Accepted Manuscript

Design, Synthesis and Biological Evaluation of (E)-3,4-Dihydroxystyryl 4-Acylaminophenethyl Sulfone, Sulfoxide Derivatives as Dual Inhibitors of HIV-1 CCR5 and Integrase

Yixing Sun, Weisi Xu, Ningning Fan, Xuefeng Sun, Xianling Ning, Liying Ma, Junyi Liu, Xiaowei Wang

PII:	S0968-0896(16)31442-0
DOI:	http://dx.doi.org/10.1016/j.bmc.2016.12.035
Reference:	BMC 13462
To appear in:	Bioorganic & Medicinal Chemistry
Received Date:	13 August 2016
Revised Date:	26 November 2016
Accepted Date:	16 December 2016

調響	ISSN ONA-DIDG						
ELSEVIER	Bioorganic & Medicinal Chemistry						
	The Tetrahedron Journal for Research at the Interface of Chemistry and Biology						
	IN THIS ISSUE:						
	The generality of kinase-catalyzed biotinylation						
	OH Sectors and ATP-bets						
	Available online at www.sciencedmict.com						
	ScienceDirect						

Please cite this article as: Sun, Y., Xu, W., Fan, N., Sun, X., Ning, X., Ma, L., Liu, J., Wang, X., Design, Synthesis and Biological Evaluation of *(E)*-3,4-Dihydroxystyryl 4-Acylaminophenethyl Sulfone, Sulfoxide Derivatives as Dual Inhibitors of HIV-1 CCR5 and Integrase, *Bioorganic & Medicinal Chemistry* (2016), doi: http://dx.doi.org/10.1016/j.bmc.2016.12.035

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Design, Synthesis and Biological Evaluation of (*E*)-3,4-Dihydroxystyryl 4-Acylaminophenethyl Sulfone, Sulfoxide Derivatives as Dual Inhibitors of HIV-1 CCR5 and Integrase

Yixing Sun^{a,}[‡], Weisi Xu^{b,}[‡], Ningning Fan^a, Xuefeng Sun^a, Xianling Ning^a,

Liying Ma^{b,*}, Junyi Liu^{a,c,*} and Xiaowei Wang^{a,*}

^a Department of Chemical Biology, School of Pharmaceutical Sciences, Peking University, Beijing 100191, China.
 ^b National Center for AIDS/STD Control and Prevention (NCAIDS), Chinese Center for Disease Control and Prevention, Beijing 102206, China

^c State Key Laboratory of Natural and Biomimetic Drugs, Peking University, Beijing 100191, China

* Corresponding authors. Tel.: +86 10 82801706 (J.L.); tel.: +86 10 82805203 (X.W.). E-mail addresses: jyliu@bjmu.edu.cn (J. Liu), xiaoweiwang@bjmu.edu.cn (X. Wang), mal@chinaaids.cn (L. Ma).

[‡] These Authors contribute equally.

ABSTRACT: Aiming at the limited effectiveness of current clinical therapeutic effect of AIDS, novel series of compounds bearing (*E*)-3,4-dihydroxystyryl sulfone (or sulfoxide) and anilide fragments were designed and synthesized as dual inhibitors of HIV-1 CCR5/IN. The biological results indicated that several target compounds showed inhibitory activity against HIV-1 Bal (R5) infection in TZM-bl cells. Besides targeting the chemokine receptor on the host cell surface, they also displayed binding affinities with HIV-1 integrase using the surface plasmon resonance (SPR) binding assays. Molecular docking studies have inferred the possible binding mode of target compounds against integrase. These data demonstrate that the structure of (*E*)-3,4-dihydroxystyryl sulfone and sulfoxide derivatives have the potential to derive potent dual inhibitors of HIV-1 Integrase and CCR5.

KEYWORDS: HIV dual inhibitors; CCR5 receptor; HIV integrase; Sulfones; Sulfoxides.

1. Introduction

In early stages of the development of highly active anti-retroviral therapy (HAART, a.k.a. "drug cocktails"), the treatments of HIV were limited to the combination therapies of reverse transcriptase (RT) inhibitors and protease inhibitors. With the application of integrase (IN) inhibitors, fusion inhibitors and co-receptor inhibitors, not only the selection scope of HAART has been widened, but the treatment outcome has also been improved.¹ Nevertheless, the poor patient compliance, the risk of drug interactions, and the emergence of resistant HIV strains brought by HAART have promoted to discovering new strategy against HIV, which include designing multi-targeted inhibitors.^{2,3}

In order to block the HIV replication cycle at different stages, we planned to develop dual inhibitors of HIV-1 CCR5 and integrase. During the integration process in HIV life cycle, chemokine receptors CCR5, along with CXCR4, are required for virus' membrane fusion with host cell.⁴ In particular, the CCR5 has a main mediating role in the initial infection of HIV.⁵ Also, with the action of integrase, cDNA strands assemble the host genomic DNA into a double-stranded DNA

to complete replication.⁶ Additionally, the integrase serves as a feasible intracellular target since it hasn't been found in mammalian cells.⁷ Accordingly, targeting both the integrase and CCR5 may provide low toxicity as well as great efficiency and timeliness.

Recent studies in the crystal structure of the CCR5-maraviroc complex have demonstrated the multiple forces between the receptor and the ligand.⁸ These include a salt bridge interaction between the protonated tropane nitrogen and E283, several hydrogen bond forces including amide nitrogen to Y251, as well as the hydrophobic contacts between the phenyl group with the aromatic amino acid residues. In the meantime, previous crystallography studies have demonstrated the binding mode of raltegravir (RAL) with the Prototype Foamy Virus IN. The di-carbonyl moiety of diketo acids (DKAs) changed the conformation of IN by coordinating to the divalent ions in the active domain of the protein.^{9,10}

Caffeic acid phenethyl ester (CAPE) has also been reported to inhibit the process of integration.^{11,12} As shown in SAR studies, diphenol and carbonyl units are necessary to maintain the IN inhibitory activity.^{13,14} Due to the susceptibility of carboxylic ester to hydrolase, bioisosteric replacement took place and sulfone (or sulfoxide) group was brought in to form the structure. Sulfone and sulfoxide groups are not only more stable than ester groups, but also more soluble in water. Moreover, Smithkline Beecham Corp. disclosed various benzanilides to acquire CCR5 inhibitory properties through generating corresponding hydrogen bonds and hydrophobic interactions.^{15,16} With these evidences in mind, we highly integrated the structures of sulfone and sulfoxide derivatives of CAPE and aroyl aniline moiety which could acquire CCR5 inhibitory properties through generating corresponding hydrogen bonds and hydrophobic interactions, and formed new series of target compounds (Fig. 1).



Figure 1. Design of dual inhibitors based on derivatives of CAPE and potent CCR5 antagonists.

Herein, we proposed to synthesize the target compounds to probe their antivirus activity against R5 HIV-1 strains and IN binding affinity. These results might provide useful information for developing novel dual inhibitors of HIV-1 IN and CCR5.

2. Results and discussion

2.1. Chemistry

For the preparation of the sulfone and sulfoxide target compounds, the synthetic routes (shown in Scheme 1) were explored respectively. The synthesis of **4**, **5**, and **6** were based on our previous report.¹⁷ Commercially available 4-nitrophenethyl bromide was reacted with mercaptoacetic acid to obtain compound **4**, which was then oxidized with H_2O_2 in different solvents to afford **5** and **6** respectively.

In the presence of β -aminopropionic acid as catalyst, the sulfonyl intermediate **7** was obtained with high yield through Knoevenagel condensation of compound **5** with acetyl protected 3,4-dihydroxybenzaldehyde. However, pyrrolidine and HOAc was used as catalysts to give the sulfinyl intermediate **8**. The change of the Knoevenagel reaction condition is due to the decreased reactivity of the methylene structure.

Following the Knoevenagel condensation reaction, the nitro group of **7** was selectively reduced with palladium-catalyzed hydrogenation to afford the aromatic amine **9**. Due to the decrease of Pd/C catalytic activity that caused by the existence of the sulfoxide group, sodium hydrosulfite was used as reductant to obtain the intermediate **10**. Reaction of **9** and **10** with various acid chlorides gave the corresponding compounds **11a-i** and **12a-f**. Final deprotections of the acetyl and MOM groups gave the target compounds **13a-i** and **14a-f**, respectively.



Scheme 1. Synthetic routes of target compounds.

Reagents & conditions: (I) NaOH, MeOH, r.t., 2h, yields 80-93%; (II) $30\%H_2O_2$, HOAc, r.t., yields 74-97%; (III) $30\%H_2O_2$, MeOH, r.t., yields 83-95%; (IV) β -aminopropionic acid, THF, 4Å MS, reflux, 12h, yield 67-80%; (V) pyrrolidine, HOAc, THF, 4Å MS, reflux, 12h, yield 51-71%; (VI) H₂, Pd/C, CH₂Cl₂, r.t., 3h, yield 64-82%; (VII) Na₂S₂O₄, EtOH, H₂O, reflux, 12h, yield 25-45%; (VIII) RCOCl, Et₃N, CH₂Cl₂, ice bath, 2h, 16-75%; (IX) K₂CO₃, MeOH, r.t., 3h, yield 56-65%; (X₁,X_{II}) RCOCl, Et₃N, CH₂Cl₂, ice bath, 2h; HCl, MeOH, reflux, 3h, total yield 10-23%.

2.2. Biological results

2.2.1. Determination of the cytotoxicity and antivirus activity against R5 HIV-1 strains

All the target compounds were sequentially tested for their cytotoxicity and their antivirus activity against R5 HIV-1 strains in TZM-bl cell line. For the *in vitro* entry inhibition assays, HIV-1 Bal (R5) virus strain was used to determine the type of chemokine receptors that mediate virus' entry.

The TZM-bl cell line was selected to complete the cell based assays. Reporter genes such as firefly luciferase gene, which is promoted by HIV long terminal repeat sequence (LTR), and *E. coli*

 β -galactosidase gene are carried and receptors including CCR5 were expressed simultaneously by this cell line. The intracellular viral load can therefore be tested by a fluorescence detector after the virus' entrance to the cell. Due to the specific mediation of CCR5 during the virus' entrance, HIV-1 Bal (R5) virus strain was used for measuring the CCR5 inhibitory activity of the target compounds.

All target compounds were tested for their cytotoxicity before testing for cell based anti-HIV activity. As shown in Table 1, the CC_{50} values of most active compounds are above 100 μ M. Meanwhile, the sulfone and sulfoxide derivatives at 50 μ M concentration displayed antiviral activity against R5 HIV-1 strains at different levels. Five compounds, including four sulfoxide derivative, showed at least 50% inhibition of HIV-1 replication in TZM-bl cells. The preliminary biological results suggested that sulfoxides showed better activity than sulfones. Besides, the difference of R substitution may affect the CCR5 inhibitory activity. Herein, 3,5-difluorophenyl group, cyclohexyl group, 4-nitrophenyl group, 1-adamantaneformyl group, and 4-fluorophenyl group are the more favored substituent groups.

 Table 1. Inhibition of HIV-1 entry through CCR5 co-receptor as analyzed in TZM-bl cells using HIV-1 Bal (R5) virus

Ö

0,0

	R R		~s	OR'	RN		ŝ/		DR' DR'	
		Target Com	pounds Sulf	ones	Targe	et Compound	ds	Sulfoxides		
Compound R	R'	Series	% Inhibition ^a	$CC_{50}^{\ b}(\mu M)$	Compoun	d R	R'	Series	% Inhibition ^a	$CC_{50}^{\ b}(\mu M)$
11a	Ac	Sulfone	<20	108	13a		Н	Sulfone	<20	238
11b	Ac	Sulfone	<20	858	13b		Н	Sulfone	<20	>1000
11c	Ac	Sulfone	26.2	162	13c	F C C	Н	Sulfone	<20	>1000
11d F	Ac	Sulfone	<20	225	13d		Н	Sulfone	33.0	414
11e	Ac	Sulfone	<20	169	13e	► A	Н	Sulfone	45.9	638
11f	Ac	Sulfone	23.7	182	13f	E,	Н	Sulfone	21.6	786
11g	Ac	Sulfone	<20	177	13g	H	Н	Sulfone	<20	168
11h 0-N	Ac	Sulfone	52.2	160	13h	O-N	Н	Sulfone	23.4	483
11i	→ Ac	Sulfone	23.0	161	13i		Н	Sulfone	<20	569
14a	Н	Sulfoxide	68.4	157	14d	Ŷ	Н	Sulfoxide	41.8	ND
14b	H	Sulfoxide	34.1	ND	14e	$\overset{r}{\bigcirc}$	Н	Sulfoxide	52.7	ND
14c F	Н	Sulfoxide	64.6	127	14f	D'	Н	Sulfoxide	64.7	ND
Maraviroc			IC50=0.2nM	>1000						

ND = Not determined.

^a % Decrease in fluorescent intensity in TZM-bl cells; data represent mean values for two wells.

 b CC₅₀ = concentration of compound required for 50% cytotoxicity against uninfected cells.

2.2.2. Assays for measuring the in vitro binding affinity of target compounds against HIV-1 IN

Having determined the antivirus activity against R5 HIV-1 strains, we carried out assays for the second target. In order to investigate the binding affinity of target compounds against HIV-1 IN, we used SPR based Biacore T200 biosensor for evaluation. The commercially available recombinant IN protein was immobilized on a CM5 chip and the binding response was recorded continuously in the response unit (RU) and graphically presented as a function of time. Preliminary screening of the 24 compounds showed that compound **11a**, **13a**, **13e**, and **14a** have better binding results (shown in Fig. 2). Then the dose-dependent relations was tested with various concentrations for the four compounds (200μ M, 100μ M, 50μ M, 25μ M, 12.5μ M, etc.). To determine the equilibrium dissociation constant (K_D), the 1:1 binding fit model (kinetics) was used. According to the affinity estimates, the K_D value of compound **14a** was 141.8 μ M. As shown in Table 2, these SPR results determined that the (*E*)-3,4-dihydroxystyryl sulfone and sulfoxide derivatives have binding affinity against HIV-1 integrase.



Table 2. K_D values of the 4 representative compounds

Figure 2. HIV-1 IN binding affinity using SPR biosensor technology. Representative sensorgram of 11a(A), 13a(B), 13e(C), 14a(D) were obtained.

It is noteworthy that compound **14a** demonstrate potency activity against R5 HIV-1 strains with relatively low toxicity to TZM-bl cells and moderate binding affinity with IN ($K_D = 141.8\mu$ M). Therefore, the preliminary results suggested that target compound **14a** could be potent dual inhibitor of HIV-1 CCR5/IN.

2.3. Molecular docking

To assess the dual inhibitory properties of the target compounds, computational docking studies were carried out to explore the binding mode of the target compounds with active sites of CCR5 and prototype foamy virus (PFV) IN respectively.

Firstly, in docking studies with CCR5 (Fig. 3) (PBD: 4MBS), the results revealed that H bonds were observed between the phenolic oxygen and Tyr37, as well as amide nitrogen and Glu283. The bonding interactions have significant similarities with results from CCR5-maraviroc complex studies which are supposed to assist the molecule's positioning in the binding pocket,⁷ and are recognized as the key points of exhibiting CCR5 inhibitory activity. The phenyl rings of inhibitors imparted π - π stacking with the CCR5 active site residues (Trp86, Tyr108, Phe109, Phe112, etc.). Additionally, hydrophobic interactions were developed by the positioning of aromatic rings into the hydrophobic pockets comprised by aromatic residues Tyr108, Phe109, Phe112, Trp248, and Tyr251.



Figure 3. Molecular docking result of compound 14a into HIV CCR5 (PBD: 4MBS). Hydrogen bonds were signed in yellow dotted line.

To further explore the potential of IN inhibitory activity of these compounds, docking works were performed using the PFV IN core domain (Fig. 4). Compound **13e** exhibited H bonding between one of the phenolic oxygen and the side chain of Glu221 residue, as well as coordination between the catechol oxygens and metal ion Mg^{2+} that results in the chelation. Meanwhile, in docking results of compound **14a**, Mg^{2+} metal ions coordinate with sulfinyl oxygens while H bonds are generated between acyl oxygen and Tyr212.



Figure 4. Molecular docking of compound **13e** (left), and **14a** (right) into PFV IN (PBD: 3OYA). The balls colored in green represents Mg²⁺ ions.

Therefore, the binding modes of the target compounds against HIV CCR5 and PFV IN were preliminarily revealed, laying a foundation for further development of these structures.

3. Conclusion

To sum up, series of (E)-3,4-dihydroxystyryl 4-acylaminophenethyl sulfones and sulfoxides were synthesized and tested for their *in vitro* HIV-1 cellular CCR5 inhibitory activity (close to 68.37% at 50µM) and binding affinity against HIV-1 integrase (K_D at around 10⁻⁴ mole). The two independent mechanism enable the target compounds to block the entry and integration stages respectively, and it would possibly reduce the generation of drug resistant viruses. Taken the molecular docking results together, our study identified that the (E)-3,4-dihydroxystyryl sulfone and sulfoxide derivatives could derive novel compounds that display dual inhibition activities for HIV-1 virus.

4. Experimental procedures

4.1. Chemistry

4.1.1. General chemistry methods

The structural characterizations of compounds were performed by NMR, which were recorded on Bruker AVANCE III-400 spectrometer (400MHz) with TMS (tetramethylsilane, Me₄Si) as an internal standard. Chemical shifts were reported in δ (ppm) while coupling constants were measured in Hz. CDCl₃ and DMSO-*d*₆ were used as the solvents for NMR experiments. The purity of target compounds was determined by HPLC (Agilent 1260 infinity HPLC system). All the assayed compounds showed >95% purity by HPLC. All reactions were monitored by TLC, which was performed on precoated silica gel GF254 plates. The detection of TLC was done by UV light irradiation (UV lamp, model UV-IIB). Column chromatography was performed by silica gel H (200-300 or 500 mesh). Melting points were measured on an X-4 apparatus without correction. THF were purified and dried by standard methods. All other organic solutions were dried over anhydrous sodium sulfate. All other reagents were purchased as reagent grade from commercial sources and without further purification unless otherwise stated.

4.1.2. Synthesis of (E)-3,4-diacetoxystyryl 4-nitrophenethyl sulfone (7)

To a solution of 2-(4-nitrophenethylsulfonyl) acetic acid (5, 3mmol) in THF (15mL), 3,4-diacetoxybenzaldehyde (2mmol) and β -aminopropionic acid (1mmol) were added and the resulting solution was heated to reflux until absence of 3,4-diacetoxybenzaldehyde (checked by TLC). The solvent was evaporated under vacuum and the mixture was diluted with H₂O and extracted with ethyl acetate. The combined organic fractions were washed with brine, dried with sodium sulfate, and concentrated in vacuo. Purification of the crude product by column chromatography (PE/EA) afforded the title compound **7**. Yield: 80%; Yellow solid; mp 86–87 °C; ¹H

NMR (400MHz, CDCl₃) δ 8.17 (d, 2H, J = 8.5 Hz), 7.55 (d, 1H, J = 15.4 Hz), 7.27-7.41 (m, 5H), 6.72 (d, 1H, J = 15.4 Hz), 3.38 (m, 2H), 3.25 (m, 2H), 2.32 (s, 6H); ¹³C NMR (100MHz, DMSO- d_6) δ 168.9, 168.7, 145.4, 145.1, 143.9, 143.2, 133.7, 132.5, 130.1, 130.1, 127.2, 124.3, 124.1, 123.9, 123.9, 122.8, 56.0, 28.3, 20.5, 20.5; MS (ESI) m/z 456.05 [M+Na]⁺.

4.1.3. Synthesis of (*E*)-1,2-bis(methoxymethoxy)-4-(2-((4-nitrophenethyl)sulfinyl)vinyl) benzene (8)

To a solution of 2-(4-nitrophenethylsulfinyl) acetic acid (**6**, 3mmol) in THF (15mL), 3,4-methoxymethoxybenzaldehyde (2mmol), pyrrolidine (catalytic amount), and acetic acid (catalytic amount) were added and the resulting solution was heated to reflux until absence of 3,4-methoxymethoxybenzaldehyde (checked by TLC). The solvent was evaporated under vacuum and the mixture was diluted with H₂O and extracted with ethyl acetate. The combined organic fractions were washed with brine, dried with sodium sulfate, and concentrated in vacuo. Purification of the crude product by column chromatography (PE/EA) afforded the title compound **8**. Yield: 71%; Yellow solid; mp 155–156 °C; ¹H NMR (400MHz, CDCl₃) δ 8.16 (d, 2H, *J* = 8.7 Hz), 7.51 (d, 1H, *J* = 15.4 Hz), 7.19-7.40 (m, 5H), 6.62 (d, 1H, *J* = 15.4 Hz), 5.27 (d, 4H, *J* = 13.2 Hz), 3.53 (d, 6H, *J* = 6.6 Hz), 3.23-3.39 (m, 4H); ¹³C NMR (100MHz, DMSO-*d*₆) δ 149.0, 146.8, 145.3, 145.0, 133.6, 132.4, 130.1, 130.1, 124.2, 123.9, 123.9, 123.3, 111.3, 108.1, 96.1, 96.1, 54.1, 53.9, 51.0, 34.3; MS (ESI) *m/z* 422.11 [M+H]⁺ 444.11 [M+Na]⁺.

4.1.4. Synthesis of (E)-3,4-diacetoxystyryl 4-aminophenethyl sulfone (9)

To a solution of *(E)*-3,4-diacetoxystyryl 4-nitrophenethyl sulfone (7, 0.5mmol) in dichloromethane (20mL), Palladium/C (20mg) were added and the resulting solution was allowed to stir under hydrogen gas for 3h at room temperature. The solvent was evaporated under vacuum and the mixture was diluted with H₂O and extracted with ethyl acetate. The combined organic fractions were washed with brine, dried with sodium sulfate, and concentrated in vacuo. The crude product of **9** was washed with petroleum ether. Yield: 82%; Yellow oil; ¹H NMR (400MHz, CDCl₃) δ 7.46 (d, 1H, *J* = 15.4 Hz), 7.35 (d, 1H, *J* = 1.6 Hz), 7.22 (d, 1H, *J* = 15.4 Hz), 6.48-7.19 (m, 6H), 4.93 (s, 2H), 3.29 (m, 2H), 2.80 (m, 2H), 2.32 (s, 6H); ¹³C NMR (100MHz, DMSO-*d*₆) δ 168.7, 168.7, 145.6, 143.7, 143.2, 133.7, 132.4, 129.3, 129.0, 129.0, 124.3, 124.1, 122.8, 115.3, 115.3, 55.7, 28.3, 20.6, 20.5; HRMS (ESI) *m/z* calcd for C₂₀H₂₁NNaO₆S [M+Na]⁺: 426.0987, found: 426.0988.

4.1.5. Synthesis of (E)-4-(2-((3,4-bis(methoxymethoxy)styryl)sulfinyl)ethyl)aniline (10)

To a solution of (E)-1,2-bis(methoxymethoxy)-4-(2-((4-nitrophenethyl)sulfinyl)vinyl)benzene (**8**, 0.5mmol) in ethanol (with 16.7% water, 20mL), sodium hydrosulfite (8mmol) were added several times and the resulting solution was heated to reflux until absence of compound **8** (checked by TLC). The solvent was evaporated under vacuum and the mixture was diluted with H₂O and extracted with ethyl acetate. The combined organic fractions were washed with brine, dried with sodium sulfate, and concentrated in vacuo. The crude product of **10** was washed with petroleum ether. Yield: 45%;

Yellow oil; ¹H NMR (400MHz, CDCl₃) δ 7.48 (d, 1H, *J* = 15.4 Hz), 6.63-7.29 (m, 7H), 6.57 (d, 1H, *J* = 15.4 Hz), 5.27 (d, 4H, *J* = 4.4 Hz), 4.84 (s, 2H), 3.53 (s, 6H), 3.01-3.31 (m, 4H); ¹³C NMR (100MHz, DMSO-*d*₆) δ 149.1, 146.9, 145.3, 134.0, 129.8, 128.9, 128.9, 128.8, 122.2, 121.3, 115.4, 115.4, 111.5, 108.2, 96.5, 96.3, 54.1, 54.0, 51.2, 34.1; MS (ESI) *m/z* 392.16 [M+H]⁺.

4.1.6. General synthesis of (*E*)-3,4-diacetoxystyryl 4-acylaminophenethyl sulfones (11a – 11i)

To a solution of (*E*)-3,4-diacetoxystyryl 4-aminophenethyl sulfone (9, 0.1mmol) in dichloromethane (10mL), trimethylamine (0.2mmol), and various acid chlorides (0.22mmol) were added under the ice bath. The resulting solution was allowed to stir for 2h. The solvent was evaporated under vacuum and the mixture was diluted with H_2O and extracted with ethyl acetate. The combined organic fractions were washed with brine, dried with sodium sulfate, and concentrated in vacuo. Purification of the crude product by column chromatography (PE/EA) afforded the target compounds **11a** to **11i**.

4.1.6.1. (*E*)-**3**,4-diacetoxystyryl 4-benzamidophenethyl sulfone (11a) 75%; Yellow solid; mp 92–93 °C; ¹H NMR (400MHz, DMSO- d_6) δ 10.21 (s, 1H), 6.79-7.95 (m, 14H), 3.42-3.46 (m, 2H), 2.94-2.98 (m, 2H), 2.33 (s, 6H); ¹³C NMR (100MHz, DMSO- d_6) δ 168.6, 168.4, 165.2, 144.3, 142.7, 141.6, 138.2, 133.7, 133.3, 131.9, 131.7, 129.9, 129.9, 129.2, 129.2, 129.0, 129.0, 127.3, 124.7, 124.0, 120.8, 120.8, 114.9, 55.8, 28.1, 20.7, 20.6; MS (ESI) *m/z* 530.31 [M+Na]⁺; HRMS (ESI) *m/z* calcd for C₂₇H₂₅NNaO₇S [M+Na]⁺: 530.1249, found: 530.1251.

4.1.6.2. *(E)*-3,4-diacetoxystyryl 4-4-methoxylbenzamidophenethyl sulfone (11b) 49%; Buff white solid; mp 106–107 °C; ¹H NMR (400MHz, DMSO- d_6) δ 10.04 (s, 1H), 7.06-7.95 (m, 13H), 3.84 (s, 3H), 3.49 (m, 2H), 2.98 (m, 2H), 2.30 (s, 6H); ¹³C NMR (100MHz, DMSO- d_6) δ 168.6, 168.5, 165.3, 162.3, 144.4, 142.8, 141.7, 138.3, 133.4, 131.8, 130.0, 130.0, 129.1, 129.1, 128.0, 127.4, 124.8, 124.1, 120.9, 120.9, 115.0, 114.0, 114.0, 55.9, 55.3, 28.1, 20.8, 20.7; MS (ESI) *m/z* 560.21 [M+Na]⁺; HRMS (ESI) *m/z* calcd for C₂₈H₂₇NNaO₈S [M+Na]⁺: 560.1355, found: 560.1356.

4.1.6.3. *(E)***-3,4-diacetoxystyryl 4-4-fluorobenzamidophenethyl sulfone (11c)** 39%; Buff white solid; mp 105–106 °C; ¹H NMR (400MHz, DMSO-*d₆*) δ 10.10 (s, 1H), 6.80-8.12 (m, 13H), 3.42 (m, 2H), 2.94 (m, 2H), 2.33 (s, 6H); ¹³C NMR (100MHz, DMSO-*d₆*) δ 168.8, 168.6, 165.4, 144.7, 142.9, 141.8, 141.2, 138.2, 133.5, 131.9, 131.8, 130.1, 130.1, 129.8, 129.8, 129.2, 129.2, 127.2, 124.9, 124.2, 120.8, 120.8, 115.1, 56.0, 28.2, 21.0, 20.8; MS (ESI) *m/z* 548.30 [M+Na]⁺; HRMS (ESI) *m/z* calcd for C₂₇H₂₄FNNaO₇S [M+Na]⁺: 548.1155, found: 548.1152.

4.1.6.4. (*E*)-**3,4-diacetoxystyryl 4-3.5-difluorobenzamidophenethyl sulfone (11d)** 36%; Yellow oil; ¹H NMR (400MHz, DMSO-*d*₆) δ 10.14 (s, 1H), 6.83-7.79 (m, 12H), 3.44 (m, 2H), 2.96 (m, 2H), 2.33 (s, 6H); ¹³C NMR (100MHz, DMSO-*d*₆) δ 164.7, 161.0, 161.0, 146.5, 145.9, 137.4, 135.1, 135.0, 134.5, 129.2, 127.9, 127.9, 124.2, 123.2, 121.5, 117.2, 115.2, 110.1, 110.1, 108.1, 60.7,

27.5, 20.9, 20.8; MS (ESI) m/z 566.08 [M+Na]⁺; HRMS (ESI) m/z calcd for C₂₇H₂₃F₂NNaO₇S [M+Na]⁺: 566.1061, found: 566.1058.

4.1.6.5. *(E)*-3,4-diacetoxystyryl 4-cyclohexylamidophenethyl sulfone (11e) 47%; Buff white solid; mp 98–99 °C; ¹H NMR (400MHz, DMSO- d_6) δ 10.12 (s, 1H), 6.74-7.85 (m, 9H), 3.44 (m, 2H), 2.95 (m, 2H), 2.29 (s, 6H), 1.20-1.76 (m, 11H); ¹³C NMR (100MHz, DMSO- d_6) δ 174.6, 149.3, 146.1, 143.9, 138.4, 130.4, 130.0, 130.0, 124.3, 125.9, 125.9, 122.5, 119.6, 119.6, 116.2, 115.7, 55.5, 45.3, 29.6, 29.6, 28.2, 25.9, 25.8, 25.7; MS (ESI) *m*/*z* 536.22 [M+Na]⁺; HRMS (ESI) *m*/*z* calcd for C₂₇H₃₁NNaO₇S [M+Na]⁺: 536.1719, found: 536.1718.

4.1.6.6. *(E)*-3,4-diacetoxystyryl 4-1-adamantaneformylamidophenethyl sulfone (11f) 18%; Yellow solid; mp 104–105 °C; ¹H NMR (400MHz, DMSO- d_6) δ 10.04 (s, 1H), 6.66-8.14 (m, 9H), 3.54 (m, 2H), 3.12 (m, 2H), 2.14 (s, 6H), 1.60-2.00 (m, 15H); ¹³C NMR (100MHz, DMSO- d_6) δ 174.6, 169.3, 169.3, 149.3, 146.1, 143.9, 148.4, 132.9, 129.2, 129.1, 129.1, 124.3, 122.5, 122.4, 119.6, 119.5, 116.2, 115.8, 55.6, 45.3, 29.6, 29.6, 29.6, 28.2, 25.9, 25.7, 25.7, 25.7, 24.7, 24.7; MS (ESI) *m*/*z* 566.24 [M+H]⁺ 588.17 [M+Na]⁺; HRMS (ESI) *m*/*z* calcd for C₃₁H₃₅NNaO₇S [M+Na]⁺: 588.2032, found: 588.2039.

4.1.6.7. (*E*)-**3,4-diacetoxystyryl 4-4-methylbenzamidophenethyl sulfone** (**11g**) 25%; White solid; mp 108–109 °C; ¹H NMR (400MHz, DMSO- d_6) δ 10.12 (s, 1H), 6.78-7.96 (m, 13H), 3.44 (m, 2H), 2.95 (m, 2H), 2.43 (s, 3H), 2.34 (s, 6H); ¹³C NMR (100MHz, DMSO- d_6) δ 168.8, 168.6, 165.4, 144.5, 142.9, 141.8, 141.2, 138.4, 133.5, 131.9, 131.8, 130.1, 130.1, 129.4, 129.4, 129.2, 129.2, 127.5, 124.9, 124.2, 121.0, 121.0, 115.1, 56.0, 28.3, 23.5, 20.9, 20.8; MS (ESI) *m/z* 544.11 [M+Na]⁺; HRMS (ESI) *m/z* calcd for C₂₈H₂₇NNaO₇S [M+Na]⁺: 544.1406, found: 544.1399.

4.1.6.8. (*E*)-**3,4-diacetoxystyryl 4-4-nitrobenzamidophenethyl sulfone (11h)** 35%; Yellow solid; mp 126–127 °C; ¹H NMR (400MHz, DMSO- d_{δ}) δ 10.21 (s, 1H), 6.80-7.95 (m, 13H), 3.43-3.46 (m, 2H), 2.94-2.98 (m, 2H), 2.33 (s, 6H); ¹³C NMR (100MHz, DMSO- d_{δ}) δ 168.6, 168.4, 165.7, 144.0, 142.7, 141.6, 138.2, 133.7, 133.3, 131.9, 131.6, 129.9, 129.9, 129.3, 129.3, 129.0, 129.0, 127.3, 124.7, 124.0, 120.8, 120.8, 114.9, 55.8, 28.1, 20.7, 20.6; MS (ESI) *m*/*z* 575.29 [M+Na]⁺; HRMS (ESI) *m*/*z* calcd for C₂₇H₂₄N₂NaO₉S [M+Na]⁺: 575.1100, found: 575.1101.

4.1.6.9. *(E)*-**3,4-diacetoxystyryl 4-2-thiopheneacetylamidophenethyl sulfone (11i)** 16%; Yellow brown solid; mp 92–93 °C; ¹H NMR (400MHz, DMSO-*d*₆) δ 7.00-8.03 (m, 12H), 6.80 (s, 1H), 3.48 (s, 2H), 3.43 (m, 2H), 2.95 (m, 2H), 2.30 (s, 6H); ¹³C NMR (100MHz, DMSO-*d*₆) δ 169.5, 168.5, 168.5, 149.3, 146.1, 143.9, 140.2, 138.4, 135.9, 133.3, 130.1, 129.2, 129.2, 129.0, 128.4, 127.3, 124.7, 124.2, 120.8, 120.8, 113.5, 55.4, 45.5, 28.0, 20.7, 20.5; MS (ESI) *m*/*z* 550.18 [M+Na]⁺; HRMS (ESI) *m*/*z* calcd for C₂₆H₂₅NNaO₇S₂ [M+Na]⁺: 550.0970, found: 550.0968.

4.1.7. General synthesis of (E)-3,4-dihydroxystyryl 4-acylaminophenethyl sulfones (13a – 13i)

To a solution of (E)-3,4-diacetoxystyryl 4-acylaminophenethyl sulfones (**11a** – **11i**, 0.06mmol) in methanol (10mL), potassium carbonate (0.18mmol) was added and the resulting solution was allowed to stir for 3h at room temperature. The solvent was evaporated under vacuum and the mixture was diluted with H₂O and extracted with ethyl acetate. The combined organic fractions were washed with brine, dried with sodium sulfate, and concentrated in vacuo. Purification of the crude product by column chromatography (PE/EA) afforded the target compounds **13a** to **13i**.

4.1.7.1. (*E*)-**3,4-dihydroxystyryl 4-benzamidophenethyl sulfone** (**13a**) 65%; White solid; mp 104–105 °C; ¹H NMR (400MHz, DMSO- d_6) δ 10.12 (s, 1H), 9.72 (s, 1H), 9.21 (s, 1H), 6.80-7.94 (m, 14H), 3.44 (m, 2H), 2.96 (m, 2H); ¹³C NMR (100MHz, DMSO- d_6) δ 165.9, 149.4, 146.1, 144.0, 138.1, 135.4, 133.8, 132.0, 129.1, 129.1, 128.8, 128.8, 128.1, 128.1, 124.4, 122.5, 122.5, 120.9, 120.9, 116.2, 115.8, 55.5, 28.2; MS (ESI) *m*/*z* 422.25 [M-H]⁻; HRMS (ESI) *m*/*z* calcd for C₂₃H₂₀NO₅S [M-H]⁻: 422.1062, found: 422.1062.

4.1.7.2. *(E)***-3,4-dihydroxystyryl 4-4-methoxylbenzamidophenethyl sulfone (13b)** 58%; Buff white solid; mp 108–109 °C; ¹H NMR (400MHz, DMSO-*d*₆) δ 10.20 (s, 1H), 9.71 (s, 1H), 9.22 (s, 1H), 6.81-7.96 (m, 13H), 3.81 (s, 3H), 3.43 (m, 2H), 2.95 (m, 2H); ¹³C NMR (100MHz, DMSO-*d*₆) δ 165.3, 162.3, 149.3, 146.1, 144.0, 138.3, 133.6, 130.4, 130.0, 120.0, 129.1, 129.1, 127.4, 123.9, 122.5, 120.9, 120.9, 116.2, 115.7, 114.1, 114.1, 55.9, 55.5, 28.2; MS (ESI) *m/z* 452.10 [M-H]⁻; HRMS (ESI) *m/z* calcd for C₂₄H₂₂NO₆S [M+Na]⁺: 452.1168, found: 452.1166.

4.1.7.3. *(E)***-3,4-dihydroxystyryl 4-4-fluorobenzamidophenethyl sulfone** (**13c**) 61%; Buff white oil; ¹H NMR (400MHz, DMSO- d_6) δ 10.21 (s, 1H), 9.71 (s, 1H), 9.20 (s, 1H), 6.79-8.12 (m, 13H), 3.43 (m, 2H), 2.97 (m, 2H); ¹³C NMR (100MHz, DMSO- d_6) δ 164.7, 149.5, 146.5, 144.5, 138.0, 135.5, 134.2, 132.1, 129.2, 129.2, 128.8, 128.8, 124.8, 123.2, 121.5, 121.5, 115.5, 115.2, 55.7, 28.5; MS (ESI) *m/z* 440.26 [M-H]⁻; HRMS (ESI) *m/z* calcd for C₂₃H₁₉FNO₅S [M-H]⁻: 440.0968, found: 440.0972.

4.1.7.4. (*E*)-**3,4-dihydroxystyryl 4-3.5-difluorobenzamidophenethyl sulfone (13d)** 56%; Yellow oil; ¹H NMR (400MHz, DMSO- d_6) δ 10.14 (s, 1H), 9.71 (s, 1H), 9.22 (s, 1H), 6.83-7.79 (m, 12H), 3.44 (m, 2H), 2.96 (m, 2H); ¹³C NMR (100MHz, DMSO- d_6) δ 164.7, 161.0, 161.0, 146.5, 145.9, 137.4, 135.1, 135.0, 134.5, 129.2, 127.9, 127.9, 124.2, 123.2, 121.5, 121.5, 117.2, 115.2, 110.1, 110.1, 108.1, 60.7, 27.5; MS (ESI) *m/z* 458.21 [M-H]⁻; HRMS (ESI) *m/z* calcd for C₂₃H₁₈F₂NO₅S [M-H]⁻: 458.0874, found: 458.0878.

4.1.7.5. (*E*)-**3,4-dihydroxystyryl 4-cyclohexylamidophenethyl sulfone** (**13e**) 65%; White solid; mp 83–84 °C; ¹H NMR (400MHz, DMSO-*d*₆) δ 10.22 (s, 1H), 9.72 (s, 1H), 9.22 (s, 1H), 6.80-7.80 (m, 9H), 3.56 (m, 2H), 2.92 (m, 2H), 1.19-2.32 (m, 11H); ¹³C NMR (100MHz, DMSO-*d*₆) δ 174.7, 149.3, 146.1, 143.9, 138.4, 130.4, 129.1, 129.1, 124.3, 123.9, 122.4, 119.6, 119.4, 116.2, 115.7, 55.5,

45.3, 29.6, 29.6, 28.2, 25.8, 25.7, 25.7; MS (ESI) *m*/*z* 428.32 [M-H]⁻; HRMS (ESI) *m*/*z* calcd for C₂₃H₂₆NO₅S [M-H]⁻: 428.1532, found: 428.1532.

4.1.7.6. (*E*)-3,4-dihydroxystyryl 4-1-adamantaneformylamidophenethyl sulfone (13f) 59%; Yellow solid; mp 122–123 °C; ¹H NMR (400MHz, DMSO- d_6) δ 9.65 (s, 1H), 9.05 (s, 1H), 9.05 (s, 1H), 6.76-8.13 (m, 9H), 3.54 (m, 2H), 3.12 (m, 2H), 1.55-2.07 (m, 15H); ¹³C NMR (100MHz, DMSO- d_6) δ 174.6, 149.3, 146.0, 143.5, 138.4, 132.9, 129.2, 129.1, 124.3, 121.5, 121.4, 119.6, 119.6, 119.5, 116.0, 115.8, 55.6, 45.3, 29.6, 29.6, 29.6, 28.1, 25.9, 25.9, 25.7, 25.7, 25.7; MS (ESI) *m/z* 480.17 [M-H]⁻; HRMS (ESI) *m/z* calcd for C₂₇H₃₀NO₅S [M-H]⁻: 480.1845, found: 480.1843.

4.1.7.7. *(E)*-3,4-dihydroxystyryl 4-4-methylbenzamidophenethyl sulfone (13g) 58%; Light brown solid; mp 119–120 °C; ¹H NMR (400MHz, DMSO- d_6) δ 10.21 (s, 1H), 9.51 (s, 2H), 6.79-7.86 (m, 13H), 3.44 (m, 2H), 2.95 (m, 2H), 2.38 (s, 3H); ¹³C NMR (100MHz, DMSO- d_6) δ 165.7, 149.4, 146.1, 144.0, 142.0, 138.2, 133.7, 132.5, 129.4, 129.4, 129.4, 129.1, 129.1, 128.1, 128.1, 124.3, 122.5, 120.9, 120.9, 116.2, 115.8, 55.5, 28.2, 21.4; MS (ESI) *m/z* 436.15 [M-H]⁻; HRMS (ESI) *m/z* calcd for C₂₄H₂₂NO₅S [M-H]⁻: 436.1219, found: 436.1219.

4.1.7.8. (*E*)-**3,4-dihydroxystyryl 4-4-nitrobenzamidophenethyl sulfone** (**13h**) 60%; White solid; mp 138–139 °C; ¹H NMR (400MHz, DMSO-*d*₆) δ 10.21 (s, 1H), 9.72 (s, 1H), 9.21 (s, 1H), 6.78-8.39 (m, 13H), 3.44 (m, 2H), 2.95 (m, 2H); ¹³C NMR (100MHz, DMSO-*d*₆) δ 164.7, 151.3, 146.5, 145.9, 136.8, 135.1, 135.0, 134.5, 129.6, 129.6, 127.9, 127.9, 124.2, 124.0, 123.2, 124.0, 121.5, 121.5, 117.2, 115.2, 60.7, 27.5; MS (ESI) *m*/*z* 467.08 [M-H]⁻; HRMS (ESI) *m*/*z* calcd for C₂₃H₁₉N₂O₇S [M-H]⁻; 467.0913, found: 467.0911.

4.1.7.9. (*E*)-**3,4-dihydroxystyryl 4-2-thiopheneacetylamidophenethyl sulfone** (**13i**) 59%; Yellow brown solid; mp 87–88 °C; ¹H NMR (400MHz, DMSO- d_6) δ 9.73 (s, 1H), 9.24 (s, 1H), 6.81-7.79 (m, 12H), 5.31 (s, 1H), 3.54 (m, 4H), 2.96 (m, 2H); ¹³C NMR (100MHz, DMSO- d_6) δ 158.8, 146.5, 145.9, 137.4, 135.1, 135.0, 134.5, 129.2, 127.9, 127.9, 124.2, 123.2, 121.5, 121.5, 117.2, 115.2, 110.1, 110.1, 108.1, 60.7, 38.0, 27.6; MS (ESI) *m/z* 442.07 [M-H]⁻; HRMS (ESI) *m/z* calcd for C₂₂H₂₀NO₅S₂ [M-H]⁻: 442.0783, found: 442.0778.

4.1.8. General synthesis of (*E*)-*N*-(4-(2-((3,4-bis(methoxymethoxy)styryl)sulfinyl)ethyl)phenyl) amides (12a – 12f)

To a solution of (E)-4-(2-((3,4-bis(methoxymethoxy)styryl)sulfinyl)ethyl)aniline (10, 0.1mmol) in dichloromethane (10mL), trimethylamine (0.2mmol), and various acid chlorides (0.22mmol) were added under the ice bath. The resulting solution was allowed to stir for 2h. The solvent was evaporated under vacuum and the mixture was diluted with H₂O and extracted with ethyl acetate. The combined organic fractions were washed with brine, dried with sodium sulfate, and

concentrated in vacuo. The crude products of **12a** to **12f** were washed with petroleum ether and were immediately added to the next step.

4.1.9. General synthesis of (E)-3,4-dihydroxystyryl 4-acylaminophenethyl sulfoxides (14a – 14f)

To a solution of (E)-N-(4-(2-((3,4-bis(methoxymethoxy)styryl)sulfinyl)ethyl)phenyl)amides (**12a** – **12f**, 0.06mmol) in methanol (10mL), hydrochloric acid (catalytic amount) was added and the resulting solution was heated to reflux for 3h. The solvent was evaporated under vacuum and the mixture was diluted with H₂O and extracted with ethyl acetate. The combined organic fractions were washed with brine, dried with sodium sulfate, and concentrated in vacuo. Purification of the crude product by column chromatography (PE/EA) afforded the target compounds **14a** to **14f**.

4.1.9.1. (*E*)-**3,4-dihydroxystyryl 4-benzamidophenethyl sulfoxide (14a)** 15%; White solid; mp 97–98 °C; ¹H NMR (400MHz, DMSO-*d*₆) δ 10.21 (s, 1H), 9.72 (s, 1H), 9.21 (s, 1H), 6.79-7.95 (m, 14H), 2.74-3.04 (m, 4H); ¹³C NMR (100MHz, DMSO-*d*₆) δ 166.0, 149.4, 146.1, 143.9, 138.1, 135.5, 133.9, 132.0, 129.1, 129.1, 128.8, 128.8, 128.1, 128.1, 124.4, 122.5, 122.5, 120.9, 120.9, 116.2, 115.7, 50.3, 36.9; MS (ESI) *m*/*z* 406.10 [M-H]⁻; HRMS (ESI) *m*/*z* calcd for C₂₃H₂₀NO₄S [M-H]⁻: 406.1113, found: 406.1112.

4.1.9.2. (*E*)-3,4-dihydroxystyryl 4-4-methoxylbenzamidophenethyl sulfoxide (14b) 11%; Yellow oil; ¹H NMR (400MHz, DMSO- d_6) δ 10.27 (s, 1H), 9.71 (s, 1H), 9.21 (s, 1H), 6.81-8.44 (m, 13H), 3.81 (s, 3H), 2.74-3.04 (m, 4H); ¹³C NMR (100MHz, DMSO- d_6) δ 165.5, 162.5, 149.5, 146.3, 144.1, 138.5, 133.8, 130.6, 130.2, 130.2, 129.3, 129.3, 127.6, 124.1, 122.6, 121.1, 121.1, 116.4, 115.9, 114.3, 114.3, 56.1, 51.6, 33.1; MS (ESI) *m/z* 436.11 [M-H]⁻; HRMS (ESI) *m/z* calcd for C₂₄H₂₂NO₅S [M+Na]⁺: 436.1219, found: 436.1216.

4.1.9.3. *(E)***-3,4-dihydroxystyryl 4-4-fluorobenzamidophenethyl sulfoxide** (**14c**) 17%; Yellow oil; ¹H NMR (400MHz, DMSO-*d*₆) δ 10.27 (s, 1H), 9.72 (s, 1H), 9.21 (s, 1H), 6.79-8.44 (m, 13H), 2.76-3.00 (m, 4H); ¹³C NMR (100MHz, DMSO-*d*₆) δ 168.2, 165.9, 149.5, 146.3, 144.2, 142.1, 138.4, 133.9, 132.7, 129.5, 129.5, 129.2, 128.3, 128.3, 124.5, 122.6, 122.6, 121.1, 121.1, 126.4, 115.9, 52.8, 31.9; MS (ESI) *m/z* 424.09 [M-H]⁻; HRMS (ESI) *m/z* calcd for C₂₃H₁₉FNO₄S [M-H]⁻: 424.1019, found: 424.1022.

4.1.9.4. (*E*)-3,4-dihydroxystyryl **4-3.5-difluorobenzamidophenethyl sulfoxide** (14d) 23%; Yellow oil; ¹H NMR (400MHz, DMSO- d_6) δ 10.27 (s, 1H), 9.72 (s, 1H), 9.22 (s, 1H), 6.80-8.43 (m, 12H), 2.76-3.04 (m, 4H); ¹³C NMR (100MHz, DMSO- d_6) δ 165.9, 161.0, 161.0, 149.4, 146.1, 144.0, 138.1, 135.5, 133.9, 132.0, 129.1, 128.9, 128.9, 122.5, 122.5, 121.0, 116.2, 115.7, 115.2, 110.6, 108.7, 108.7, 50.4, 36.9; MS (ESI) *m*/*z* 442.08 [M-H]⁻; HRMS (ESI) *m*/*z* calcd for C₂₃H₁₈F₂NO₄S [M-H]⁻: 442.0925, found: 442.0928.

4.1.9.5. (*E*)-**3,4-dihydroxystyryl 4-cyclohexylamidophenethyl sulfoxide** (**14e**) 10%; Yellow oil; ¹H NMR (400MHz, DMSO-*d*₆) δ 10.27 (s, 1H), 9.72 (s, 1H), 9.22 (s, 1H), 6.80-7.94 (m, 9H), 2.74-3.04 (m, 4H), 1.19-1.76 (m, 11H); ¹³C NMR (100MHz, DMSO-*d*₆) δ 174.8, 149.5, 146.3, 144.1, 138.6, 133.1, 129.4, 129.3, 124.5, 122.7, 122.6, 119.8, 119.7, 116.4, 116.0, 52.3, 45.5, 34.1, 29.8, 29.8, 26.1, 25.9, 25.9; MS (ESI) *m/z* 412.15 [M-H]⁻; HRMS (ESI) *m/z* calcd for C₂₃H₂₆NO₄S [M-H]⁻: 412.1583, found: 412.1582.

4.1.9.6. (*E*)-**3,4-dihydroxystyryl 4-1-adamantaneformylamidophenethyl sulfoxide (14f)** 15%; Yellow solid; mp 111–112 °C; ¹H NMR (400MHz, DMSO- d_6) δ 10.05 (s, 1H), 9.65 (s, 1H), 9.05 (s, 1H), 6.79-7.96 (m, 9H), 2.74-3.04 (m, 4H), 1.60-2.00 (m, 15H); ¹³C NMR (100MHz, DMSO- d_6) δ 174.7, 149.3, 146.1, 143.9, 138.4, 130.4, 129.1, 129.1, 124.3, 123.9, 122.5, 122.5, 119.6, 119.6, 116.2, 115.7, 52.4, 35.7, 35.4, 35.4, 34.3, 34.3, 32.9, 32.5, 32.1, 32.1, 25.9; MS (ESI) *m/z* 464.18 [M-H]⁻; HRMS (ESI) *m/z* calcd for C₂₇H₃₀NO₄S [M-H]⁻: 464.1896, found: 464.1893.

4.2. Biology

4.2.1. Cytotoxicity assays

The cytotoxicity of all target compounds were assessed *via* measurement of the amount of TZM-bl cells after exposure to the compounds for 2 to 3 days. In short, the target compounds were dissolved in DMSO cell freezing medium. TZM-bl cells were seeded in 96-well culture plates at 1×10^5 cells per mL (100µL per well). The cells were treated with various concentrations (1000µM, 200µM, 40µM, 8µM, and 1.6µM) of each compounds and were cultivated under 5% CO₂ at 37°C. Three wells were used in parallel for each concentration. Cells treated with culture medium were used as a negative control. Two days later, the culture medium was discarded and 10µL of coloring agent CCK-8 solution was added to each well before incubation was allowed to last for another three to four hours. The absorbance (*A*, at 450nm) was measured by a microplate reader. The (%) survival of TZM-bl cells was calculated using the following formula: (*A*₄₅₀ of the experimental group/*A*₄₅₀ of the negative control group) × 100%.

4.2.2. Determination of antivirus activity against R5 HIV-1 strains

TZM-bl cells, the same type used in cytotoxicity assays, were seeded in 96-well plates. Complete medium were added to all sample wells, viral control wells and cell control wells. Twelve hours later, the culture medium was discarded before the solutions of target compounds were diluted to 50μ M and added to sample wells. Two wells were used in parallel for each compound. HIV-1 Bal (R5) virus (200TCID50) were added into all sample wells and virus control wells and DEAE was added to a final concentration of 10μ g/mL. The infected cells were cultivated for 3 days under 5% CO₂ at 37° C. The luciferase assay substrate were dissolved by luciferase lysis buffer and were added (100μ L per well) into all experiment wells which had 100μ L of the culture medium discarded beforehand. The luciferase assay system was shielded from light and was allowed to react for 2 minutes at room temperature. The mixtures in all wells were blown to well-distributed and

transferred to a black test plate for 150μ L for each well. The results were measured by a fluorescence detector and the % inhibition of virus' infection was represented with the decrease in fluorescent intensity.

4.2.3. SPR binding assays

The SPR binding assays were performed as previously described by using SPR biosensor technology. The experiments were carried out on a Biacore T200 instrument that contains dual flow cells. Immobilization of IN proteins to sensor chip CM5 (Biacore) was carried out by primary amine coupling method resulting in an immobilization level of approximately 10000RU. By using automatic corrections of Biacore T200 Evaluation Software, all the raw sensorgrams were processed along with solvent corrected. The specific binding figures of compounds to immobilized IN were obtained after subtracting response signal from control flow cell. All Biacore data were collected at 25°C with PBST as running buffer at a constant flow of 30μ L/min. The equilibrium dissociation constants (K_D) evaluating the binding affinity between IN and compounds were determined using 1:1 binding model and the curve fitting efficiency was checked by χ 2 and residual plots.

Acknowledgements

This study was supported by the National Natural Science Foundation of China (Grants 20972011, 21042009, and 21172014) and Ministry of Science and Technology of China (Grant 2009ZX09301-010).

References and notes

- 1. Barton, K. M.; Burch, B. D.; Soriano-Sarabia, N.; Margolis, D. M. Clin. Pharmacol Ther. 2013, 93, 46.
- 2. Latham, C. F.; La, J.; Tinetti, R. N.; Chalmers, D. K.; Tachedjian, G. Curr. Top. Med. Chem. 2016, 16, 1135.
- 3. Zimmermann, G. R.; Leha'r, J.; Keith, C. T. Drug Discov. Today 2007, 12, 34.
- 4. Oppermann, M. Cell Signal 2004, 16, 1201.
- 5. Connor, R. I.; Sheridan, K. E.; Ceradini, D.; Choe, S.; Landau, N. R. J. Exp. Med. 1997, 185, 621.
- 6. Dubey, S.; Satyanarayana, Y. D.; Lavania, H. Eur. J. Med. Chem. 2007, 42, 1159.
- 7. Di Santo, R. J. Med. Chem. 2014, 57, 539.
- Tan, Q.; Zhu, Y.; Li, J.; Chen, Z.; Han, G. W.; Kufareva, I.; Li, T.; Ma, L.; Fenalti, G.; Li, J.; Zhang, W.; Xie, X.; Yang, H.; Jiang, H.; Cherezov, V.; Liu, H.; Stevens, R.; Zhao, Q.; Wu, B. *Science* **2013**, *341*, 1387.
- 9. Hare, S.; Gupta, S. S.; Valkov, E.; Engelman, A.; Cherepanov, P. Nature 2010, 464, 232.
- Hare, S.; Vos, A. M.; Clayton, R. F.; Thuring, J. W.; Cummings, M. D.; Cherepanov, P. Proc. Natl. Acad. Sci. U. S. A. 2010, 107, 20057.
- Burke, T. R.; Fesen, Jr., M. R.; Mazumder, A.; Wang, J.; Carothers, A. M.; Grunberger, D.; Driscoll, J.; Kohn, K.; Pommier, Y. J. Med. Chem. 1995, 38, 4171.
- 12. Yan, A.; Xuan, S.; Hu, X. Comb. Chem. High Throughput Screen. 2012, 15, 792.

- Artico, M.; Di Santo, R.; Costi, R.; Novellino, E.; Greco, G.; Massa, S.; Tramontano, E.; Marongiu, M.E.; De Montis, A.; La Colla, P. J. Med. Chem. 1998, 41, 3948.
- 14 Kawasuji, T.; Johns, B. A.; Yoshida, H.; Weatherhead, J. G.; Akiyama, T.; Taishi, T.; Taoda, Y.; Mikamiyama-Iwata, M.; Murai, H.; Kiyama, R.; Fuji, M.; Tanimoto, N.; Yoshinaga, T.; Seki, T.; Kobayashi, M.; Sato, A.; Garvey, E. P.; Fujiwara, T. J. Med. Chem. 2013, 56, 1124.
- Trkola, A.; Ketas, T. J.; Nagashima, K. A.; Zhao, L.; Cilliers, T.; Morris, L.; Moore, J. P.; Maddon, P. J.; Olson, W. C. *J. Virol.* 2001, *75*, 579.
- 16. Bondinell, W. E.; Ku, T. W.; Wang, N. PCT Int. Appl. 2000, WO 2000040239 A1 20000713.
- Ning, X.; Guo, Y.; Wang, X.; Ma, X.; Tian, C.; Shi, X.; Zhu, R.; Cheng, C.; Du, Y.; Ma, Z.; Zhang, Z.; Liu, J. J. Med. Chem. 2014, 57, 4302.

A CERTIFIC MANUS

