The synthesis of estriol 16- and 17monoglucuronide from estriol

Yinqiu Wu and Leonard F. Blackwell Ph.D.

Department of Chemistry & Biochemistry, Massey University, Palmerston North, New Zealand

An efficient and convenient procedure for the synthesis of estriol 16- and 17-monoglucuronides from estriol is described. This is achieved by the selective protection and deprotection of the hydroxy groups in estriol, Koenigs-Knorr reactions with methyl 1-bromo-1-deoxy-2,3,4-tri-O-acetyl- α -D-glucopyranuronate and subsequent hydrolysis. The products have been characterized by proton nuclear magnetic resonance (¹H NMR), two-dimensional ¹H homonuclear shift-correlated spectra (2D-COSY) and mass spectra. The selective Koenigs-Knorr reaction of the alcoholic hydroxyl group in the presence of a phenolic hydroxyl group is also reported. (Steroids **58:**452–456, 1993)

Keywords: steroids; estriol; estriol 16- and 17-glucuronides; selective protection; Koenigs-Knorr reaction; synthesis

Introduction

As part of our current research program into the development of enzyme immunoassay systems for the home measurement of steroid glucuronides,¹ we require a supply of estriol monoglucuronides. Estriol 16,17-diacetate is easily prepared from estriol by acetylation with acetic anhydride (Ac₂O) and a catalytic amount of boron trifluoride etherate in tetrahydrofluran (THF) at room temperature,² or in our laboratory* by simply refluxing in acetic acid (HAc) catalyzed with copper(II) acetate monohydrate.³ Hence estriol 3-glucuronide is comparatively easy to synthesize after coupling of estriol 16,17-diacetate with methyl 1-bromo-1-deoxy-2,3,4-tri-O-acetyl- α -D-glucopyranuronate and subsequent hydrolysis. However, estriol 16- or 17-glucuronides are less readily available because of the difficulty in their synthesis. The usual synthesis of estriol 16and 17-glucuronides from estrone involves many steps including the glucuronidation of 16α -hydroxy-17-oxoderivatives and reduction of the 17-carbonyl group. The reduction of the 17-carbonyl group often produces significant quantities of the 17α -isomer, which significantly decreases the yield. For example, during synthesis of estriol 16-glucuronide from estrone according to a literature procedure,⁴ we obtained about 30% of methyl 3,17 α -dihydroxyl-1,3,5(10)-estratrien-16 α yl-2,3,4-tri-O-acetyl- β -D-glucopyranosiduronate (17 α isomer)† during reduction of the 17-carbonyl group.

We now report an alternative method for the synthesis of either estriol 16- or 17-glucuronide from estriol based on the selective protection and deprotection of the hydroxyl groups, glucuronidation, and hydrolysis. The methodology provides a simple procedure and good yields of pure products, which have been characterized by spectroscopy.

Experimental

Melting points (mp) were determined on a Bausch and Lomb microscope hot plate and are uncorrected. The infrared spectra (IR) were obtained on a Bio-Rad FTS spectrophotometer. Proton nuclear magnetic resonance (¹H NMR) and 2D-COSY spectra were measured on a JEOL GX270 spectrometer (270 MHz) using CDCl₃ or CD₃OD as solvent and tetramethylsilane as internal standard. Values are given in ppm (δ) (s, single, d, double, t,

Address reprint requests to Dr. Leonard F. Blackwell, Department of Chemistry & Biochemistry, Massey University, Palmerston North, New Zealand.

Received January 15, 1993; accepted May 28, 1993.

^{*,} In accordance with reference 3, we refluxed estriol in acetic acid catalyzed with copper(II) acetate monohydrate to get estriol 16α , 17β -diacetate in 60% yield. IR ν_{max} 3,414 cm⁻¹ (3-OH), 1,742 and 1,713 cm⁻¹ (C=O, C-16, 17-diacetate); ¹H NMR (CDCl₃) δ , 0.84 (s, 3H, 18-CH₃), 2.06 (s, 3H, OCOCH₃), 2.10 (s, 3H, OCOCH₃), 5.00 (d, 1H, J = 5.59 Hz, 17\alpha-H), 5.18 (m, 1H, 16 β -H), 5.36 (s, 1H, 3-OH), 6.57–7.14 (m, 3H, aromatic).

^{†,} The reduction of methyl 2,4-dibromo-17-oxo-1,3,5(10)-estratrien-16α-yl-2,3,4-tri-O-acetyl-β-D-glucopyranosiduronate gave 30% of methyl 3,17α-dihydroxyl-1,3,5(10)-estratrien-16α-yl-2,3,4-tri-O-acetyl-β-D-glucopyranosiduronate (17α-isomer). High EIMS m/z, 604.2563 (M)⁺ (calculated for $C_{31}H_{40}O_{12}$, 604.2520); ¹H NMR 8, 0.72 (s, 3H, 18-CH₃), 2.04 (s, 6H, 2OCOCH₃), 2.06 (s, 3H, OCOCH₃), 3.73 (d, 1H, J = 4.39 Hz, 17β-H), 3.77 (s, 3H, COOCH₃), 4.07 (d, 1H, J = 4.39 Hz, 5'-H), 4.38 (m, 1H, 16β-H), 4.46 (d, 1H, J = 8.06 Hz, 1'-H), 5.05 (t, 1H, 2'-H), 5.18-5.33 (m, 2H, 3' and 4'-H), 6.55-7.17 (m, 3H, aromatic).





(a) Selective protection and deprotection of estriol and its derivatives

16α - (*tert* - Butyldimethylsilyloxy) - 1,3,5(10) - estratrien - 3,17βdiol (2) and 3,17β-diacetoxy-16α-(*tert*-butyldimethylsilyloxy)-1,3,5(10)-estratrien (3). Compounds 2 and 3 were prepared from estriol 1 by selective protection of the 16α-hydroxyl group with *tert*-butyldimethylsilyl chloride, followed by acetylation with Ac₂O and pyridine according to established literature procedures⁵ in 82% and 93% yield, respectively. For compound 2 (white solid), mp, 193–194 C; reported mp, 194 C (lit. 5). IR ν_{max} 3,563 cm⁻¹ (OH), 3,316 cm⁻¹ (OH); ¹H NMR (CDCl₃) δ , 0.06 (s, 6H, -SiMe₂), 0.73 (s, 3H, 18-CH₃), 0.87 (s, 9H, -SiCMe₃), 3.30 (s, 1H, 17β-OH), 3.46 (d, 1H, J = 5.49 Hz, 17α-H), 4.06 (m, 1H, 16β-H), 4.66 (s, 1H, 3-OH), 6.49–7.08 (m, 3H, aromatic); For compound 3 (fine crystals) mp, 110–112 C; (no melting point

Synthesis of estriol 16- and 17-glucuronides: Wu and Blackwell

reported⁵). IR ν_{max} 1,766 and 1,744 cm⁻¹ (C=O, C-3,17-diacetate); ¹H NMR (CDCl₃) δ , 0.03 (d, 6H, -SiMe₂), 0.77 (s, 3H, 18-CH₃), 0.88 (s, 9H, -SiCMe₃), 2.09 (s, 3H, OCOCH₃), 2.28 (s, 3H, OCOCH₃), 4.30 (m, 1H, 16 β -H), 4.84 (d, 1H, J = 5.59 Hz, 17 α -H), 6.78-7.28 (m, 3H, aromatic).

3-Acetoxy-16a-(tert-butyldimethylsilyloxy)-1,3,5(10)-estratrien-17 β -ol (9). Protection of the 3-hydroxyl group in compound 2 was performed by an acetylation procedure similar to that used in the preparation of compound 3, expect that the reaction was stirred at 0 C (ice-cooled bath)² for 0.5 hours instead of at room temperature for 40 hours. After extracting with ethyl acetate (EtOAc) (2 \times 20 ml), the organic layer was dried over magnesium sulfate (MgSO₄) overnight. Removal of solvent under reduced pressure afforded a white solid that was purified by flash chromatography using hexane/EtOAc (3:1) as eluent to give compound 9 as a white solid (48 mg, 87% yield). mp, 190-192 C; Analysis calculated: C₂₆H₄₀O₄Si · 1/2H₂O: C, 68.83; H, 9.11; Found: C, 68.93; H, 8.91; IR, ν_{max} 3,617 and 3,594 cm⁻¹ (17 β -OH), 1,754 cm⁻¹ (C = 0, 3-acetate); ¹H NMR (CDCl₃) δ , 0.06 (s, 3H, -SiMe), 0.07 (s, 3H, -SiMe), 0.75 (s, 3H, 18-CH₃), 0.89 (s, 9H, -SiCMe₃), 2.25 (s, 3H, 3-OCOCH₃), 3.44 (s, 1H, 17β-OH), 3.54 (d, J = 5.86 Hz, 1H, 17 α -H), 4.06 (m, 1H, 16 β -H), 6.75-7.26 (m, 3H, aromatic).

3,17 β **-Diacetoxy-1,3,5(10)-estratrien-16\alpha-ol (4).** A typical experimental procedure for the desilylation of the 16α -tertbutyldimethylsilyl group was as follows:⁶ Compound **3** (239 mg) was dissolved in 15 ml of solvent mixture (HAc/H₂O/THF, 3:1:1), and stirred for 40 hours at room temperature. After adding 100 ml of ether (Et₂O), the organic layer was separated, washed with ice-cooled NaHCO₃ and water, and dried over MgSO₄. Removal of solvent under reduced pressure afforded a white solid that was purified by flash chromatography using hexane/EtOAc (2:1) as eluent to give compound **4** as a white solid (177 mg, 97% yield). mp, 140 C; reported mp, 132 C (lit. 5). IR ν_{max} 3,503 cm⁻¹ (16 β -OH), 1,751 and 1,719 cm⁻¹ (C=O, 3,17-diacetate); ¹H NMR (CDCl₃) δ , 0.86 (s, 3H, 18-CH₃), 2.15 (s, 3H, OCOCH₃), 2.28 (s, 3H, OCOCH₃), 3.76 (s, 1H, 16 α -OH), 4.16 (m, 1H, 16 β -H), 4.28 (d, J = 4.48 Hz, 1H, 17 α -H), 6.80–7.28 (m, 3H, aromatic).

Methyl 3,16 α -dihydroxy-1,3,5(10)-estratrien-17 β -yl-2,3,4-tri-Oacetyl- β -D-glucopyranosiduronate (8). The desilylation reaction was performed by a procedure similar to that used in the preparation of compound 4. Compound 8 was obtained from compound 7 (220 mg) as colorless crystals (160 mg, 85% yield). mp, 216–218 C; reported mp, 207–212 C (lit. 7). Analysis calculated, C₃₁H₄₀O₁₂ · H₂O: C, 59.79; H, 6.79; Found: C, 59.71; H, 6.60; 'H NMR (CDCl₃) δ , 0.74 (s, 3H, 18-CH₃), 2.04 (s, 3H, OCOCH₃), 2.05 (s, 3H, OCOCH₃), 2.08 (s, 3H, OCOCH₃), 3.34 (d, J = 5.22 Hz, 1H, 17 α -H), 3.71 (s, 1H, 16 α -OH), 3.77 (s, 3H, COOCH₃), 4.11 (d, J = 9.33 Hz, 1H, 5'-H), 4.22 (m, 1H, 16 β -H), 4.59 (d, J = 7.83 Hz, 1H, 1'-H), 5.09 (t, 1H, 2'-H), 5.25–5.30 (m, 2H, 3' and 4'-H), 5.77 (s, 1H, 3-OH), 6.56–7.11 (m, 3H, aromatic); EIMS m/z, 604 (M)⁺.

Methyl 3-acetoxy- 16α -hydroxyl-1,3,5(10)-estratrien- 17β -yl-2,3,4-tri-O-acetyl- β -D-glycopyranosiduronate (11). The desilylation reaction was again performed by a procedure similar to that used in the preparation of 4. Compound 11 was obtained from compound 10 (50 mg) as a white solid (33 mg, 78% yield). mp, 225–227 C; reported mp, 222–224 C (lit. 8). ¹H NMR (CDCl₃) δ , 0.76 (s, 3H, 18-CH₃), 2.04 (s, 3H, OCOCH₃), 2.05 (s, 3H, OCOCH₃), 2.08 (s, 3H, OCOCH₃), 2.28 (s, 3H, 3-OCOCH₃), 3.34 (d, J = 5.13 Hz, 1H, 17 α -H), 3.49 (s, 1H, 16 α -OH), 3.78 (s, 3H,

Papers

COOCH₃), 4.12 (d, J = 9.53 Hz, 1H, 5'-H), 4.26 (m, 1H, 16 β -H), 4.60 (d, J = 8.06 Hz, 1'-H), 5.09 (t, 1H, 2'-H), 5.23-5.31 (m, 2H, 3'- and 4'-H), 6.78-7.27 (m, 3H, aromatic).

(b) Glucuronidation reactions

Methyl 3,17*B*-diacetoxy-1,3,5(10)-estratrien-16*a*-yl-2,3,4-tri-Oacetyl-B-D-glucopyranosiduronate (5). A typical experimental procedure for the coupling reaction of estriol 3.17β -diacetate 4 with methyl 1-bromo-1-deoxy-2,3,4-tri-O-acetyl-a-D-glucopyranuronate was as follows.9 Dried CdCO₃ (315 mg, 1.83 mmol) was added to a solution of compound 4 (170 mg, 0.46 mmol) in anhydrous toluene (50 ml), and the suspension was concentrated to approximately 30 ml by distillation to remove the moisture. After distillation, the solution of methyl 1-bromo-1-deoxy-2,3,4tri-O-acetyl-α-p-glucopyranuronate (725 mg, 1.83 mmol) in anhydrous toluene (20 ml) was added dropwise by syringe, keeping the addition rate the same as the rate of distillation. Finally, the whole reaction mixture was refluxed for 2.5 hours. The precipitate was then removed by filtration and washed with CH₂Cl₂. The filtrate and washings were combined and evaporated under reduced pressure. Purification of the oily residue by flash chromatography using hexane/EtOAc (1:1) as eluent gave compound 5 as a white solid (250 mg, 80% yield). mp, 170-173 C; Analysis calculated: $C_{35}H_{44}O_{14} \cdot 2/3H_2O$: C, 59.99; H, 6.52; Found: C, 59.96; H, 6.24; ¹H NMR (CDCl₃) 8, 0.75 (s, 3H, 18-CH₃), 2.02 (s, 3H, OCOCH₃), 2.03 (s, 3H, OCOCH₃), 2.05 (s, 3H, OCOCH₃), 2.06 (s, 3H, OCOCH₃), 2.28 (s, 3H, 3-OCOCH₃), 3.77 (s, 3H, COOCH₁), 3.96 (d, J = 9.70 Hz, 1H, 5'-H), 4.25 (m, 1H, 16 β -H), 4.56 (d, J = 7.46 Hz, 1H, 1'-H), 4.95-4.98 (m, 2H, 2' and 17α -H), 5.22-5.25 (m, 2H, 3' and 4'-H), 6.77-7.26 (m, 3H, aromatic).

Methyl 3-hydroxy-16a-(tert-butyldimethylsilyloxy)-1,3,5-(10)-estratrien - 17 B-yl - 2,3,4 - tri - O - acetyl - B - D - glucopyranosiduronate (7). The coupling reaction was performed by a procedure similar to that used in the preparation of 5, except that a 2:1 ratio of methyl 1-bromo-1-deoxy-2,3,4-tri-O-acetyl-a-D-glucopyranuronate to the compound 2 was used instead of a 4:1 mole ratio as for compound 4 and there was no refluxing time after addition of the methyl 1-bromo-1-deoxy-2,3,4-tri-O-acetyl- α -D-glucopyranuronate. Compound 7 was obtained from compound 2 (370 mg, 0.92 mmol) as a light-yellow solid (285 mg, 47%). Attempts to crystallize compound 7 were unsuccessful because the tert-butyldimethylsilyl group of compound 7 was easily removed in solvent methanol (MeOH) by heating gently and standing at room temperature for 24 hours. Hence compound 8 was obtained as pure crystals in this way. The IR, ¹H NMR, and melting point were identical with product 8 made by deprotection of 7 under weak acid conditions (HAc/H₂O/THF, 3:1:1).

Methyl 3-acetoxy-16 α -(*tert*-butyldimethylsilyloxy)-1,3,5-(10)-estratrien -17 β -yl-2,3,4-tri-O-acetyl- β -D-glucopyranosiduronate (10). The coupling reaction was performed by a procedure similar to that used in the preparation of 5. Compound 10 was obtained from compound 9 (45 mg, 0.10 mmol) as a white solid (53 mg, 69%). mp, 212–214 C; Analysis calculated, C₃₉H₅₆O₁₃Si: C, 61.55; H, 7.42; Found: C, 61.39; H, 7.37; ¹H NMR (CDCl₃) δ , 0.04 (s, 3H, -SiMe), 0.08 (s, 3H, -SiMe), 0.72 (s, 3H, 18-CH₃), 0.88 (s, 9H, -SiCMe₃), 2.02 (s, 3H, OCOCH₃), 2.03 (s, 3H, OCOCH₃), 2.06 (s, 3H, OCOCH₃), 2.28 (s, 3H, 3-OCOCH₃), 3.66 (d, J = 5.5 Hz, 17 α -H), 3.73 (s, 3H, COOCH₃), 3.98 (d, J = 9.89 Hz, 1H, 5'-H), 4.10 (m, 1H, 16 β -H), 4.64 (d, J = 7.7 Hz, 1'-H), 5.02 (t, 1H, 2'-H), 5.22–5.26 (m, 2H, 3' and 4'-H), 6.78–7.28 (m, 3H, aromatic). FABMS m/z, 761 (M)⁺.

(c) Hydrolysis

3.17B-Dihydroxy-1.3.5(10)-estratrien-16a-yl-B-D-glucopyranosiduronic acid (Estriol 16-glucuronide) (6). A typical hydrolysis experimental procedure for the preparation of estriol 16-glucuronide was as follows. NaOH (1 ml; 0.5 M) was added to the solution of compound 5 (48 mg, 0.07 mmol) in MeOH (10 ml), and stirred for 24 hours at room temperature. When TLC showed no starting material 5, the reaction mixture was titrated to pH = 8 and concentrated to approximately 3 ml by distillation under reduced pressure. On standing overnight a white solid (sodium salt) was obtained. The crystals were filtered and washed with a little MeOH, and dissolved in MeOH/H₂O (3:2). Titration to pH = 2.5 and standing overnight gave compound 6 as fine crystals (free acid) (27 mg, 65%). mp, 223-224 C; reported mp, 223-224 C (lit. 10); Analysis calculated: $C_{24}H_{32}O_9 \cdot 4H_2O$: C, 53.72; H, 7.51; Found: C, 53.77; H, 6.82; FABMS m/z, 464 $(M)^+$. ¹H NMR chemical shift values: see Table 1.

3,16 α -Dihydroxy-1,3,5(10)-estratrien-17 β -yl- β -D-glucopyranosiduronic acid (Estriol 17-glucuronide) (12). The hydrolysis reaction was performed by a procedure similar to that used in the preparation of 6. Compound 12 was obtained from compound 8 and 11 as fine crystals in 65% yield respectively. mp, 245–247 C; reported mp, 241 C (lit. 8); Analysis calculated: C₂₄H₃₂O₉ · 3.5H₂O: C, 54.64; H, 7.45; Found: C, 54.19; H, 7.37; FABMS m/z, 465 (MH)⁺. ¹H NMR chemical shift values: see Table 1.

Hydrolysis of the estriol glucuronides (6) and (12) with β -glucuronidase. A solution of the estriol glucuronide 6 or 12 (15 mg) and β -glucuronidase (E.C.3.2.1.31 Sigma Co., 5 mg, 14000 Fishman Units) in 0.1-M acetate buffer (pH = 4.6, 30 ml) was incubated at 38 C for 24 hours. The incubated mixture was extracted with EtOAc (3 × 30 ml), washed with water, dried (MgSO₄), and evaporated to give a white solid. The solid was crystallized from aqueous MeOH (1:1) to give estriol (7 mg) and (6 mg) as colorless plates, mp, 279–284 C, from enzymatic digestion of 6 and 12 respectively. The products were identical with an authentic sample of estriol in every respect.

Results and discussion

The procedure described here (see Scheme 1) represents a simple way to synthesize estriol 16- and 17monoglucuronides in good overall yields (39% for estriol 16-glucuronide, 25% for estriol 17-glucuronide) and purity. The glucuronidation reactions with protected estriol derivatives give higher yields than from comparable 16a-hydroxy-17-oxo-derivatives, because there is no adjacent carbonyl group in the substrate; hence the free hydroxyl group is more nucleophilic in the coupling reactions.^{7,11} Since the stereochemistry at carbon-16 and carbon-17 is known in the D-ring when estriol is used as starting material, the problem of obtaining the 17α -isomer during reduction of the 17-carbonyl group is avoided. Also, it should be noted that determination of the absolute stereochemistry at these positions in the 5-membered D-ring is difficult by spectroscopic techniques.12

The present method has advantages over previous procedures using estriol as a starting material. For example, the direct glucuronidation of estriol 3-benzoate or the 3-benzyl ether forms a mixture of 16- and 17glucuronides, from which the 16-glucuronide can be

н	Estriol 16-glucuronide (6) ppm	Estriol 17-glucuronide (12) ppm	Difference in ppm
18-CH	0.79 (s. 3H)	0.87 (s. 3H)	+ 0.08
17 <i>α</i> -H	3.64 (d, J = 5.49 Hz)	3.48 (d, J = 4.76 Hz)	- 0.16
16 <i>B</i> -H	4.10 (m. 1H)	4.18 (m, 1H)	+ 0.08
1.2.4-3H	6.47-7.07 (m)	6.46-7.06 (m)	—
(aromatic)			
1'-H	4.36 (d, $J = 7.69$ Hz)	4.43 (d, J = 7.69 Hz)	+ 0.07
2'-H	3.25-3.34 (m)	3.25-3.34 (m)	
3'-H	3.41 (t)	3.41 (t)	0
4'-H	3.54 (t)	3.54 (t)	0
5'-H	3.79 (d, J = 9.53 Hz)	3.88 (d, J = 9.53 Hz)	+ 0.09

Table 1 ¹H NMR chemical shift values for the Estriol 16- and 17-glucuronides

separated by fractional crystallization. However, this method produces a low overall yield of the 16-glucuronide together with some contamination by the 17-isomer.^{8,13} Also, it is difficult to get high yields of 3-protected estriols.² Although estriol 3-benzoate undergoes selective monoglucuronidation in CHCl₃ activated by Ag_2CO_3 , mainly affording the 17-glucuronide,⁸ the low yield of the 3-protected estriol is still a problem. Scheme 1 provides a very simple alternative procedure, which gave the 16- or 17-glucuronide in comparatively high yields with preservation of the stereochemistry and location of the glucuronide linkage. By selectively protecting and deprotecting the hydroxyl groups of estriol in a controlled sequence, this method also provides an opportunity for further manipulation of estriol and opens the way for the synthesis of a wide range of estriol derivatives.

It is more convenient to use CdCO₃ as a promoter in the Koenigs-Knorr reactions than Ag₂CO₃, because all the compounds and the solvent can be easily and thoroughly dried by azeotropic distillation to remove moisture before starting the reaction. It is thus easier to control the reaction under scrupulously anhydrous conditions and the reaction shows greater selectivity. For example, the Koenigs-Knorr reaction with 2,3-protected 2-hydroxyestriol using CdCO₃ gave a mixture of the 16 α - and 17 β -coupling products in a ratio of 7:1.14 However, there is no previous report about the preferential selection of the Koenigs-Knorr reaction for an alcoholic hydroxyl group in the presence of a phenolic hydroxyl group. In the present work, we synthesized estriol 17-glucuronide by direct glucuronidation of compound 2 (Scheme 1), in which both types of hydroxyl group were available. The result showed that the alcoholic hydroxyl group is much more reactive in this compound than the phenolic hydroxyl group despite the fact that there was a bulky tert-butyldimethylsilyl group at the 16 α -OH position. Hence estriol 17glucuronide can be prepared without the necessity to protect the 3-OH group.

An interesting observation made during our synthesis was the ease of deprotection of the *tert*-butyldimethylsilyl group in some of these compounds. The usual reagent for the cleavage of a silyl ether is tetra-*n*-butylammonium fluoride. However in the current work, removal of the *tert*-butyldimethylsilyl group using weak acid (HAc/H₂O/THF) gave a much higher yield than that by fluoride ion as described in the literature.⁵ It is possible that tetra-*n*-butyl-ammonium fluoride functions as a strong base in THF and causes 1,2-migration of the acyl group.¹⁵ We also noted that the *tert*-butyldimethylsilyl group in compound 7 could be removed simply by dissolution in MeOH and gentle heating.

Although estriol monoglucuronides have been reported as the human metabolites of estradiol, and synthesized by different methods, spectroscopic determination of the exact position and the configuration of the glycosidic linkage still awaits the availability of appropriate standards. In the present work the ¹H NMR and mass spectra unambiguously identified estriol 16- and 17-glucuronides. The conjugated position and configuration of the glucuronide linkage in the two isomers were established on the basis of ¹H NMR and 2D-COSY spectral data. The 2D-COSY spectra allowed the assignment of the 1'-H to 5'-H of the glucuronic acid moiety and unambiguously identified the 16 β - and 17 α -protons. Figures 1 and 2 clearly show the correlations of 1'-H/2'-H, 2'-H/3'-H, 3'-H/4'-H, 4'-H/ 5'-H and 16 β -H/17 α -H. The 17 α - and 16 β -protons signals in the 16 α -glucuronide (compound 6, Scheme 1) appeared as a doublet at 3.64 ppm and as a multiplet at 4.10 ppm, respectively, while those in the 17β -glucuronide (Compound 12, Scheme 1) appeared as a doublet at 3.48 ppm and as a multiplet at 4.18 ppm. These chemical shift values are in good agreement with those reported for the 16 α - and 17 β -glucuronides of 2,3-protected 2-hydroxyestriol.¹⁴ The transrelationship of the 1'-H and 2'-H in the glucuronic acid was demonstrated by the large coupling constants for both isomers $(J_{1,2} = 7.69 \text{ Hz})^{16}$ and subsequent hydrolysis of the glucuronides with β -glucuronidase confirmed the β linkage of the glucuronic acid with estriol. There were only small differences in ¹H NMR chemical shift values between the two isomers (Table 1). The major differences were that the position of the 16*B*-H, 18-CH₃, 1'-H, and 5'-H signals were shifted downfield nearly 0.1



Figure 1 2D-COSY spectrum of estriol 16-glucuronide. The figure shows the proton correlations of 1'/2', 2'/3', 3'/4', 4'/5', $16\beta/17\alpha$; and in the sequence $1'-16\beta-5'-17\alpha-4'-3'-2'$.



Figure 2 2D-COSY spectrum of estriol 17-glucuronide. The figure shows the proton correlations of 1'/2', 2'/3', 3'/4', 4'/5', $16\beta/17\alpha$; and in the sequence $1'-16\beta-5'-4'-17\alpha-3'-2'$.

ppm in the 17β -isomer compared with the 16α -isomer. However, the 17α -proton was shifted upfield by about 0.2 ppm as clearly shown in Figures 1 and 2. In the 16α -glucuronide, the 17α -proton occurred as a doublet situated downfield of the 4'-H position (Figure 1), but in the 17 β -glucuronide, the 17 α -proton was upfield of the 4'-H position (Figure 2). This small difference is useful in distinguishing between the two isomers.

Acknowledgments

The authors are especially indebted to Dr. D.L. Officer for helpful discussions. We also thank J. Hastie for NMR measurements and J. Allen (The Horticulture & Food Research Institute of New Zealand Ltd.) for mass spectra.

References

- Brown JB, Blackwell LF, Cox RI, Holmes JM, Smith MA (1988). Chemical and homogeneous enzyme immunoassay methods for the measurement of estrogens and pregnanediol and their glucuronides in urine. *Prog Clin Biol Res* 285: 119-138.
- Nagao Y, Fuita E, Kohno T, Yagi M (1981). An efficient method for selective acetylation of alcoholic hydroxy groups. *Chem Pharm Bull (Tokyo)* 29:3202-3207.
- Horiuchi CA, Haga A, Satoh JY (1986). Novel regioselective iodination of estradiol 17β-acetate. Bull Chem Soc Jpn 59: 2459-2462.
- Numazawa M, Nagaoka M, Tsuji M, Osawa Y (1983). Novel and efficient synthesis of estriol and its 16-glucuronide via 2,4,16α-tribromestrone. J Chem Soc Perkin Trans I 121-125.
- 5. Laurent H, Bittler D, Beier S, Elger W (1985). Estriol esters. Eur Pat Appl EP 163,596 (Chem Abstr 105:P134239e).
- Corey EJ, Venkateswarlu A (1972). Protection of hydroxyl groups as *tert*-butyldimethylsilyl derivatives. J Am Chem Soc 94:6190-6191.
- Nambara T, Imai K (1967). Syntheses of estriol monoglucuronides. Chem Pharm Bull (Tokyo) 15:1232–1238.
- Nambara T, Kawaeada Y, Shibata K, Abe T (1972). Studies on steroid conjugates. IX. New synthesis of estriol 16- and 17monoglucuronides. *Chem Pharm Bull (Tokyo)* 20:1988–1992.
- Conrow RB, Bernstein S (1971). Steroid conjugates. VI. An improved Koenigs-Knorr synthesis of aryl glucuronides using cadmium carbonate, a new and effective catalyst. J Org Chem 36:863-870.
- Neeman M, Hashimoto Y (1962). The structure of estriol monoglucosiduronic acid from human pregnancy urine. J Am Chem Soc 84:2972-2978.
- 11. Elce JS, Carpenter JGP, Kellie AE (1967). The synthesis of estrogen monoglucuronides. J Chem Soc (C) 542-550.
- 12. Boucheau V, Renaud M, de Ravel MR, Mappus E, Cuilleron CY (1990). Proton and carbon-13 nuclear magnetic resonance spectroscopy of diastereoisomeric 3- and 17β -tetrahydropy-ranyl ether derivatives of estrone and estradiol. *Steroids* **55**:209-221.
- Joseph JP, Dusza JP, Cantrall EW, Bernstein S (1969). Steroid conjugate. V. The synthesis of a sulfoglucuronide derivative of estriol. *Steroids* 14:591-601.
- Ohkubo T, Wakasawa T, Nambara T (1990). Synthesis of 2hydroxyestriol monoglucuronides and monosulfates. *Steroids* 55:128-132.
- 15. Dodd GH, Golding BI, Ioannou PV (1975). Limitations of *t*butyldimethylsilyl as a protecting group for hydroxy-functions. J Chem Soc (C) 249-250.
- Lemieux RU, Kullnig RK, Bernstein HJ, Schneider WG (1958). Configurational effects on the proton magnetic resonance spectra of six-membered ring compounds. J Am Chem Soc 80:6098-6105.