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Synthesis of Dissymmetric Indolocarbazole Glycosides Using the Mitsunobu Reaction at the Glycosylation Step

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Abstract: A novel method for the synthesis of N-glycosylated dissymmetric indolo[2,3-a]pyrrolo-[3,4-c]carbazole derivatives was developed by applying the Mitsunobu reaction to the N-glycosylation reaction of substituted indole substrates. © 1997 Elsevier Science Ltd.

INTRODUCTION

Indolo[2,3-*a*]pyrrolo[3,4-*c*]carbazole compounds such as staurosporine $(1)^{11}$, K-252a $(2)^{21}$ and rebeccamycin $(3)^{31}$ are known to have various biological effects. We recently reported that NB-506 $(4)^{41}$ is a potent antitumor agent that has a wider therapeutic index in mice than currently available anticancer drugs against murine and human solid tumors.

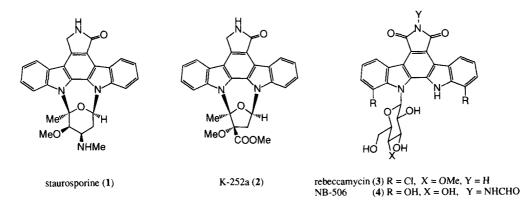
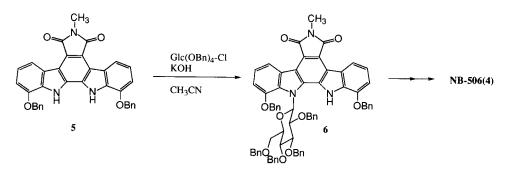


Fig. 1

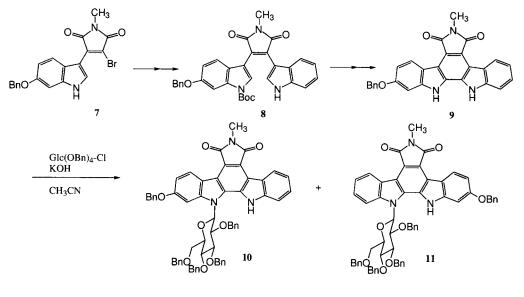
To discover anticancer agents more potent than NB-506, the synthesis of dissymmetric indolocarbazole glycosides was planned. However, the method of synthesizing NB-506⁴⁰ shown in Scheme 1 could not be applied to the synthesis of an dissymmetric indolocarbazole glycoside because of lack of regioselectivity (10:11

= 1:1) (Scheme 2). The glycosylation of compound 7 or 8^{5} was also unsuccessful because these compounds were unstable to KOH. Glycosylation reactions proceeding under neutral conditions were subsequently investigated, and the Mitsunobu reaction was found to be applicable to the glycosylation of compounds 7 or 8.

This communication reports the details of glycosylation by the Mitsunobu reaction.





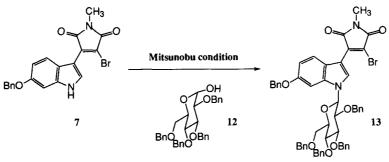


Scheme 2

RESULTS AND DISCCUSSION

The Mitsunobu reaction⁶⁾ is widely used for the alkylation of various acids (or nucleophiles; HA) utilizing diethyl azodicarboxylate (DEAD) - triphenylphosphine (PPh₃) with the Walden inversion. However, few applications to a glycosylation reaction have been reported⁷⁾ probably because the pKa of HA is required to be

lower than 13⁸). Since the pKa value of compound 7 was supposed⁹ to be less than 13, we tried to apply the Mitsunobu reaction to the glycosylation of compound 7. We tested several reagents⁸ which were developed to improve the Mitsunobu reaction. As shown in Table 1, the glycosylation of compound 7 with 2,3,4,6-tetra-O-benzyl- α -D-glucose 12 in the presence of both DIAD and PPh₃ yielded the desired β -glucoside 13 in 79-82% yields without formation of an α -anomer.



Scheme 3

Table 1 Glycosylations of 7 with 12 under several Mitsunobu conditions^{a)}

Mitunobu reagent (eq.) ^{b)}		Phosphine (eq.)		12 (eq.)	yield (%) ^{c)}
DEAD	(1.5)	PPh ₃	(1.5)	1.5	73
DEAD	(1.5)	PBu ₂	(1.5)	1.5	48
DIAD	(1.5)	PPh	(1.5)	1.5	79
DBAD	(1.5)	PPh ₃	(1.5)	1.5	41 ^d
TMAD	(1.5)	PBu ₃	(1.5)	1.5	56 ^{d)}
ADDP	(1.5)	PBu ₃	(1.5)	1.5	34
DIAD	(3.0)	PPh ₃	(3.0)	3.0	82

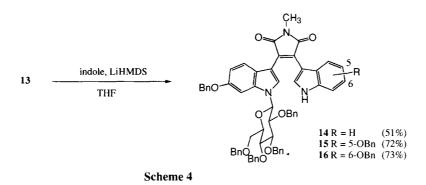
a) The reaction was carried out at -40-0 °C for 1-2 h.

 b) DEAD (diethylazodicarboxylate), DIAD (diisopropylazodicarboxylate), DBAD (di-tert-butylazodicarboxylate), TMAD (N,N,N',N'-tetramethylazocarboxamide), ADDP [1,1'-(azodicarbonyl)dipiperazine]

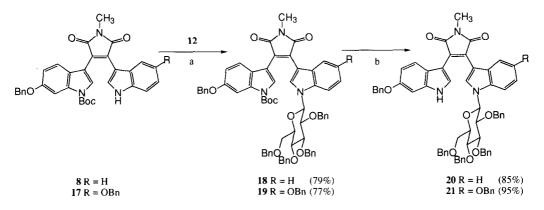
c) The yield of β -glucoside 10; α -anomer was not observed.

d) The reaction mixture was left overnight, but the reaction was almost over after 2 h.

A second indolylation reaction of bromide 13 with three kinds of indole using a two-fold excess of Litium hexamethyldisilazide (LiHMDS) afforded dissymmetric compounds 14 and 15 as well as a symmetric compound 16 in good yields. (Scheme 4).



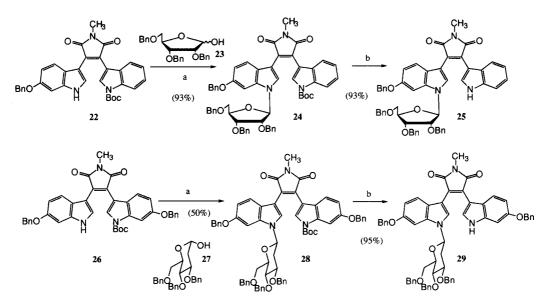
As shown in Scheme 5, the Mitsunobu reaction was also applicable to the glycosylation of dissymmetric bisindolyl compounds 8 and 17^{5} to respectively give β -glucosides 18 and 19 in good yields; in these cases, α -anomers were not detected. The removal of the Boc group of compounds 18 and 19 using methylamine afforded glycosylated bisindolylmaleimides 20 and 21, respectively, in good yields.



Scheme 5 reagents; (a) DIAD, PPh₃; (b) 40% MeNH₂-MeOH

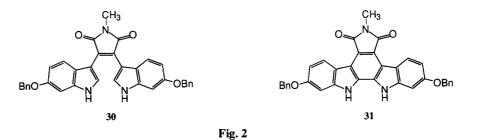
This glycosylation was utilized for synthesizing other sugar derivatives. As shown in Scheme 6, the glycosylation of compound 22 with ribofuranose 23 ($\alpha:\beta = 1:1$) proceeded at -40 °C to provide only β -ribofuranoside 24. The yield of 24 depended on the number of equivalents of 23 that were used: 1.0, 2.0 and 3.0 equiv of 23 gave 24 in 20, 84 and 93% yields, respectively. The anomeric configuration of 24 was determined by ROE studies in the C1-C4 region as well as 2D-COSY experiments. Furthermore, the glycosylation of 26⁵ with 2-deoxyglucose 27(α -anomer) afforded β -2-deoxyglucoside 28 in 50% yield; the configuration was determined by analysis of the ¹H NMR coupling constants and a 2D-COSY experiment.

Removal of the Boc group of 24 and 28 with methylamine afforded deprotected compounds 25 and 29, respectively, in good yields.

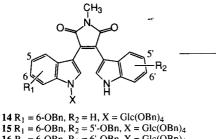


Scheme 6 reagents; (a) DIAD, PPh3; (b) 40% MeNH2-MeOH

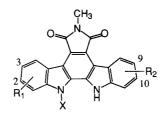
We also tried to glycosylate bisindolylmaleimide 30 and indolocarbazole 31^{5} in a similar manner, but the reactions did not proceed. The absence of an electron-withdrawing group such as Br or Boc in these compounds may have caused the decrease in reactivity.



The oxidative cyclization of glycosylated bisindolyl compounds with $CuCl_2$ and $PdCl_2$ gave the corresponding indolopyrrolocarbazole glycosides in good yields (Table 2). In contrast, the use of DDQ⁵ was unsuccessful.



16 $R_1 = 6$ -OBn, $R_2 = 6$ -OBn, $X = Glc(OBn)_4$ 20 $R_1 = H, R_2 = 6$ -OBn, $X = Glc(OBn)_4$ 21 $R_1 = 5$ -OBn, $R_2 = 6$ -OBn, $X = Glc(OBn)_4$ 25 $R_1 = H, R_2 = 6$ -OBn, $X = Rib(OBn)_3$ 29 $R_1 = 6$ -OBn, $R_2 = 6$ -OBn, X = 2-deoxy-Glc(OBn)_3



 $\begin{array}{l} \textbf{32} \ R_1 = 2\text{-OBn}, \ R_2 = H, \ X = Glc(OBn)_4 \\ \textbf{33} \ R_1 = 2\text{-OBn}, \ R_2 = 9\text{-OBn}, \ X = Glc(OBn)_4 \\ \textbf{34} \ R_1 = 2\text{-OBn}, \ R_2 = 10\text{-OBn}, \ X = Glc(OBn)_4 \\ \textbf{35} \ R_1 = H, \ R_2 = 10\text{-OBn}, \ X = Glc(OBn)_4 \\ \textbf{36} \ R_1 = 3\text{-OBn}, \ R_2 = 10\text{-OBn}, \ X = Glc(OBn)_4 \\ \textbf{37} \ R_1 = H, \ R_2 = 10\text{-OBn}, \ X = Glc(OBn)_3 \\ \textbf{38} \ R_1 = 2\text{-OBn}, \ R_2 = 10\text{-OBn}, \ X = 2\text{-deoxy-Glc}(OBn)_3 \\ \textbf{38} \ R_1 = 2\text{-OBn}, \ R_2 = 10\text{-OBn}, \ X = 2\text{-deoxy-Glc}(OBn)_3 \\ \textbf{38} \ R_1 = 2\text{-OBn}, \ R_2 = 10\text{-OBn}, \ X = 2\text{-deoxy-Glc}(OBn)_3 \\ \textbf{38} \ R_1 = 2\text{-OBn}, \ R_2 = 10\text{-OBn}, \ X = 2\text{-deoxy-Glc}(OBn)_3 \\ \textbf{38} \ R_1 = 2\text{-OBn}, \ R_2 = 10\text{-OBn}, \ X = 2\text{-deoxy-Glc}(OBn)_3 \\ \textbf{38} \ R_1 = 2\text{-OBn}, \ R_2 = 10\text{-OBn}, \ X = 2\text{-deoxy-Glc}(OBn)_3 \\ \textbf{38} \ R_1 = 2\text{-OBn}, \ R_2 = 10\text{-OBn}, \ X = 2\text{-deoxy-Glc}(OBn)_3 \\ \textbf{38} \ R_1 = 2\text{-OBn}, \ R_2 = 10\text{-OBn}, \ X = 2\text{-deoxy-Glc}(OBn)_3 \\ \textbf{38} \ R_1 = 2\text{-OBn}, \ R_2 = 10\text{-OBn}, \ X = 2\text{-deoxy-Glc}(OBn)_3 \\ \textbf{38} \ R_1 = 2\text{-OBn}, \ R_2 = 10\text{-OBn}, \ X = 2\text{-deoxy-Glc}(OBn)_3 \\ \textbf{38} \ R_1 = 2\text{-OBn}, \ R_2 = 10\text{-OBn}, \ X = 2\text{-deoxy-Glc}(OBn)_3 \\ \textbf{38} \ R_1 = 2\text{-OBn}, \ R_2 = 10\text{-OBn}, \ X = 2\text{-deoxy-Glc}(OBn)_3 \\ \textbf{38} \ R_1 = 2\text{-OBn}, \ R_2 = 10\text{-OBn}, \ X = 2\text{-deoxy-Glc}(OBn)_3 \\ \textbf{38} \ R_1 = 2\text{-OBn}, \ R_2 = 10\text{-OBn}, \ X = 2\text{-deoxy-Glc}(OBn)_3 \\ \textbf{38} \ R_1 = 2\text{-OBn}, \ R_2 = 10\text{-OBn}, \ R_2 = 10\text{-OBn}$

Scheme 7

Table 2 Oxidative cyclization of glycosylated bisindolylmaleimide

bisindole	reagent	solvent ^{a)}	temp (°C)	product	yield (%)
14	CuCl ₂	MEK	70	32	80
15	$CuCl_2$	MEK	60	33	80
16	$CuCl_2$	MEK	30	34	84
20	$PdCl_2$	DMF	100	35	78
21	PdCl ₂	DMF	110	36	77
25	CuCl ₂	MEK	80	37	98
29	$CuCl_2$	MEK	60	38	71

a) MEK (methylethylketone), DMF (N,N-dimethylformamide)

SUMMARY

The synthesis of dissymmetric indolocarbazole glycosides was achieved by applying the Mitsunobu reaction at the glycosylation step, which made it possible to evaluate the influence of differences in the position of a hydroxyl group on antitumor activities

EXPERIMENTAL

¹H and ¹³C NMR were recorded on a Varian VXR-300, Jeol JNM-EX 400 or JNM-A 500 instrument. Infrared spectra were recorded on a Horiba FT-200 spectrometer. High-resolution mass spectra (HRMS) were recorded on a Jeol JMS-SX 102A instrument, and optical rotations were measured on a Fisons Model EA 1108 polarimeter. Melting points were determined on a Mettler FP 62 or Yanako Model MP-S3 melting point apparatus and are uncorrected.

General Procedure for the Mitsunobu Glycosylation: DIAD was added to a solution of indole (0.01-0.1 M), PPh₃ and sugar in dry THF at -78 to 0 °C under a nitrogen atmosphere. The reaction mixture was stirred at -40 to 0 °C until TLC indicated complete consumption of the indole (15-60 min). The reaction mixture

was poured into 0.2 M aqueous hydrochloric acid, and the mixture was extracted with ethyl acetate. The organic phase was successively washed with aqueous NaHCO₃, H₂O and saturated brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was chromatographed on silica gel to yield the indolyl β -glycoside.

4-Bromo-2,5-dihydro-3-[6-benzyloxy-1-(2,3,4,6-tetra-*O***-benzyl**-β**-D-glucopyranosyl**)-1*H***indol-3-yl**]-1-methyl-1*H*-pyrrolo-2,5-dione (13): From 7 (200 mg, 0.49 mmol), PPh₃ (382 mg, 1.47 mmol), 2,3,4,6-tetra-*O*-benzyl-D-glucose 12 (α- dominant, Sigma, 787 mg, 1.47 mmol) and DIAD (0.28 mL, 1.47 mmol), 13 (373 mg, 0.40 mmol) was obtained as a yellow amorphous substrate (82%) after chromatography (toluene-ethyl acetate (50:1-15:1)). $[α]^{20}_{D}$ -46.4° (c 0.16, DMSO); IR (KBr) v_{max} 1767, 1707, 1603, 1454, 1379, 1090 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.92 (1H, d, J = 8.9 Hz), 7.93 (1H, s), 7.17-7.41 (20H, m), 6.97-7.17 (5H, m), 6.71 (2H, dd, J = 1.3, 8.9 Hz), 5.29 (1H, d, J = 8.4 Hz), 5.00 (1H, d, J = 7.8 Hz), 4.97 (1H, d, J = 7.8 Hz), 4.93 (1H, d, J = 11.9 Hz), 4.92 (1H, d, J = 10.8 Hz), 4.90 (1H, d, J = 11.9 Hz), 4.52 (1H, d, J = 11.9 Hz), 4.17 (1H, d, J = 10.0 Hz), 3.75-3.96 (5H, m), 3.71 (1H, br d, J = 9.6 Hz), 3.56 (1H, br d, J = 9.6 Hz), 3.56 (1H, d, J = 10.0 Hz), 3.16 (3H, s); HRMS (FAB) calcd for C₅₄H₄₉N₂O₅Br 932.2627, found 932.2694.

General Procedure for the indolylation: LiHMDS (1 M in THF, 2.1-2.4 equiv) was added to a solution of 1.0-1.2 equiv of the appropriate indole (0.05 M) in THF at -20 °C under a nitrogen atmosphere and stirred for 30-45 min. A solution of the bromide (0.1 M) in THF was then added by drip over 30-60 min, followed by stirring for 15 min at 0 °C. The reaction mixture was poured into 0.2 M aqueous hydrochloric acid, and the mixture was extracted with ethyl acetate. The organic phase was successively washed with aqueous NaHCO₃, H_2O and saturated brine, dried over Na_2SO_4 and concentrated in vacuo. The residue was chromatographed to obtain the glycosylated bisindolylmaleimide.

2,5-Dihydro-3-[6-benzyloxy-1-(2,3,4,6-tetra-*O***-benzyl**-*β***-D-glucopyranosyl**)-1*H***-indol-3yl**]-4-(1*H***-indol-3-yl**)-1-**methyl**-1*H***-pyrrolo-2,5-dione** (14): From indole (27 mg, 0.21 mmol), LiHMDS (1 M in THF, 0.51 mL, 0.51 mmol) and 13 (200 mg, 0.21 mmol), the bisindolyl maleimide 14 (104 g, 0.11 mmol) was obtained as an orange amorphous substrate (51%) after chromatography on silica gel (toluene-ethyl acetate (50:1-15:1)). $[\alpha]_{D}^{20}$ -61.6° (c 0.50, DMSO); IR (KBr) ν_{max} 3411, 1697, 1540, 1456, 1093, 1027 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 8.40 (1H, s), 7.95 (1H, s), 7.61 (1H, d, J = 1.5 Hz), 7.03-7.40 (26H, m), 6.79 (2H, dd, J = 1.5, 7.8 Hz), 6.65 (1H, d, J = 8.7 Hz), 6.63 (1H, dd, J = 1.8, 9.0 Hz), 6.42 (1H, dd, J = 2.1, 9.0 Hz), 5.32 (1H, d, J = 8.4 Hz), 4.83-4.93 (5H, m), 4.66 (1H, d, J = 10.2 Hz), 4.57 (2H, q, J = 11.7 Hz), 4.12 (1H, d, J = 9.6 Hz), 3.93 (2H, t, J = 9.3 Hz), 3.80-3.83 (4H, m), 3.68 (2H, d, J = 10.2 Hz), 3.19 (3H, s); HRMS (FAB) calcd for C₆₉H₆₁N₃O₉ 969.3989, found 969.3990.

2,5-Dihydro-3-[6-benzyloxy-1-(2,3,4,6-tetra-O-benzyl- β -D-glucopyranosyl)-1H-indol-3yl]-4-(5-benzyloxy-1H-indol-3-yl)-1-methyl-1H-pyrrolo-2,5-dione (15): From 5-benzyloxyindole (49 mg, 0.21 mmol), LiHMDS (1 M in THF, 0.51 mL, 0.51 mmol) and 13 (200 mg, 0.21 mmol), the bisindolyl maleimide 15 (163 mg, 0.15 mmol) was obtained as an orange amorphous substrate (72%) after chromatography on silica gel (toluene-ethyl acetate (50:1-25:1)). $[\alpha]_{D}^{20}$ -13.6° (c 0.50, DMSO); IR (KBr) ν_{max} 3412, 1697, 1456, 1218, 1093 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 8.36 (1H, s), 7.78 (1H, d, J = 1.8 Hz), 7.75 (1H, s), 7.03-7.35 (31H, m), 6.92 (1H, d, J = 9.0 Hz), 6.75 (1H, dd, J = 2.1, 8.4 Hz), 6.70 (2H, d, J = 8.4 Hz), 6.56 (1H, dd, J = 2.4, 9.0 Hz), 6.54 (1H, d, J = 2.4 Hz), 5.25 (1H, d, J = 9.0 Hz), 4.87 (2H, d, J = 8.1 Hz), 4.84 (2H, s), 4.62 (1H, d, J = 11.1 Hz), 4.42 (1H, d, J = 12.3 Hz), 4.33 (1H, d, J = 11.4 Hz), 4.32 (1H, d, J = 11.4 Hz), 4.24 (1H, d, J = 12.3 Hz), 4.13 (1H, d, J = 10.2 Hz), 3.89 (2H, dt, J = 3.0, 9.3 Hz), 3.77 (1H, d, J = 9.3 Hz), 3.70 (1H, dd, J = 3.1, 11.1 Hz), 3.62 (1H, d, J = 10.2 Hz), 3.52 (1H, m), 3.40 (1H, d, J = 9.9 Hz), 3.19 (3H, s); HRMS (FAB) calcd for $C_{69}H_{61}N_3O_9$ 1075.4408, found 1075.4392.

2,5-Dihydro-3-[6-benzyloxy-1-(2,3,4,6-tetra-*O***-benzyl-***β***-D-glucopyranosyl)-1***H***-indol-3-yl]-4-(6-benzyloxy-1***H***-indol-3-yl]-1-methyl-1***H***-pyrrolo-2,5-dione** (16): From 6-benzyloxyindole (360 mg, 1.61 mmol), LiHMDS (1 M in THF, 3.45 mL, 3.45 mmol) and **13** (1480 mg, 1.59 mmol), the bisindolyl maleimide **16** (1234 mg, 1.15 mmol) was obtained as an orange amorphous substrate (73%) after chromatography on silica gel (hexane-ethyl acetate (4:1)). $[\alpha]^{20}_{D}$ -67.2° (c 0.25, DMSO); IR (KBr) v_{max} 3311, 1697, 1621, 1533, 1454, 1385, 1090 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 8.29 (1H, d, J = 2.7 Hz), 7.96 (1H, s), 7.52 (1H, d, J = 2.7 Hz), 7.19-7.40 (25H, m), 7.03-7.16 (5H, m), 6.78-6.84 (3H, m), 6.67 (1H, d, J = 8.8 Hz), 6.45 (1H, dd, J = 2.2, 8.8 Hz), 6.34 (1H, dd, J = 2.2, 8.8 Hz), 5.34 (1H, d, J = 8.7 Hz), 4.82-4.94 (7H, m), 4.67 (1H, d, J = 10.7 Hz), 4.62 (1H, d, J = 12.2 Hz), 4.53 (1H, d, J = 12.2 Hz), 4.12 (1H, d, J = 10.2 Hz), 3.67-3.98 (7H, m), 3.18 (3H, s); HRMS (FAB) calcd for C₆₉H₆₁N₃O₉ 1075.4408, found 1075.4392.

2,5-Dihydro-3-[6-benzyloxy-1-(2,3,4,6-tetra-*O*-benzyl- β -D-glucopyranosyl)-1*H*-indol-3yl]-4-[1-(*tert*-butoxycarbonyl)-1*H*-indol-3-yl]-1-methyl-1*H*-pyrrolo-2,5-dione (18): From 8 (4.0 g, 7.3 mmol), PPh₃ (5.74 g, 21.9 mmol), 2,3,4,6-tetra-*O*-benzyl-D-glucose 12 (11.8 g, 21.9 mmol) and DIAD (4.3 mL, 21.9 mmol), 18 (6.2g, 5.8 mmol) was obtained as a yellow amorphous substrate (79%) after chromatography (toluene-ethyl acetate (50:1)). $[\alpha]^{20}_{\ D}$ -27.2° (c 1.00, DMSO); IR (KBr) v_{max} 1734, 1701, 1585, 1541, 1456, 1363, 1217, 1153, 1093 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 8.13 (1H, s), 7.95 (1H, s), 7.78 (1H, br s), 7.52 (1H, d, J = 8.7 Hz), 6.96-7.45 (24H, m), 6.92 (1H, d, J = 8.3 Hz), 6.65-6.85 (4H, m), 6.23 (1H, dd, J = 2.6, 8.7 Hz), 5.43 (1H, d, J = 8.9 Hz), 4.78-4.92 (5H, m), 4.48-4.70 (3H, m), 4.14 (1H, d, J = 9.9 Hz), 3.60-4.03 (7H, m), 3.20 (3H, s), 1.64 (9H, s); HRMS (FAB) calcd for C₆₇H₆₃N₃O₁₀ 1069.4514, found 1069.4513.

$2,5-Dihydro-3-[6-benzyloxy-1-(2,3,4,6-tetra-O-benzyl-\beta-D-glucopyranosyl)-1H-indol-3-yl]-4-[5-benzyloxy-1-(tert-butoxycarbonyl)-1H-indol-3-yl]-1-methyl-1H-pyrrolo-2,5-dione$

(19): From 17 (8.50 g, 13.0 mmol), PPh₃ (8.52 g, 32.5 mmol), 2,3,4,6-tetra-*O*-benzyl-D-glucose 12 (17.57 g, 32.5 mmol) and DIAD (6.4 mL, 32.5 mmol), 10 (11.71 g, 9.96 mmol) was obtained as a yellow amorphous substrate (77%) after chromatography (hexane-ethylacetate(3:1), toluene-ethyl acetate (50:1)). $[\alpha]_{D}^{20}$ -35.2° (c 1.00, DMSO); IR (KBr) ν_{max} 1734, 1701, 1616, 1454, 1369, 1213, 1153, 1091 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 8.24 (1H, s), 7.84 (1H, s), 7.81 (1H, br s), 7.42 (1H, d, J = 9.1 Hz), 7.04-7.37 (28H, m), 7.00 (1H, d, J = 8.8 Hz), 6.72-6.81 (3H, m), 6.34-6.40 (2H, m), 5.38 (1H, d, J = 8.8 Hz), 4.80-4.93 (5H, m), 4.65 (1H, d, J = 10.7 Hz), 4.62 (1H, d, J = 12.1 Hz), 4.54 (1H, d, J = 12.1 Hz), 3.62-4.18 (10H, m), 3.21 (3H, s), 1.51 (9H, s); HRMS (FAB) calcd for $C_{74}H_{69}N_3O_{11}$ 1175.4932, found 1175.4969.

General Procedure for Boc deprotection: 40% methylamine in methanol was added to the Bocprotected compound and the reaction mixture was stirred for 0.5-2 h at room temp. The solvent was removed in vacuo and the residue was purified to afford the deprotected compound.

2,5-Dihydro-3-[6-benzyloxy-1-(2,3,4,6-tetra-*O***-benzyl-***β***-D-glucopyranosyl)-1***H***-indol-3-yl]-4-(1***H***-indol-3-yl)-1-methyl-1***H***-pyrrolo-2,5-dione** (**20**): From **18** (6.2 g, 5.79 mmol) and 40% methylamine in methanol (100 mL), compound **20** (4.78 g, 4.93 mmol) was obtained as an orange amorphous substrate (85%) after chromatography (hexane-ethyl acetate (2:1)). $[\alpha]_{D}^{20}$ -28.0° (c 0.25, DMSO); IR (KBr) ν_{max} 3241, 1697, 1539, 1456, 1385, 1159, 1093, 1028 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) & 8.24 (1H, br s), 8.06 (1H, s), 7.48-7.58 (3H, m), 6.99-7.39 (24H, m), 6.67-6.85 (5H, m), 6.30 (1H, dd, J = 2.3, 8.3 Hz), 5.45 (1H, d, J = 8.9 Hz), 4.80-4.92 (5H, m), 4.66 (1H, d, J = 10.9 Hz), 4.61 (1H, d, J = 12.5 Hz), 4.53 (1H, d, J = 12.5 Hz), 4.16 (1H, d, J = 8.1 Hz), 3.60-4.08 (7H, m), 3.19 (3H, s); HRMS (FAB) calcd for C₆₂H₅₃N₃O₈ 969.3989, found 969.3956.

2,5-Dihydro-3-[6-benzyloxy-1-(2,3,4,6-tetra-*O***-benzyl**- β **-D-glucopyranosyl**)-1*H***-indol-3-yl**]-**4**-(**5-benzyloxy-1***H***-indol-3-yl**]-**1-methyl**-1*H***-pyrrolo-2,5-dione** (**21**): From **19** (122 mg, 0.10 mmol) and 40% methylamine in methanol (1 mL), compound **21** (105 mg, 0.097 mmol) was obtained as an orange amorphous substrate (95%) after chromatography (hexane-ethyl acetate (2:1)). $[\alpha]_{D}^{20}$ -41.6° (c 0.25, DMSO); IR (KBr) v_{max} 3402, 1697, 1541, 1456, 1387, 1157, 1093, 1026 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 8.33 (1H, d, J = 2.3 Hz), 8.15 (1H, s), 7.05-7.45 (31H, m), 6.46 (1H, dd, J = 2.3, 9.8 Hz), 6.23 (1H, d, J = 2.3 Hz), 5.39 (1H, d, J = 8.8 Hz), 4.83-4.92 (3H, m), 4.80 (2H, s), 4.64 (1H, d, J = 10.8 Hz), 4.59 (1H, d, J = 12.2 Hz), 4.52 (1H, d, J = 12.2 Hz), 4.00-4.15 (4H, m), 3.46-3.90 (5H, m), 3.18 (3H, s); HRMS (FAB) calcd for C_{6.9}H_{6.1}N₃O₉ 1075.4408, found 1075.4349.

2,5-Dihydro-3-(6-benzyloxy-1H-indol-3-yl)-4-[1-(*tert***-butoxycarbonyl)-1H-indol-3-yl]-1**methyl-1H-pyrrolo-2,5-dione (22): From 6-benzyloxyindole (1.78 g, 8.00 mmol), LiHMDS (1M in THF, 17.6 mL, 17.6 mmol) and 4-bromo-2,5-dihydro-3-[(1-*tert*-butoxycarbonyl)-1H-indol-3-yl]-1-methyl-1H- pyrrol - 2,5-dione (2.95 g, 7.28 mmol), the bisindolyl maleimide 22 (3.42 g, 6.25 mmol) was obtained as an orange solid (86%) after chromatography on silica gel (hexane-ethyl acetate (3:1)). mp 129-132 °C: IR (KBr) v_{max} 3363, 1734, 1697, 1541, 1456, 1387, 1373, 1252, 1236, 1153, 1066 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 8.37 (1H, br s), 8.14 (1H, d, J = 8.7 Hz), 8.05 (1H, s), 7.70 (1H, d, J = 2.1 Hz), 7.10-7.42 (6H, m), 6.72-6.99 (4H, m), 6.55 (1H, dd, J = 2.4, 8.9 Hz), 5.01 (2H, s), 3.19 (3H, s), 1.68 (9H, s); HRMS (FAB) calcd for C₁₃H₂₀N₃O₅ 547.2107, found 547.2098.

2,5-Dihydro-3-[6-benzyloxy-1-(2,3,5-tri-*O*-benzyl- β -D-ribofuranosyl)-1*H*-indol-3-yl]-4-[1-(*tert*-butoxycarbonyl)-1*H*-indol-3-yl]-1-methyl-1*H*-pyrrolo-2,5-dione (24): From 22 (2.19 g, 4.00 mmol), PPh₃ (3.12 g, 11.9 mmol), 2,3,5-tri-*O*-benzyl-D-ribofuranose 23 ($\alpha/\beta = 1/1$, 5.0 g, 11.9 mmol) and DIAD (2.34 mL, 11.9 mmol), 24 (3.49 g, 3.68 mmol) was obtained as a yellow amorphous powder (93%) after chromatography (toluene-ethyl acetate (30:1)). [α]²⁰_D -70.0° (c 1.00, DMSO); IR (KBr) v_{max} 1734, 1701, 1568, 1541, 1456, 1371, 1209, 1153 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 8.14 (1H, d, J = 8.2 Hz), 8.03 (1H, s), 7.89 (1H, s), 7.10-7.40 (22H, m), 7.00 (1H, d, J = 8.9 Hz), 6.86 (1H, d, J = 7.9 Hz), 6.75 (1H, d, J = 7.3 Hz), 6.58 (1H, dd, J = 2.2, 9.1 Hz), 6.08 (1H, d, J = 5.6 Hz), 4.93 (2H, s), 4.33-4.62 (6H, m), 4.26-4.32 (1H, m), 4.17 (1H, t, J = 5.1 Hz), 3.99 (1H, t, J = 4.4 Hz), 3.58 (1H, dd, J = 3.2, 10.6 Hz), 3.45 (1H, dd, J = 3.2, 10.6 Hz), 3.18 (3H, s), 1.65 (9H, s); HRMS (FAB) calcd for $C_{59}H_{55}N_3O_9$ 949.3938, found 949.3934

2,5-Dihydro-3-[6-benzyloxy-1-(2,3,5-tri-*O*-benzyl- β -D-ribofuranosyl)-1*H*-indol-3-yl]-4-(1*H*-indol-3-yl)-1-methyl-1*H*-pyrrolo-2,5-dione (25): From 24 (3.49 g, 3.68 mmol) and 40% methylamine in methanol (40 mL), compound 25 (2.90 g, 3.41 mmol) was obtained as an orange amorphous powder (93%) after chromatography (hexane-ethyl acetate (2:1)). $[\alpha]_{D}^{20}$ -34.4° (c 0.50, DMSO); IR (KBr) ν_{max} 3362, 1697, 1541, 1456, 1387, 1209, 1124 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 8.39-8.45 (1H, m), 7.83 (1H, s), 7.68 (1H, d, J = 2.8 Hz), 7.00-7.40 (24H, m), 6.91 (1H, d, J = 8.8 Hz), 6.74 (1H, t, J = 7.1 Hz), 6.54 (1H, dd, J = 2.2, 8.8 Hz), 6.10 (1H, d, J = 5.6 Hz), 4.95 (2H, s), 4.28-4.62 (7H, m), 4.21 (1H, t, J = 5.4 Hz), 3.99 (1H, dd, J = 4.1, 5.0 Hz), 3.60 (1H, dd, J = 3.3, 10.7 Hz), 3.48 (1H, dd, J = 3.3, 10.8 Hz), 3.17 (3H, s); HRMS (FAB) calcd for C₅₄H₄₇N₃O₇ 849.3414, found 849.3424.

2,5-Dihydro-3-[6-benzyloxy-1-(2-deoxy-3,4,6-tri-*O*-benzyl- β -D-glucopyranosyl)-1*H*-indol-**3-yl]-4-[6-benzyloxy1-1-(***tert*-butoxycarbonyl)-1*H*-indol-**3-yl]-1-methyl-1***H***-pyrrolo-2,5dione (28): From 26 (1.0 g, 1.53 mmol), PPh₃ (790 mg, 3.0 mmol), 2-deoxy-3,4,6-tri-***O***-benzyl-\alpha-Dglucopyranose 27 (1.3 g, 3.0 mmol) and DIAD (0.58 mL, 3.0 mmol), 28 (818 mg, 0.76 mmol) was obtained as a yellow amorphous powder (50%) after chromatography (toluene-ethyl acetate (30:1)). [\alpha]_{D}^{20} -22.2° (c 0.25, DMSO); IR (KBr) v_{max} 1733, 1701, 1540, 1456, 1369, 1218, 1153 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) \delta 7.95 (1H, s), 7.84 (1H, s), 7.71 (1H, s), 7.18-7.39 (25H, m), 6.98 (1H, d, J = 9.0 Hz), 6.93 (1H, d, J = 2.4 Hz), 6.75 (1H, d, J = 8.4 Hz), 6.57 (2H, dt, J = 2.4, 9.0 Hz), 5.40 (1H, dd, J = 1.5, 10.8 Hz), 5.00 (2H, s), 4.95 (2H, s), 4.69 (1H, d, J = 11.7 Hz), 4.62 (2H, d, J = 11.1 Hz), 4.59 (1H, d, J = 11.1 Hz), 4.52 (2H, d, J = 11.7 Hz), 3.77-3.88 (1H, m), 3.75 (2H, s), 3.65-3.69 (2H, m), 3.18 (3H, s), 2.44 (1H, dd, J = 1.5, 10.8 Hz), 2.17 (1H, q, J = 11.7 Hz), 1.65 (9H, s); HRMS (FAB) calcd for C₆₇H₆₃N₃O₁₀ 1069.4513, found 1069.4496.**

2,5-Dihydro-3-[6-benzyloxy-1-(2-deoxy-3,4,6-tri-*O***-benzyl**- β **-D-glucopyranosyl**)-1*H***-indol-3-yl**]-**4**-[**1**-(*tert*-**butoxycarbonyl**)-1*H***-indol-3-yl**]-**1**-methyl-1*H*-pyrrolo-2,5-dione (29): From 28 (818 mg, 0.77 mmol) and 40% methylamine in methanol (50 mL), compound 29 (705 mg, 0.73 mmol) was obtained as an orange amorphous powder (95%) after chromatography (hexane-ethyl acetate (2:1)). [α]²⁰_D -16.0° (c 0.12, DMSO); IR (KBr) v_{max} 3368, 1693, 1616, 1538, 1454, 1097 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 8.32 (1H, s), 7.69 (1H, s), 7.56 (1H, d, J = 1.5 Hz), 7.17-7.40 (25H, m), 6.96 (1H, d, J = 9.0 Hz), 6.87 (1H, d, J = 9.0 Hz), 6.85 (1H, d, J = 2.4 Hz), 6.58 (1H, dd, J = 2.4, 9.0 Hz), 6.53 (1H, dd, J = 2.1, 9.0 Hz), 5.41 (1H, d, J = 9.6 Hz), 4.99 (2H, s), 4.95 (2H, s), 4.95 (1H, d, J = 10.8 Hz), 4.70 (1H, d, J = 10.8 Hz), 4.62 (1H, d, J = 10.8 Hz), 4.60 (1H, 11.7 Hz), 4.52 (1H, d, J = 10.2 Hz), 4.48 (1H, d, J = 10.5 Hz), 3.75-3.90 (2H, m), 3.76 (1H, s), 3.67 (2H, d, J = 9.0 Hz), 3.17 (3H, s), 2.50 (1H, dd, J = 1.8, 9.6 Hz), 2.19 (1H, q, J = 10.8 Hz); HRMS (FAB) calcd for C₆₂H₅₅N₃O₈ 969.3989, found 969.3983.

12,13-Dihydro-2-benzyloxy-13-(2,3,4,6-tetra-O-benzyl- β -D-glucopyranosyl)-5*H*-indolo-[2,3-*a*]pyrrolo[3,4-*c*]carbazole-6-methyl-5,7(6*H*)-dione (32): Cupric chloride (1.4 g, 10 mmol) was added to a solution of 14 (1.0 g, 1.03 mmol) in the presence of molecular sieves 4A (1.0 g) in MEK (200mL), followed by stirring at 70 °C for 2 h. The precipitate was filtered off and the filtrate was concentrated. The residue was dissolved in ethyl acetate and the organic mixture was successively washed with 1M HCl, aqueous NaHCO₃, H₂O and saturated brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by chromatography on silica gel (hexane-acetone (3:1)) to yield **32** (800 mg, 0.83 mmol) as a yellow powder (80%). $[\alpha]^{2^0}_{D}$ +98.4° (c 1.00, CHCl₃); IR (KBr) ν_{max} 3334, 1749, 1697, 1618, 1577, 1496, 1454, 1375, 1153, 1079 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 10.68 (1H, s), 9.25 (2H, d, J = 8.4 Hz), 7.13-7.50 (27H, m), 7.00 (1H, t, J = 8.7 Hz), 6.87 (1H, t, J = 8.7 Hz), 6.20 (2H, d, J = 8.4 Hz), 5.87 (2H, d, J = 8.9 Hz), 5.20 (2H, d, J = 2.5 Hz), 5.00 (1H, d, J = 10.6 Hz), 4.88 (2H, s), 4.75 (2H, t, J = 12.3 Hz), 4.63 (1H, d, J = 10.6 Hz), 4.38 (1H, d, J = 9.8 Hz), 3.93-4.06 (4H, m), 3.86 (2H, d, J = 9.8 Hz), 3.33 (3H, s); HRMS (FAB) calcd for C₆₂H₅₃N₃O₈ 967.3833, found 967.3808.

12,13-Dihydro-10-benzyloxy-13-(2,3,4,6-tetra-O-benzyl-β-D-glucopyranosyl)-5H-indolo-

[2,3-*a*]pyrrolo[3,4-*c*]carbazole-6-methyl-5,7(6*H*)-dione (35): Palladium chloride (53 mg, 0.30 mmol) was added to a solution of 20 (50 mg, 0.052 mmol) in DMF (2.5 mL), followed by stirring at 100 °C for 16 h. After cooling, the reaction mixture was poured into aqueous NaHCO₃ and extracted with ethyl acetate. The organic layer was washed with saturated brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by chromatography on silica gel (hexane-ethyl acetate (4:1)) to yield **35** (39 mg, 0.040 mmol) as a yellow amorphous powder (78%). $[\alpha]_{D}^{20}$ +63.2° (c 1.00, CHCl₃); IR (KBr) v_{max} 3343, 1749, 1697, 1456, 1375, 1124, 1074 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 10.76 (1H, s), 9.38 (1H, d, J = 8.2 Hz), 9.15 (1H, d, J = 9.6 Hz), 7.10-7.67 (25H, m), 6.98 (1H, t, J = 7.7 Hz), 6.86 (2H, t, J = 7.6 Hz), 6.07 (2H, d, J = 6.9 Hz), 5.99 (1H, d, J = 8.9 Hz), 5.18 (1H, d, J = 11.7 Hz), 5.08 (1H, d, J = 11.7 Hz), 4.97 (1H, d, J = 10.7 Hz), 4.80-4.92 (2H, m), 4.75 (1H, d, J = 13.3 Hz), 4.67 (1H, d, J = 10.7 Hz), 4.57 (1H, d, J = 13.3 Hz), 4.32 (1H, t, J = 9.1 Hz), 3.89-4.09 (4H, m), 3.83 (1H, d, J = 9.6 Hz), 3.80 (1H, d, J = 9.1 Hz), 3.34 (3H, s), 2.90 (1H, d, J = 9.6 Hz); HRMS (FAB) calcd for C₆₂H₅₃N₃O₈ 967.3833, found 967.3831.

12,13-Dihydro-3,10-dibenzyloxy-13-(2,3,4,6-tetra-*O***-benzyl-***β***-D-glucopyranosyl)-5Hindolo[2,3-***a***]pyrrolo[3,4-***c***]carbazole-6-methyl-5,7(6H)-dione** (**36**): Palladium chloride (42.5 mg, 0.24 mmol) was added to a solution of **21** (52 mg, 0.048 mmol) in DMF (2.0 mL), followed by stirring at 110 °C for 1 h. After cooling, the precipitate was filtered off and the filtrate was poured into 2 M HCl and extracted with ethyl acetate. The organic layer was washed with saturated brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by chromatography on silica gel (hexane-ethyl acetate (3:1)) to yield **36** (40 mg, 0.037 mmol) as a yellow powder (77%). $[\alpha]_{D}^{20}$ +65.2° (c 1.00, CHCl₃); IR (KBr) v_{max} 3334, 1749, 1697, 1616, 1456, 1375, 1252, 1080 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 10.71 (1H, s), 9.14 (1H, d, J = 9.5 Hz), 9.08 (1H, d, J = 2.6 Hz), 6.80-7.65 (34H, m), 6.14 (2H, dd, J = 1.4, 6.9 Hz), 5.81 (1H, d, J = 8.8 Hz), 5.35 (2H, s), 5.17 (1H, d, J = 11.5 Hz), 5.07 (1H, d, J = 11.8 Hz), 4.96 (1H, d, J = 10.7 Hz), 4.85 (2H, d, J = 2.3 Hz), 4.74 (1H, d, J = 13.0 Hz), 4.66 (1H, d, J = 10.6 Hz), 4.56 (1H, d, J = 13.0 Hz), 4.31 (1H, t, J = 9.5 Hz), 3.75-4.05 (5H, m), 3.34 (3H, s); HRMS (FAB) calcd for C₆₉H₅₉N₃O₉ 1073.4251, found 1073.4279.

12,13-Dihydro-2,9-dibenzyloxy-13-(2,3,4,6-tetra-O-benzyl- β -D-glucopyranosyl)-5H-

indolo[2,3-a]pyrrolo[3,4-c]carbazole-6-methyl-5,7(6H)-dione (33): Cupric chloride (29 mg, 0.14 mmol) was added to a solution of 15 (100 mg, 0.093 mmol) in MEK (2 mL), followed by stirring at 60 °C for 1 h. The precipitate was filtered off and the filtrate was concentrated in vacuo. The residue was purified by

chromatography on silica gel (toluene:ethyl acetate (50:1)) to yield **33** (79 mg, 0.074 mmol) as a yellow powder (80%). $[\alpha]_{D}^{20}$ +141.6° (c 1.00, CHCl₃); IR (KBr) ν_{max} 3336, 1749, 1697, 1456, 1203, 1079 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 10.50 (1H, s), 9.23 (1H, d, J = 8.7 Hz), 8.94 (1H, d, J = 1.8 Hz), 7.60 (2H, d, J = 7.8 Hz), 7.00-7.45 (28H, m), 6.87 (2H, t, J = 8.4 Hz), 6.19 (2H, d, J = 8.7 Hz), 5.84 (1H, d, J = 9.3 Hz), 5.31(2H, s), 5.18 (2H, d, J = 2.4 Hz), 5.00 (1H, d, J = 10.2 Hz), 4.86 (2H, s), 4.74 (1H, d, J = 10.2 Hz), 4.71 (1H, d, J = 12.0 Hz), 4.60 (1H, d, J = 12.0 Hz), 4.35 (1H, t, J = 9.6 Hz), 4.03 (2H, t, J = 9.0 Hz), 3.95 (1H, d, J = 9.6 Hz), 3.93 (1H, d, J = 9.3 Hz), 3.86 (2H, m), 3.34 (3H, s), 2.99 (1H, d, J = 9.3 Hz); HRMS (FAB) calcd for C₆₉H₅₉N₃O₉ 1073.4251, found 1073.4277.

12,13-Dihydro-2,10-dibenzyloxy-13-(2,3,4,6-tetra-O-benzyl-β-D-glucopyranosyl)-5H-

indolo[2,3-*a*]pyrrolo[3,4-*c*]carbazole-6-methyl-5,7(6*H*)-dione (34): Cupric chloride (29 mg, 0.14 mmol) was added to a solution of 16 (52 mg, 0.048 mmol) in the presence of molecular sieves 4A (50 mg) in MEK (1 mL), followed by stirring at 25 °C for 2 h. The precipitate was filtered off and the filtrate was concentrated in vacuo. The residue was purified by chromatography on silica gel (dichloromethane) to yield 34 (42 mg, 0.024 mmol) as a yellow powder (84%). $[\alpha]_{D}^{20}$ +109.6° (c 1.00, CHCl₃); IR (KBr) v_{max} 3332, 1749, 1621, 1496, 1284 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 10.06 (1H, s), 9.24 (1H, d, J = 9.5 Hz), 9.13 (1H, d, J = 9.5 Hz), 7.07-7.50 (29H, m), 6.98-7.03 (1H, m), 6.83-6.91 (2H, m), 6.18 (2H, m), 5.84 (1H, d, J = 8.9 Hz), 5.12-5.22 (2H, m), 5.18 (1H, d, J = 11.5 Hz), 5.08 (1H, d, J = 11.5 Hz), 4.97 (1H, d, J = 10.7 Hz), 4.89 (1H, d, J = 10.7 Hz), 4.84 (1H, d, J = 10.7 Hz), 4.74 (1H, d, J = 13.0 Hz), 4.67 (1H, d, J = 10.7 Hz), 4.56 (1H, d, J = 13.0 Hz), 4.32 (1H, t, J = 9.6 Hz), 3.98-4.07 (2H, m), 3.82-3.97 (3H, m), 3.79 (1H, dd, J = 2.7, 10.2 Hz), 3.33 (3H, s), 3.00 (1H, d, J = 9.7 Hz); HRMS (FAB) calcd for C₆₉H₅₉N₃O₉ 1073.4251, found 1073.4237.

12,13-Dihydro-10-benzyloxy-13-(2,3,5-tri-O-benzyl- β -D-ribofuranosyl)-5*H*-indolo[2,3-*a*]pyrrolo[3,4-*c*]carbazole-6-methyl-5,7(6*H*)-dione (37): Cupric chloride (47.4 mg, 0.35 mmol) was added to a solution of 25 (100 mg, 0.12 mmol) in the presence of calcium carbonate (400 mg) in MEK (20 mL), followed by stirring at 80 °C for 1 h. The reaction mixture was poured into 0.2 M HCl and extracted with ethyl acetate. The organic phase was successively washed with aqueous NaHCO₃, H₂O and saturated brine, dried over Na₂SO₄ and concentrated in vacuo to yield 37 (98 mg, 0.12 mmol) as a yellow amorphous powder (98%).

 $[\alpha]_{D}^{20}$ +116.8° (c 1.00, CHCl₃); IR (KBr) ν_{max} 3358, 1749, 1697, 1578, 1456, 1377, 1329, 1190, 1120, 1047 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 10.06 (1H, s), 9.23 (1H, d, J = 7.8 Hz), 9.21 (1H, d, J = 9.0 Hz), 7.50 (1H, d, J = 6.9 Hz), 7.26-7.45 (20H, m), 7.14 (1H, dd, J = 1.8, 9.0 Hz), 6.96 (1H, t, J = 7.3 Hz), 6.85 (2H, t, J = 7.3 Hz), 6.57 (2H, d, J = 7.3 Hz), 6.30 (1H, d, J = 7.3 Hz), 5.19 (2H, s), 4.70-4.90 (3H, m), 4.56 (1H, t, J = 7.1 Hz), 4.46 (1H, d, J = 11.5 Hz), 4.22-4.30 (2H, m), 3.98-4.09 (3H, m), 3.80 (1H, dd, J = 2.0, 11.6 Hz), 3.35 (3H, s); HRMS (FAB) calcd for C₅₄H₄₅N₃O₇ 847.3258, found 847.3240.

12,13-Dihydro-2,10-dibenzyloxy-13-(2-deoxy-3,4,6-tri-O-benzyl- β -D-glucopyranosyl)-5Hindolo[2,3-a]pyrrolo[3,4-c]carbazole-6-methyl-5,7(6H)-dione (37): Cupric chloride (82 mg, 0.30 mmol) was added to a solution of 29 (150 mg, 0.15 mmol) in the presence of molecular serves 4A (200 mg) in MEK (10 mL), followed by stirring at 60 °C for 0.5 h. The precipitate was filtered off and the filtrate was concentrated. The residue was dissolved in ethyl acetate and the organic mixture was successively washed with 1 M HCl, aqueous NaHCO₃, H₂O and saturated brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by chromatography on silica gel (toluene-acetone (50:1)) to yield **37** (107 mg, 0.11 mmol) as a yellow amorphous powder (71%). $[\alpha]_{D}^{20}$ +97.4° (c 1.00, CHCl₃); IR (KBr) v_{max} 3334, 1749, 1697, 1454, 1375, 1026 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 10.76 (1H, s), 9.22 (1H, d, J = 8.7 Hz), 9.10 (1H, d, J = 8.7 Hz), 7.02-7.53 (30H, m), 5.94 (1H, dd, J = 4.0, 9.8 Hz), 5.23 (2H, s), 5.13 (1H, d, J = 11.4 Hz), 5.03 (1H, d, J = 10.8 Hz), 5.02 (1H, d, J = 11.7 Hz), 4.78 (1H, d, J = 13.2 Hz), 4.67 (1H, d, J = 10.5 Hz), 4.60 (1H, d, J = 13.2 Hz), 4.53 (2H, q, J = 11.7 Hz), 4.23 (1H, t, J = 9.2 Hz), 4.03 (1H, d, J = 10.0 Hz), 3.91 (1H, dd, J = 5.8, 9.2, 9.8 Hz), 3.85 (1H, d, J = 9.2 Hz), 3.78 (1H, dd, J = 2.5, 10.0 Hz), 3.31 (3H, s), 2.21 (1H, dd, J = 4.0, 9.8 Hz); HRMS (FAB) calcd for C₆₇H₃₃N₃O₁₀ 967.3833, found 967.3803.

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