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Letter

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Discovery of a Tetrahydrobenzisoxazole Series of γ-Secretase Modulators

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KEYWORDS: Alzheimer's disease; y-Secretase modulators; tetrahydrobenzisoxazole.

ABSTRACT: The design and synthesis of a new series of tetrahydrobenzisoxazoles and their structure-activity relationship (SAR) as modulators of γ -secretase activity will be detailed. Several compounds are active γ -secretase modulators (GSMs) with good to excellent selectivity for the reduction of A β_{42} in the cellular assay. Compound **14a** was tested *in vivo* in a non-transgenic rat model and was found to significantly reduce A β_{42} in the CNS compartment compared to vehicle-treated animals (up to 58% reduction of cerebrospinal fluid A β_{42} as measured 3h after an acute oral dosing at 30 mg/kg).

Alzheimer's disease (AD) is the sixth leading cause of death in the United States. An estimated 5.5 million Americans have AD, costing the nation more than \$230 billion in 2016.¹ As the elderly population continues to grow, the prevalence of AD will be increasing greatly. Therefore, there is an urgent need to develop an effective therapy to prevent Alzheimer's disease.

Aβ proteins, which are metabolites of the amyloid precursor protein (APP), are considered to be involved in the degeneration and loss of neurons as well as the onset of AD.^{2,3} It is known that $A\beta_{40}$ is the most abundant species within the soluble pool of A β peptides; however, deposited A β plaques primarily consist of $A\beta_{42}$,^{4,5} which is the species that is most prone to aggregation, forming aggregates of insoluble fibrils in the brain.⁶ The higher aggregation potential of $A\beta_{42}$ may due to the distinct initial folding properties of $A\beta_{40}$ and $A\beta_{42}$.⁷ These $A\beta$ fragments are produced when APP is first cleaved by BACE (β-amyloid cleaving enzyme) and subsequently by y-secretase.8 Based on these facts, y-secretase inhibitors (GSIs) and BACE inhibitors have been proposed as treatments for the purpose of reducing the production of $A\beta$ fragments.⁹ However, notch-related adverse effects are known as a major obstacle for the development of GSIs for the treatment of AD.¹⁰ Therefore, γ -secretase modulators (GSMs) have attracted much attention in recent years.^{11,12}

GSMs selectively inhibit the production of the most amyloidogenic and neurotoxic $A\beta_{42}$ by shifting the position of γ secretase cleavage toward the formation of shorter, more soluble peptides, without interfering with the overall γ -secretase function, as measured by the formation of total amyloid ($A\beta_{to}$ t_{tal}).¹³ A larger ratio of $A\beta_{total}/A\beta_{42}$ represents a more selective GSM. More selective GSMs do not shut down the processing of Notch by γ -secretase, and thus offer a potentially improved side effect profile compare to GSIs. Due to this advantage, GSMs have attracted great interest from researchers.¹⁴ There are two major classes of GSMs, the nonsteroidal anti-**Public** inflammatory drugs (NSAIDs)^{15,16} and the non-NSAIDs class, such as **1** from Eisai,¹⁷ **2** and **3** from Merck.^{18,19} In addition, natural product-based GSMs have also been developed.^{20,21} Following our efforts²²⁻²⁷ to develop in the non-NSAID GSMs, we developed a new series, represented by structure **4** (Figure 1),²⁸ where the replacement of the double bond present in structures **1**– **3** to mitigiate a potential metabolic. Therefore, the double bond and the lactam carbonyl in **1** was replaced by a small heterocyclic fused ring such as tetrahydrobenzisoxazole in **4**.

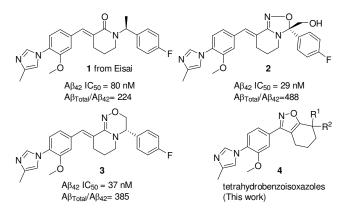
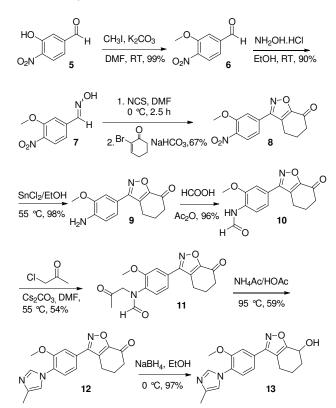


Figure 1. Design of GSMs from known structures

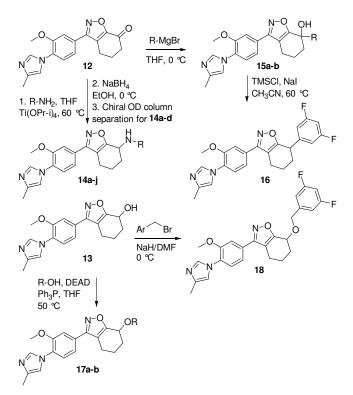
The tetrahydrobenzisoxazole was selected as a linker to lock the side chain in the same conformation as the double bond did in the lead structures 1-3. We synthesized a series of compounds with a core structure of 4. Our chemical synthesis involves key intermediates 12 and 13 which is summarized in Scheme 1. The *O*-methylation of 5 afforded 6, which was converted to the oxime compound 7. The key step is a [3 + 2] cycloaddition to afford isoxazole **8**. After optimization, we were able to develop a one pot reaction to produce intermediate **8** with a 67% yield (two steps). The nitro group in **8** was reduced to the corresponding amine to afford **9**. The amine group in **9** was converted to formamide **10**. The *N*-alkylation of **10** with 1-chloropropan-2-one, followed by treatment with ammonium acetate afforded **12**, which was reduced by sodium borohydride to deliver **13**.

Scheme 1. Synthesis of intermediates 12 and 13



From 12 and 13 a variety of final targets were prepared through traditional chemistry: such as reductive amination (12 to 14a-j), Grignard reaction (12 to 15a-b), Mitsunobu reaction (13 to 17a-b), dehydroxylation²⁹ (15a to 16) and alkylation (13 to 18) as illustrated in Scheme 2.

Scheme 2. Synthesis of final GSM compounds



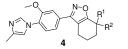
The biological test results are summarized in Table 1. Human embryonic kidney (HEK) 293 cells were treated with a variable concentration of GSM compounds for 5 h. $A\beta_{42}$ and $A\beta_{total}$ in conditioned media were measured using MesoScale Discovery (MSD) sandwich immunoassays.³⁰ $A\beta_{42}$ was measured using a pair of labeled antibodies, TAG-G2-11 and biotin-4G8; total $A\beta$ was measured using antibodies TAG-W02 and biotin-4G8. The electrochemiluminescence (ECL) signal was measured using an ECLM8 instrument (IGEN International, Inc.) according to the manufacturer's instructions.

From previously known SAR, the left side 3methoxyphenyl-4-methyl-1*H*-imidazole is required for GSM activity. Therefore, we focused our efforts on the SAR development of the right side of the molecule. The SAR is summarized in Table 1. When R^2 is *N*-methylamino, the A β_{42} inhibition activity is only 12 μ M with no selectivity (14h, clogD = 1.07, pKa = 8.10). It is known that increasing lipophilicity often results in improved in vitro activity and improved blood brain barrier permeability.³¹ Controlling the pKa was a successful approach to improve the CNS penetration.³² We then introduced fluorine to modulate both lipophilicity and basicity of the target molecules. Substitution with Ntrifluoroethylamino led to improved A β_{42} inhibition activity and selectivity (464 nM and 43-fold for 14i, clogD = 2.77, pKa = 5.42). When the *N*-trifluoroethylamino (14i) was replaced by an *N*-para-fluorobenzylamino (14e, clogD = 3.71, pKa = 7.08), the A β_{42} activity of **14e** increased more than 3 fold. This result suggests that the phenyl ring can increase activity. When the *N*-cyclohexanamino (14j, clogD = 2.86, pKa = 8.42) was replaced by an N-(4-fluorophenyl) amino, the $A\beta_{42}$ inhibition activity and selectivity are greatly improved (39 nM and 499-fold for 14a, clogD = 4.20, pKa = 5.33). This result is probably due to the strong electron withdrawing effect and the lipophilic effect of the fluorine introduced on the phenyl ring. When a tertiary amino (14f) or N-alpha-methyl parafluorobenzylamino was introduced (14g), the compounds lost some activity when compared to 14e. When nitrogen was replaced by oxygen (17a, b and 18) the compound also lost



some activity relative to **14e**. Nevertheless, we found that the enantiomerically pure compounds **14a** and **14c** are two compounds in the series with both good activity and high selectivity. To this point, the double bond and the lactam carbonyl present in **1** was successfully replaced by a tetrahydrobenzisoxazole ring. The new GSM compounds, **14a** and **14c**, are two-fold more active than GSM **1**, and have similar activity to **2** and **3**. Additionally, **14a** and **14c** are more selective than **1** and **3**, and have similar selectivity to **2**.

Table 1. Cellular IC₅₀ of Tetrahydrobenzisoxazole Analogs



Comp- ound	\mathbf{R}^{1}	R^2	$\begin{array}{c} A\beta_{42} \ IC_{50}, \\ (\mu M)^{*} \end{array}$	$A\beta_{tot}/A\beta_{42}$
14a, b	Н		3.9×10 ⁻² (14a)	499
			0.23 (14b)	86
14c, d	Н		2.9×10 ⁻² (14c)	575
			0.64 (14d)	28
14e	Н		0.13	32
14f	Н		0.25	79
14g	Н		0.26	77
14h	Н		1.2×10	2
14i	Н		0.46	43
14j	Н		0.31	64

15a	ОН	0.19	105
15b	ОН	0.15	102
16	Н	0.11	178
17a	Н	0.29	68
17b	Н	0.20	98
18	Н	0.51	39

* n = 3

Mass spectrometric analysis of A β peptides secreted by HEK293 cells carrying the London-Swedish double mutation after treatment with vehicle (DMSO) or 10 μ M solutions of compounds **14a**, **14c**, **14e** is shown in Figure 2. As expected for γ -secretase modulators, compounds **14a**, **14c**, and **14e** blocked the production of A β_{42} and A β_{40} , while the relative proportion of A β_{38} and A β_{37} is increased in comparison to A β_{40} (numbers in spectra are amyloid- β numbering). The A β profile in conditioned medium was analyzed using surface enhanced laser desorption/ionization (SELDI) mass spectrometry as described previously.³³

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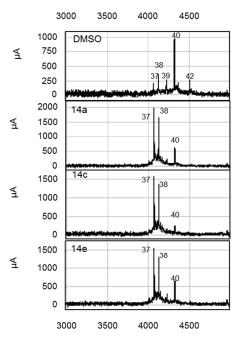


Figure 2. Mass spectrometric analysis of Aß profile

Compounds **14a**, **14c**, **14d** and **15a** were then tested *in vivo* rat model. Compound **14a** significantly reduced cerebrospinal fluid (CSF) $A\beta_{42}$ by 58% (p = 0.0066, n = 5) 3h after a single oral dose at 30 mg/kg in rats compared to the vehicle-treated animals (Figure 3).

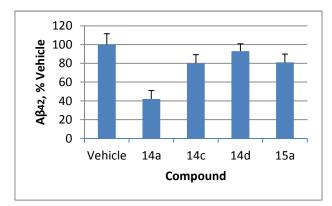


Figure 3. CSF $A\beta_{42}$ reduction in rat^{*}

* Measured 3 h after an acute oral dosing at 30 mg/kg compared to the vehicle

Compound **14a** was the most efficacious among compounds tested, albeit it was not as potent as **14c** *in vitro*. To get an insight into this discrepancy, plasma and brain drug levels were assessed in the rat tissues collected after dosing. We concluded that the efficacy of **14a** was driven by its excellent penetration into the CNS compartment, which provided a ~6fold higher exposure compared to **14c** (Table 2). This conclusion was supported by the fact that **14c** is a PGP efflux substrate ($P_{a-b} = 68 \text{ nm/s}$, $P_{b-a} = 462 \text{ nm/s}$, ER = 6.8). The efflux ratio of **14c** is 4-fold higher than **14a** ($P_{a-b} = 376 \text{ nm/s}$, $P_{b-a} = 600 \text{ nm/s}$, and ER= 1.6). The higher efflux ratio of **14c** could be the cause for its low brain penetration and poor *in vivo* efficacy.

	Plasma Concentration	Brain Concentration
ound	(µM)*	(µM)*
14a	10.4	14.3
14c	6.01	2.19

"measured 3 h after the dosing (n = 3)

In summary, we designed and synthesized a series of GSM compounds containing a unique tetrahydrobenzisoxazole ring. The unique tetrahydrobenzisoxazole ring successfully replaced the double bond in previously published GSMs. This series contained several compounds which are potent GSMs with good to excellent selectivity. Specifically, **14a** and **14c**, are two-fold more active than **1** and have similar activity to GSM **2** and **3**. In addition, **14a** and **14c** are more selective than GSM **1** and **3**, and have similar selectivity to **2**. Selected compounds were tested *in vivo*. Among them, compound **14a** significantly reduced CSF A β_{42} by 58% after 3 h, dosed orally at 30 mg/kg in rats compared to vehicle-treated animals.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website.

Biological assay protocols, general experimental descriptions, procedures and characterization of final compounds. (File type, PDF)

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ABBREVIATIONS

Abbreviations: AD, Alzheimer's disease; A β , amyloid β ; ; AE, adverse effects; APP, amyloid precursor protein; BACE, β -amyloid cleaving enzyme; CSF, cerebrospinal fluid; DEAD, diethyl azodicarboxylate; GSM, γ -secretase modulators; GSI, γ -secretase inhibitor; NCS, N-chlorosuccinimide; TMS, trimethylsilyl.

Notes

The authors are or were employees of Merck.

REFERENCES



2	
3	
4	
5	(1) Alzheimer's Association. 2017 Alzheimer's Disease Facts and Figures. Alzheimer's & Dementia 2017; 13, 325-373.
6	(2) Esparza, T.J.; Zhao. H.; Cirrito, J.R.; Cairns, N. J.; Bateman, R. J.; Holtzman, D. M.; Brody, D. L. Amyloid-beta Oligomerization in
7	Alzheimer Dementia vs. High Pathology Controls. Ann Neurol. 2013, 73(1), 104–119.
8	(3) Sadigh-Eteghad, S.; Sabermarouf, B.; Majdi, A.; Talebi, M.; Farhoudi, M.; Mahmoudi, J. Amyloid-Beta: A Crucial Factor in
	Alzheimer's Disease. <i>Med. Princ. Pract.</i> 2015 , 24(1), 1–10.
9	(4) Kreft, A. F.; Martone, R.; Porte, A. Recent Advances in the Identification of γ -Secretase Inhibitors to Clinically Test the A β Oligomer
10	Hypothesis of Alzheimer's Disease. J. Med. Chem. 2009, 52, 6169–6188.
11	(5) Irvine, G. B.; El-Agnaf, O. M.; Shankar, G. M.; Walsh, D. M.; Protein Aggregation in the Brain: The Molecular Basis for Alzheimer's
12	and Parkinson's Diseases, <i>Mol Med.</i> 2008, 14(7-8): 451–464.
13	(6) Gu, L. and Guo, Z.; Alzheimer's Aβ42 and Aβ40 Peptides form Interlaced Amyloid Fibrils, J. Neurochem. 2013, 126(3), 305–311.
14	(7) Chen, Y. and Glabe, C. G. Distinct Early Folding and Aggregation Properties of Alzheimer Amyloid-β Peptides Aβ40 and Aβ42. J. of
15	<i>Biological Chem.</i> 2006 , 281(34), 24414-24422.
16	(8) Jonsson, T.; Atwal, J. K.; Steinberg, S.; Snaedal, J.; Jonsson, P. V.; Bjornsson, S.; Stefansson, H.; Sulem, P.; Gudbjartsson, D.; Malo-
17	ney, J.; Hoyte, K.; Gustafson, A.; Liu, Y.; Lu, Y.; Bhangale, T.; Graham, R. R.; Huttenlocher, J.; Bjornsdottir, G.; Andreassen, O. A.;
18	Joensson, E. G.; Palotie, A.; Behrens, T. W.; Magnusson, O. T.; Kong, A.; Thorsteinsdottir, U.; Watts, R. J.; Stefansson, K. A Mutation in
19	APP Protects Against Alzheimer's Disease and Age-Related Cognitive Decline. Nature 2012, 488, 96–99.
20	(9) Yan, R.; Vassar, R. Targeting the β-Secretase BACE1 for Alzheimer's Disease Therapy. <i>Lancet Neurol.</i> 2014 , <i>13</i> , 319-329
20	(10) Choi, S. H.; Norstrom, E. Moderate Reduction of γ-Secretase: Is There a Therapeutic Sweet Spot? J. Neurosci. 2007, 27, 13579-
22	(11) Bursavich, M. G.; Harrison, B. A.; Blain, J-F. Gamma Secretase Modulators: New Alzheimer's Drugs on the Horizon? J. Med. Chem.
23	2016 , 59, 7389-7409. (12) Xia, W.; Wong, S. T.; Hanlon, E.; Morin, P. γ-Secretase Modulator in Alzheimer's Disease: Shifting the End. J. Alzheimer's Dis.
24	(12) Ala, W., Wong, S. T., Hamon, E., Morin, T. y-Secretase Modulator in Alzheimer's Disease. Sinthing the End. J. Alzheimer's Dis. 2012 , 31, 685-696.
25	(13) Weggen, S., Eriksen, J. L., Das, P., Sagi, S. A., Wang, R., Pietrzik, C. U., Findlay, K. A., Smith, T. E., Murphy, M. P., Bulter, T.,
26	Kang, D. E., Marquez-Sterling, N., Golde, T. E., and Koo, E. H. A Subset of NSAIDs Lower Amyloidogenic Aβ42 Independently of Cy-
27	clooxygenase Potency. <i>Nature</i> 2011 , 414, 212–216.
28	(14) Crump, C. J.; Johnson, D. S. Li, Y-M.; Development and Mechanism of γ -Secretase Modulators for Alzheimer's Disease, <i>Biochemis</i> -
29	<i>try</i> 2013 , 52, 3197 – 3216.
30	(15) Joo, Y.; Kim, HS.; Woo, RS.; Park, C. H.; Shin, KY.; Lee, JP.; Chang, KA.; Kim, S.; Suh, YH. Mefenamic Acid Shows
31	Neuroprotective Effects and Improves Cognitive Impairment in in vitro and in vivo Alzheimer's Disease Models. Mol. Pharmacol. 2006,
32	69, 76–84.
33	(16) Davis, K. L. NSAID and Alzheimer's Disease; Possible Answers and New Questions. Mol. Psychiatry 2002, 7, 925–926.
34	(17) Kimura, T.; Kawano, K.; Doi, E.; Kitazawa, N.; Shin, K.; Miyagawa, T.; Kaneko, T.; Ito, K.; Takaishi, M.; Sasaki, T.; Hagiwara, H.
35	Preparation of Cinnamide, 3-Benzylidenepiperidin-2-one, Phenylpropynamide Compounds as Amyloid β Production Inhibitors. PCT Int.
36	<i>Appl.</i> 2005 , WO 2005115990 A1 20051208.
37	(18) Sun, ZY.; Asberom, T.; Bara, T.; Bennett, C.; Burnett, D.; Chu, I.; Clader, J.; Cohen-Williams, M.; Cole, D.; Czarniecki, M.;
38	Durkin, J.; Gallo, G.; Greenlee, W.; Josien, H.; Huang, X.; Hyde, L.; Jones, N.; Kazakevich, I.; Liu, X.; Lee, J.; MacCoss, M.;
	Mandal, M. B.; McCracken, T.; Nomeir, A.; Mazzola, R.; Palani, A.; Parker, E. M.; Pissarnitski, D. A.; Qin, J.; Song, L.; Terracina, G.;
39	Vicarel, M.; Voigt, J.; Xu, R.; Zhang, L.; Zhang, Q.; Zhao, Z.; Zhu, X.; Zhu, Z. Cyclic Hydroxyamidines as Amide Isosteres: Discovery of Oreglians and Oreglians and Uichly Effection on Secretary Medulators in vivo. 1 Med. Chem. 2012, 55, 480, 502
40	Oxadiazolines and Oxadiazines as Potent and Highly Efficacious γ-Secretase Modulators <i>in vivo. J. Med. Chem.</i> 2012 , 55, 489–502. (19) Huang, X.; Zhou, W.; Liu, X.; Li, H.; Sun, G.; Mandal, M.; Vicarel, M.; Zhu, X.; Bennett, C.; McCraken, T.; Pissarnitski, D.; Zhao,
41	Z.; Cole, D.; Gallo, G.; Zhu, Z.; Palani, A.; Aslanian, R.; Clader, J.; Czarniecki, M.; Greenlee, W.; Burnett, D.; Cohen-Williams, M.;
42	L; Cole, D., Galo, G., Zhu, Z., Falan, A., Asianan, K., Cladel, J., Czamecki, M., Oreemec, w., Burnett, D., Cohen-winnans, M., Hyde, L.; Song, L.; Zhang, L.; Chu, I.; and Buevich. A. Synthesis and SAR Studies of Fused Oxadiazines as γ-Secretase Modulators for
43	Treatment of Alzheimer's Disease. ACS Med. Chem. Lett. 2012, 3, 931–935.
44	(20) Haugabook, S. J.; Yager, D. M.; Eckman, E. A.; Golde, T. E.; Younkin, S. G.; Eckman, C. B. High Throughput Screens for the Identi-
45	fication of Compounds That Alter the Accumulation of The Alzheimer's Amyloid Beta Peptide (Abeta). J. Neurosci. Methods 2001, 108,
46	171–179.
47	(21) Findeis, Mark A.; Schroeder, F. C.; Creaser, S. P.; McKee, T. D. and Xia, W. Natural Product and Natural Product-Derived Gamma
48	Secretase Modulators from Actaea Racemosa Extracts. Medicines 2015, 2, 127 -140.
49	(22) Huang, X.; Aslanian, R.; Zhou, W.; Zhu, X.; Qin, J.; Greenlee, W.; Zhu, Z.; Zhang, L.; Hyde, L.; Chu, I.; Cohen-Williams, M.; Palani,
50	A. The Discovery of Pyridone and Pyridazone Heterocycles as γ-Secretase Modulators. ACS Med. Chem. Lett. 2010, 1, 184–187.
51	(23) Qin, J.; Zhou, W.; Huang, X.; Dhondi, P.; Palani, A.; Aslanian, R.; Zhu, Z.; Greenlee, W.; Cohen-Williams, M.; Jones, N.; Hyde, L.;
52	Zhang, L. Discovery of a Potent Pyrazolopyridine Series of γ-Secretase Modulators. ACS Med. Chem. Lett. 2011, 2, 471–476.
52 53	(24) Qin, J.; Dhondi, P.; Huang, X.; Mandal, M.; Zhao, Z.; Pissarnitski, D.; Zhou, W.; Aslanian, R.; Zhu, Z.; Greenlee, W.; Clader, J.;
	Zhang, L.; Cohen-Williams, M.; Jones, N.; Hyde, L.; Palani, A. Discovery of Fused 5,6-Bicyclic Heterocycles as γ-Secretase Modulators.
54	Bioorg. Med. Chem. Lett. 2011, 21, 664–669.
55	(25) Caldwell, J. P.; Bennett, C. E.; McCracken, T. M.; Mazzola, R. D.; Bara, T.; Buevich, A. V.; Burnett, D. A.; Chu, I.; Cohen-Williams,
56	M.; Jones, N. T.; Josien, H.; Hyde, L. A.; Lee, J.; McKittrick, B.; Song, L.; Terracina, G.; Voigt, J. H.; Zhang, L.; Zhu, Z. Iminoheterocy-
57	cles as Gamma-Secretase Modulators. <i>Bioorg. Med. Chem. Lett.</i> 2010 , 20, 5380–5384.
58	(26) Zhu, Z.; Greenlee, W. J.; Li, H.; Vicarel, M. L.; Qin, J.; Dhondi, P. K.; Huang, X.; Palani, A.; Liu, X.; Sun, ZY.; Josien, H. B.; Xu,

(26) Zhu, Z.; Greenlee, W. J.; Li, H.; Vicarel, M. L.; Qin, J.; Dhondi, P. K.; Huang, X.; Palani, A.; Liu, X.; Sun, Z.-Y.; Josien, H. B.; Xu, R.; Cole, D. J.; Burnett, D. A.; Bennett, C. E.; Mccracken, C. M.; Maccoss, M. Heterocyclic compounds as gamma-secretase modulators and their preparation and use in the treatment of CNS diseases. *PCT Int. Appl.* **2010**, WO 2010056722 A1 20100520.



59 60

- (27) Zhu, Z.; Greenlee, W. J.; Cole, D. J.; Pissarnitski, D. A.; Gallo, G. V.; Li, H.; Josien, H. B.; Qin, J.; Knutson, C. E.; Mandal, M.; Vicarel, M. L.; Rajagopalan, M.; Dhondi, P. K.; Xu, R.; Sun, Z.-Y.; Bara, T. A.; Huang, X.; Zhu, X.; Zhao, Z.; Clader, J. W.; Palani, A.;
- Asberom, T.; Mccracken, T.; Bennett, C. E. Gamma Secretase Modulators. *PCT Int. Appl.* **2010**, WO 2010054078 A1 20100514.
- (28) Greenlee, W. J.; Pissarnitski, D. A.; Zhao, Z.; Zhu, Z. Gamma Secretase Modulators. *PCT Int. Appl.* **2013**, WO 2013066740 A1 20130510.
- (29) Sakai, T.; Miyata,K.; Utaka,M.; and Takeda, A. Me₃SiCl-NaI-CH₃CN as an Efficient and Practical Reducing Agent for Benzylic Alcohols. *Tetrahedron Lett.* **1987**, 28(33), 3817-3818.
- (30) Please see the supporting information for details.
- (31) Rankovic, Z. CNS Drug Design: Balancing Physicochemical Properties for Optimal Brain Exposure, *J. Med. Chem.* 2015, 58, 2584-2608.
- (32) Charifson, S. P. and Walters, W. P. Acidic and Basic Drugs in Medicinal Chemistry. J. Med. Chem. 2014, 57, 9701–9717.
- (33) Lee, J.; Song, L.; Terracina, G.; Bara, T.; Josien, H.; Asberom, T.; Sasikumar, T. K.; Burnett, D. A.; Clader, J.; Parker, E. M.; Zhang, L. Identification of Presenilin 1-Selective γ-Secretase Inhibitors With Reconstituted γ-Secretase Complexes. *Biochemistry* 2011, 50, 4973–4980.



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 $\begin{array}{l} A\beta_{42} \mbox{ IC}_{50} = 39 \mbox{ nM}; \mbox{ } A\beta_{Total} / A\beta_{42} = 499 \\ 58\% \mbox{ reduction of rat CSF } A\beta_{42} \mbox{ at 30 mpk} \\ \mbox{ oral dosing} \end{array}$