Asymmetric Total Synthesis of Cladosporin and Isocladosporin

Huaiji Zheng,[†] Changgui Zhao,[†] Bowen Fang,[†] Peng Jing,[†] Juan Yang,[†] Xingang Xie,[†] and Xuegong She^{*,†,‡}

[†]State Key Laboratory of Applied Organic Chemistry, College of Chemistry and Chemical Engineering, Lanzhou University, Lanzhou, Gansu 730000, P.R. China

[‡]State Key Laboratory for Oxo Synthesis and Selective Oxidation, Lanzhou Institute of Chemical Physics, Chinese Academy of Sciences, Lanzhou, Gansu, 730000, P.R. China

Supporting Information

ABSTRACT: The first asymmetric total syntheses of cladosporin and isocladosporin were accomplished in 8 steps with 8% overall yield and 10 steps with 26% overall yield, respectively. The relative configuration of isocladosporin was determined via this total synthesis.



INTRODUCTION

Cladosporin ((-)-1) (also known as asperentin) is an antifungal antibiotic and plant growth inhibitor produced by various fungal sources.^{1,2} Vederas and other researchers at Merck assigned its absolute stereochemistry,^{3,2e} and Grove et al. reported the general features of its biosynthesis in *Aspergillus flavus* by administration of $[1-^{13}C]$ acetate and $[2-^{14}C]$ -malonate.⁴ Isocladosporin (2 or 3), whose structure was not determined on the basis of the available characterization data, was isolated from the fungus *Cladosporium cladosporioides* in 1993 (Figure 1).⁵ Cladosporin ((-)-1) and isocladosporin (2



Figure 1. Structures of cladosporin (1) and isocladosporin (2 or 3).

or 3) both feature a 2,6-disubstituted tetrahydropyran (THP) ring and a δ -valerolactone with a fused 1,3-dihydroxybenzene ring. Their favorable bioactivities as well as interesting structures inspired our interest in their syntheses. As part of our ongoing program toward the syntheses of bioactive pyranand pyranone-type natural products,^{6,7} we herein report the first asymmetric total synthesis of cladosporin and isoclado-sporin and also the efforts to determine the relative configuration of isocladosporin (given the lack of optical data for natural isocladosporin, the absolute configuration of the natural product cannot be deduced).

RETROSYNTHETIC ANALYSIS

The stereodivergent construction of cyclic ethers remains an important area of synthetic interest,⁸ in which 2,6-disubstituted

tetrahydropyran systems are present in a large number of biologically active natural products such as leucascandrolide,⁹ phorboxazole,¹⁰ and spirastrellolide.¹¹ Based on the chemistry involving intramolecular tandem cyclization—addition¹² and reductive etherification¹³ reactions, we envisioned that cladosporin and isocladosporin could be synthesized in short steps from tetrahydropyran (THP) oxocarbenium ions **6** and **10**, respectively (Scheme 1). In addition, the δ -valerolactone could be synthesized through a similar reaction sequence involving oxa-Pictet—Spengler cyclization¹⁴ and Jones oxidation.¹⁵

RESULTS AND DISCUSSION

Our synthesis of (-)-1 thus began with 7, which was derived from ring-opening of (S)-propylene oxide 8 in one step (Scheme 2). Hydroxy dioxane 7 was treated with allyltrimethylsilane in the presence of a catalytic amount of TMSOTf to give 16 with 2,6-trans-THP selectivity. We then accessed the enantioenriched epoxide (-)-17 using the Jacobsen hydrolytic kinetic resolution $(HKR)^{16}$ from the corresponding epoxide which was obtained as a 1:1 mixture by *m*-CPBA epoxidation of alkene 16. Alcohol trans-4 was obtained by reaction of epoxide 17 with the Grignard reagent and CuI at low temperature. Subsequently, the lactone 19 was furnished from trans-4 by a two-step process involving oxa-Pictet-Spengler cyclization and Jones oxidation. Initially, the oxa-Pictet-Spengler reaction was found to be unsuccessful between *trans*-4 and MOMCl/ZnCl₂, DMM/BF₃·Et₂O, or DMM/TMSOTf (DMM: dimethoxymethane). Finally, condensation of trans-4 with trimethyl orthoformate under pTSA conditions yielded the desired acetal 18 in 89% yield, which was characterized to be a single diastereomer of *trans*-pyran.¹⁷ Jones oxidation of 18 led to the

Received: April 20, 2012 **Published:** June 4, 2012

Scheme 1. Retrosynthetic Analysis of Cladosporin (1) and Isocladosporin (2 or 3)



Scheme 2. Total Synthesis of Cladosporin ((-)-1)



formation of lactone **19**, which was converted to the natural product (-)-**1** by removal of both aromatic methyl ether functions under Maier's conditions¹⁸ (AlI₃, TBAI, and phloroglucinol). The crystal structure determination established the stereochemistry of the 2,6-*trans*-tetrahydropyran ring at positions C10 and C14 and the 1,3-*anti*-diol at positions C3 and C10 (for details, see the Supporting Information).¹⁹

Based on the successful synthesis of cladosporin, we then turned our attention to the synthesis of isocladosporin (Scheme 3). According to the structure of (-)-2, the more plausible configuration of isocladosporin according to the original literature,⁵ aldehyde 13 was selected as the starting material. Homologation of 13 was performed via a Wittig reaction, and the resulting enol ether was directly treated with 1,3propanedithiol and BF₃·Et₂O to give dithiane 20 in satisfactory yield.²⁰ Treatment of 20 with ^tBuLi in THF was followed by the ring-opening of (S)-epichlorohydrin and concurrent epoxide formation to afford 21. The ring-opening of epoxide 21, deprotection of the 1,3-dithiane group with I_{2} , and $Me_4NBH(OAc)_3$ reduction²¹ of β -hydroxy ketone 12 gave the desired 1,3-anti-diol 11. Then we turned to the reductive cyclization of hydroxy dioxolanes developed by Colobert and co-workers.²² Treatment of 11 with an excess of Et₃SiH (5 equiv), followed by TMSOTf (1.3 equiv) in CH₂Cl₂ at 0 °C, led to the rapid (within 30 min) formation of the 2,6-cis-THP derivative cis-4 as a sole diastereomer. The remaining steps involving oxa-Pictet-Spengler reaction, Jones oxidation, and deprotection of both aromatic methyl ether functions were conducted as depicted as the synthesis of cladosporin. The characterization data of (-)-2 (¹H, ¹³C NMR determined in CDCl₃, and melting point) were consistent with the structure reported for isocladosporin (the solvent was not mentioned in

Article

Scheme 3. Total Synthesis of (-)-2





the original reference; however, the comparative analysis of ${}^{1}\text{H}$ and ${}^{13}\text{C}$ chemical shifts between Jacyno and Springer led us to believe that the data were collected in CDCl₃).

To further confirm the configuration of isocladosporin, the enantiomer of the other possible structure (-)-3 was also synthesized (Scheme 4). Alcohol *cis*-4 was first converted to alcohol **25** through Mitsunobu reaction²³ and deprotection of the ester by K₂CO₃ in MeOH. Subsequently, (-)-3 was synthesized from **25** in three steps according to the previous route.

The principal differences between 2 and 3 were observed in ¹³C NMR spectra and melting point (see Table 1). Specifically, the ¹³C resonances at lower field for 3 ranging from 172.0 to 141.0 ppm and 78.0–73.0 ppm appears fairly diagnostic between 3 and 2. It is worth mentioning that the NMR data of 3 were collected in CD₃OD solvent because of its insolubility in CDCl₃. In addition, the melting point of 196–202 °C for 3 was significantly different from the value of isocladosporin obtained by Jacyno or the value of 2. Therefore, we confirmed that the configuration of isocladosporin is 2.

CONCLUSION

In conclusion, we have accomplished the first asymmetric total synthesis of cladosporin ((-)-1) and isocladosporin ((-)-2) from commercially available (S)-propylene oxide and (S)-epichlorohydrin as chiral sources, respectively. The strategy was developed on the basis of a notion of concise enantioselective assembly of the tetrahydropyran ring. The relative configuration of isocladosporin was defined as the structure of 2 compared with 3 through this total synthesis.

EXPERIMENTAL SECTION

Materials and Methods. All chemicals were used as received. Solvent THF was refluxed with Na, and CH_2Cl_2 was refluxed with CaH₂ and freshly distilled prior to use. All reactions under standard conditions were monitored by thin-layer chromatography (TLC) on gel F254 plates. The silica gel (200–300 meshes) was used for column chromatography, and the distillation range of petroleum was 60–90 °C. ¹H and ¹³C NMR spectral data were reported in ppm relative to tetramethylsilane (TMS) as internal standard. High-resolution mass

								¹³ C NMR	(CDCl ₃)									() du
I -(-)	Jacyno et al. ⁵	169.68	164.34	162.34	141.85	106.48	102.04	101.96	76.31	67.83	66.35	39.47	33.66	30.95	30.86	18.92	18.19	
	Springer et al. ^{2e}	169.5	164.4	163.9	141.2	106.8	101.4	100.4	76.0	6.99	66.2	38.7	33.5	31.1	30.5	19.2	18.2	180-184
	Vederas et al. ³	169.7	164.5	162.5	141.9	106.6	102.0	128.4	76.4	67.9	66.5	39.5	33.7	31.00	30.97	19.0	18.3	187.5-188.5
	our work	170.1	164.1	163.7	141.6	106.9	101.8	101.0	76.3	67.8	66.6	38.8	33.4	30.9	30.6	19.0	18.0	175-177
2	Jacyno et al. ⁵	170.10	164.30	162.84	141.94	106.67	101.92	101.69	75.94	74.02	73.16	41.79	33.64	33.17	31.86	23.47	22.02	164 - 166
	our work	170.3	164.2	163.3	141.9	106.8	101.9	101.3	76.0	74.1	73.2	41.7	33.6	33.1	31.6	23.4	22.0	159-160
3^a		171.7	166.4	165.8	143.6	108.1	102.3	101.7	78.0	75.5	75.2	42.2	34.5	33.8	32.5	24.7	22.6	196-202
'NMR (data was collected i	n CD ₃ OD.																
		5																

Table 1. Analysis data of (-)-1, 2, and 3

spectra (HRMS) were recorded using a Fourier transform ion cyclotron resonance (FT-ICR) mass spectrometer.

Preparation of (S)-5-(1,3-Dioxan-2-yl)pentan-2-ol (7). To a suspension of Mg (1.26 g, 0.052 g atom) in THF (10 mL) was added I₂ to activate Mg, and a solution of the bromide (9.75 g, 50.0 mmol) in THF (100 mL) was added dropwise. After the addition (about 1 h), the mixture was stirred at rt for 2 h and then cooled to -30 °C.

To this cold Grignard reagent solution was added CuI (0.95 g, 5 mmol), followed by the addition of (*S*)-propylene oxide (5.8 g, 100 mmol) in THF (30 mL). The mixture was stirred at -30 °C overnight and then quenched by addition of saturated aqueous NH₄Cl solution (50 mL). After 30 min of stirring, the mixture was extracted with Et₂O, and the combined organic layers were dried over Na₂SO₄, filtered, and concentrated in vacuo. The resulting yellowish oil was purified by column chromatography to afford alcohol 7 as a colorless oil (7.22 g, 83%): [α]²⁰_D = +4.0 (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 4.53 (t, *J* = 5.2 Hz, 1H), 4.10 (dd, *J* = 4.4, 11.2 Hz, 2H), 3.81–3.73 (m, 3H), 2.13–2.03 (m, 1H), 1.76–1.33 (m, 8H), 1.18 (d, *J* = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 102.2, 67.8, 66.8, 39.0, 34.9, 25.8, 23.4, 20.1; HRMS (EIS) calcd for C₉H₁₉O₃ [M + H]⁺ 175.1329, found 175.1335.

Preparation of (2R,6S)-2-Allyltetrahydro-6-methyl-2H-pyran (16). To a cooled (-40 °C) solution of 7 (10.0 g, 57.5 mmol) and allyltrimethylsilane (18.2 mL, 115 mmol) in CH_2Cl_2 (100 mL) was added TMSOTf (2.1 mL, 11.5 mmol). The reaction mixture was then warmed to -20 °C and stirred for 1 h. After quenching by addition of saturated aqueous NaHCO₃ solution, the resulting mixture was diluted with Et₂O. The layers were separated, and the aqueous layer was extracted with Et₂O. The combined organic layers were dried over anhydrous Na₂SO₄ and concentrated in vacuo. The residue was purified by column chromatography to afford 16 as a colorless oil (5.8 g, 72%): $[\alpha]_{D}^{20} = -40.0$ (c = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 5.86–5.76 (m, 1H), 5.10–5.03 (m, 2H), 3.95–3.90 (m, 1H), 3.84-3.79 (m, 1H), 2.46-2.38 (m, 1H), 2.24-2.17 (m, 1H), $1.69-1.61 (m, 4H), 1.38-1.26 (m, 2H), 1.18 (d, J = 6.4 Hz, 3H); {}^{13}C$ NMR (100 MHz, CDCl₃) δ 135.5, 116.4, 70.6, 67.0, 37.9, 31.5, 29.2, 19.7, 18.3; HRMS (EIS) calcd for $C_9H_{17}O [M + H]^+$ 141.1274, found 141.1265.

Preparation of (25,6*R*)-Tetrahydro-2-methyl-6-(((5)-oxiran-2-yl)methyl)-2*H*-pyran (17). To a solution of alkene 16 (1.4 g, 10 mmol) in CH_2Cl_2 (15 mL) at 0 °C was added 70% (m/m) *m*-CPBA (3.94 g, 16 mmol) in one portion. The reaction mixture was warmed to room temperature and stirred for 10 h. The reaction mixture was quenched with 1 N aqueous NaOH and saturated aqueous $Na_2S_2O_3$ and extracted with CH_2Cl_2 . The organic layer was dried over anhydrous Na_2SO_4 , filtered, and concentrated under vacuum to give a yellow liquid which was purified by column chromatography to afford the corresponding epoxide as a colorless liquid (1.27 g, 79%).

A flask open to air was charged with (R,R)-Co^{II} salen (24 mg, 0.04 mmol) and AcOH (27 mg, 0.45 mmol) in toluene (10 mL) at room temperature. The solution, initially red, turned brown within a few seconds. The solution was stirred under air atmosphere for 1 h and then concentrated under vacuum. The above epoxide was added neat. The solution was cooled to 0 $^\circ\text{C}\textsc{,}$ and water (80 mg, 5 mmol) was added dropwise while the temperature was controlled. After the addition, the reaction mixture was allowed to warm to room temperature (water bath was used to ensure temperature remains around 20 °C). The reaction mixture was stirred for 18 h and then directly purified by column chromatography to afford pure epoxide 17 (0.53 g, 42%) as a colorless oil: $[\alpha]_{D}^{20} = -59.0$ (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 4.04–3.88 (m, 2H), 3.05–3.01 (m, 1H), 2.79 (t, J = 4.4 Hz, 1H), 2.49–2.47 (m, 1H), 1.99–1.93 (m, 1H), $1.73-1.59 (m, 4H), 1.48-1.28 (m, 3H), 1.19 (d, J = 6.4 Hz, 3H); {}^{13}C$ NMR (100 MHz, CDCl₃) δ 68.7, 67.2, 50.0, 47.2, 36.8, 31.3, 30.3, 19.5, 18.3; HRMS (EIS) calcd for $C_9H_{16}NaO_2 [M + Na]^+$ 179.1043, found 179.1050.

Preparation of (*R*)-1-((2*R*,6*S*)-Tetrahydro-6-methyl-2*H*-pyran-2-yl)-3-(3,5-dimethoxyphenyl)propan-2-ol (*trans*-4). To a cooled (-30 °C) THF solution of 10, which had been prepared from Mg (0.6 g, 0.025 g atom), 1-bromo-3,5-dimethoxybenzene (5.0 g, 23

mmol), I₂ (cat.), and THF in a similar manner as described above, was added CuI (0.44 g, 2.31 mmol) followed by the addition of epoxide 17 (0.83 g, 5.32 mmol) in THF (10 mL). The mixture was stirred at -30 °C overnight and then quenched by addition of saturated aqueous NH₄Cl solution (10 mL). After 30 min of stirring, the mixture was extracted with Et₂O, and the combined organic layers were dried over Na₂SO₄, filtered, and concentrated in vacuo. The resulting yellowish oil was purified by column chromatography to afford alcohol *trans*-4 as a colorless oil (1.52 g, 97%): $[\alpha]^{20}_{D} = -19.5$ (c = 2.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 6.38 (d, J = 2.0 Hz, 2H), 6.33 (d, J = 2.0 Hz, 1H), 4.10–4.06 (m, 2H), 3.97–3.95 (m, 1H), 3.77 (s, 6H), 2.80 (br s, 1H), 2.71 (d, J = 6.4 Hz, 2H), 1.89–1.83 (m, 1H), 1.70–1.26 (m, 8H), 1.20 (d, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 160.7, 141.2, 107.2, 98.2, 69.5, 67.5, 67.5, 55.1, 44.3, 39.6, 30.8, 30.5, 18.7, 18.2; HRMS (EIS) calcd for C₁₇H₂₇O₄ [M + H]⁺ 295.1904, found 295.1913.

Preparation of (1R,3R)-3,4-Dihydro-3-(((2R,6S)-tetrahydro-6methyl-2H-pyran-2-yl)methyl)-1,6,8-trimethoxy-1H-isochromene (18). To a solution of trans-4 (0.96 g, 3.26 mmol) and trimethyl orthoformate (8 mL) in CH₂Cl₂ (20 mL) was added pTSA (62 mg, 0.326 mmol). After being stirred for 1 h, the reaction mixture was quenched with addition of saturated aqueous NaHCO₃ solution, and the resulting mixture was diluted with Et2O. The layers were separated, and the aqueous layer was extracted with Et₂O. The combined organic layers were dried over anhydrous Na2SO4 and concentrated in vacuo. The residue was purified by column chromatography to afford 18 (0.98 g, 89%) as a white solid: $[\alpha]^{20}$ = -35.0 (*c* = 2.0, CHCl₃); mp 62 °C; ¹H NMR (400 MHz, CDCl₃) δ 6.29 (s, 1H), 6.20 (s, 1H), 5.52 (s, 1H), 4.40-4.33 (m, 1H), 4.16-4.11 (m, 1H), 3.94-3.91 (m, 1H), 3.80 (s, 3H), 3.77 (s, 3H), 3.55 (s, 3H), 2.62 (d, J = 7.6 Hz, 2H), 1.99–1.93 (m, 1H), 1.74–1.56 (m, 5H), 1.42–1.30 (m, 2H), 1.21 (d, J = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 160.3, 158.0, 136.6, 115.9, 103.7, 96.7, 95.3, 67.0, 66.9, 63.0, 55.6, 55.2, 55.1, 39.7, 34.6, 31.5, 30.7, 19.4, 18.4; HRMS (EIS) calcd for $C_{19}H_{28}NaO_5$ [M + Na]⁺ 359.1829, found 359.1822.

Preparation of (R)-3,4-Dihydro-3-(((2R,6S)-tetrahydro-6methyl-2H-pyran-2-yl)methyl)-6,8-dimethoxyisochromen-1one (19). To a cooled (0 °C) solution of 18 (0.80 g, 2.37 mmol) in acetone (12 mL) was added 3.0 M Jones oxidant (2.4 mL, 7.2 mmol). The reaction mixture was then warmed to rt and stirred for 1 h. After quenching by addition of water, the resulting mixture was diluted with EtOAc. The layers were separated, and the aqueous layer was extracted with EtOAc. The combined organic layers were dried over anhydrous Na₂SO₄ and concentrated in vacuo. The residue was purified by column chromatography to afford 19 as a foamy solid (0.57 g, 75%): $[\alpha]^{20}_{D} = +76.5 \ (c = 2.0, \text{ CHCl}_3); \ ^1\text{H NMR} \ (400 \text{ MHz}, \text{ CDCl}_3) \ \delta \ 6.34$ (s, 1H), 6.24 (s, 1H), 4.53-4.51 (br m, 1H), 4.02 (br m, 1H), 3.85 (br m, 1H), 3.85 (s, 3H), 3.79 (s, 3H), 2.86-2.73 (m, 2H), 1.87-1.55 (m, 6H), 1.27 (br m, 2H), 1.13 (d, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 164.1, 162.8, 162.6, 143.8, 106.8, 103.7, 97.5, 74.2, 67.3, 65.9, 55.9, 55.3, 39.3, 35.3, 30.8, 30.7, 18.7, 18.1; HRMS (EIS) calcd for $C_{18}H_{24}NaO_5$ [M + Na]⁺ 343.1516, found 343.1512.

Preparation of Cladosporin ((-)-1). A suspension of Al powder (1.81 g, 67 mmol) in dry benzene (10 mL) was treated with I₂ (6.35 g, 100 mmol)25 mmol) under Ar, and the violet mixture was stirred under reflux for 30 min until the color had changed to a colorless mixture. After the mixture was cooled to 0 °C, a few crystals of TBAI (58 mg, 0.157 mmol) and phloroglucinol (0.99 g, 7.85 mmol) were added before a solution of lactone 19 (0.5 g, 1.56 mmol) in dry benzene (3 mL) was added in one portion. The resulting green-brown suspension was stirred for 30 min at 0 °C before saturated Na₂S₂O₃ solution (15 mL) and ethyl acetate (15 mL) were added. After separation of the layers, the aqueous phase was extracted with ethyl acetate. The combined organic layers were washed with brine, dried over Na2SO4, filtered, and concentrated in vacuo. Purification by column chromatography to afford 1 (0.29 g, 63%) as a white solid: $[\alpha]_{D}^{20} = -14.0$ (c = 1.0, EtOH); mp 175–177 °C; ¹H NMR (400 MHz, CDCl₃) δ 10.99 (d, J = 1.6 Hz, 1H), 8.46 (m, 1H), 6.29 (d, J = 2.0 Hz, 1H), 6.16 (d, J = 1.6Hz, 1H), 4.71-4.66 (m, 1H), 4.13-4.10 (m, 1H), 3.97 (br s, 1H), 2.88-2.75 (m, 2H), 2.02-1.95 (m, 1H), 1.84-1.78 (m, 1H), 1.681.53 (m, 4H), 1.37–1.32 (m, 2H), 1.20 (d, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 170.1, 164.1, 163.7, 141.6, 106.9, 101.8, 101.0, 76.3, 67.8, 66.6, 38.8, 33.4, 30.9, 30.6, 19.0, 18.0; HRMS (EIS) calcd for C₁₆H₂₁O₅ [M + H]⁺ 293.1384, found 293.1389.

Preparation of 2-(3,5-Dimethoxybenzyl)-1,3-dithiane (20). To a suspension of methoxymethylphosphonium chloride (20.5 g, 60 mmol) in 150 mL of THF was added LDA (2.0 M, 30 mL, 60 mmol) at 0 °C, and the mixture was stirred for 30 min at 0 °C. Aldehyde **13** (6.64 g, 40 mmol) in 50 mL of THF was added to the mixture. After being stirred for 1 h, the reaction mixture was quenched by addition of saturated aqueous NH_4Cl solution, and the resulting mixture was diluted with Et_2O . The layers were separated, and the aqueous layer was extracted with Et_2O . The combined organic layers were dried over anhydrous Na_2SO_4 and concentrated in vacuo. The residue was dissolved in petroleum ether; after removal of $Ph_3P=O$, the filtrate was concentrated in vacuo.

The crude residue was dissolved in CH₂Cl₂ (100 mL), and 1,3propanedithiol (5.3 g, 48 mmol) was added at -10 °C. To the resultant solution was added BF₃·Et₂O (10 mL, 80 mol) dropwise and the solution stirred at -10 °C for 2 h. The reaction was then carefully quenched with saturated NaHCO₃ solution and extracted with Et₂O. The combined organic extracts were dried over anhydrous Na₂SO₄ and concentrated in vacuo. MeOH was added to this residue; **20** (10.8 g, 79%) was then obtained as a white solid after filtration: mp 75–76 °C; ¹H NMR (400 MHz, CDCl₃) δ 6.41 (d, *J* = 2.4 Hz, 2H), 6.36 (t, *J* = 2.4 Hz, 1H), 4.25 (t, *J* = 7.6 Hz, 1H), 3.78 (s, 6H), 2.96 (d, *J* = 7.6 Hz, 2H), 2.90–2.78 (m, 4H), 2.15–2.08 (m, 1H), 1.91–1.80 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 160.6, 139.4, 107.2, 98.8, 55.1, 48.3, 42.0, 30.5, 25.7; HRMS (EIS) calcd for C₁₃H₁₉O₂S₂ [M + H]⁺ 271.0821, found 271.0832.

Preparation of (S)-2-((2-(3,5-Dimethoxybenzyl)-1,3-dithian-2-yl)methyl)oxirane (21). To a solution of 1,3-dithiane 20 (3.80 g, 14 mmol) in anhydrous THF (10 mL) at -30 °C was added ^tBuLi (1.7 M in pentane, 12.4 mL, 21 mmol). After 30 min, (S)epichlorohydrin (1.95 g, 21 mmol) was added dropwise, and the reaction mixture was stirred for a further 1 h at -30 °C and then allowed to warm to rt and stirred overnight. The reaction was quenched by saturated aqueous NH4Cl solution and extracted with Et₂O. The combined organic layers were dried over anhydrous Na₂SO₄ and concentrated in vacuo. The crude product was purified by column chromatography to give epoxide 21 as a white solid (3.52 g, 77%): $[\alpha]_{D}^{20} = +12.0$ (c = 1.0, CHCl₃); mp 62 °C; ¹H NMR (400 MHz, CDCl₃) δ 6.51 (d, J = 2.0 Hz, 2H), 6.34 (d, J = 2.0 Hz, 1H), 3.74 (s, 6H), 3.27-3.17 (m, 3H), 2.86-2.84 (m, 4H), 2.83-2.77 (m, 1H), 2.50–2.48 (m, 1H), 2.19 (dd, J = 4.4, 15.2 Hz, 1H), 2.01–1.91 (m, 3H); 13 C NMR (100 MHz, CDCl₃) δ 159.9, 137.2, 109.2, 98.6, 55.0, 52.0, 49.0, 46.4, 45.8, 40.5, 26.2, 24.5; HRMS (EIS) calcd for $C_{16}H_{22}NaO_{3}S_{2}$ [M + Na]⁺ 349.0903, found 349.0905.

Preparation of (R)-1-(2-(3,5-Dimethoxybenzyl)-1,3-dithian-2-yl)-5-(2-methyl-1,3-dioxolan-2-yl)pentan-2-ol (22). To a cooled (-30 °C) THF solution of 15, which had been prepared from Mg (0.40 g, 0.017 g atom), 2-(2-bromoethyl)-2-methyl-1,3dioxolane (3.0 g, 15 mmol), I₂ (cat.), and THF in a similar manner as described above, was added CuI (0.286 g, 1.5 mmol) followed by the addition of epoxide $\mathbf{21}$ (1.63 g, 5 mmol) in THF (10 mL). The mixture was stirred at -30 °C overnight and then quenched by addition of saturated aqueous NH4Cl solution (10 mL). After 30 min of stirring, the mixture was extracted with Et₂O, and the combined organic layers were dried over Na2SO4, filtered, and concentrated in vacuo. The resulting yellowish oil was purified by column chromatography to afford alcohol 22 as a colorless oil (2.05 g, 93%): $[\alpha]_{D}^{20} = -31.0$ (c = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 6.47 (s, 2H), 6.37 (s, 1H), 4.08 (m, 1H), 3.93 (m, 4H), 3.77 (s, 6H), 3.50 (br s, 1H), 3.21 (dd, J = 14.0, 32.4 Hz, 2H), 3.08-3.02 (m, 1H), 2.96-2.90 (m, 1H), 2.83 (m, 2H), 2.28-2.21 (m, 1H), 2.02 (m, 1H), 1.93-1.85 (m, 2H), 1.65-1.37 (m, 6H), 1.32 (s, 3H); $^{13}\mathrm{C}$ NMR (100 MHz, CDCl_3) δ 160.1, 137.2, 109.9, 109.3, 98.8, 68.5, 64.5, 55.2, 52.4, 46.2, 43.9, 39.0, 37.7, 26.8, 26.3, 24.5, 23.6, 20.0; HRMS (EIS) calcd for $C_{22}H_{34}NaO_5S_2$ [M + Na]⁺ 465.1740, found 465.1744.

Preparation of (R)-4-Hydroxy-1-(3,5-dimethoxyphenyl)-7-(2methyl-1,3-dioxolan-2-yl)heptan-2-one (12). Dithiane 22 (1.50 g, 3.39 mmol) was dissolved in THF/H2O (4:1, 35 mL) and CaCO3 (3.39 g, 33.9 mmol) and then cooled to 0 °C, whereupon iodine (2.60 g, 10.2 mmol) was added portionwise. The reaction mixture was then allowed to stir for 30 min before quenching with saturated Na₂S₂O₃ (20 mL) solution. The aqueous phase was separated and extracted with EtOAc. The combined organic extracts were washed with brine, dried over anhydrous Na2SO4, and concentrated in vacuo. The crude residue was purified by column chromatography to give hydroxy ketone 12 (1.05 g, 88%) as a colorless oil: $[\alpha]_{D}^{20} = -33.0$ (c = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 6.37 (s, 1H), 6.33 (d, J = 2.0 Hz, 2H), 4.01 (m, 1H), 3.95-3.86 (m, 4H), 3.78 (s, 6H), 3.63 (s, 2H), 3.01 (s, 1H), 2.66-2.52 (m, 2H), 1.67-1.33 (m, 6H), 1.30 (s, 3H); $^{13}\mathrm{C}$ NMR (100 MHz, CDCl_3) δ 209.3, 161.0, 135.7, 109.9, 107.4, 99.0, 67.4, 64.5, 55.2, 51.0, 48.1, 38.8, 36.4, 23.7, 19.9; HRMS (EIS) calcd for $C_{10}H_{28}NaO_6$ [M + Na]⁺ 375.1778, found 375.1779

Preparation of (2R,4R)-1-(3,5-Dimethoxyphenyl)-7-(2-methyl-1,3-dioxolan-2-yl)heptane-2,4-diol (11). To a stirred solution of Me₄NBH(OAc)₃ (4.2 g, 16 mmol) in acetonitrile (10 mL) was added acetic acid (12 mL), and the mixture was stirred at room temperature for 30 min. The mixture was cooled to -30 °C, and an acetonitrile solution of compound 12 (0.7 g, 2 mmol in 5 mL of acetonitrile) was added. The mixture was stirred at the same temperature for 12 h. The reaction was quenched with 1 N NaOH, and the resulting mixture was diluted with EtOAc. The layers were separated, and the aqueous layer extracted with EtOAc. The combined organic layers were dried over Na2SO4 and concentrated in vacuo. The residue was purified by column chromatography to give diol 11 (0.64 g, 91%) as a colorless oil: $[\alpha]_{D}^{20} = +3.5$ (c = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 6.35 (s, 2H), 6.31 (d, J = 1.6 Hz, 1H), 4.16– 4.11 (m, 1H), 3.96-3.86 (m, 5H), 3.75 (s, 6H), 2.80 (br s, 2H), 2.69 (d, J = 6.8 Hz, 2H), 1.63–1.34 (m, 8H), 1.28 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 160.8, 140.7, 109.9, 107.2, 98.3, 69.8, 68.8, 64.5, 55.2, 44.3, 42.0, 38.9, 37.4, 23.6, 20.1; HRMS (EIS) calcd for C₁₉H₃₁O₆ [M + H]⁺ 355.2115, found 355.2111.

Preparation of (R)-1-((2R,6R)-Tetrahydro-6-methyl-2Hpyran-2-yl)-3-(3,5-dimethoxyphenyl)propan-2-ol (cis-4). To a cooled (0 °C) solution of 11 (0.4 g, 1.13 mmol) and triethylsilane (0.66 g, 5.65 mmol) in CH₂Cl₂ was added TMSOTf (0.33 g, 1.49 mmol). After being stirred for 30 min at 0 °C, the reaction was quenched with saturated aqueous NaHCO3 solution, the resulting mixture was diluted with Et₂O. The layers were separated, and the aqueous layer was extracted with Et₂O. The combined organic layers were dried over anhydrous Na2SO4 and concentrated in vacuo. The residue was purified by column chromatography to afford cis 4 as a colorless oil (0.31 g, 93%): $[\alpha]^{20}_{D} = -0.5$ (c = 2.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 6.40 (d, J = 2.0 Hz, 2H), 6.34 (dd, J = 2.0, 2.4 Hz, 1H), 4.19-4.13 (m, 1H), 3.79 (s, 6H), 3.72-.367 (m, 1H), 3.50-3.43 (m, 1H), 3.13 (d, J = 3.6 Hz, 1H), 2.72 (d, J = 6.4 Hz, 2H), 1.84-1.79 (m, 1H), 1.73-1.62 (m, 2H), 1.57-1.45 (m, 3H), 1.40-1.30 (m, 1H), 1.24–1.14 (m, 1H), 1.17 (d, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 160.6, 141.2, 107.2, 98.1, 75.4, 74.0, 69.4, 55.1, 44.2, 41.5, 33.0, 30.7, 23.5, 22.1; HRMS (EIS) calcd for C₁₇H₂₇O₄ [M + H] 295.1904, found 295.1915.

Preparation of (1*R*,3*R*)-3,4-Dihydro-3-(((2*R*,6*R*)-tetrahydro-6methyl-2*H*-pyran-2-yl)methyl)-1,6,8-trimethoxy-1*H*-isochromene (23). The same procedure as described above for the preparation of 18 was used in the preparation of 23. To a solution of *cis*-4 (0.282 g, 0.96 mmol) and trimethyl orthoformate (2.5 mL) in CH₂Cl₂ (10 mL) was added pTSA (19 mg, 0.096 mmol). After being stirred for 1 h, the reaction mixture was quenched by addition of saturated aqueous NaHCO₃ solution, and the resulting mixture was diluted with Et₂O. The layers were separated, and the aqueous layer was extracted with Et₂O. The combined organic layers were dried over anhydrous Na₂SO₄ and concentrated in vacuo. The residue was purified by column chromatography to afford 23 (0.306 g, 95%) as a white solid: $[\alpha]^{20}_{D} = -31.0$ (*c* = 1.0, CHCl₃); mp 104–105 °C; ¹H NMR (400 MHz, CDCl₃): δ 6.29 (d, *J* = 2.0 Hz, 1H), 6.20 (d, *J* = 2.0 Hz, 1H), 5.53 (s, 1H), 4.53–4.47 (m, 1H), 3.81 (s, 3H), 3.77 (s, 3H), 3.72–3.67 (m, 1H), 3.51 (s, 3H), 3.48–3.41 (m, 1H), 2.60 (d, J = 7.2 Hz, 2H), 1.86–1.50 (m, 6H), 1.30–1.18 (m, 2H), 1.17 (d, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 160.3, 157.9, 136.8, 115.9, 103.8, 96.7, 95.1, 73.9, 73.7, 62.2, 55.6, 55.2, 54.5, 42.4, 34.5, 33.3, 31.8, 23.8, 22.0; HRMS (EIS) calcd for C₁₉H₂₈NaO₅ [M + Na]⁺ 359.1829, found 359.1826.

Preparation of (R)-3,4-Dihydro-3-(((2R,6R)-tetrahydro-6methyl-2H-pyran-2-yl)methyl)-6,8-dimethoxyisochromen-1one (24). The same procedure as described for the preparation of 19 was used in the preparation of 24. To a cooled (0 $^{\circ}$ C) solution of 23 (0.166 g, 0.494 mmol) in acetone (2.5 mL) was added 3.0 M Jones oxidant (0.5 mL, 1.482 mmol). The reaction mixture was then warmed to rt and stirred for 1 h. After quenching by addition of water, the resulting mixture was diluted with EtOAc. The layers were separated, and the aqueous layer was extracted with EtOAc. The combined organic layers were dried over anhydrous Na2SO4 and concentrated in vacuo. The residue was purified by column chromatography to afford 24 as a yellowish oil (0.117 g, 74%): $[\alpha]^{20}_{D} = +126.0$ (c = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 6.39 (s, 1H), 6.28 (s, 1H), 4.69–4.64 (m, 1H), 3.91 (s, 3H), 3.84 (s, 3H), 3.72-3.67 (m, 1H) 3.46-3.39 (m, 1H), 2.90–2.76 (m, 2H), 1.89–1.77 (m, 2H), 1.73–1.66 (m, 1H), 1.58-1.51 (m, 3H), 1.25-1.15 (m, 2H), 1.10 (d, J = 6.4 Hz, 3H); ${}^{13}C$ NMR (100 MHz, CDCl₃) δ 164.3, 163.0, 162.9, 144.1, 107.1, 103.8, 97.7, 73.8, 73.6, 73.0, 56.1, 55.5, 41.7, 35.5, 33.2, 31.6, 23.5, 22.0; HRMS (EIS) calcd for $C_{18}H_{25}O_5 [M + H]^+$ 321.1697, found 321.1705.

Preparation of Isocladosporin ((-)-2). The same procedure as described for the preparation of 1 was used in the preparation of (-)-2. A suspension of Al powder (0.38 g, 14.1 mmol) in dry benzene (5 mL) was treated with I_2 (1.34 g, 5.27 mmol) under Ar, and the violet mixture was stirred under reflux for 30 min until the color had changed to a colorless mixture. After the mixture was cooled to 0 °C, a few crystals of TBAI (12 mg, 0.033 mmol) and phloroglucinol (0.21 g, 1.67 mmol) were added before a solution of lactone 24 (0.105 g, 0.328 mmol) in dry benzene (1 mL) was added in one portion. The resulting green-brown suspension was stirred for 30 min at 0 °C before saturated Na₂S₂O₃ solution (5 mL) and ethyl acetate (5 mL) were added. After separation of the layers, the aqueous phase was extracted with ethyl acetate. The combined organic layers were washed with brine, dried over Na2SO4, filtered, and concentrated in vacuo. Purification by column chromatography to afford (-)-2 (0.085 g, 89%) as a white solid: $[\alpha]^{20}_{D} = -5.0$ (c = 1.0, CHCl₃); mp 158–159 °C; ¹H NMR (400 MHz, CDCl₃): δ 11.07 (d, J = 2.0 Hz, 1H), 7.83 (m, 1H), 6.34 (d, J = 2.0 Hz, 1H), 6.17 (s, 1H), 4.83-4.78 (m, 1H),3.72-3.66 (m, 1H), 3.49-3.45 (m, 1H), 2.84-2.71 (m, 2H), 1.92-1.71 (m, 3H), 1.60–1.50 (m, 3H), 1.27–1.20 (m, 2H), 1.15 (d, J = 6.4 Hz, 3H); ^{13}C NMR (100 MHz, CDCl₃) δ 170.3, 164.2, 163.3, 141.9, 106.8, 101.9, 101.3, 76.0, 74.1, 73.2, 41.7, 33.6, 33.1, 31.6, 23.4, 22.0; HRMS (EIS) calcd for $C_{16}H_{21}O_5 [M + H]^+$ 293.1384, found 293.1394.

Preparation of (*S*)-1-((*2R*,*6R*)-Tetrahydro-6-methyl-2*H*pyran-2-yl)-3-(3,5-dimethoxyphenyl)propan-2-ol (25). To a stirred, cooled (0 °C) mixture of *cis*-4 (0.50 g, 1.7 mmol), triphenylphosphine (0.50 g, 2.57 mmol), and 4-nitrobenzoic acid (0.43 g, 2.57 mmol) in THF (20 mL) was added dropwise diisopropyl azodicarboxylate (0.41 g, 2.57 mmol), and the mixture was stirred at rt overnight. After removal of the solvent, the residue was loaded directly onto a silica gel column, and elution with PE and EtOAc yielded the desired 4-nitrobenzoate as a yellow oil (0.58 g, 78%).

To a MeOH solution of the 4-nitrobenzoate obtained above was added K₂CO₃ (0.36 g, 2.6 mmol), and the resulting mixture was stirred at rt for 1 h before water and EtOAc were added. After separation of the layers, the aqueous phase was extracted with ethyl acetate. The combined organic layers were washed with brine, dried over Na₂SO₄, filtered, and concentrated in vacuo. Purification by column chromatography to afford **25** (0.354 g, 92%) as a colorless oil: $[\alpha]^{20}_{D} = -18.0 (c = 1.0, CHCl_3); {}^{1}H NMR (400 MHz, CDCl_3) \delta 6.39$ (d, *J* = 2.0 Hz, 2H), 6.32 (d, *J* = 2.0 Hz, 1H), 4.31 (s, 1H), 4.12–4.05 (m, 1H), 3.78 (s, 6H), 3.59–3.44 (m, 2H), 2.79 (dd, *J* = 6.4, 13.2 Hz, 1H), 2.58 (dd, *J* = 6.4, 13.2 Hz, 1H), 1.80–1.75 (m, 1H), 1.58–1.43 (m, SH), 1.27–1.17 (m, 2H), 1.15 (d, *J* = 6.0 Hz, 3H); {}^{13}C NMR (100 MHz, CDCl₃) δ 160.6, 141.2, 107.4, 98.1, 79.3, 74.0, 73.2, 55.2, 44.3,

41.8, 32.8, 31.6, 23.3, 22.1; HRMS (EIS) calcd for $C_{17}H_{27}O_4\,[M+H]^+$ 295.1904, found 295.1901.

Preparation of (15,35)-3,4-Dihydro-3-(((2R,6R)-tetrahydro-6methyl-2H-pyran-2-yl)methyl)-1,6,8-trimethoxy-1H-isochromene (26). The same procedure as described for the preparation of 18 was used in the preparation of 26. To a solution of 25 (0.235 g, 0.8 mmol) and trimethyl orthoformate (1.6 mL) in CH₂Cl₂ (5 mL) was added pTSA (16 mg, 0.08 mmol). After being stirred for 1 h, the reaction mixture was quenched with addition of saturated aqueous NaHCO₃ solution, and the resulting mixture was diluted with Et₂O. The layers were separated, and the aqueous layer was extracted with Et₂O. The combined organic layers were dried over anhydrous Na_2SO_4 and concentrated in vacuo. The residue was purified by column chromatography to afford 26 (0.261 g, 97%) as a white solid: $[\alpha]_{D}^{20}$ = +5.0 (c = 1.0, CHCl₃); mp 92 °C; ¹H NMR (400 MHz, $CDCl_3$: δ 6.30 (d, J = 2.0 Hz, 1H), 6.22 (d, J = 2.0 Hz, 1H), 5.51 (s, 1H), 4.36-4.29 (m, 1H), 3.81 (s, 3H), 3.77 (s, 3H), 3.66-3.59 (m, 1H), 3.52 (s, 3H), 3.49-3.44 (m, 1H), 2.65 (d, J = 6.8 Hz, 2H), 2.05-1.97 (m, 1H), 1.87-1.84 (m, 1H), 1.74-1.48 (m, 4H), 1.27-1.22 (m, 2H), 1.17 (d, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 160.4, 158.1, 136.6, 115.9, 103.8, 96.8, 95.5, 74.5, 74.0, 63.3, 55.6, 55.3, 55.2, 42.3, 34.1, 33.3, 31.1, 23.7, 22.2; HRMS (EIS) calcd for C19H29O5 [M + H]⁺ 337.2010, found 337.2016.

Preparation of (S)-3,4-Dihydro-3-(((2R,6R)-tetrahydro-6methyl-2H-pyran-2-yl)methyl)-6,8-dimethoxyisochromen-1one (27). The same procedure as described for the preparation of 19 was used in the preparation of 27. To a cooled (0 °C) solution of 26 (0.18 g, 0.53 mmol) in acetone (3 mL) was added 3.0 M Jones oxidant (0.55 mL, 1.65 mmol). The reaction mixture was then warmed to rt and stirred for 1 h. After quenching by addition of water, the resulting mixture was diluted with EtOAc. The layers were separated, and the aqueous layer was extracted with EtOAc. The combined organic layers were dried over anhydrous Na2SO4 and concentrated in vacuo. The residue was purified by column chromatography to afford 27 as a white solid (0.139 g, 81%): $[\alpha]_{D}^{20} = -70.0$ (c = 1.0, CHCl₃); mp 123 °C; ¹H NMR (400 MHz, CDCl₃) δ 6.40 (s, 1H), 6.32 (s, 1H), 4.58-4.55 (m, 1H), 3.92 (s, 3H), 3.86 (s, 3H), 3.66-3.61 (m, 1H), 3.46-3.42 (m, 1H), 2.97 (dd, J = 11.2, 16.0 Hz, 1H), 2.84 (dd, J = 2.4, 16.0 Hz, 1H), 2.13-2.05 (m, 1H), 1.83-1.73 (m, 2H), 1.64-1.47 (m, 3H), 1.28-1.14 (m, 2H), 1.13 (d, J = 6.0 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 164.3, 163.1, 162.7, 144.1, 107.0, 103.9, 97.7, 74.3, 73.8, 73.4, 56.1, 55.5, 40.8, 34.7, 33.1, 30.9, 23.5, 22.1; HRMS (EIS) calcd for $C_{18}H_{25}O_5 [M + H]^+$ 321.1697, found 321.1690.

Preparation of (S)-3,4-Dihydro-3-(((2R,6R)-tetrahydro-6methyl-2H-pyran-2-yl)methyl)-6,8-dihydroxyisochromen-1one ((–)-3). The same procedure as described for the preparation of (-)-1 was used in the preparation of (-)-3. A suspension of Al powder (0.36 g, 13.4 mmol) in dry benzene (5 mL) was treated with I₂ (1.27 g, 5 mmol) under Ar, and the violet mixture was stirred under reflux for 30 min until the color had changed to a colorless mixture. After the mixture was cooled to 0 °C, a few crystals of TBAI (12 mg, 0.033 mmol) and phloroglucinol (0.197 g, 1.56 mmol) were added before a solution of lactone 27 (0.10 g, 0.313 mmol) in dry benzene (1 mL) was added in one portion. The resulting green-brown suspension was stirred for 30 min at 0 °C before saturated Na₂S₂O₃ solution (5 mL) and ethyl acetate (5 mL) were added. After separation of the layers, the aqueous phase was extracted with ethyl acetate. The combined organic layers were washed with brine, dried over Na₂SO₄, filtered, and concentrated in vacuo. Purification by column chromatography to afford (-)-3 (0.079 g, 86%) as a white solid: $[\alpha]_{D}^{20} = -46.0$ (c = 1.0, EtOH); mp 196–202 °C; ¹H NMR (400 MHz, CD₃OD) δ 6.22 (s, 1H), 6.20 (s, 1H), 4.73-4.66 (m, 1H), 3.63-3.57 (m, 1H), 3.51-3.44 (m, 1H), 2.98-2.86 (m, 2H), 2.07-2.00 (m, 1H), 1.85-1.77 (m, 2H), 1.66-1.56 (m, 3H), 1.28-1.53 (m, 2H), 1.13 (d, J = 6.0 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 171.7, 166.4, 165.8, 143.6, 108.1, 102.3, 101.7, 78.0, 75.5, 75.2, 42.2, 34.5, 33.8, 32.5, 24.7, 22.6; HRMS (EIS) calcd for $C_{16}H_{21}O_5$ [M + H]⁺ 293.1384, found 293.1390.

ASSOCIATED CONTENT

S Supporting Information

Spectroscopic data for all new compounds and X-ray data for compounds 1-3 (CIF). This material is available free of charge via the Internet at http://pubs.acs.org.

AUTHOR INFORMATION

Corresponding Author

*Email: shexg@lzu.edu.cn.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

This research was supported by the MOST (2010CB833200), the NSFC (21125207, 21072086, 21102062), and Program 111.

REFERENCES

(1) (a) Scott, P. M.; Van Walbeek, W. J. Antibiot. 1971, 24, 747–755.
(b) Grove, J. F. J. Chem. Soc., Perkin Trans. 1 1972, 2400–2406.

(2) (a) For a review, see: Scott, P. M. In Mycotoxins: Production, Isolation, Separation and Purification; Betina, V., Ed.; Elsevier: Amsterdam, 1984; pp 457-461. (b) Anke, H.; Zähner, H.; König, W. A. Arch. Microbiol. 1978, 116, 253-257. (c) Podojil, M.; Sedmera, P.; Vokoun, J.; Betina, V.; Baráthová, H.; Duracková, Z.; Horáková, K.; Nemec, P. Folia Microbiol. (Prague) 1978, 23, 438-443. (d) Anke, H. J. Antibiot. 1979, 32, 952-958. (e) Springer, J. P.; Cutler, H. G.; Crumley, F. G.; Farrist, G.; Cox, R. H.; Davis, E. E.; Thean, J. E. J. Agric. Food Chem. 1981, 29, 853-855. (f) Grove, J. F.; Pople, M. Mycopathologia 1981, 76, 65-67.

(3) Reese, P. B.; Rawlings, B. J.; Ramer, S. E.; Vederas, J. C. J. Am. Chem. Soc. 1988, 110, 316-318.

(4) Cattel, L.; Grove, J. F.; Shaw, D. J. Chem. Soc., Perkin Trans. 1 1973, 2626–2629.

(5) Jacyno, J. M.; Harwood, J. S.; Cutler, H. G.; Lee, M.-K. J. Nat. Prod. 1993, 56, 1397–1401.

(6) (a) Zheng, H.; Zheng, J.; Yu, B.; Chen, Q.; Wang, X.; He, Y.; Yang, Z.; She, X. J. Am. Chem. Soc. 2010, 132, 1788. (b) Yu, B.; Jiang, T.; Li, J.; Su, Y.; Pan, X.; She, X. Org. Lett. 2009, 11, 3442. (c) He, J.; Tang, S.; Liu, J.; Su, Y.; She, X.; Pan, X. Tetrahedron 2008, 64, 8797. (d) He, J.; Zheng, J.; Liu, J.; She, X.; Pan, X. Org. Lett. 2006, 8, 4637. (7) (a) Li, J.; Zheng, H.; Su, Y.; Xie, X.; She, X. Synlett 2010, 2283. (b) Wang, X.; Wang, W.; Zheng, H.; Su, Y.; Jiang, T.; He, Y.; She, X. Org. Lett. 2009, 11, 3136. (c) Su, Y.; Xu, Y.; Han, J.; Zheng, J.; Qi, J.; Jiang, T.; Pan, X.; She, X. J. Org. Chem. 2009, 74, 2743. (d) Xu, Y.; Huo, X.; Li, X.; Zheng, H.; She, X.; Pan, X. Synlett 2008, 1665. (e) Zhang, J.; Li, Y.; Wang, W.; She, X.; Pan, X. J. Org. Chem. 2006, 71, 2918. (f) Wang, Q.; Huang, Q.; Chen, B.; Lu, J.; Wang, H.; She, X.; Pan, X. Angew. Chem., Int. Ed. 2006, 45, 3651.

(8) (a) Lewis, M. D.; Cha, J. K.; Kishi, Y. J. Am. Chem. Soc. **1982**, 104, 4976–4978. (b) Eliel, E. L.; Wilen, S. H.; Mander, L. N. Stereochemistry of Organic Compounds; Wiley: New York, 1994; pp 686–740. (c) Romero, J. A. C.; Tabacco, S. A.; Woerpel, K. A. J. Am. Chem. Soc. **2000**, 122, 168–169. (d) Ayala, L.; Lucero, C. G.; Romero, J. A. C.; Tabacco, S. A.; Woerpel, K. A. J. Am. Chem. Soc. **2003**, 125, 15521–15528.

(9) D'Ambrosio, M.; Guerriero, A.; Debitus, C.; Pietra, F. Helv. Chim. Acta 1996, 79, 51.

(10) Searle, P. A.; Molinski, T. F. J. Am. Chem. Soc. 1995, 117, 8126.
(11) (a) Williams, D. E.; Roberge, M.; Van Soest, R.; Andersen, R. J. J. Am. Chem. Soc. 2003, 125, 5296. (b) Williams, D. E.; Lapawa, M.; Feng, X.; Tarling, T.; Roberge, M.; Andersen, R. J. Org. Lett. 2004, 6, 2607–2610. (c) Warabi, K.; Williams, D. E.; Patrick, B. O.; Roberge, M.; Andersen, R. J. J. Am. Chem. Soc. 2007, 129, 508.

(12) Evans, P. A.; Cui, J.; Gharpure, S. J.; Hinkle, R. J. J. Am. Chem. Soc. 2003, 125, 11456-11457.

(13) (a) Komatsu, N.; Ishida, J.-y.; Suzuki, H. Tetrahedron Lett. 1997, 38, 7219–7222. (b) Evans, P. A.; Cui, J.; Gharpure, S. J.; Polosukhin, A.; Zhang, H.-R. J. Am. Chem. Soc. 2003, 125, 14702–14703. (c) Evans, P. A.; Cui, J.; Gharpure, S. J. Org. Lett. 2003, 5, 3883–3885. (d) Evans, P. A.; Andrews, W. J. Tetrahedron Lett. 2005, 46, 5625–5627.

(14) For intramolecular oxa-Pictet-Spengler cyclization, see:
(a) Giles, R. G. F.; Rickards, R. W.; Senanayake, B. S. J. Chem. Soc., Perkin Trans. 1 1997, 3361-3370. (b) Xu, Y.-C.; Kohlman, D. T.; Liang, S. X.; Erikkson, C. Org. Lett. 1999, 1, 1599-1602. (c) Bianchi, D. A.; Rua, F.; Kaufman, T. S. Tetrahedron Lett. 2004, 45, 411-415.
(d) Choukchou-Braham, N.; Mostefa-Kara, B.; Cheikh, N.; Didi, M. A. Synth. Commun. 2005, 35, 169-178. (e) Giles, R. G. F.; McManus, J. D. Tetrahedron Lett. 2009, 50, 6361-6363. (f) Sawant, R. T.; Jadhav, S. G.; Waghmode, S. B. Eur. J. Org. Chem. 2010, 4442-4449.

(15) Bowden, K.; Heilbron, I. M.; Jones, E. R. H.; Weedon, B. C. L. J. Chem. Soc. 1946, 39-45.

(16) (a) Tokunaga, M.; Larrow, J. F.; Kakiuchi, F.; Jacobsen, E. N. *Science* **1997**, 936–938. (b) Schaus, S. E.; Brandes, B. D.; Larrow, J. F.; Tokunaga, M.; Hansen, K. B.; Gould, A. E.; Furrow, M. E.; Jacobsen, E. N. *J. Am. Chem. Soc.* **2002**, *124*, 1307–1315.

(17) Bender, C. F.; Yoshimoto, F. K.; Paradise, C. L.; De Brabander, J. K. J. Am. Chem. Soc. 2009, 131, 11350-11352.

(18) Rink, C.; Sasse, F.; Zubriene, A.; Matulis, D.; Maier, M. E. Chem.—Eur. J. 2010, 16, 14469–14478.

(19) Crystallographic data for the structures of 1-3 reported in this paper have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication nos. CCDC-872667, -872668, and -881449.

(20) The most common approach was treatment of aldehyde with 1,3-propanedithiol in the presence of Lewis acid. The direct method could obviously reduce the amount of byproduct during hydrolysis.

(21) Evans, D. A.; Chapman, K. T.; Carreira, E. M. J. Am. Chem. Soc. 1988, 110, 3560-3578.

(22) (a) Carreño, M. C.; Mazery, R. D; Urbano, A.; Colobert, F.; Solladié, G. J. Org. Chem. 2003, 68, 7779–7787. (b) Colobert, F.; Choppin, S.; Ferreior-Mederos, L.; Obringer, M.; Arratta, S. L.; Urbano, A.; Carreño, M. C. Org. Lett. 2007, 9, 4451–4454.

(23) (a) Mitsunobu, O.; Yamada, M. Bull. Chem. Soc. Jpn. 1967, 40, 2380–2382. (b) Mitsunobu, O.; Yamada, M.; Mukaiyama, T. Bull. Chem. Soc. Jpn. 1967, 40, 935–939.