167. Catalytic Effect of Tetrabutylammonium Fluoride in the Preparation of Secoribo-nucleosides

by Gholam H. Hakimelahi* and Ali Khalafi-Nezhad

Chemistry Department, Shiraz University, Shiraz, Iran

(31.III.89)

A general and rapid procedure is developed for the preparation of secoribo-nucleoside analogues. Bu_4NF has a marked effect on the condensation of guanine (5) with 2-(chloromethoxy)ethyl benzoate (2) or 1-halo-2-(chloromethoxy)ethanes 12 and 16. Condensation and deprotection of the acyl protecting group and nucleophilic displacement of the halogen atoms to yield 9-[(2-hydroxyethoxy)methyl] guanine (9) occurred in one step.

Recently, we have reported the exclusive preparation of N(9)-alkylated secoribonucleosides which are potential antiviral drugs by condensation of silylated derivatives of nucleobases with chloromethyl ethers using an equimolar amount of Bu_4NF [1] [2]. This condensation and the deprotection of an acyl-protecting group in the ether moiety yielding the secoribo-nucleoside analogues occurred in one step. In this paper, we wish to describe a general procedure for the same reaction but involving the use of a catalytic amount of Bu_4NF .

Our first approach involved the condensation of adenine (1) with 2-(chloromethoxy)ethyl benzoate (2; *Scheme 1*). Thus, when bis(trimethylsilyl)adenine was prepared from 1 and hexamethyldisilazan (HMDS) and condensed with 2 in THF using a small amount of Bu_4NF , a fairly rapid reaction occurred. The desired product 3 was isolated in 85% yield. Treatment of 3 with MeOH/NH₃ at 25° gave the adenine derivative 4 (95%), identical with an authentic sample [1].



Similarly, condensation of guanine (5) (as its tris(trimethylsilyl) derivative) with chloromethyl ether 6 in the presence of Bu_4NF as catalyst gave 7 (98%), and deprotection by catalytic transfer reduction [3] [4] afforded 8 (= BIOLF-62 [5]; 90%) (Scheme 2). Using ether 2, compound 9 (= Zovirax [6]; 92%) was prepared directly from 5, after silylation and condensation (3 h) using Bu_4NF as catalyst (Scheme 2). In the case of



Bz = COPh, $BzI = CH_2Ph$

adenine (1), hydrolysis of the ester function occurred, after 30 h, only when 1 equiv. of Bu_4NF was used for the condensation [1].

Considering that even dried Bu_4NF contains 1–2 mol-equiv. of H_2O , the one-step condensation and deprotection of the acyl group is plausible in the presence of an equimolar amount of Bu_4NF (1 \rightarrow 4) but rather surprising in the case of a catalytic amount of Bu_4NF (5 \rightarrow 9). A reasonable explication would be the involvement of the NH_2 function of the guanine moiety in the ester hydrolysis.

In an attempt to clarify the scope of the Bu_4NF -catalyzed condensation, the following experiments were carried out. Guanine (5), hypoxanthine (10), and 8-azaguanine (11) were silylated with HMDS and coupled with 1-bromo-2-(chloromethoxy)ethane (12) in refluxing THF to give the secoribo-nucleosides 9 (95%), 13 ca. 80%, and 14 (ca. 80%), after 3 h, by means of catalytic Bu_4NF (Scheme 3). Compound 9 was found to be identical with an authentic sample [6] [7] and was further characterized by preparing its (tert-butyl)dimethylsilyl derivative 15. Moreover, the ¹H-NMR spectrum of 9 clearly indicates an A_2B_2 pattern for the OCH₂CH₂O moiety while the corresponding signals of 13–15 (R¹CH₂CH₂O) exhibit an A_2X_2 pattern.



In another experiment, guanine (5) was silvlated and condensed to 1-chloro-2-(chloromethoxy)ethane (16) using Bu_4NF as catalyst (*Scheme 4*). The condensation proceeded smoothly giving a 1:1 mixture 9/17 in high yield after 4 h. However, by prolonging the reaction time to 30 h, 9 was found to be the exclusive product. When special care was taken to run the reaction in H₂O-free solvent under a stream of N₂, 9 and 17 were obtained in a 1:3 ratio in high yield after 4 h. This ratio did not change much, even after 30 h.



The reaction of adenine (1) with chloromethyl ethers 12 and 16 in the presence of Bu_4NF as catalyst gave the expected products 18 and 19 (*ca.* 95%), respectively, the latter being identical with an authentic sample [8]. Finally, treatment of the silylated derivatives of uracil (20) and thymine (21) with chloromethyl ethers 12 or 16 in the presence of a small amount of Bu_4NF gave the corresponding pyrimidine secoribo-nucleosides 22–25 (*ca.* 80–90%; Scheme 5).



When the reactions of *Schemes* 1-5 were conducted in the absence of the F⁻ ion or in the presence of Bu₄NCl, they failed to produce the desired compounds. Therefore, Bu₄NF has to be considered as a novel catalyst for the high-yield preparation of secoribonucleoside analogues.

This work was supported by the Shiraz University Research Council. We are grateful to *Radja Chemical and Pharmaceutical Company* for their financial support.

Experimental Part

General. See [8].

General Procedure for Condensation of Nucleobases 1, 5, 10, 11, 20, and 21 with Chlormethyl Ethers 2, 6, 12, and 16 Yielding 3, 7, 9, 13, 14, 17–19, and 22–25 (Data: Table). Representative Procedure (yields 80–98%): Adenine (1 13.5 g, 0.1 mol) and $(NH_4)_2SO_4$ (500 mg) were suspended in hexamethyldisilazane (HMDS, 500 ml) and refluxed for 24 h. The solvent was evaporated and the residue dissolved in THF (650 ml). Bu₄NF (1 g) was dried by azeotrope distillation in benzene (80 ml) and added dropwise within 10 min at boiling temp. after the volume of the benzene soln. had been reduced to 10 ml. Then, 2 (21.5 g, 0.1 mol) was added dropwise within 15 min at the same

'Ta'	ble.	Pro	perties	of	Nuci	leosid	e Anai	logues
------	------	-----	---------	----	------	--------	--------	--------

Compound	M.p. [°]	λ_{\max} [nm] (EtOH)	$R_{\rm f}({ m TLC})$
3	120	260	0.85 ^a)
4	150	259	0.69 ^a)
7	182	252, 270 (sh)	0.42 ^b)
8	> 285 (dec.)	253, 272 (sh)	0.44 ^c)
9	> 280 (dec.)	253, 273 (sh)	0.39 ^b)
13	160	250	0.90 ^a)
14	> 275 (dec.)	254, 274 (sh)	0.50 ^b)
15	220	252, 271 (sh)	0.88 ^b)
17	260	252, 271 (sh)	0.63^{b})
18	215	257.5	0.06^{d})
19	225	257	0.04^{d})
22	100	257	0.33 ^e)
23	110	264	0.30^{e})
24	foam	257	0.61°)
25	87	264	0.53 ^e)

^a) $CH_2Cl_2/MeOH 8:2.$

^b) AcOEt/MeOH 3:1.

^c) i-PrOH/NH₄OH/H₂O 7:1:2.

d) AcOEt.

e) AcOEt/Et₂O 1:1.

temp. After 3 h, the soln. was diluted with AcOEt (600 ml) and H₂O (400 ml). The org. layer was separated, washed with H₂O (2 × 300 ml), dried (Na₂SO₄), filtered, and evaporated. The residue was chromatographed on silica gel, and the impurities were eluted with CH₂Cl₂. Elution with AcOEt/acetone 2:1 afforded 2-[(adenin-9-yl)-methoxy]ethyl benzoate (3; 85%). Data: Table. IR (KBr): 3400 (br.), 1725s, 1630s, 1585m, 1100s. ¹H-NMR ((D₆)DMSO): 3.85 (br. t, CH₂O); 4.35 (br. t, CH₂OCO); 5.56 (s, OCH₂N); 7.25–7.85 (m, Ph, NH₂); 8.05 (s, H–C(2)); 8.25 (s, H–C(8)). Anal. calc. for C₁₅H₁₅N₅O₃ (313.21): C 57.51, H 4.79, N 22.36; found: C 57.50, H 4.80, N 22.38.

9-[(2-Hydroxyethoxy)methyl]guanine (9): From 5 and 2 (92%), from 5 and 12 (95%), and from 5 and 16 (40%, along with 17 (40%) after 4 h, and 90% after 30 h). IR (nujol): 3100-3520 (br.), 1700-1720s (br.), 1610s,

1630s, 1110s (br.). ¹H-NMR ((D_6)DMSO): 3.62–3.92 (br. *s*, OCH₂CH₂O); 5.35 (*s*, OCH₂N); 6.52 (br. *s*, OH, NH₂, exchange with D_2 O); 7.60 (br., NH, exchange with D_2 O); 7.81 (*s*, H–C(8)). Anal. calc. for C₈H₁₁N₅O₃ (225.21): C 42.66, H 4.88, N 31.11; found: C 42.56, H 5.01, N 31.22.

9-[(2-Chloroethoxy)methyl]guanine (17): ¹H-NMR ((D₆)DMSO): 3.43–4.02 (*m*, ClCH₂CH₂O); 5.45 (*s*, OCH₂N); 6.61 (br., NH₂, exchange with D₂O); 7.71 (br., NH, exchange with D₂O); 7.90 (*s*, H–C(8)). Anal. calc. for C₈H₁₀ClN₄O₂ (243.71): C 39.42, H 4.11, Cl 14.58, N 28.75; found: C 39.38, H 4.05, Cl 14.60, N 28.81.

 $9 - \{\{2-(Benzyloxy)-1-[(benzyloxy)methyl\}ethoxy\}methyl\}guanine (7): ^{1}H-NMR ((D_6)DMSO): 3.32-3.61 (br., 2 CH₂O); 3.90-4.19 (m, CHO); 4.31 (s, 2 PhCH₂O); 5.55 (s, OCH₂N); 6.45 (br., NH₂, exchange with D₂O); 7.25 (br. s, 2 Ph); 7.88 (br., NH, exchange with D₂O); 7.93 (s, H-C(8)). Anal. calc. for C₂₃H₂₅N₅O₄ (435.32): C 63.45, H 5.75, N 16.09; found: C 63.43, H 5.85, N 16.18.$

 $9-[(2-Bromoethoxy)methyl]hypoxanthine (13): {}^{1}H-NMR ((D_6)DMSO): 3.20-3.50 (m, CH_2Br); 3.65-3.96 (m, CH_2O); 5.81 (s, OCH_2N); 7.80 (br., NH, exchanged with D_2O); 8.62 (s, H-C(2)); 8.78 (s, H-C(8)). Anal. calc. for C_8H_9BrN_4O_2 (273.32): C 35.16, H 3.30, Br 29.30, N 20.51; found: C 35.20, H 3.41, Br 29.18, N 20.61.$

9-[(2-Bromoethoxy)methyl]-8-azaguanine (14): ¹H-NMR ((D₆)DMSO): 3.31–3.65 (*m*, CH₂Br); 3.70–4.08 (*m*, CH₂O); 5.83 (*s*, OCH₂N); 7.95 (br., NH, NH₂ exchange with D₂O). Anal. calc. for C₇H₉BrN₆O₂ (289.33): C 29.06, H 3.11, Br 27.68, N 29.06; found: C 29.26, H 3.18, Br 27.64, N 29.21.

1-[(2-Bromoethoxy)methyl]uracil(22): ¹H-NMR (CDCl₃): 3.22–3.60 (m, CH₂Br); 3.70–4.05 (m, CH₂O); 5.20 (s, OCH₂N); 5.75 (d, J = 8, H–C(5)); 7.31 (d, J = 8, H–C(6)); 10.33 (br. s, NH, exchange with D₂O). Anal. calc. for C₇H₉BrN₂O₃ (249.31): C 33.73, H 3.61, Br 32.13, N 11.24; found: C 33.85, H 3.60, Br 32.01, N 11.20.

l-[(2-Bromoethoxy)methyl]thymine (23): ¹H-NMR (CDCl₃): 1.91 (s, CH₃); 3.23–3.58 (m, CH₂Br); 3.69–4.01 (m, CH₂O); 5.21 (s, OCH₂N); 7.20 (s, H–C(6)); 10.43 (br. s, NH, exchange with D₂O). Anal. calc. for C₈H₁₁BrN₂O₃ (263.32): C 36.50, H 4.18, Br 30.42, N 10.65; found: C 36.61, H 4.19, Br 30.53, N 10.68.

 $I-[(2-Chloroethoxy)methyl]uracil (24): {}^{1}H-NMR (CDCl_3): 3.41-4.01 (m, ClCH_2CH_2O); 5.21 (s, OCH_2N); 5.74 (d, J = 8, H-C(5)); 5.32 (d, J = 8, H-C(6)); 10.34 (br. s, NH, exchange with D_2O). Anal. calc. for C₇H₉ClN₂O₃ (204.63): C 41.07, H 4.40, Cl 17.36, N 13.69; found: C 41.12, H 4.51, Cl 17.40, N 13.71.$

1-[(2-Chloroethoxy)methyl]thymine (25): ¹H-NMR (CDCl₃): 1.90 (s, CH₃): 3.50–4.11 (m, ClCH₂CH₂O); 5.22 (s, OCH₂N); 7.21 (s, H–C(6)); 9.99 (br. s, NH, exchange with D₂O). Anal. calc. for C₈H₁₁ClN₂O₃ (218.64): C 43.93, H 5.03, Cl 16.25, N 12.81; found: C 43.91, H 5.13, Cl 16.30, N 12.95.

11-Amino-6,7-dihydro-4H-[1,3,5] $oxadiazepino[3,4,5-cd]purin-8-ium Bromide (18): {}^{1}H-NMR ((D_6)DMSO): 3.79 (s, NCH_2CH_2O); 5.67 (s, OCH_2N); 7.68 (br. s, NH_2, exchange with D_2O); 8.30 (s, H-C(2)); 8.40 (s, H-C(8)). Anal. calc. for C₈H₁₀BrN₅O (272.32): C 35.29, H 3.67, Br 29.41, N 25.73; found: C 35.32, H 3.70, Br 29.31, N 25.90.$

19: Identical with the authentic sample [8].

9-[(2-Hydroxyethoxy)methyl]adenine (4): To a soln. of 3 (3.13 g, 0.01 mol) in MeOH (50 ml), 150 ml of sat. NH₃/MeOH were added. The flask was sealed and maintained at 25° for 24 h. The mixture was concentrated to 5 ml, and Et₂O (50 ml) was added to afford 4 as a white precipitate. Filtration gave 4 (95%), identical with an authentic sample [1]. Data: *Table*.

 $9 - \{[2-Hydroxy-1-(hydroxymethyl)ethoxy]methyl\}guanine (8)$. To a soln. of 7 (5.20 g, 0.012 mol) in refluxing EtOH (280 ml), cyclohexene (200 ml) and PdCl₂ (3.5 g) were added, and refluxing was continued for 5 h. The soln. was collected by filtration and the catalyst washed with hot DMF (5 ml) and 3% aq. NaHCO₃ soln. (10 ml). The filtrate and washings were evaporated, and the residue was dissolved in hot EtOH/H₂O (100 ml) and filtered. The solvent was evaporated and the residue crystallized from EtOH/H₂O 4:1 to afford 8 (90%), identical with an authentic sample [9]. Data: *Table*.

 $9-\{\{2-[(\text{tert-Butyl}) dimethylsilyloxy] ethoxy\} methyl\} guanine (15).$ To a soln. of 9 (2.25 g, 0.01 mol) in DMF (50 ml), imidazol (2.04 g, 0.03 mol) and (*t*-Bu)Me₂SiCl (3.20 g, 0.02 mol) were added, the mixture was stirred for 48 h at 25° and then poured into H₂O (100 ml). The aq. soln. was extracted with AcOEt, and the extract was dried (Na₂SO₄), filtered, and evaporated. The residue was applied to a column of silica gel and 15 (97%) was eluted with AcOEt/MeOH 9:1 as a white precipitate. Data: *Table*. ¹H-NMR ((D₆)DMSO): 0.12 (*s*, (CH₃)₂Si); 0.91 (*s*, (CH₃)₃C); 3.30–3.78 (*m*, CH₂O); 3.80–4.28 (*m*, CH₂OSi); 5.45 (*s*, OCH₂N); 6.70–7.0 (br., NH₂, NH, exchange with D₂O); 7.90 (*s*, H–C(8)).

REFERENCES

- [1] G.H. Hakimelahi, A. Khalafi-Nezhad, F. Mohanazadeh, submitted to Helv. Chim. Acta.
- [2] G.H. Hakimelahi, F. Mohanazadeh, A. Khalafi-Nezhad, M. Zakerinia, Med. J. I.R. Iran 1989, to be published.
- [3] E.A. Braude, R.P. Linstead, P.W. Mitchell, K.R.H. Woolridge, J. Chem. Soc. 1954, 3595; V.S. Rao, A.S. Perlin, Carbohydr. Res. 1980, 83, 175.
- [4] S. Hanessian, T. J. Liak, B. Vanasse, Synthesis 1981, 396.
- [5] K.K. Ogilvie, U.O. Cheriyan, B.K. Radatus, K.O. Smith, K.S. Galloway, W.L. Kennell, Can. J. Chem. 1982, 60, 3005.
- [6] H.J. Schaeffer, L. Beauchamp, P. de Miranda, G.B. Elion, D.J. Bauer, P. Collins, Nature (London) 1978, 272, 583.
- [7] R. T. Walker, E. de Clercq, F. Eckstein, Eds., 'Nucleoside Analogues', Plenum Press, New York, 1978; G. H. Hakimelahi, Iran. Pat. 23291, Sept. 6, 1986.
- [8] G. H. Hakimelahi, M. Zarrinehzad, A. A. Jarrahpour, H. Sharghi, Helv. Chim. Acta 1987, 70, 219.