## FLAVONOIDS OF Juniperus zeravschanica

## M. P. Yuldashev<sup>1</sup> and L. Kh. Rasulova<sup>2</sup>

Isoquercitrin and the new flavonglycoside zeravschanoside, 5,6,8,3',4'-pentahydroxy-7-O- $\beta$ -D-glucopyranosylflavone, the structure of which was established from chemical transformations and spectral data, were isolated from Juniperus zeravschanica Kom.

Key words: Juniperus zeravschanica, isoquercitrin, 5,6,8,3',4'-pentahydroxy-7-O- $\beta$ -D-glucopyranosylflavone.

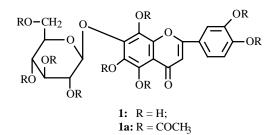
Many species of *Juniperus* L. (Cupressaceae) are rich sources of flavonoids [1] and are widely used in folk and official medicine [2].

Zeravschan juniper (*Juniperus zeravschanica* Kom.) is a dioecious evergreen tree 20 m in height that grows on rocky, gravelly, and thinly soiled slopes of mountains in Central Asia [3].

Compounds 1 and 2 were isolated from the alcohol extract of fruit collected in autumn on mountain slopes near Kumyshkan (Tashkent district).

The UV spectrum ( $\lambda_{max}$ , nm, 276, 343) of the new flavonoid **1** is characteristic of flavone derivatives [4]. The PMR of **1** contains signals for H-3, H-5', H-6', H-2', an anomeric proton, and other protons of a carbohydrate. The chromatographic mobility and PMR spectrum are consistent with a glycoside. This is confirmed by formation of 5,6,7,8,3',4'-hexahydroxyflavone and D-glucose monosaccharide upon acid hydrolysis of **1**. The aglycone of **1** gives a positive qualitative reaction with SrCl<sub>2</sub>, which indicates the presence of *ortho*-dihydroxyls in the C-5 and C-6 positions [5], and a positive gossypetin test, which is consistent with hydroxyls on C-5 and C-8 [6].

Acetylation of **1** gave the nonaacetyl derivative **1a**,  $C_{39}H_{38}O_{22}$ , the mass spectrum of which exhibits a peak for the molecular ion with m/z 858 and strong peaks for fragments of tetraacetylhexose with m/z 331, 329, 271, and 169 [7] and the aglycone with m/z 318.



The absence of a bathochromic shift in the UV spectrum of **1** in the presence of NaOAc indicates that the 7-OH of the aglycone was glycosylated [4]. The anomeric proton of D-glucose resonates at 5.53 ppm as a doublet with SSCC 7.0 Hz. Therefore, there is a  $\beta$ -glycosidic bond between the carbohydrate and the aglycone [4].

Thus, **1** has the structure 5,6,8,3',4' -pentahydroxy-7-O- $\beta$ -D-glucopyranosylflavone.

Compound **2** is a flavonol glycoside according to spectral data. Acid hydrolysis produces quercetin (3,5,7,3',4'- pentahydroxyflavone) [1, 8] and D-glucose. The PMR, UV, and mass spectra, chemical transformations, and comparison of the physicochemical properties with those in the literature identify **2** as isoquercitrin (quercetin-3-O- $\beta$ -D-glucoside) [1, 8].

<sup>1)</sup> S. Yu. Yunusov Institute of the Chemistry of Plant Substances, Academy of Sciences of the Republic of Uzbekistan, Tashkent, fax (99871) 120 64 75; 2) Tashkent Pharmaceutical Institute, Tashkent, fax (99871) 256 08 18. Translated from Khimiya Prirodnykh Soedinenii, No. 3, pp. 195-196, May-June, 2001. Original article submitted June 25, 2001.

## EXPERIMENTAL

We used solvent systems CHCl<sub>3</sub>—CH<sub>3</sub>OH (9:1) (1) and *n*-butanol—pyridine—water (6:4:3) (2).

Preparative TLC used LSL 5/40 µm silica gel (Chemapol, Czech Rep.). TLC was performed on Silufol UV-254 plates. Compounds were visualized on TLC in UV light using ammonia vapor. Sugar was detected on PC (Filtrak No. 12) by spraying with acidic anilinium phthalate with heating for 3-5 min at 90-100°C.

Conditions for recording spectra have been published [9].

**Extraction and Isolation of Flavonoids.** Dried and ground fruit (0.2 kg) of zeravschan juniper that was collected in autumn 1996 on slopes near Kumyshkan (Tashkent) was extracted with ethanol (95%) in a Soxhlet apparatus for 4 h. The alcohol extract was condensed until dry. Then, recrystallization from methanol gave **1** (0.09 g). Preparative TLC of the mother liquor in system 1 gave **2** (0.07 g).

**Zeravschanoside (1).**  $C_{21}H_{20}O_{13}$ , mp 328-330°C (methanol). UV spectrum (EtOH,  $\lambda_{max}$ , nm): 276, 343; CH<sub>3</sub>CO<sub>2</sub>Na 275, 345. IR spectrum (KBr,  $v_{max}$ , cm<sup>-1</sup>): 3445 (OH), 1665 (C=O of  $\gamma$ -pyrone), 1600, 1560, 1510 (aromatic C=C), 1095, 1030 (C–O of glycosides).

PMR spectrum ( $\delta$ , ppm, J/Hz, C<sub>5</sub>D<sub>5</sub>N): 3.50-4.50 (carbohydrate protons), 5.53 (1H, d, J = 7.0, H-1"), 6.86 (1H, s, H-3), 7.12 (1H, d, J = 9.0, H-5'), 7.63 (1H, dd, J = 9.0, H-6'), 8.06 (1H, d, J = 2.0, H-2').

**Zeravschanoside Nonaacetate (1a).** Glycoside **1** (15 mg) was dissolved in pyridine (1 mL) and acetic anhydride (2 mL), worked up as usual after 3 h, and recrystallized from ethanol to produce **1a** (14 mg),  $C_{39}H_{38}O_{22}$ , mp 248-250°C. Mass spectrum, *m/z*: M<sup>+</sup> 858, 331, 329, 318, 271, 169, 109, etc.

Acid Hydrolysis of 1. Glycoside 1 (18 mg) was hydrolyzed by aqueous-methanolic HCl (20 mL, 5%) for 4 h on a boiling-water bath. Then, the methanol was evaporated in vacua. The precipitated aglycone was filtered off and recrystallized from methanol. Yield of 5,6,7,8,3',4'-hexahydroxyflavone, 8 mg, mp >360°C,  $C_{15}H_{10}O_8$  (M<sup>+</sup> 318). D-Glucose was observed in the hydrolysate by PC using system 2.

**Isoquercitrin 2.**  $C_{21}H_{20}O_{12}$ , mp 238-239°C. UV spectrum (EtOH,  $\lambda_{max}$ , nm): 255, 265<sup>\*</sup>, 362. IR spectrum (KBr,  $v_{max}$ , cm<sup>-1</sup>): 3450 (OH), 1660 (C=O of  $\gamma$ -pyrone), 1618, 1575, 1518 (aromatic C=C), 1075, 1026, 1009 (C–O of glycosides). PMR spectrum ( $\delta$ , ppm, J/Hz, C<sub>5</sub>D<sub>5</sub>N): 3.65-4.44 (sugar protons), 5.47 (1H, d, J = 7.0, H-1"), 6.51 (1H, d, J = 2.0, H-6), 6.59 (1H, d, J = 2.0, H-8), 7.21 (1H, d, J = 8.5, H-5'), 7.86 (1H, dd, J = 2.0, J = 8.5, H-6'), 8.11 (1H, br.s, H-2'), 13.78 (1H, br.s, 5-OH).

Acid hydrolysis of **2** (5% HCl, 4 h) formed D-glucose and quercetin (3,5,7,3',4'-pentahydroxyflavone),  $C_{15}H_{10}O_7$  (M<sup>+</sup> 302), mp 313-315°C, UV spectrum (MeOH,  $\lambda_{max}$ , nm): 257, 268, 371; CH<sub>3</sub>CO<sub>2</sub>Na 270, 405.

Acetylation of 2 (acetic anhydride and pyridine) gave the octaacetate, mp 200-202°C (M<sup>+</sup> 770 and peaks for fragments of tetraacetylhexose with m/z 331, 271, and 169).

## REFERENCES

- 1. L. K. Klyshev, V. A. Bandyukova, and L. S. Alyukina, *Plant Flavonoids* [in Russian], Nauka, Alma-Ata (1978).
- 2. Kh. Kh. Kholmatov, I. A. Kharlamov, and P. K. Alimbaeva, *Principal Medicinal Plants of Middle Asia* [in Russian], Meditsina, Tashkent (1984).
- 3. Flora of Uzbekistan [in Russian], Tashkent (1941), Vol. 1, p. 111.
- 4. T. J. Mabry, K. R. Markham, and M. B. Thomas, *The Systematic Identification of Flavonoids*, Springer-Verlag, Berlin (1970), p. 146, 329.
- 5. Y. Kikuchi, Y. Miyaichi, Y. Yamaguchi, H. Kisu, and T. Tomimori, *Chem. Pharm. Bull.*, **39**, 1047 (1991).
- 6. K. R. Markham, *Isolation Techniques for Flavonoids*, in: *The Flavonoids*, J. B. Harborne, ed., Chapman and Hall, London (1975), p. 2-44.
- 7. N. K. Kochetkov and O. S. Chizhov, *Mass Spectrometry of Carbohydrates* [in Russian], in: *Methods of Carbohydrate Research* [in Russian], Mir, Moscow (1975).
- 8. Sh. V. Abdullaev, A. Sattikulov, E. Kh. Batirov, Yu. V. Kurbatov, and V. M. Malikov, *Khim. Prir. Soedin.*, 104 (1983).
- 9. M. P. Yuldashev, *Khim. Prir. Soedin.*, 193 (2001).