outlined in Scheme 1. The phenylalkylisoxazole aldehydes were prepared from the substituted phenylaldehydes (**1**). Reaction of hydroxylamine HCl with aldehydes **1** in ethanolic aqueous solution (EtOH:H₂O=2/1) gave the corresponding oximes in 90–100% yields. Oximes **2** were treated with NCS(N-chlorosuccinimide) and a catalytic amount of pyridine in THF at 60 °C under nitrogen atmosphere. The reaction mixtures were stirred for 30 min at room temperature and then 5-hexyn-1-ol (for $n = 4$) was added slowly. The reaction mixtures were treated with Et₃N in THF over 10 min. After executing the aqueous workup, the alcohols **3** were obtained in 55–80% yields. Reaction of alcohols **3** with PCC/SiO₂ in CH₂Cl₂ gave aldehydes **4** in 45–85% yields (Scheme 1).

Construction of solution phase combinatorial library was described in Scheme 2. The combinatorial synthesis was accomplished by the reductive amination of the prepared aldehydes **4** with commercially available phenylpiperazine

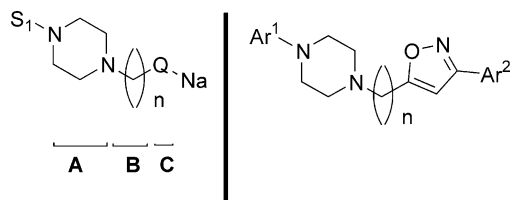
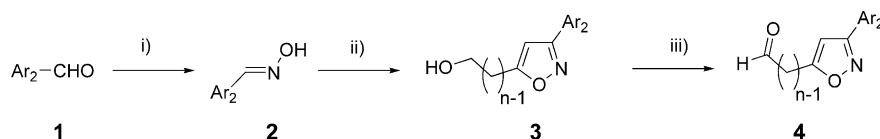
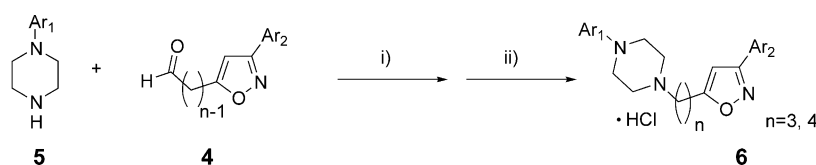


Figure 1.



Scheme 1. Reagents and reaction conditions: (i) NH₂OH·HCl, Na₂CO₃, 60 °C, 1 h, EtOH/H₂O (2/1), 90–100%; (ii) pyridine (cat.), NCS, 60 °C, 0.5 h, THF/4-pentyn-1-ol (for $n = 3$) or 5-hexyn-1-ol (for $n = 4$), Et₃N, 50 °C–rt, 2 h, 55–80%; (iii) PCC, silica gel, CH₂Cl₂, 5–12 h, 45–85%.



Scheme 2. Reagents and reaction conditions: (i) NaBH(OAc)₃ (3 equiv), molecular sieves (3 beads), CH₂Cl₂, rt, 5–12 h; (ii) After executing aqueous workup, the reaction mixture in 2 mL of diethyl ether was treated with ethereal HCl. The precipitant was washed with diethyl ether and dried, 85–100%.

derivatives **5** using NaBH(OAc)₃.¹⁹ Solutions of amines **5** and aldehydes **4** in CH₂Cl₂ was added NaBH(OAc)₃ (3 equiv) and molecular sieve (3 beads). The reaction mixtures were stirred for 5–12 h at room temperature. After executing of the aqueous workup, reaction mixtures dissolved in 2 mL of diethyl ether followed by treatment with 1M HCl solution in diethyl ether gave HCl salts of the products **6** as solid. The solvent was filtered off and the precipitant was washed with diethyl ether, and dried in vacuo. All the compounds were obtained in good yields (85–100%) and high purities ranging from 85–96%. Purity and identity of the products were confirmed through ¹H NMR, HPLC and HRMS after the solid products were converted into the corresponding free bases. A small library with well-characterized 300 compounds was constructed by using reductive amination, precipitation sequence (Scheme 2).

Results and Discussion

The generated phenylpiperazinylalkylisoxazole analogues were evaluated in vitro for dopamine D₂–D₄ receptors binding affinity by measuring their ability to displace radioligands ([³H]spiperone for D₂ and D₄, [³H]YM-09151–2 for D₃) from the cloned human dopamine receptors D_{2long}, D₃ and D_{4.2} which were expressed in CHO cells, respectively. The affinity and selectivity of this class of compounds for the dopamine receptors were largely dependent on substitution pattern at *o*-position on phenylpiperazinyl group and the length of the alkyl chain linker. The extension of alkyl chain from $n = 3$ to $n = 4$ affected the binding affinity greatly since the affinities of compounds with a three atom tether were very low and showed no selectivity among receptors in the primary screening. Table 1 shows the binding data of the selected compounds that showed good activity and selectivity among dopamine D₂, D₃, and D₄ receptors. With a methoxy group at *o*-position of phenyl group of Ar₁(Ar₁3), the binding affinity to dopamine D₃ receptor increased (compounds **6k–6q**). Extension of methoxy group at *o*-position of phenyl group (Ar₁4) to ethoxy group (compounds **6r–6u**) gave the further improvement of affinity for the D₃ receptor (2.6–19 nM). On the other hand, with a methyl group (compounds **6h–**

Table 1. Binding affinity (K_i , nM)²⁰ for n = 4

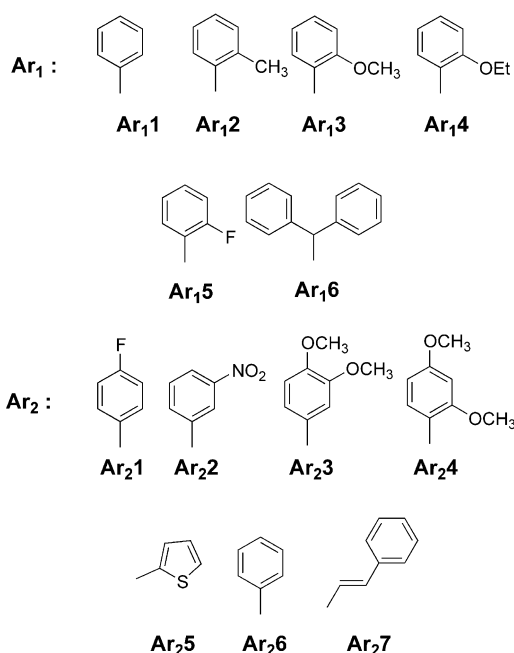
Entry ^a	Substituent		Receptor			Selectivity	
	Ar ₁	Ar ₂	D ₂	D ₃	D ₄	D ₂ /D ₃	D ₄ /D ₃
6a	Ar ₁ 1	Ar ₂ 1	2530	12	96	211	8
6b	Ar ₁ 1	Ar ₂ 2	1470	60	1697	25	28
6c	Ar ₁ 1	Ar ₂ 3	3388	24	774	141	32
6d	Ar ₁ 1	Ar ₂ 4	2196	16	76	137	5
6e	Ar ₁ 1	Ar ₂ 5	4136	15	117	276	8
6f	Ar ₁ 1	Ar ₂ 6	3366	18	541	187	30
6g	Ar ₁ 1	Ar ₂ 7	9724	27	346	360	13
6h	Ar ₁ 2	Ar ₂ 3	1401	38	413	37	11
6i	Ar ₁ 2	Ar ₂ 5	2108	163	465	13	3
6j	Ar ₁ 2	Ar ₂ 6	6512	292	683	22	2
6k	Ar ₁ 3	Ar ₂ 1	321	23	33	14	1.4
6l	Ar ₁ 3	Ar ₂ 2	295	6	50	49	8
6m	Ar ₁ 3	Ar ₂ 3	418	5.2	257	80	49
6n	Ar ₁ 3	Ar ₂ 4	440	6.5	34	68	5
6o	Ar ₁ 3	Ar ₂ 5	726	4.7	21	154	4
6p	Ar ₁ 3	Ar ₂ 6	1727	9.5	198	181	21
6q	Ar ₁ 3	Ar ₂ 7	964	15	88	64	6
6r	Ar ₁ 4	Ar ₂ 2	95	19	51	5	3
6s	Ar ₁ 4	Ar ₂ 3	119	2.6	28	46	11
6t	Ar ₁ 4	Ar ₂ 5	194	3.9	12	50	3
6u	Ar ₁ 4	Ar ₂ 6	341	9	34	38	4
6v	Ar ₁ 5	Ar ₂ 3	5148	20	233	257	12
6w	Ar ₁ 5	Ar ₂ 7	585	10	58	59	6
6x	Ar ₁ 6	Ar ₂ 1	1461	30	1346	49	45
6y	Ar ₁ 6	Ar ₂ 2	1522	19	2314	80	121
6z	Ar ₁ 6	Ar ₂ 3	2618	20	2438	130	122
6aa	Ar ₁ 6	Ar ₂ 4	3828	16	1118	239	70
6bb	Ar ₁ 6	Ar ₂ 5	620	52	2145	12	41
6cc	Ar ₁ 6	Ar ₂ 6	486	15	2243	32	150
6dd	Ar ₁ 6	Ar ₂ 7	491	40	603	12	15
Haloperidol			18	25	42	0.72	1.7

^aAll compounds gave satisfactory analytical and/or mass spectral data.

6j) at *o*-position of phenylpiperazine, the binding affinities decreased (38–292 nM). *o*-Fluorophenylpiperazine analogues (compounds **6v** and **6w**) also gave good binding affinities (10 and 20 nM). Among phenylpiperazine analogues, compounds **6s** and **6t** showed high binding affinity with K_i values of 2.6 and 3.9 nM, respectively. Though diphenylmethylpiperazine analogues (compounds **6x–6dd**) showed rather lower binding affinity (15–52 nM) than arylpiperazines for the D₃, compounds **6x** to **6dd** exhibited high selectivity for dopamine D₃ receptor over the D₂ and D₄ receptors regardless of Ar₂. For compounds **6z** and **6aa**, 130- to 239-fold higher selectivity of the D₃ receptor over the D₂ receptor was observed. Compounds **6e**, **6g** and **6v** showed the high selectivity for the D₃ over the D₂ receptor (276-, 360- and 257-fold, respectively). Compound **6x–6aa** showed a good selectivity for the D₃ over the D₄ receptor. Generally, high affinity appeared to be guaranteed by 3,4-dimethoxyphenyl- (Ar₂3), 2,4-dimethoxyphenyl- (Ar₂4) and 2-thienyl- (Ar₂5) groups and high selectivity was exerted either by *o*-alcoxy group or an extra phenyl group (Table 1; Fig. 2).

Summary

A small library of piperazinylbutyl(propyl)isoxazoles designed as GPCR ligands was constructed through solution phase combinatorial synthesis. The variation imposed on the two aromatic groups and the length of

**Figure 2.**

the spacer that connected two parts generated an array of activity and selectivity for the dopamine receptors. Also, the isoxazole ring as an alternative of bioisosteres play a important role in binding with receptors. With the chain length of four-carbon tether, most of the compounds showed good binding affinity and selectivity at the desirable target receptors, the D₃ and D₄ receptors over D₂ receptor. Compounds **6s** and **6t** showed K_i values of 2.6 and 3.9 nM for the D₃ receptor with 46-fold and 50-fold selectivity over the D₂ receptor, respectively. Further QSAR (Quantitative Structure and Activity Relationships) studies of these series are in progress.

Acknowledgements

This work was supported by the Korea Ministry of Science and Technology.

References and Notes

- Kang, K. H.; Pae, A. N.; Choi, K. I.; Cho, Y. S.; Chung, B. Y.; Lee, J. E.; Jung, S. H.; Koh, H. Y.; Lee, H.-Y. *Tetrahedron Lett.* **2001**, *42*, 1057.
- Watson, S.; Arkininstall, S. *The G-Protein Linked Receptor*; Academic Press: San Diego, 1994.
- Kebabian, J.; Greengard, P. *Science* **1971**, *174*, 1346.
- Onali, P.; Plianas, M. C.; Gessa, G. L. *Mol. Pharmacol.* **1985**, *28*, 138.
- Deary, J. R.; Gingrich, J. A.; Falardeau, R. T.; Fremeau, R. T.; Bates, M. D.; Caron, M. *Nature* **1990**, *347*, 72.
- Sunahara, R. K.; Guan, H.-C.; O'Dowd, B. F.; Seeman, P.; Laurier, L. G.; George, S. R.; Torchia, J.; Van Tol, H. M.; Niznik, H. B. *Nature* **1991**, *350*, 614.
- Grandy, D. K.; Marchionni, M. A.; Makam, H.; Stofko, R. E.; Alfano, M.; Frothingham, L.; Fischer, J. B.; Burke-Howie, K. J.; Bunzow, J. R.; Server, A. C.; Civelli, O. *Proc. Natl. Acad. Sci. U.S.A.* **1989**, *86*, 9762.

8. Sokoloff, P.; Giros, B.; Martres, M.-P.; Bouthenet, M.-L.; Schwartz, J.-C. *Nature* **1990**, *347*, 146.
9. Van Tol, H. M.; Bunzow, J. R.; Guan, H. C.; Sunahara, R. K.; Seeman, P.; Niznik, H. B.; Civelli, O. *Nature* **1991**, *30*, 610.
10. Baldessarini, R. J.; Tarsy, D. *Ann. Rev. Neurosci.* **1980**, *3*, 23.
11. Haadsma-Svensson, S. R.; Cleek, K. A.; Dinh, D. M.; Duncan, J. N.; Haber, C. L.; Huff, R. M.; Lajiness, M. E.; Nichols, N. F.; Smith, M. W.; Svensson, K. A.; Zaya, M. J.; Carlsson, A.; Lin, C.-H. *J. Med. Chem.* **2001**, *44*, 4716 and references therein.
12. (a) Rowley, M.; Collins, I.; Broughton, H. B.; Davey, W. B.; Baker, R.; Emms, F.; Marwood, R.; Patel, S.; Patel, S.; Ragan, C. I.; Freedman, S. B.; Ball, R.; Leeson, P. D. *J. Med. Chem.* **1997**, *40*, 2374. (b) Patchett, A. A.; Nargund, R. P. *Ann. Rep. Med. Chem.* **2000**, *35*, 289.
13. (a) Austin, N. E.; Avenell, K. Y.; Boyfield, I.; Branch, C. L.; Hadley, M. S.; Jeffrey, P.; Johnson, C. N.; Macdonald, G. J.; Nash, D. J.; Riley, G. J.; Smith, A. B.; Stemp, G.; Thewlis, K. M.; Vong, A. K. K.; Wood, M. D. *Bioorg. Med. Chem. Lett.* **2001**, *11*, 685. (b) Stemp, G.; Ashmeade, T.; Branch, C. L.; Hadley, M. S.; Hunter, A. J.; Johnson, C. N.; Nash, D. J.; Thewlis, K. M.; Vong, A. K. K.; Austin, N. E.; Jeffrey, P.; Avenell, K. Y.; Boyfield, I.; Hagan, J. J.; Middelemis, D. N.; Reavill, C.; Riley, G. J.; Routledge, C.; Wood, M. *J. Med. Chem.* **2000**, *43*, 1878.
14. De Oliveira, I. R.; De Sena, E. P.; Pereira, E. L.; Miranda, A. M. A.; De Pliveira, N. F.; Ribeiro, M. G.; De Castro-Silva, E.; Dardennes, R. M.; Samuel-Lajeunesse, B.; Marcilio, C. *J. Clin. Pharm. Ther.* **1996**, *21*, 229.
15. (a) Oshiro, Y.; Sato, S.; Kurahashi, N.; Tanaka, T.; Kikuchi, T.; Tottori, K.; Uwahodo, Y.; Nishi, T. *J. Med. Chem.* **1998**, *41*, 658. (b) Mewshaw, R. E.; Husvands, M.; Gildersleeve, E. S.; Webb, M. B.; Shi, X.; Mazandarani, H.; Cockett, M. I.; Ochalski, R.; Brennan, J. A.; Abou-Gharbia, M.; Marquis, K.; McGaughey, G. B.; Coupet, J.; Andree, T. H. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 295. (c) Dutta, A. K.; Coffey, L. L.; Reith, M. E. A. *J. Med. Chem.* **1998**, *41*, 699.
16. Jaen, J. C.; Caprathe, B. W.; Pugsley, T. A.; Wise, L. D.; Akunne, H. *J. Med. Chem.* **1993**, *36*, 3929.
17. (a) Löber, S.; Hübner, H.; Utz, W.; Gmeiner, P. *J. Med. Chem.* **2001**, *44*, 2691. (b) Einsiedel, J.; Thomas, C.; Hübner, H.; Gmeiner, P. *Bioorg. Med. Chem. Lett.* **2000**, *10*, 2041. (c) Thurkauf, A.; Tuan, J.; Chen, X.; He, S. H.; Wasley, J. W. F.; Hutchison, A.; Woodruff, K. H.; Meade, R.; Hoffman, D. C.; Donovan, H.; Jones-Hertzog, D. K. *J. Med. Chem.* **1997**, *40*, 1. (d) Gazi, L.; Sommer, B.; Nozulak, J.; Schoeffter, P. *Eur. J. Pharmacol.* **1999**, *372*, R9. (e) Rowley, M.; Bristow, L. J.; Huston, P. H. *J. Med. Chem.* **2001**, *44*, 477. (f) Gazi, L.; Bobirnac, I.; Danzeisen, M.; Schüpbach, E.; Langenegger, D.; Sommer, B.; Hoyer, D.; Tricklebank, M.; Schoeffter, P. *Br. J. Pharmacol.* **1999**, *128*, 613.
18. Baldino, C. M. *J. Comb. Chem.* **2000**, *2*, 89.
19. Abdel-Magid, A. F.; Carson, K. G.; Harris, B. D.; Maryanoff, C. A.; Shah, R. D. *J. Org. Chem.* **1996**, *61*, 3849.
20. Assay protocol: Sf-9 membranes expressing either dopamine hD_{2L}, dopamine rD₃ or hD_{4,2} receptors were purchased from PerkinElmer Life Sciences (Boston, MA). Radioligands used were [³H]Spiperone (D₂ and D₃ dopamine binding assays, 1 nM), and [³H] YM-09151-2 (D₄ dopamine binding assays, 0.06 nM). [³H]Spiperone and [³H] YM-09151-2 bindings were performed by the protocol provided by supplier of Sf-9 membranes for both hD_{2L} and rD₃, and hD_{4,2} receptors, respectively. Briefly, the buffer used in hD_{2L} or rD₃ receptor binding assay was 50 mM Tris-HCl (pH 7.4), 10 mM MgCl₂, 1 mM EDTA, or 50 mM Tris-HCl (pH 7.4), 5 mM MgCl₂, 5 mM EDTA, 5 mM KCl, 1.5 mM CaCl₂, 120 mM NaCl, respectively. In [³H] YM-09151-2 receptor binding assays, the buffer containing 50 mM Tris-HCl (pH 7.4), 5 mM MgCl₂, 5 mM EDTA, 5 mM KCl and 1.5 mM CaCl₂ was used. Non-specific binding was determined with haloperidol (10 μM) and clozapine (10 μM) for D₂ and D₃, and D₄ receptors, respectively. Competition binding studies were carried out with 8 concentrations of the test compound run in duplicate tubes, and isotherms from three assays were calculated by computerized nonlinear regression analysis (GraphPad Prism Program, San Diego, CA) to yield inhibition constant (K_i) values.