

Research paper

Salicylimine-based fluorescent chemosensor for magnesium ions in aqueous solution

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ABSTRACT

A new fluorescent schiff base chemosensor (H_2L) was prepared for the sensitive and selective sensing of Mg^{2+} ions based on multiple mechanisms. H_2L is a weak fluorescent ($f = 0.031$) due to PET process and C=N isomerization. Upon addition of Mg^{2+} , the complex $[MgL]$ was formed and a remarkable fluorescence enhancement ($f = 0.182$) was produced. H_2L had no such significant effect on the fluorescence in the presence of metal ions, such as Na^+ , Ag^+ , K^+ , Ca^{2+} , Mg^{2+} , Hg^{2+} , Mn^{2+} , Co^{2+} , Ni^{2+} , Cu^{2+} , Zn^{2+} , Cd^{2+} , Pb^{2+} , Cr^{3+} , Fe^{3+} , and In^{3+} . The satisfactory linear relationship was observed between the added concentrations of Mg^{2+} and the fluorescence intensity of H_2L . The detection limit was 3.04×10^{-9} M with a rapid response time. Chemical inputs of Mg^{2+} and Fe^{3+} ions satisfy the conditions of INHIBIT molecular logic gate.

1. Introduction

Magnesium is one of the vital cofactors that participates in a lot of biochemical reactions including protein synthesis, muscle and nerve function, blood glucose control and blood pressure regulation [1]. Magnesium is also necessary for the synthesis of DNA and RNA, the antioxidant glutathione, oxidative phosphorylation, normal heart rhythm, muscle contraction, the structural development of bone, glycolysis and energy production [2]. Diseases such as Alzheimer, diabetes, hypertension, and epilepsy have relation with abnormal concentrations of magnesium in the cytosol and subcellular regions [3–6]. Therefore, finding an efficient analytical and simple technique for imaging and detection of magnesium is of great interest [7]. Fluorescence chemosensors offer a number of outstanding characteristics such as real-time response and simple handling allowing dynamic measurements. In recent years, considerable efforts have been spent on developing of new suitable fluorescent probes matching all desired properties such as easy preparation, selectivity, high sensitivity, low-cost synthesis and aqueous solubility [8–17]. Designing of chemosensors for Mg^{2+} have been attracting increasing interest based on the fluorescence signaling mechanisms such as intramolecular charge transfer (ICT) [18], C=N isomerization [19], photoinduced electron transfer (PET) [20], appropriate cavities [21], excited-state intra-/intermolecular proton transfer (ESIPT) [22]. The photoinduced electron transfer (PET) mechanism is the process by which an electron falls from

an excited site to a site with lower energy. The matching of their oxidation–reduction potentials of the two sites and the closeness in space are necessary requirements for this transport. The mechanism of isomerization related to molecules which are easily isomerize in the excited state but highly stable in the ground state. The isomerization mechanism has been observed for several classes of molecules. Schiff base compounds are one of the classical examples for isomerization mechanism. Furthermore, because of the strong binding abilities to various metal ions and the individual photophysical properties, Schiff base derivatives have been extensively explored in the field of the field of chemosensors for detection of metal ions [23–27]. In this work, a π -conjugated Schiff base receptor (H_2L) was designed and synthesized. The photophysical studies showed that H_2L can be used as a selective and sensitive chemosensor for Mg^{2+} ions by enhancement of fluorescence emission intensity in EtOH/ H_2O (9:1, v/v, pH = 7.40). H_2L exhibits rapid response, low detection limit (3.04×10^{-9} M), excellent selectivity and sensitivity to Mg^{2+} .

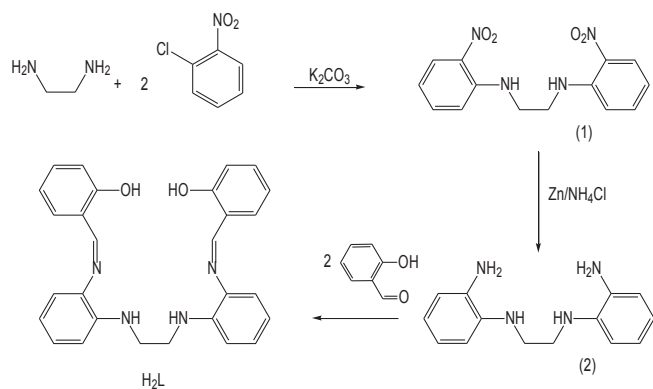
2. Experimental

2.1. Materials and instruments

All solvents commercially purchased and were of reagent grade quality. 1,2-diamino ethane, 1-chloro-2-nitrobenzene and 2-hydroxybenzaldehyde were obtained from Merck and was used without any

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Scheme 1. The synthetic route of H₂L.

further purification. A Bruker A V300 MHz spectrometer was used to obtain NMR spectra. A BIO-RAD FTS-40A spectrophotometer was used to record infrared spectra in KBr pellets (4000–400 cm⁻¹). A Kratos-MS-50 spectrometer was used to record positive ion FAB mass spectra with 3-nitrobenzyl alcohol as the matrix solvent. A Varian Cary Eclipse 300 spectrophotometer was used to obtain UV-Vis absorption spectra. The fluorescence spectra were recorded on a Varian spectrofluorometer. Both emission and excitation bands were set at 5 nm.

2.2. *N,N'*-bis(2-nitrophenyl)-1,2-ethanediamine (1)

Potassium carbonate (2.76, 20 mmol), 1-chloro-2-nitrobenzene (3.14 g, 20 mmol) and 1,2-diaminoethane (0.64 g, 10 mmol) were completely mixed in a round-bottomed flask. The reaction was heated at 175 °C in an oil bath for 3 h. After cooling, the yellow solid was collected and washed by 3:1 H₂O/EtOH (×3). Yield (82%), Scheme 1. Anal. Calc. for C₁₄H₁₄N₄O₄: C, 55.63; H, 4.67; N, 18.53. Found: C, 55.82; H, 4.70; N, 18.49%. IR (KBr, cm⁻¹) 1350, 1570 m (NO₂), 3360(NH); ¹H NMR δ_H (CDCl₃, ppm) 3.40 (t, 4H, CH₂-CH₂), 6.70 (m, 4H, Ar), 7.35(t, 2H, Ar), 7.90 (d, 2H, Ar), 7.35 (t, 2H, NH).

2.3. *N,N'*-bis(2-aminophenyl)-1,2-ethanediamine (2)

A mixture of ammonium chloride (4 g), 2-nitro-N-(2-(2-nitrophenylamino)propyl)benzenamine (3.16 g, 10 mmol), and H₂O (2 ml) in 100 ml of ethanol was heated to boiling and then 3 g of zinc dust were gradually added over a period of 0.5 h. When the color of the solution changed from brown to pale yellow, it was filtered and washed. Distilled water (1000 ml) was added to the filtrated solution and the pH was adjusted to 12 with potassium hydroxide. The product as brown powder participates were appeared in the solution. Yield (1.3 g, 62%). Anal. Calc. for C₁₄H₁₆N₄: C, 69.39; H, 7.46; N, 23.12. Found: C, 69.20; H, 7.34; N, 23.32%. IR (KBr, cm⁻¹) 3452 and 3464 m (NH₂); 3344 m (NH); ¹H NMR δ_H (CDCl₃, ppm) 3.33 (br, 10H, CH₂-CH₂ and NH), 6.64 (m, 8H, Ar); ¹³C NMR δ_C (CDCl₃, ppm) 40.0, 135.9, 134.7, 120.1, 118.0, 113.4, 109.1

2.4. Synthesis of H₂L

To a solution of 2-hydroxybenzaldehyde (0.24 g, 2 mmol) in absolute ethanol, *N,N'*-bis(2-aminophenyl)-1,2-ethanediamine (0.23 g, 1 mmol) was added and the reaction was gently refluxed approximately for 3 h. The product precipitating was filtered off and washed with ethanol, and dried in air at room temperature. Yield (70%). Anal. Calc. for C₃₆H₃₀N₄O₂: C, 74.65; H, 5.82; N, 12.44. Found: C, 74.88; H, 5.70; N, 12.61%. IR (KBr, cm⁻¹) 3412 (OH), 1621 (C=N); ¹H NMR δ_H (DMSO-*d*₆, ppm) 3.52 (s, 4H, CH₂), 4.59 (s, 2H, NH) 6.74–7.37 (4 signals, 16H, Ar), 8.51 (s, CH = N, 2H), 12.82 (s, OH, 2H); ¹³C NMR δ_C (DMSO-*d*₆, ppm) 43.08, 110.79, 117.10, 117.36, 118.31, 119.18,

128.32, 132.30, 132.99, 135.66, 141.83, 160.69 and 162.51. The mass spectrum shows peak at *m/z* = 450.2 corresponding to the H₂L (Figs. S1–S4).

3. Results and discussion

3.1. Synthesis

H₂L was readily synthesized in 74% yield by condensation of *N,N'*-bis(2-aminophenyl)-1,2-ethanediamine and salicylaldehyde in ethanol (scheme 1). Its structure was investigated by mass, ¹H NMR, ¹³C NMR spectra (see Figs. S1–S4, in the supporting information).

The main band appearing in the FTIR spectrum was related to stretching vibration of C=N bands and no bands attributable to C=O were observed. The stretch vibration of secondary amines (N–H) appear at ca 3396 cm⁻¹. In ¹H NMR spectrum, a singlet signal at 3.52 ppm and a broad signal at 4.59 ppm were appeared related to methylene protons and amine protons, respectively. A single ¹H imine resonance at 8.51 ppm was appeared demonstrating the equivalence of the two imine environments. The signal at 12.82 ppm can be attributed to the hydroxyl protons (the protons related to the intramolecular hydrogen bonds were appeared in very low-field proton resonance). Four signals have been observed in the aromatic region (6.74–7.37 ppm). In ¹³C NMR spectrum, one signal at 43.08 ppm was observed attributed to methylene carbons. There is one type for the imine carbon atoms (162.51 ppm) demonstrating the equivalence of the two imine environments. Eleven peaks have been observed in the aromatic region (110.79–160.69 ppm).

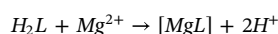
From reaction if H₂L with MgCl₂, the complex of [MgL] was synthesized. The IR spectra of H₂L and [MgL] were prepared and compared. As can be seen from Fig. S3, two differences have been observed: (i) The absorption band at 3397 cm⁻¹ corresponding to the N–H stretching vibration shifted to 3381 cm⁻¹ (ii) the stretch vibration of imine group (C=N) at 1609 cm⁻¹ shifted to 1624 cm⁻¹. These observations suggest that the imine groups and the amine groups may participate in the coordination interaction to Mg²⁺ ions.

3.2. Spectral properties of fluorescent H₂L

H₂L (10 μM) exhibits a weak fluorescence intensity around 490 nm when excited at 423 nm. The fluorescence emission measurements of H₂L were made at room temperature in different solvents: water, ethanol, methanol, chloroform, acetonitrile and dimethylformamide (DMF). Investigation of the solvent-dependent fluorescence emission displayed that the fluorescence is strongly quenched in protic solvents. The magnitude of the quenching depends on the H bond-donating ability of the solvent (Fig. 1).

The salicylimine- derivatives are known to be good ligands for fluorescence studies and used to develop chemosensors. The excitation and emission wavelengths of the H₂L were found at higher wavelength, in compared to naphthalene groups. This red-shift is related to the delocalization the imine π-electrons on the aromatic amines results in a red-shift of the excitation and emission wavelengths of the related aromatic rings. The emission spectra of H₂L on excitation at 423 nm shows a broad band at 490 nm. The selective binding behavior of H₂L towards different metal nitrates (Na⁺, K⁺, Cs⁺, Mg²⁺, Ba²⁺, Ca²⁺, Al³⁺, Pb²⁺, Mn²⁺, Fe²⁺, Fe³⁺, Co²⁺, Ni²⁺, Cu²⁺, Zn²⁺, Cd²⁺, Hg²⁺ and Ag⁺) was studied by fluorescence spectroscopies. All of the titration experiments were carried out in EtOH/H₂O (9/1, v/v) upon adding the respective metal ions. Fig. 2 exhibits the changes in the fluorescence spectra of H₂L by addition of the various tested metal ions.

The complexation reaction occurs with formation of neutral complex [MgL] coupling with release of H⁺ ions:



It should be noted that H₂L molecules are functioning as charged

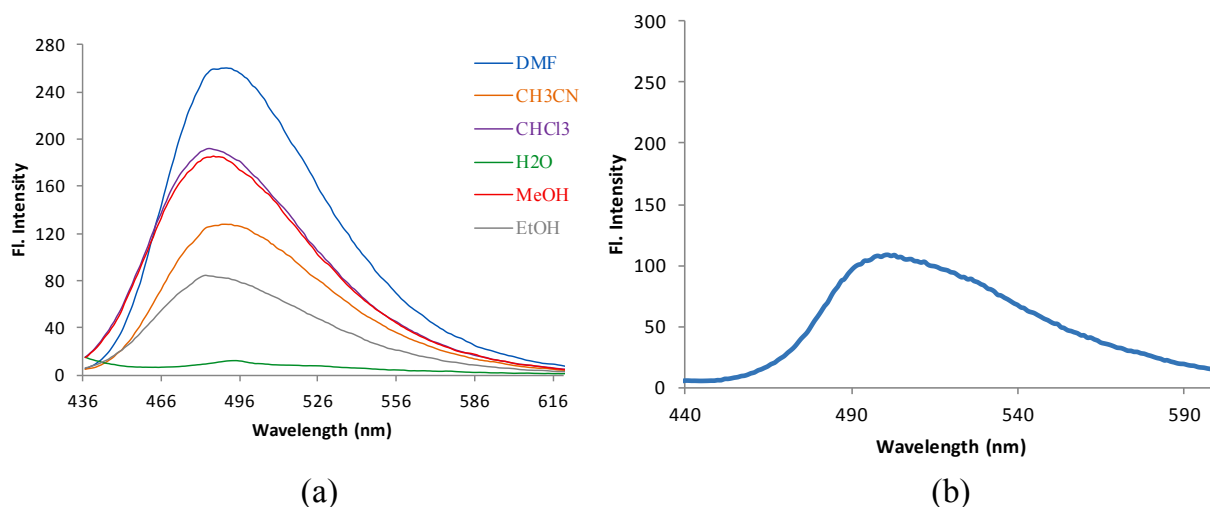


Fig. 1. (a) Changes of fluorescence spectrum of H₂L in different solvents, (b) solid-state fluorescence of H₂L. Excitation wavelength was 423 nm.

ionophore, thus taking into account stability constant of the complex the stability constant of [MgL] is pH dependent. Therefore, a few drops of triethylamine were added to the solution and then the selective binding behavior of H₂L towards different cations was studied (Fig. 3).

Among the tested metal ions (5.0 equiv), no significant spectral changes were observed in the presence of Na⁺, K⁺, Cs⁺, Ba²⁺, Ca²⁺ and Zn²⁺, while a fluorescence quenching was seen upon the addition of Al³⁺, Pb²⁺, Mn²⁺, Fe²⁺, Fe³⁺, Co²⁺, Ni²⁺, Cu²⁺, Cd²⁺, Hg²⁺ and Ag⁺. However, a large enhancement in fluorescence intensity was only observed in the presence of Mg²⁺ ions.

The FEF (fluorescence enhancement factor) values of (H₂L + triethylamine) corresponding to different metal ions are shown in Fig. 4. The FEF ($I_x - I_{H_2L} / I_{H_2L}$) was calculated using minimal (I_{H_2L}) and maximal (I_x) fluorescence intensities recorded before and after addition of metal ions at 490 nm, respectively. The highest fluorescence enhancement for (H₂L) has been observed in the presence of Mg²⁺ ions (Fig. 4).

To test practical applicability of H₂L as a fluorescence chemosensor for Mg²⁺, competition experiments were carried out in the presence of other metal cations (Fig. 5). The results show that the fluorescence of H₂L upon complexation to Mg²⁺ did not significantly change in the presence of Na⁺, K⁺, Mg²⁺, Ca²⁺, Ba²⁺, Al³⁺, Co²⁺, Ni²⁺, Zn²⁺, Cd²⁺, Pb²⁺, Cd²⁺, and Hg²⁺ with a concentration up to 100 mM. In

the presence of Mn²⁺, Fe²⁺, Cu²⁺, Fe³⁺ and Ag⁺ ions had a disturbance under their much excess.

The coordination mode of H₂L to Mg²⁺ was investigated by mixing various amounts of Mg²⁺ with a constant concentration of H₂L (Fig. 6). In the fluorescence titration experiments, the fluorescence emission intensity was gradually increased when 1 equivalent of Mg²⁺ was added, it no longer changed when the concentration of Mg²⁺ increased (Fig. 6, inset), indicating the 1:1 binding of H₂L to Mg²⁺ (Fig. 6).

To determine the binding stoichiometry of H₂L and Mg²⁺ ion, the job plot method was also used. The molar ratio was given by $[Mg^{2+}] / ([Mg^{2+}] + [H_2L])$ and measured from 0 to 1. The total concentrations were kept at 10 μ M. As seen in Fig. 7, the maximum value of the molar ratio was 0.5. This means that the molar ratio of the H₂L–Mg²⁺ complex was 1:1 in the binding stoichiometry.

Free H₂L shows a weak fluorescence emission because of existence the photo-induced electron transfer (PET) of the lone pair electrons from imine to the fluorophore in the excited state and the C=N isomerization. When Mg²⁺ was added a stable chelation of Mg²⁺ to H₂L inhibited PET process and C=N isomerization, resulting in a significant fluorescence emission.

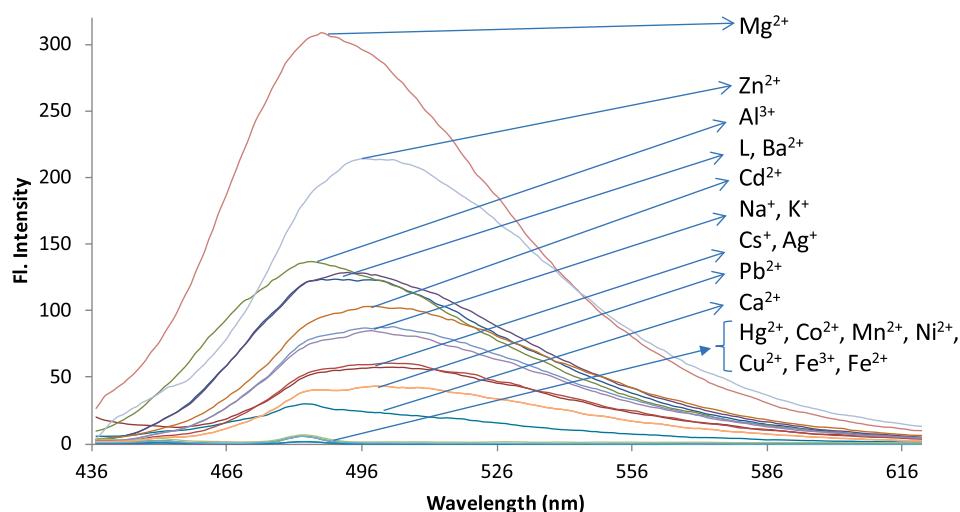


Fig. 2. Changes of fluorescence emission intensity of H₂L (10 μ M) upon addition of various metal ions (10 equiv) in EtOH/H₂O (9:1, v/v) containing HEPES buffer (10 mM, pH 7.4).

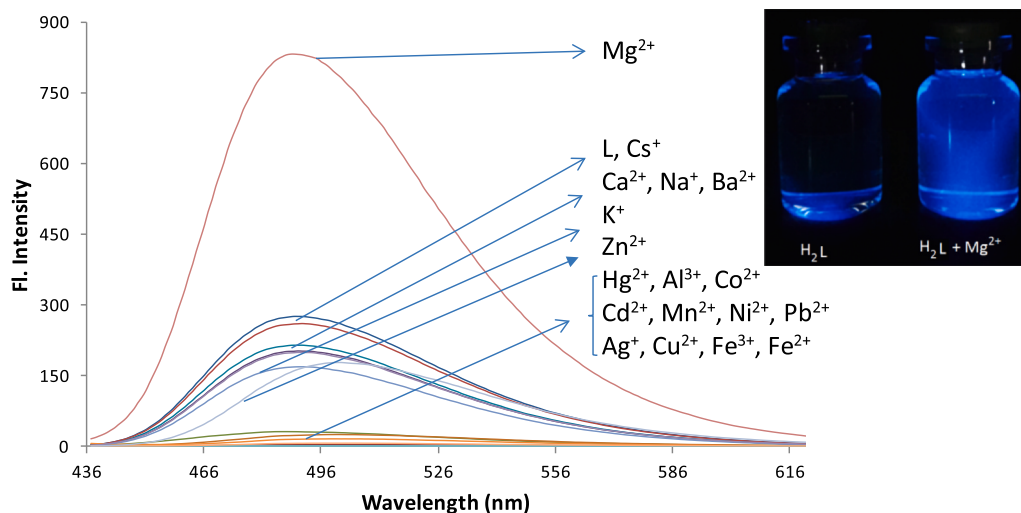


Fig. 3. Changes of fluorescence emission intensity of H_2L + trimethylamine in EtOH/ H_2O (9:1, v/v; HEPES, pH 7.4) upon addition of various metal ions (10 equiv) with an excitation wavelength of 423 nm.

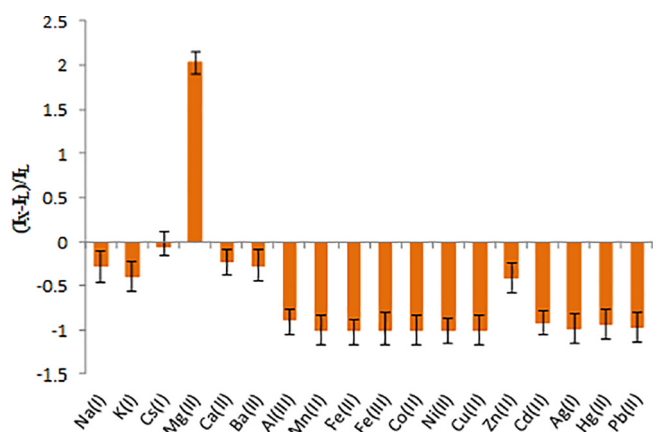


Fig. 4. The fluorescence enhancement factor (FEF) of (H_2L + triethylamine) upon addition of various metal ions in EtOH/ H_2O (9:1, v/v; HEPES, pH 7.4) at 25 °C. Both the excitation and emission slit widths were 5.0 nm.

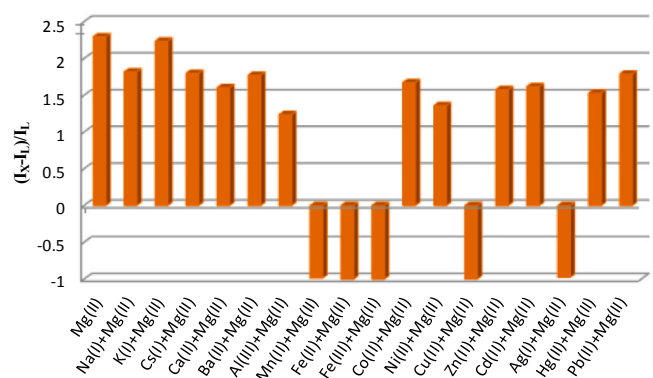


Fig. 5. The fluorescence enhancement factor (FEF) of (H_2L + triethylamine) upon addition of Mg^{2+} ions in the presence of various metal ions in EtOH/ H_2O (9:1, v/v; HEPES, pH 7.4) at 25 °C. Both the excitation and emission slit widths were 5.0 nm.

3.3. NMR studies

The 1H NMR spectra of H_2L were recorded in the absence and presence of and Mg^{2+} ions (Fig. 8) As shown in Fig. 8, upon the addition of Mg^{2+} , the resonance signals corresponding to imine protons

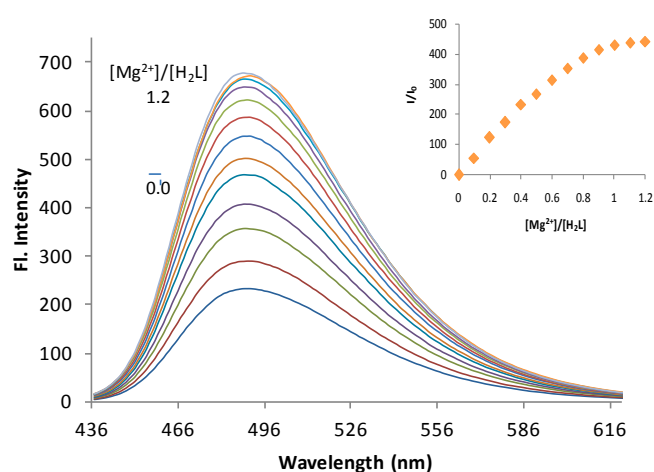


Fig. 6. The titration of (H_2L) (10 μM) as a function of added Mg^{2+} concentration in EtOH/ H_2O (9:1, v/v). Excitation wavelength was 423 nm. Both the emission and excitation slit widths were 5.0 nm.

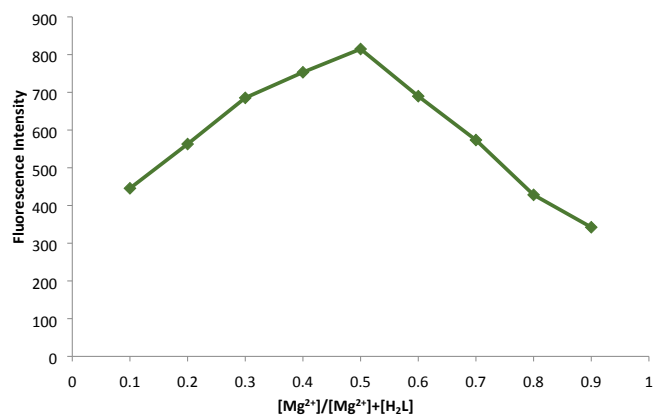


Fig. 7. Job's plot analysis of H_2L and Mg^{2+} .

shift downfield from 8.51 to 10.31 ppm, and the resonance signals corresponding to methylene protons shift downfield from 3.52 to 4.39 ppm. When Mg^{2+} was added, the signals corresponding to hydroxyl protons ($-OH$) were disappeared due to deprotonation of the hydroxyl groups to strongly coordinate to Mg^{2+} . The proton signals of

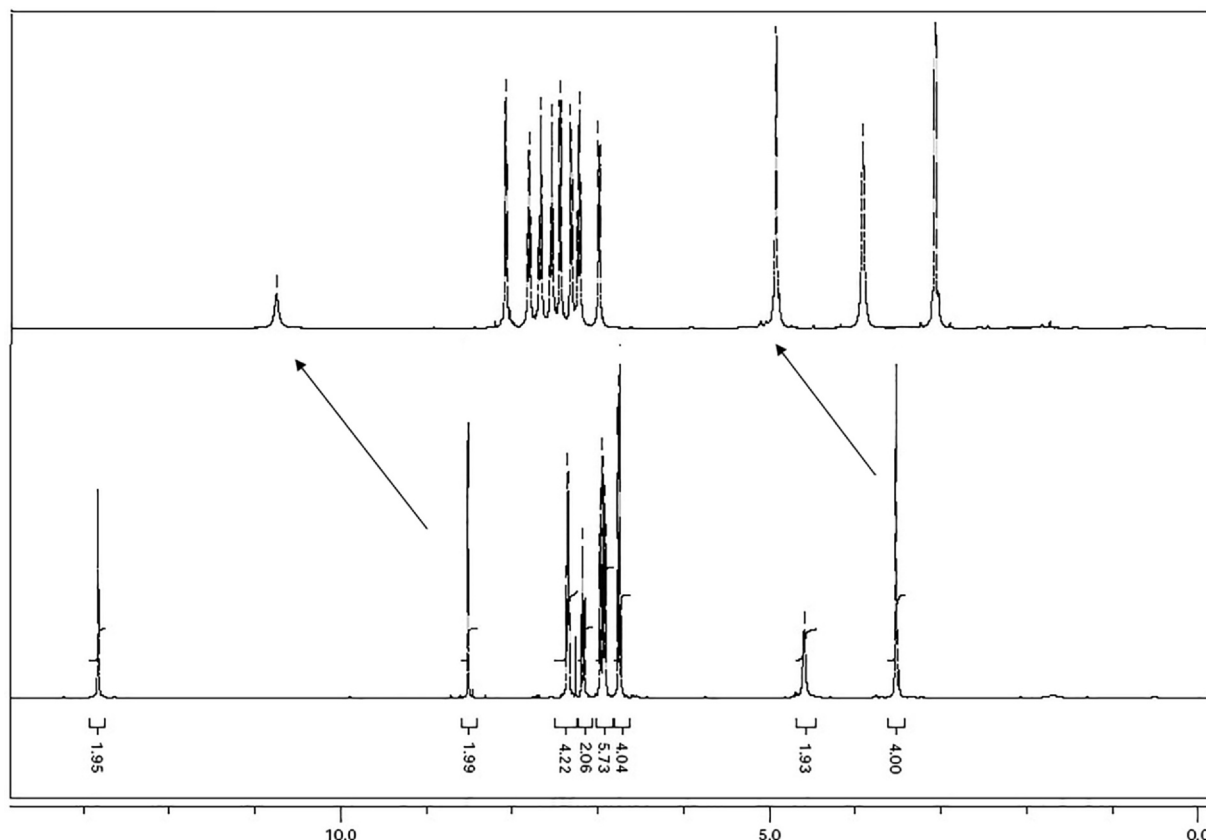


Fig. 8. ^1H NMR spectra of compound H_2L and its complex with Mg^{2+} ($[\text{MgL}]$) at room temperature.

NH (4.59 ppm) are exchanged with deuterium atoms of $\text{DMSO-}d_6$. This result indicates that a more efficient coordination of Mg^{2+} ion occurs with the imine nitrogen ($\text{C}=\text{N}$), the nitrogen ($-\text{NH}$) of ethylene diamine moiety and hydroxyl groups. When Mg^{2+} ion is coordinated to H_2L , the charge transfer was arrested due to the strong complexation of Mg^{2+} with H_2L . In addition, Mg^{2+} ion as a soft acid, preferentially interacts with the hydroxyl group and nitrogen atoms according to Pearson's HSAB theory.

3.4. Infrared studies

The IR spectrum of H_2L shows the bands at 1620 and 3381 cm^{-1} related to stretching vibrations of $\nu(\text{C}=\text{N})$ and $\nu(\text{N}-\text{H})$, respectively (Fig. 9). The band related to azomethine has been shifted to lower frequency 1609 cm^{-1} in the spectrum of the complex. This shift can be attributed to the coordination of nitrogen atoms of the azomethine group to the metal atom. This can be explained by the donation of electrons from bonding orbital $\text{C}=\text{N}$ to the empty d-orbitals of the metal ion. The band related to secondary amine has been shifted to higher frequency 3391 cm^{-1} in the spectrum of the complex. The IR spectra of L also exhibited a band at 3263 cm^{-1} , which was attributed to the stretching vibration of $\nu(\text{OH})$. In the spectrum of the complex, this absorption was disappeared, that expressed deprotonation and participation of the enol oxygen in coordination (Fig. 9). These evidences indicate that the nitrogen of azomethine and secondary groups and the oxygen of ligand are involved in chelation with metal ion.

3.5. Limit of detection and response time

The fluorescence titration was used to determine of the limit of detection. For this means, the equation of $3\sigma/s$ was applied where s is the slope of the calibration curve and σ is the standard deviation of the

blank solution. As shown in Fig. 10, H_2L has a detection limit of $3.04 \times 10^{-9}\text{ M}$ for Mg^{2+} . The affinity and sensitivity of the probe towards Mg^{2+} in the presence of Ca^{2+} ions is a challenge for in vivo and in vitro monitoring of Mg^{2+} under intracellular conditions.

The concentration of Mg^{2+} in intracellular conditions is of the order $0.1\text{--}6\text{ mmol L}^{-1}$ whereas the intracellular concentration of Ca^{2+} may vary from $0.1\text{ }\mu\text{mol L}^{-1}$ to 1 mmol L^{-1} . This study shows that H_2L is a highly selective and sensitive fluorescent chemosensor to recognize of Mg^{2+} without the interference of other metal ions, especially Ca^{2+} . The response time is a necessary and important factor for fluorescence chemosensors. The response time of the chemosensor was studied by the kinetics of fluorescence enhancement at 358 nm . The response time to Mg^{2+} is less than 5 s . Fast changes in the fluorescence intensity of the chemosensor were appeared and remained quite stable when H_2L was titrated with magnesium ions.

3.6. Binding constants measurements

The association constant (K_a) of H_2L with Mg^{2+} was calculated from the Benesi-Hildebrand plot. The fluorescent titration spectra of H_2L ($10\text{ }\mu\text{M}$) in the presence of various concentrations of Mg^{2+} ($0\text{--}1.2\text{ }\mu\text{M}$) were conducted. When Mg^{2+} ions were increased, the fluorescence intensity emission of the solution was increased (Fig. 11). It should be noted that at more than $10\text{ }\mu\text{M}$ of Mg^{2+} , the fluorescence intensity reached to the constant value.

$$\frac{1}{I - I_0} = \frac{1}{K_a(I_{\text{max}} - I_0)[\text{Mg}^{2+}]} - \frac{1}{I_{\text{max}} - I_0}$$

where I and I_0 represent the emission intensity of H_2L in the presence and absence of Mg^{2+} , respectively, I_{max} is the saturated emission of H_2L in the presence of excess amount of Mg^{2+} ; $[\text{Mg}^{2+}]$ is the concentration of Mg^{2+} ion added. The measured emission intensity $[1/(I - I_0)]$ varied as a function of $1/[\text{Mg}^{2+}]$ in a linear relationship ($R^2 = 0.99$),

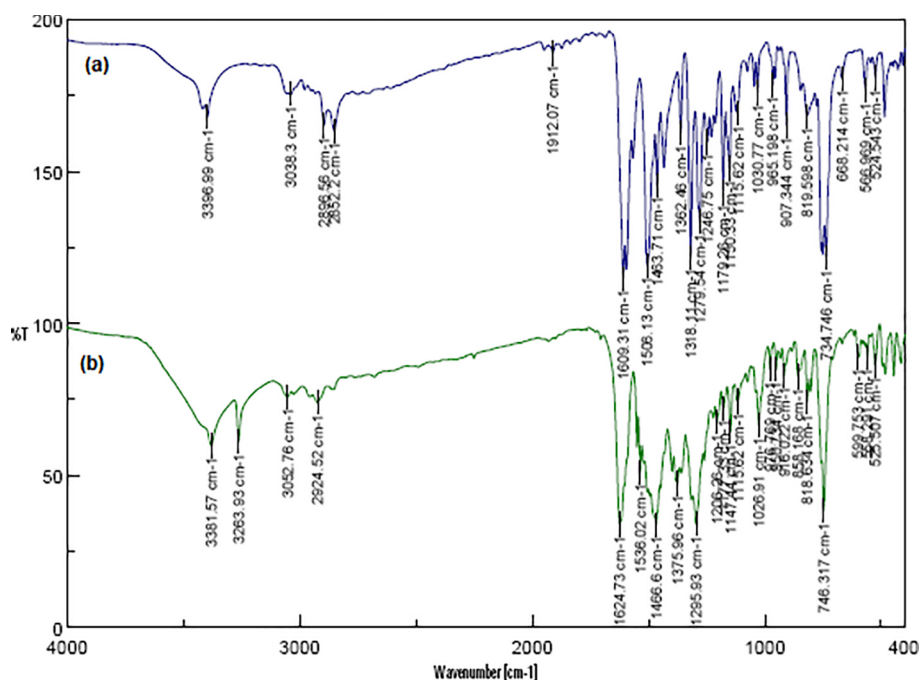


Fig. 9. The comparison between the IR spectra of (a) [MgL], (b) H₂L.

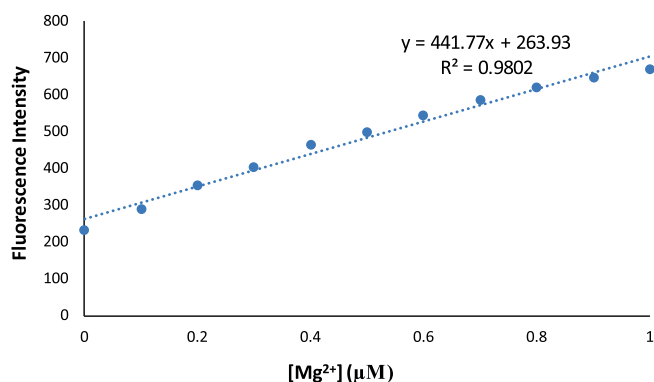


Fig. 10. Linear response curve of H₂L (10 μM) at 490 nm depending on Mg²⁺ concentration.

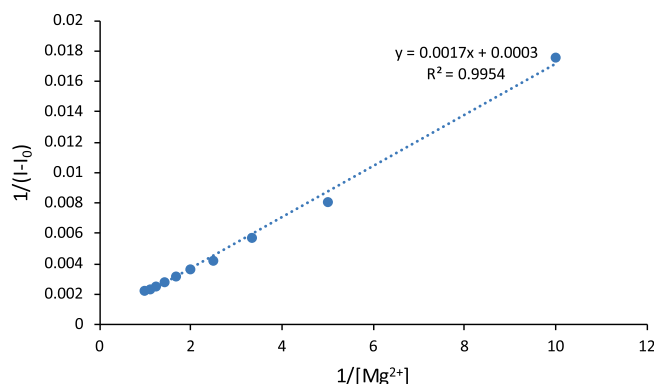


Fig. 11. Benesi-Hildebrand plot of H₂L (10 μM) with Mg²⁺ in EtOH/H₂O (9:1, v/v) solution.

The resulting Benesi-Hildebrand plot, shown in Fig. 8, had a K_a of $1.47 \times 10^6 \text{ M}^{-1}$. The K_a obtained illustrates a strong binding ability of Mg²⁺ with the H₂L.

Some of the reported LOD values as well as the related LOD in the present work for Mg²⁺ were collected and compared in Table 1. The

results shown in Table 1 indicate on exclusive situation of H₂L among the other chemosensors in terms of lowest LOD value [28–38]. Besides high selectivity, a short response time is necessary for a fluorescent chemosensor. To investigate the response time of the chemosensor to Mg²⁺, the kinetics of fluorescence enhancement at 490 nm were carried out. The results showed that the response time of H₂L to Mg²⁺ is very fast. When H₂L titrated with Mg²⁺ ions, fast changes in fluorescence intensity were appeared and remained quite stable.

3.7. Absorption spectra and ground state structure

The absorption spectrum of H₂L shows three peaks at ~270 and ~320 nm were attributed to $\pi \rightarrow \pi^*$ transitions associated with the aromatic rings, whereas the broad band from 360 to 460 nm with its maximum centered at 408 nm, which is assigned to the C=N isomerization of the Schiff base.⁵

The UV-Vis absorption spectrum of H₂L in the presence of Mg²⁺ is almost the same as that of sensor H₂L, as shown in Fig. 12. As there is no shift in the band maxima positions, it can be concluded that the change in fluorescence due to the binding of Mg²⁺ to sensor H₂L is a consequence of excited state phenomena.

3.8. Application

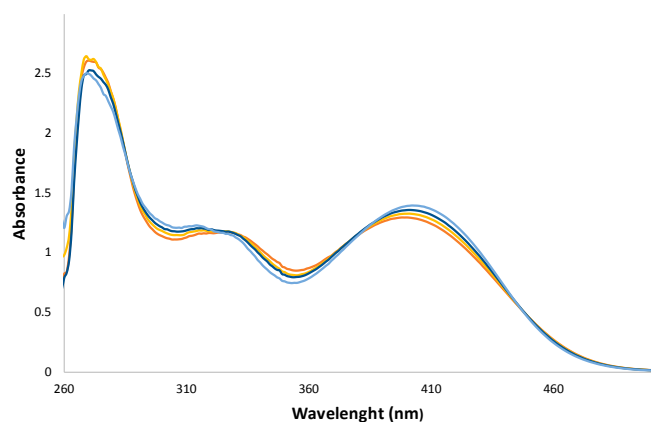
H₂L was employed as a fluorescent chemosensor for the determination of Mg²⁺ in drinking water. Results were satisfactory, agreeing with the atomic absorption spectroscopic (AAS) method as shown in Table 2. The percentages of relative error were less than 10%.

3.9. Logic gate operation

H₂L can be applied as a molecular logic gate. The input signals were selected Mg²⁺ (input1) and Fe³⁺ (input 2) and the fluorescence emission at 490 nm as output. When H₂L coordinated with Mg²⁺ (input 1), the fluorescence emission intensity was increased at 490 nm attributing to the typical PET process. The fluorescence intensity was completely quenched upon the continuous addition of Fe³⁺ (input 2). The fluorescence band of H₂L was also quenched when titration was carried out with Fe³⁺ alone. In a positive logic convention, if the fluorescence

Table 1Comparison of the limit of detection found in the present work with the reported values for Mg^{2+} chemosensors in some of the literatures.

Probe	Medium	LOD (M)	Reference
[2-(quinolin-8'-yloxy) acetyl]hydrazone	Acetonitrile	6.8×10^{-7}	28
2-hydroxy-1-naphthaldehyde derivatives	Aqueous ethanol	2.28×10^{-9}	29
pyridyl-hydrazono-coumarin	Aqueous ethanol	1.05×10^{-7}	30
(E)-N'-((8-hydroxyquinolin-2-yl) methylene) -4-methylbenzohydrazide	DMSO	4.2×10^{-7}	31
(E)-2-(5-allyl-2-hydroxy-3-methoxybenzylidene)-N-phenylhydrazinecarbothioamide	Acetonitrile	8.91×10^{-8}	32
calix[4]arene diamide derivatives	aqueous DMSO	1.3×10^{-5}	33
Schiff base derivatives	DMF	1.0×10^{-7}	34
2-((Z)-1-(2-(2-(1-((Z)-1-(2-hydroxynaphthyl)ethylideneamino)propan-2-yl)disulfanyl)ethylimino)ethyl)naphthalen-1-ol	aqueous DMF	5.0×10^{-8}	35
Schiff base derivatives	Ethanol	1.47×10^{-6}	36
β -Hydroxy- α -naphthaldehyde [2-(quinolin-8'-yloxy) acetyl] hydrazone	Acetonitrile	1.0×10^{-7}	37
naphthalene-based	Acetonitrile	1.47×10^{-6}	38
H ₂ L	Aqueous ethanol	3.0×10^{-9}	Present work

**Fig. 12.** Changes in the absorption spectra of H₂L (50 μ M) as a function of added Mg^{2+} concentration in EtOH/H₂O (9:1, v/v).**Table 2**Determination of Mg^{2+} in drinking water samples.

Drinking waters	AAS method (mg/L)	Proposed method (mg/L)	Relative error (%)
Mineral water	4.4 ± 0.12	4.6 ± 0.12	4.4
Tap water	1.9 ± 0.09	2.1 ± 0.09	10

intensity of 250 was selected as the threshold values, output = 0 when the intensity is lower than 250 and output = 1 when the emission intensity is higher than 250. As discussed above, the intensity of the fluorescence is high enough (output 1 = 1) only under the addition of Mg^{2+} (input 1 = 1 and input 2 = 0). According to the truth table of

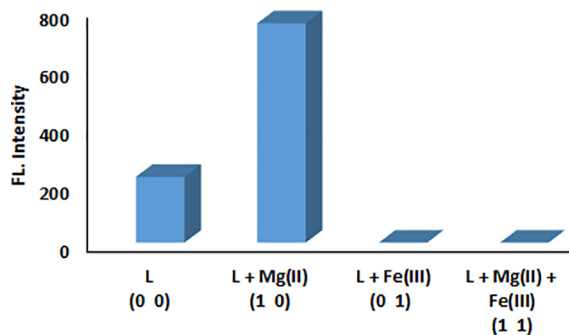


Fig. 13. the fluorescence responses of H₂L with the two chemical inputs Mg^{2+} and Fe^{3+} satisfy the INHIBIT type logic gate function at the molecular level (**Fig. 13**).

4. Conclusion

We have developed a new naphthol-based receptor which acts as a fluorescent chemosensor for Mg^{2+} ions. The stoichiometry of [H₂L- Mg^{2+}] complex is 1:1. The association constant of the [H₂L- Mg^{2+}] complex is $1.47 \times 10^6 M^{-1}$. The lower detection limit of Mg^{2+} is $3.0 \times 10^{-9} M$ and indicate on exclusive situation of H₂L among the other chemosensors in terms of lowest LOD value. Molecular logic gates INHIBIT is proposed using Mg^{2+} and Fe^{3+} as inputs.

CRediT authorship contribution statement

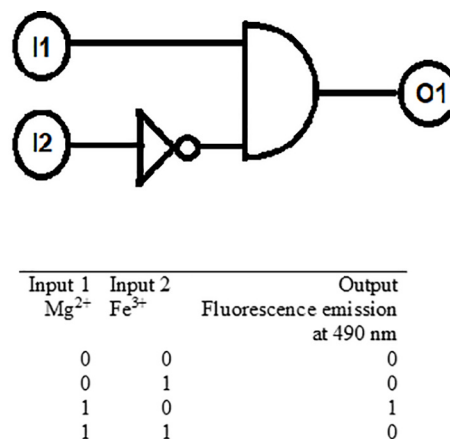
Reza Azadbakht: Conceptualization, Project administration, Writing - original draft. **Mostafa Koolivand:** Investigation, Formal analysis. **Saeid Menati:** Validation, Writing - review & editing.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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**Fig. 13.** Logic scheme and the truth table for the proposed INHIBIT type logic gate.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.ica.2020.120021>.

References

- [1] R. Swaminathan, Magnesium metabolism and its disorders, *Clin. Biochem. Rev.* 24 (2003) 47–66.
- [2] R.K. Rude, Magnesium, in: A.C. Ross, B. Caballero, R.J. Cousins, K.L. Tucker, T.R. Ziegler (Eds.), *Modern Nutrition in Health and Disease*, 11th ed., Lippincott Williams & Wilkins, Baltimore, Mass, 2012, pp. 159–175.
- [3] S.C. Larsson, M.J. Virtanen, S. Männistö, P. Pietinen, D. Albanes, M.J. Virtamo, *Arch. Intern. Med.* 168 (2008) 459–465.
- [4] N.-E.-L. Saris, E. Mervaala, H. Karppanen, J.A. Khawaja, A. Lewenstam, Magnesium, *Clin. Chim. Acta* 294 (2000) 1–26.
- [5] M. Barbagallo, M. Belvedere, D. Sprini, L.J. Dominguez, *Diet and Nutrition in Dementia and Cognitive Decline*, Elsevier, 2015.
- [6] A.W.C. Yuen, J.W. Sander, *EpilepsyRes.* 100 (2012) 152–156.
- [7] V. Trapani, G. Farruggia, C. Marraccini, S. Iotti, A. Cittadini, F.I. Wolf, *Analyst.* 135 (2010) 1855–1866.
- [8] J. Yin, Y. Hu, J. Yoon, *Chem. Soc. Rev.* 44 (2015) 4619–4644.
- [9] G. Men, C. Chen, S. Zhang, C. Liang, Y. Wang, M. Deng, H. Shang, B. Yang, S. Jiang, *Dalton Trans.* 44 (2015) 2755–2762.
- [10] V.K. Gupta, N. Mergu, L.K. Kumawat, A.K. Singh, *Sens. Actuators B Chem.* 207 (2015) 216–223.
- [11] Z. Liu, H. Xu, S. Chen, L. Sheng, H. Zhang, F. Hao, *Spectrochim. Acta. A Mol. Biomol. Spectrosc.* 149 (2015) 83–89.
- [12] V.K. Gupta, B. Sethi, R.A. Sharma, S. Agarwal, A. Bharti, *J. Molecular Liquids* 177 (2013) 114–118.
- [13] V.K. Gupta, H. Karimi-Maleh, R. Sadegh, *Int. J. Electrochem. Sci.* 10 (2015) 303–316.
- [14] V.K. Gupta, A.K. Singh, L.K. Kumawat, *Sens. Actuators B* 195 (2014) 98–108.
- [15] M.H. Dehghani, D. Sanaei, I. Ali, A. Bhatnagar, *J. Molecular Liquids* 215 (2016) 671–679.
- [16] H. Karimi-Maleh, F. Tahernejad-Javazmi, N. Atar, M. Lutfi Yola, V.K. Gupta, A.A. Ensafi, *Ind. Eng. Chem. Res.* 54 (2015) 3634–3639.
- [17] A. Asfaram, M. Ghaedi, S. Agarwal, I. Tyagi, V.K. Gupta, *RSC Adv.* 5 (2015) 18438–18450.
- [18] (a) Y. Suzuki, H. Komatsu, T. Ikeda, N. Saito, S. Araki, D. Citterio, H. Hisamoto, Y. Kitamura, T. Kubota, J. Nakagawa, K. Oka, K. Suzuki, *Anal. Chem.* 74 (2002) 1423–1428;
(b) H. Komatsu, T. Miki, D. Citterio, T. Kubota, Y. Shindo, Y. Kitamura, K. Oka, K. Suzuki, *J. Am. Chem. Soc.* 127 (2005) 10798–10799;
(c) H.M. Kim, P.R. Yang, M.S. Seo, J.S. Yi, J.H. Hong, S.-J. Jeon, Y.-G. Ko, K.J. Lee, B.R. Cho, *J. Org. Chem.* 72 (2007) 2088–2096;
(d) E. Brunet, M.T. Alonso, O. Juannes, R. Sedano, J.C. Rodriguez-Ubis, *Tetrahedron Lett.* 38 (1997) 4459–4462;
(e) L.F. Capitan-Vallvey, M.D. Fernandez-Ramos, A. Lapresta-Fernandez, E. Brunet, J.C. Rodriguez-Ubis, O. Juannes, *Talanta* 68 (2006) 1663–1670.
- [19] (a) D. Ray, P.K. Bharadwaj, *Inorg. Chem.* 47 (2008) 2252–2254;
(b) Z.D. Liu, H.J. Xu, C.F. Song, D.Q. Huang, L.Q. Sheng, R.H. Shi, *Chem. Lett.* 40 (2011) 75–77;
(c) L.N. Wang, W.W. Qin, X.L. Tang, W. Dou, W.S. Liu, *J. Phys. Chem. A* 115 (2011) 1609–1616.
- [20] (a) T. Shoda, K. Kikuchi, H. Kojima, Y. Urano, H. Komatsu, K. Suzuki, T. Nagano, *Analyst* 128 (2003) 719–723;
(b) H. Komatsu, N. Iwasawa, D. Citterio, Y. Suzuki, T. Kubota, K. Tokuno, Y. Kitamura, K. Oka, K. Suzuki, *J. Am. Chem. Soc.* 126 (2004) 16353–16360;
(c) G. Farruggia, S. Iotti, L. Prodi, M. Montalti, N. Zaccheroni, P.B. Savage, V. Trapani, P. Sale, F.I. Wolf, *J. Am. Chem. Soc.* 128 (2006) 344–350;
(d) H. Hama, T. Morozumi, H. Nakamura, *Tetrahedron Lett.* 48 (2007) 1859–1861;
(e) X.H. Dong, J.H. Han, C.H. Heo, H.M. Kim, Z.H. Liu, B.R. Cho, *Anal. Chem.* 84 (2012) 8110–8113;
(f) S.C. Schwartz, B. Pinto-Pacheco, J.P. Pitteloud, D. Buccella, *Inorg. Chem.* 53 (2014) 3204–3209.
- [21] (a) X.M. He, E.C.C. Cheng, N.Y. Zhu, V.W.W. Yam, *Chem. Commun.* 27 (2009) 4016–4018;
(b) X. Zhu, C. He, D.P. Dong, Y. Liu, C.Y. Duan, *Dalton Trans.* 39 (2010) 10051–10055;
(c) Y. Dong, J.F. Li, X.X. Jiang, F.Y. Song, Y.X. Cheng, C.J. Zhu, *Org. Lett.* 13 (2011) 2252–2255;
(d) M. Tian, H. Ihmels, S. Ye, *Org. Biomol. Chem.* 10 (2012) 3010–3018;
(e) C. Marraccini, G. Farruggia, M. Lombardo, L. Prodi, M. Sgarzi, V. Trapani, C. Trombini, F.I. Wolf, N. Zaccheroni, S. Iotti, *Chem. Sci.* 3 (2012) 727–734;
(f) L. Zhao, Y. Liu, C. He, J. Wang, C. Duan, *Dalton Trans.* 43 (2014) 335–343;
(g) A. Sargentini, G. Farruggia, E. Malucelli, C. Cappadone, L. Merolle, C. Marraccini, G. Andreani, L. Prodi, N. Zaccheroni, M. Sgarzi, C. Trombini, M. Lombardo, S. Iotti, *Analyst* 139 (2014) 1201–1207.
- [22] (a) K.C. Song, M.G. Choi, D.H. Ryu, K.N. Kim, S.K. Chang, *Tetrahedron Lett.* 48 (2007) 5397–5400;
(b) N. Singh, N. Kaur, R.C. Mulrooney, J.F. Callan, *Tetrahedron Lett.* 49 (2008) 6690–6692.
- [23] K. Rout, A.K. Manna, M. Sahu, G.K. Patra, *Inorg. Chim. Acta* 486 (2019) 733–741.
- [24] M.S. Kim, T. GeunJo, M. Yang, J. Han, M.H. Lim, C. Kim, *Spectrochim. Acta. A Mol. Biomol. Spectrosc.* 211 (2019) 34–43.
- [25] V. Raju, R. Selva Kumar, Y. Tharakeswar, S.K. Ashok Kumar, *Inorg. Chim. Acta* 493 (2019) 49–56.
- [26] R. Azadbakht, M. Koolivand, J. Khanabadi, *Anal. Methods* 9 (2017) 4688–4694.
- [27] L.K. Kumawat, N. Mergu, M. Asif, V.K. Gupta, *Sens. Actuators B Chem.* 231 (2016) 847–859.
- [28] J.-C. Qin, Z.-Y. Yang, L. Fan, B.-D. Wang, *Spectrochim. Acta. A Mol. Biomol. Spectrosc.* 140 (2015) 21–26.
- [29] Z. Liu, H. Xu, S. Chen, L. Sheng, H. Zhang, F. Hao, P. Su, W. Wang, *Spectrochim. Acta. A Mol. Biomol. Spectrosc.* 149 (2015) 83–89.
- [30] J. Orrego-Hernández, N. Nuñez-Dallos, J. Portilla, *Talanta* 152 (2016) 432–437.
- [31] P. Marimuthu, A. Ramu, *Sens. Actuators B Chem.* 266 (2018) 384–391.
- [32] M. Patil, K. Keshav, M.K. Kumawat, S. Bothra, S.K. Sahoo, R. Srivastava, J. Rajput, R. Bendre, A. Kuwar, *J. Photoch. Photobio. A* 364 (2018) 758–763.
- [33] K.C. Song, M.G. Choi, D.H. Ryu, K.N. Kim, S.-K. Chang, *Tetrahedron Lett.* 48 (2007) 5397–5400.
- [34] P.S. Hariharan, S.P. Anthony, *RSC Adv.* 4 (2014) 41565–41571.
- [35] G. Tamil Selvan, V. Chitra, Israel V.M.V. Enocha, P. Mosae Selvakumar, *New J. Chem.* 42 (2018) 902–909.
- [36] T. Yu, P. Sun, Y. Hu, Y. Ji, H. Zhou, B. Zhang, Y. Tian, J. Wu, *Biosensors Bioelectronics* 86 (2016) 677–682.
- [37] J.-C. Qin, Z.-Y. Yang, L. Fan, B.-D. Wang, *Spectrochim. Acta. A Mol. Biomol. Spectrosc.* 140 (2015) 21–26.
- [38] J.-C. Qin, Z.-Y. Yang, G.-Q. Wang, *Inorg. Chim. Acta* 435 (2015) 194–199.