Enantioselective Total Synthesis of (+)-Scyphostatin, a Potent and Specific Inhibitor of Neutral Sphingomyelinase

Munenori Inoue,^a Wakako Yokota,^a Tadashi Katoh*^b

- ^a Sagami Chemical Research Center, 2743-1 Hayakawa, Ayase, Kanagawa 252-1193, Japan
- ^b Department of Chemical Pharmaceutical Science, Tohoku Pharmaceutical University, 4-4-1 Komatsushima, Aoba-ku, Sendai 981-8558, Japan
- Fax +81(22)7270135; E-mail: katoh@tohoku-pharm.ac.jp

Received 4 August 2006; revised 31 October 2006

Abstract: The total synthesis of (+)-scyphostatin, a specific and potent neutral sphingomyelinase inhibitor from a microorganism, was accomplished for the first time starting from D-arabinose and both enantiomers of methyl 3-hydroxy-2-methylpropionate. The method involves (a) stereoselective aldol coupling of a methyl 1,3-dioxolane-4-carboxylate with the Garner aldehyde to establish the asymmetric quaternary carbon center at C4, (b) ring-closing metathesis of the resultant diene to construct the cyclohexene ring, (c) Negishi coupling of a vinyl iodide and an alkyl iodide to form the requisite trisubstituted *E*-alkene, (d) formation of an amide from the cyclohexene segment and the trisubstituted *E*-alkene fatty acid segment, and (e) stereospecific formation of an epoxide ring from the mesylate of the amide formed from the two segments.

Key words: scyphostatin, sphingomyelinase, inhibitors, total synthesis, aldol reaction, Negishi coupling

Introduction

Scyphostatin (1, Figure 1), isolated from the mycelia extract *Trichopeziza mollissima* SANK 13892 by Ogita et al. in 1997, has been shown to be a powerful and specific inhibitor of membrane-bound neutral sphingomyelinase (N-SMase).^{1,2} It has been reported that 1 inhibits N-SMase and acidic SMase (A-SMase) with IC₅₀ values of 1.0 μ M and 49.3 μ M, respectively.^{1,2} Remarkably, of the many low-molecular-weight N-SMase inhibitors of natural³ or synthetic⁴ origin, this natural product is the most potent and specific one known to date.

SMase is the enzyme that specifically cleaves the phosphoester linkage of sphingomyelin (SM) to generate phosphocholine and ceramide. The SM-derived ceramide has been known as an intracellular lipid second messenger in mammalian cell membranes and plays important roles in the cellular signal transmission pathway, in particular, as a signal transduction factor in cell differentiation and apoptosis induction.⁵ Consequently, SMase inhibitors are considered as valuable chemical tools to explore the biological function of the enzyme and the catabolite ceramide in signal transduction. Additionally, potent and selective SMase inhibitors are highly anticipated to be promising candidates for the treatment of ceramide-mediated patho-

SYNTHESIS 2007, No. 4, pp 0622–0637 Advanced online publication: 18.01.2007 DOI: 10.1055/s-2007-965893; Art ID: F12106SS © Georg Thieme Verlag Stuttgart · New York genic states such as AIDS,⁶ inflammation,⁷ and immunological and neurological disorders.⁸

The gross structure of scyphostatin (1) was revealed by extensive spectroscopic analyses and derivatization studies.¹ It consists of a novel, highly oxygenated cyclohexenone ring attached to an aminopropanol side chain substituted with a 20-carbon unsaturated fatty acid. This initial structure elucidation only established the relative and absolute stereochemistry of the cyclohexenone portion of 1. Subsequently, Kogen et al. determined the relative and absolute configurations of the three stereogenic centers at C8', C10', and C14' in the fatty acid side chain.⁹ At almost the same time, Hoye et al. achieved an enantioselective synthesis of the 20-carbon unsaturated fatty acid moiety for the first time, and provided alternative proof of its stereostructure, including the absolute configuration.¹⁰



Figure 1 Structure of (+)-scyphostatin (1)

The remarkable biological properties and unique structural features have made **1** an exceptionally intriguing and timely target for total synthesis. So far, many synthetic efforts toward scyphostatin (**1**) have been reported.¹¹ We have already reported our own preliminary results concerning the enantioselective synthesis of the epoxycyclohexenone substructure.¹² Subsequently, we have also disclosed an efficient method for the introduction of a fatty acid side chain at the aminopropanol moiety.¹³ More recently, we have communicated the first total synthesis of (+)-scyphostatin (**1**) in an enantiomerically pure form.¹⁴ In this article, we wish to describe the full details of our successful total synthesis of (+)-**1**.

Results and Discussion

Synthetic Plan for (+)-Scyphostatin (1)

Our synthetic plan for (+)-scyphostatin (1), outlined in Scheme 1, is based on our intensive model studies.¹² Since the highly oxygenated cyclohexenone system in 1 was expected to be fairly labile under acidic and/or basic conditions,^{1,2} we planned the construction of the epoxycyclohexenone system to be at a late stage of the synthesis. The target molecule 1 would be produced from the cyclohexenone segment 2 and the 20-carbon fatty acid segment 3 via an amide coupling of these segments, followed by enone and epoxide-ring formations.

The segment 2 would be derived from diene 4 through ring-closing metathesis (RCM). The RCM substrate 4 would be prepared through aldol coupling of ester 5 with the Garner aldehyde (6),¹⁵ where we envisioned that 6 approaches exclusively from the less hindered α -face of the enolate of 5, leading to establishment of the requisite asymmetric quaternary carbon center at the C4-position (scyphostatin numbering). This type of coupling reaction is considerably challenging at the synthetic chemistry level, because substrate 5 possesses unusual trihydroxy functionalities at the C4-, C5-, and C6-positions. Ester 5, in turn, could be accessible from D-arabinose (7) that has the correct stereochemistries for 5.

On the other hand, segment 3 could be derived from the Hoye intermediate 8¹⁰ through Horner-Wadsworth-Emmons (HWE) olefination. The key feature of our synthesis of 8 is Negishi coupling between vinyl iodide 9 and alkyl iodide 10 to generate the requisite trisubstituted Eolefin 8. Intermediates 9 and 10 could be accessed from the two commercially available isomers methyl (R)-3-hydroxy-2-methylpropionate (11) and its S-isomer ent-11, respectively.

Biographical Sketches



Munenori Inoue was born in 1971 in Hiroshima, Japan. He received his PhD in 1999 from the University of Tokyo under the direction of Prof. Takeshi Kitahara. After spending a postdoctoral year (1999–2001) with Prof. John L. Wood at Yale Uni-

Wakako Yokota was born in 1975 in Kochi, Japan. She studied pharmaceutical science at Tokushima University and obtained her BS degree in 1999 and MS degree in 2001 under the guidof Prof. Kozo

versity, he joined the group of Prof. Tadashi Katoh at Sagami Chemical Research Center as a research fellow (2001-2004). In 2004 he became a junior research fellow and started his own research at the same institute. He is also a visiting

Shishido. In 2001 she joined the group of Prof. Tadashi Katoh at Sagami Chemical Research Center as a research fellow (2001-2003). In 2003 she was transferred to the group of Dr. Kenji Hirai at the same institute, associate professor at the Tokyo Institute of Technology. His current research interests include the synthesis of biologically active compounds, especially natural products, and the development of new chemical reactions.

where she was engaged in the synthesis of new agrochemicals (2003 - 2006).Currently she works as a pharmacist at Kitasato Pharmacy in Kanagawa, Japan.





postdoctoral year (1988-

1989) with Prof. Philip D.

Magnus at Indiana Univer-

sity and the University of

Texas, he joined Sagami

Chemical Research Center as a research fellow in 1989, where he enjoyed working with Dr. Shiro Terashima (1989-1998). In 1998 he was appointed as a visiting professor at the Tokyo Institute of Technology, and started his independent research career. He received a Progress Award (1998) and a Pfizer Award (1999) in synthetic organic chemistry in Japan. In 2004 he moved to Tohoku Pharmaceutical University as full professor. His research interests include development of new synthetic methodologies for structurally unique and biologically significant natural products.



Scheme 1 Synthetic plan for (+)-scyphostatin (1)

Synthesis of Methyl Ester 5

At first, we pursued the synthesis of methyl ester **5** (Scheme 2), the substrate for the crucial aldol coupling reaction, starting from the known compound **12**,¹⁶ readily derived from D-arabinose (**7**) by *O*-benzylglycosylation and acetonide formation. After *p*-methoxybenzyl protection of the hydroxy group in **12** (70% yield), the benzyl protecting group of the resulting ether **13** was removed by hydrogenolysis to furnish hemiacetal **14** (2:1 mixture of α -and β -anomers) in 86% yield. Wittig olefination of **14** afforded alcohol **15** (86% yield), which was further converted into substrate **5** in 74% overall yield over a three-step sequence involving Swern oxidation, oxidation with sodium chlorite, and methyl esterification with diazomethane.

Synthesis of Diene 4

Having obtained intermediate **5**, we next carried out the synthesis of diene **4**, the substrate for the key RCM reaction, as shown in Scheme 3. Initial attempts to achieve the aldol coupling between **5** and the Garner aldehyde $(6)^{15}$ with a lithium base such as lithium diisopropylamide or

lithium hexamethyldisilazide proved to be problematic: the reaction rate was very slow and the yield of the coupling product 18 was low (ca. 30%). After several experiments, we were delighted to find that the use of a sodium base such as sodium hexamethyldisilazide was fairly effective for this coupling reaction. Thus, treatment of 5 with sodium hexamethyldisilazide (1.2 equiv) in tetrahydrofuran at -78 °C, followed by reaction with the Garner aldehyde (6) (1.1 equiv) at the same temperature for four hours provided the desired coupling product 18 in 69% yield as a single diastereomer. The newly formed C4 stereochemistry of the product 18 was completely controlled, as we expected; the assignment was later confirmed by an NOESY experiment of the transformed compound 19. The C3 stereochemistry in 18 was tentatively assigned on the basis of the well-known Felkin-Anh model. As shown in intermediate 17 (Scheme 3), the electrophile 6 (E^+) would be accessed exclusively from the less hindered α face of the sodium enolate under the influence of the adjacent C5 stereocenter, leading to establishment of the requisite asymmetric quaternary carbon center at C4 in **18**.



Scheme 2 Synthesis of intermediate 5. *Reagents and conditions*: (a) PMBCl, NaH, DMSO, r.t., 70%; (b) H_2 , Raney Ni, EtOH, r.t., 86%; (c) Ph₃MeP⁺Br⁻, *t*-BuOK, benzene, reflux, 86%; (d) DMSO, (COCl)₂, CH₂Cl₂, -78 °C, then *i*-Pr₂NEt, -78 to 0 °C, 95%; (e) NaClO₂, NaH₂PO₄, DMSO-H₂O (4:1), r.t.; (f) CH₂N₂, Et₂O, MeOH, 0 °C, 78% (2 steps).

The secondary hydroxy group in **18** was eliminated by use of the Barton–McCombie protocol,¹⁷ to give the deoxygenated product **19** in 53% overall yield (92% yield based on recovery of **18**). At this stage, the C4 stereochemistry could be unambiguously confirmed by 500-MHz ¹H NMR NOESY experiments of **19**, in which a clear NOE interaction between C3-H and C5-H was observed.

To continue the synthesis, methyl ester **19** was reduced with diisobutylaluminum hydride in dichloromethane at -100 °C, providing the corresponding aldehyde **20** in 88% yield. Subsequent Grignard reaction of **20** with vinylmagnesium bromide in tetrahydrofuran at 0 °C furnished diene **4** in 93% yield as the single diastereomer. This reaction proceeded presumably through the usual Felkin– Anh model. The C9 stereochemistry of **4** was later confirmed by a NOESY experiment of the transformed compound **2** (see below).

Synthesis of Cyclohexenone Segment 2

We next conducted the synthesis of the cyclohexene segment **2** as shown in Scheme 4. The critical RCM reaction¹⁸ of diene **4** proceeded smoothly and cleanly in refluxing dichloromethane in the presence of the firstgeneration Grubbs catalyst [RuCl₂(=CHPh)(PCy₃)₂]¹⁹ (10 mol%) for 12 hours, and the desired cyclohexene **21** was produced in 96% yield. After protection of the secondary hydroxy group in **21** (93% yield), the *N*,*O*-isopropylidene group of the resulting *tert*-butyldimethylsilyl ether **22** was selectively cleaved by treatment with pyridinium *p*-toluenesulfonate in ethanol at 60 °C; this gave the requisite cyclohexene segment **2** in 57% yield (84% yield based on



Scheme 3 Synthesis of intermediate 4. *Reagents and conditions*: (a) NaHMDS, THF, -78 °C, then Garner aldehyde (6), THF, -78 °C, 69%; (b) NaHMDS, THF, 0 °C, then CS₂, then MeI, 0 °C to r.t.; (c) *n*-Bu₃SnH, AIBN, toluene, reflux, 53% (2 steps) (92% based on recovery of **18**); (d) DIBAL-H, CH₂Cl₂, -100 °C, 88%; (e) H₂C=CH₂MgBr, THF, 0 °C, 93%.

recovery of **22**). At this stage, the C9 configuration of the Grignard reaction product **4** (cf. Scheme 3) was determined to be *R* by an NOESY experiment of **2**; the selected NOE correlation is depicted in the formula **2A** (Scheme 4): clear NOE interactions between a C3-H and C5-H, a C3-H and C9-H, an -OSiMe group and an isopropylidene Me group, and an -OSiMe group and C6-H are observed; this revealed an *R*-configuration at C9.



Scheme 4 Synthesis of intermediate 2. *Reagents and conditions*: (a) $[RuCl_2(=CHPh)(PCy_3)_2]$ (10 mol%), CH_2Cl_2 , reflux, 96%; (b) TBSOTf, 2,6-lutidine, CH_2Cl_2 , r.t., 93%; (c) PPTS, EtOH, 60 °C, 57% (84% based on recovery of **22**).

FEATURE ARTICLE

Syntheses of Vinyl Iodide 9 and Alkyl Iodide 10

Having established the synthetic pathway to the cyclohexene segment 2 possessing the requisite multifunctional groups and all the correct stereochemistries, we next undertook the syntheses of vinyl iodide 9 and alkyl iodide 10, both key intermediates for the synthesis of the 20-carbon fatty acid segment 3. As shown in Scheme 5, vinyl iodide 9 was prepared from the known aldehyde 23,²⁰ easily derived from commercially available methyl (R)-3-hydroxy-2-methylpropionate (11). Thus, aldehyde 23 was subjected to Corey-Fuchs acetylene formationmethylation²¹ to afford methylacetylene **24** in 94% yield. Subsequent hydrozirconation-iodination²² (Cp₂ZrHCl, benzene, 40 °C; I₂, r.t.) of 24 produced the desired vinyl iodide 9 in 70% yield as a mixture of 2- and 3-iodo regioisomers (2-I/3-I, 87:17 by 500-MHz ¹H NMR) that were very difficult to separate.



Scheme 5 Synthesis of intermediates 9 and 10. *Reagents and conditions*: (a) PPh₃, CBr₄, CH₂Cl₂, 0 °C; (b) *n*-BuLi, THF, -78 °C, then MeI, 0 °C, 94% (2 steps); (c) Cp₂ZrHCl, benzene, 40 °C, then I₂, r.t., 70%; (d) O₃, CH₂Cl₂–MeOH (3:1), -78 °C, then NaBH₄, r.t., 84%; (e) MsCl, Et₃N, CH₂Cl₂, r.t.; (f) NaI, acetone, reflux, 95% (2 steps).

Alkyl iodide **10**, the coupling partner of **9**, was prepared from the known olefin 25^{23} that was accessed from the commercially available *S*-isomer of **11** (*ent*-**11**) (Scheme 5). Thus, ozonolysis of **25** followed by reductive workup with sodium borohydride provided the corresponding alcohol **26** in 84% yield. Compound **26** was then converted into alkyl iodide **10** in 95% overall yield in a two-step sequence involving conventional mesylation and iodination.

Synthesis of the C-20 Fatty Acid Segment 3

With both vinyl iodide **9** and alkyl iodide **10** in hand, our next efforts were devoted to the synthesis of the 20-carbon fatty acid segment **3**, as shown in Scheme 6. Our initial at-

tempts to realize the crucial Negishi coupling reaction of 10 and 9 under the original conditions^{24,25} [e.g., *t*-BuLi (2.0 equiv), $ZnCl_2$ (1.0 equiv), THF or Et_2O , -78 °C to r.t. for preparation of the zinc reagent of 10; Pd(PPh₃)₄, THF or Et₂O, r.t. to 50 °C for the coupling reaction with 9] resulted in failure; the coupling product 28 was not obtained. The preparation of the zinc reagent of 10 with activated zinc, Rieke zinc, and a zinc-copper couple was also unsuccessful. After several experiments, to our delight, we found that the use of the modified conditions developed by Smith et al.²⁶ gave satisfactory results. Thus, a mixture of 10 and one equivalent of zinc(II) chloride in diethyl ether was treated with three equivalents of tertbutyllithium to generate the putative mixed zinc reagent 27^{26} which was allowed to react with vinyl iodide 9 in the presence of tetrakis(triphenylphosphine)palladium (10 mol%) in tetrahydrofuran at room temperature for 13 hours, leading to the desired coupling product 28 in 84% yield. In this reaction, the minor inseparable 3-iodo regioisomer of 9, as we expected, did not give the corresponding coupling product and was recovered unchanged.

In the next stage, we examined the conversion of the coupling product **28** into the Hoye intermediate **8** (Scheme 6). Initial attempts to remove the benzyl protecting group from **28** under Birch conditions [Li (20 equiv), NH₃(l), tetrahydrofuran, -30 °C] turned out to be fruitless; unexpectedly, the benzene rings of the *tert*-butyldiphenylsilyl group were predominantly reduced to form the corresponding cyclohexa-1,4-diene rings. After several trials, we were pleased to find that lithium 4,4'-di-*tert*-butylbiphenylide (LiDBB)²⁷ was quite effective for this purpose; the desired alcohol **29** was obtained in 90% yield. Subsequent tosylation of **29** under standard conditions delivered the Hoye intermediate **8**.¹⁰

Further transformation of tosylate 8 into aldehyde 32 (Scheme 6) proceeded via intermediates 30 and 31 by sequential methylation with Me₂CuLi (97% yield), deprotection of the *tert*-butyldiphenylsilyl group in 30 with tetrabutylammonium fluoride (91% yield), and oxidation of **31** with *n*-Pr₄NRuO₄.²⁸ Aldehyde **32** was then subjected to the HWE reaction with the known dimethyl phosphonate 33^{29} to provide methyl ester 34 (46% yield from 31) and its 6'Z-isomer (17% yield from 31); these products were separated by HPLC. Spectral data (¹H NMR and ¹³C NMR, MS) of 34 were in good agreement with those reported by Kogen et al.9 Finally, the desired acid chloride 3 was synthesized from 34 via carboxylic acid 35 by alkali hydrolysis of 34 with two molar potassium hydroxide followed by reaction with oxalyl chloride in the presence of *N*,*N*-dimethylformamide.

Completion of the Total Synthesis of (+)-Scyphostatin (1)

The final route that led to completion of the total synthesis of (+)-scyphostatin (1) is summarized in Scheme 7. At first, we investigated the coupling reaction between the cyclohexene segment 2 and the fatty acid segment 3.



Scheme 6 Synthesis of the intermediate 3. *Reagents and conditions*: (a) *t*-BuLi, ZnCl₂, Et₂O, -78 to 0 °C, then 9, Pd(PPh₃)₄, THF, r.t., 81%; (b) LiDBB, THF, -78 °C, 90%; (c) TsCl, Et₃N, DMAP, CH₂Cl₂, r.t.; (d) Me₂CuLi, Et₂O, -40 °C, 97% (2 steps); (e) TBAF, THF, r.t., 91%; (f) *n*-Pr₄NRuO₄, NMO, MS, CH₂Cl₂, r.t.; (g) 33, LDA, THF, -78 to -30 °C, 46% (2 steps); (h) 2 M aq KOH, MeOH-THF, r.t.; (i) (COCl)₂, CH₂Cl₂, DMF, r.t.

Thus, the tert-butoxycarbonyl and p-methoxybenzyl protecting groups of 2 were simultaneously removed by treatment with trimethylsilyl triflate³⁰ in the presence of 2,6lutidine to produce amine **36**, wherein the two hydroxy groups were partially protected as the trimethylsilyl ethers. The liberated amine 36 was immediately treated with acid chloride 3 in the presence of triethylamine to give the amide coupling product 37 (73% yield from 2) upon treatment with aqueous acetic acid. Amide 37 was further converted into mesylate 39 in 67% overall yield by selective acetylation of the primary hydroxy group and subsequent mesylation of the secondary hydroxy group in the resulting acetate 38. Deprotection of the tert-butyldimethylsilyl group in 39 followed by Dess-Martin oxidation³¹ furnished enone **40** in 98% yield for the two steps.

The remaining task to complete the projected synthesis was cleavage of the *N*,*O*-isopropylidene moiety in **40** followed by epoxide-ring formation and final deprotection of the acetyl group. In our preliminary model studies,¹² the epoxide-ring formation was successfully achieved by hydrolysis of an *N*,*O*-isopropylidene moiety with aqueous trifluoroacetic acid followed by treatment with one molar sodium hydroxide at 0 °C. Upon applying these conditions to **40**, however, partial isomerization of the C12'–C13' double bond in the side chain was observed; this is presumably due to the use of a strong acid such as trifluoroacetic acid. Less strong acids such as *p*-toluenesulfonic acid, pyridinium *p*-toluenesulfonate, hydrogen chloride, boron trifluoride–diethyl ether complex, iron(III) chlo-

ride–silica gel, and acetic acid turned out to be ineffective; the corresponding diol was not obtained and the starting material **40** was recovered unchanged. Eventually, we overcame this problem by using aqueous trichloroacetic acid; this, followed by treatment with two molar sodium hydroxide at ambient temperature, produced the desired epoxide **41** in 45% overall yield (82% yield based on recovery of **40**) without appreciable isomerization of the C12'–C13' double bond (Scheme 7).

The final step was deprotection of the acetyl group in **41**. Initial attempts to achieve this under conventional conditions (e.g., K₂CO₃, NaOMe, KOH in MeOH, aq KOH in THF or CH₂Cl₂, DBU or NH₃ in THF) resulted in failure; disappearance of the starting material 41 could be ascertained by TLC analysis, while unidentified decomposition products were generated in the reaction mixture. We reasoned that this failure was due to the sensitivity of the epoxyenone moiety to these basic conditions. Therefore, we decided to attempt the acetyl deprotection by an enzymatic method in the hope that the sensitive epoxyenone system would remain intact under such relatively mild conditions. After the screening of several enzymes such as lipases PS, AY-30G, and F-AP15, we were pleased to find that the desired acetyl deprotection was best achieved by the use of lipase PS. Thus, exposure of 41 to lipase PS in phosphate buffer-acetone at room temperature for ten hours provided (+)-scyphostatin (1) in 60% yield (Scheme 7); the spectroscopic properties (IR, ¹H NMR and ¹³C NMR, and MS) of this product were identical with those of natural 1. The optical rotation of a synthetic sam-



Scheme 7 Synthesis of (+)-scyphostatin (1). *Reagents and conditions*: (a) TMSOTf, 2,6-lutidine, CH_2Cl_2 , r.t., then MeOH; (b) 3, Et₃N, CH_2Cl_2 , r.t., then aq AcOH, 73% (2 steps); (c) Ac₂O, py, DMAP, CH_2Cl_2 , r.t., 72%; (d) MsCl, Et₃N, CH_2Cl_2 , r.t., 93%; (e) TBAF, THF, r.t.; (f) DMP, CH_2Cl_2 , r.t., 98% (2 steps); (g) Cl_3CCO_2H , H_2O , CH_2Cl_2 , reflux, then 2 M NaOH, r.t., 45% (82% based on recovery of **40**); (h) lipase PS, pH 7 phosphate buffer, acetone, r.t., 60%.

ple of $\mathbf{1} \{ [\alpha]_D^{25} + 61.0 (c \ 0.04, MeOH) \}$ was identical to that of natural $\mathbf{1} \{ [\alpha]_D^{25} + 66.4 (c \ 0.09, MeOH) \}$.

Conclusion

We have accomplished the first total, enantioselective synthesis of naturally occurring (+)-scyphostatin (1), starting from D-arabinose derivative 12 in 22 steps (linear sequence of reactions). The key steps of the synthesis were (a) a stereoselective aldol-type coupling reaction of ester 5, derived from D-arabinose (7), with Garner aldehyde (6) to establish the requisite asymmetric quaternary carbon center at C4; (b) RCM reaction of diene 4 to construct the cyclohexene ring 21; (c) Negishi coupling between alkyl iodide 10 and vinyl iodide 9 to give trisubstituted *E*-olefin 28; (d) amide formation to connect

the cyclohexene segment 2 and the fatty acid segment 3; and (e) stereospecific epoxide-ring formation of mesylate 40 to deliver the fully functionalized cyclohexene 2. Importantly, the explored synthetic route has the potential for producing scyphostatin analogues possessing a variety of fatty acid side chains in enantiomerically pure forms. These efforts are currently underway.

Routine monitoring of reactions was carried out on glass-supported Merck silica gel 60 F254 TLC plates. Flash column chromatography was performed on Kanto Chemical Silica Gel 60N (spherical, neutral 40-50 µm) with the solvents indicated at the pertinent procedures. All solvents and reagents were used as supplied, with the following exceptions. THF and Et2O were freshly distilled from Na/ benzophenone under argon. Toluene was distilled from Na under argon. CH₂Cl₂, DMSO, DMF, and benzene were distilled from CaH2 under argon. Optical rotations were measured on a JASCO P-1020 automatic digital polarimeter. Melting points were determined on a Yanaco MP-3 micro melting point apparatus and are uncorrected. ¹H NMR and ¹³C NMR spectra were obtained on a Bruker DRX-500 (500 MHz) spectrometer. Chemical shifts are expressed in ppm relative to TMS ($\delta = 0$) as internal standard. IR spectra were measured on a JASCO FT/IR-5300 spectrometer. HRMS spectra were obtained on a JEOL MStation JMS-700 mass spectrometer.

Benzyl 3,4-O-Isopropylidene-2-O-(4-methoxybenzyl)- β -D-arabinoside (13)

A soln of **12**¹⁶ (5.00 g, 18 mmol) in anhyd DMSO (10 mL) was added dropwise to a stirred suspension of NaH (60% dispersion in mineral oil; 1.00 g, 25 mmol) in anhyd DMSO (15 mL) at 0 °C under argon. After the mixture had stirred for 1 h at r.t., PMBCl (3.10 mL, 23 mmol) was added, and stirring was continued for 15 h at r.t. The reaction was quenched with sat. aq NH₄Cl (10 mL) at 0 °C, and the resulting mixture was extracted with Et₂O (2 × 50 mL). The combined extracts were washed with sat. aq NaHCO₃ (2 × 30 mL) and brine (30 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane–EtOAc, 3:1) to give a white solid; yield: 5.00 g (70%).

White powder (recrystallization, hexane–CHCl₃, 3:1); mp 52.0–53.0 °C; $[\alpha]_D^{20}$ –160.6 (*c* 1.04, CHCl₃).

IR (KBr): 744, 1030, 1091, 1178, 1255, 1383, 1516, 1612, 2926, 2951, 3024 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): $\delta = 1.35$ (s, 3 H), 1.43 (s, 3 H), 3.52 (dd, J = 3.4, 7.8 Hz, 1 H), 3.79 (s, 3 H), 3.92 (dd, J = 0.7, 3.3 Hz, 1 H), 3.97 (dd, J = 2.7, 3.3 Hz, 1 H), 4.22 (m, 1 H), 4.36 (dd, J = 5.7, 7.8 Hz, 1 H), 4.52 (d, J = 12.4 Hz, 1 H), 4.56 (d, J = 12.1 Hz, 1 H), 4.65 (d, J = 12.1 Hz, 1 H), 4.72 (d, J = 12.4 Hz, 1 H), 4.81 (d, J = 3.4 Hz, 1 H), 6.83 (d, J = 8.7 Hz, 2 H), 7.23 (d, J = 8.7 Hz, 2 H), 7.28–7.40 (m, 5 H).

¹³C NMR (125 MHz, CDCl₃): δ = 26.3, 28.2, 55.3, 58.9, 69.2, 71.8, 73.5, 75.4, 76.4, 95.8, 108.8, 113.7 (2 C), 127.8, 127.9 (2 C), 128.4 (2 C), 129.4 (2 C), 130.4, 137.3, 159.2.

Anal. Calcd for $C_{23}H_{28}O_6$: C, 68.98; H, 7.05. Found C, 68.87; H, 7.18.

3,4-O-Isopropylidene-2-O-(4-methoxybenzyl)-D-arabinose (14) A mixture of **13** (5.00 g, 12.5 mmol) and Raney Ni (W-2, 5.00 g) in EtOH (50 mL) was stirred for 12 h under H₂ (1 atm) at r.t. The mixture was diluted with EtOAc (50 mL), and then the catalyst was removed by filtration of the mixture through a small pad of Celite. Concentration of the filtrate in vacuo afforded a residue, which was purified by column chromatography (hexane–EtOAc, 2:1 to 1:1); this gave a mixture of $\alpha\text{-}$ and $\beta\text{-}anomers$ 14 (a/\beta, 1:2) as a white semisolid.

Yield: 2.50 g (86%); $[\alpha]_D^{20}$ -47.0 (*c* 1.00, CHCl₃).

Recrystallization of the anomeric mixtures (hexane–CHCl₃, 3:1) gave pure β -anomer 14 as white needles.

Mp 115.5–116.5 °C; [α]_D²⁰–49.4 (*c* 1.00, CHCl₃).

IR (KBr): 778, 1035, 1066, 1177, 1244, 1377, 1510, 1612, 2932, 2959, 3013, 3323 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ (*α*-anomer) = 1.36 (s, 3 H), 1.48 (s, 3 H), 3.45 (d, J = 6.7 Hz, 1 H), 3.47 (m, 1 H), 3.80 (s, 3 H), 3.80 (ddd, J = 0.8, 3.4, 13.1 Hz, 1 H), 4.06 (dd, J = 2.1, 13.0 Hz, 1 H), 4.20-4.24 (m, 2 H), 4.69 (d, J = 11.4 Hz, 1 H), 4.71 (m, 1 H), 4.72 (d, J = 11.4 Hz, 1 H), 6.88 (d, J = 8.6 Hz, 2 H), 7.31 (dd, J = 8.6 Hz, 2 H).

¹³C NMR (125 MHz, CDCl₃): δ (α-anomer) = 25.8, 27.6, 55.3, 62.5, 72.70, 72.71, 77.2, 78.8, 95.5, 109.9, 113.8 (2 C), 129.7 (2 C), 130.0, 159.4.

¹H NMR (500 MHz, CDCl₃): δ (β-anomer) = 1.36 (s, 3 H), 1.46 (s, 3 H), 3.09 (d, J = 5.7 Hz, 1 H), 3.60 (dd, J = 3.4, 5.9 Hz, 1 H), 3.80 (s, 3 H), 3.83 (dd, J = 2.0, 13.0 Hz, 1 H), 4.12 (dd, J = 3.0, 13.0 Hz, 1 H), 4.23 (m, 1 H), 4.37 (t, J = 6.0 Hz, 1 H), 4.61 (d, J = 11.7 Hz, 1 H), 4.74 (d, J = 11.7 Hz, 1 H), 5.11 (dd, J = 3.4, 5.7 Hz, 1 H), 6.88 (d, J = 8.6 Hz, 2 H), 7.27 (dd, J = 8.6 Hz, 2 H).

¹³C NMR (125 MHz, CDCl₃): δ (β-anomer) = 25.9, 27.7, 55.3, 60.0, 72.4, 72.6, 74.4, 75.6, 90.9, 109.2, 113.9 (2 C), 129.71 (2 C), 129.73, 159.5.

HRMS–FAB: m/z [M + H]⁺ calcd for C₁₆H₂₃O₆: 311.1495; found: 311.1509.

(2R,3S,4R)-2,3-O-Isopropylidene-4-(4-methoxybenzyloxy)hex-5-ene-1,2,3-triol (15)

t-BuOK (3.10 g, 28 mmol) was added to a stirred suspension of Ph₃MeP⁺Br⁻ (10.4 g, 29 mmol) in anhyd benzene (80 mL), and the resulting mixture was heated at reflux for 3 h under argon. A soln of **14** (1.75 g, 5.6 mmol) in anhyd THF (40 mL) was added to the phosphorane soln, and the resulting mixture was heated at reflux for 1 h. After cooling, the reaction mixture was quenched with sat. aq NH₄Cl (40 mL) at 0 °C, and then extracted with Et₂O (2 × 60 mL). The organic layer was washed successively with sat. aq NaHCO₃ (2 × 40 mL) and brine (40 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane–EtOAc, 4:1).

Yield: 1.50 g (86%); colorless oil; $[\alpha]_D^{20}$ –19.4 (*c* 1.04, CHCl₃).

IR (neat): 756, 933, 1037, 1248, 1379, 1514, 1612, 2839, 2986, 3076, 3474 $\rm cm^{-1}.$

¹H NMR (500 MHz, CDCl₃): $\delta = 1.37$ (s, 3 H), 1.50 (s, 3 H), 2.60 (dd, J = 5.3, 7.9 Hz, 1 H), 3.66 (dt, J = 11.8, 5.6 Hz, 1 H), 3.72 (ddd, J = 4.9, 7.9, 11.8 Hz, 1 H), 3.80 (s, 3 H), 3.96 (dd, J = 5.3, 7.9 Hz, 1 H), 4.18 (dt, J = 5.1, 6.0 Hz, 1 H), 4.24 (dd, J = 5.4, 6.2 Hz, 1 H), 4.38 (d, J = 11.5 Hz, 1 H), 4.61 (d, J = 11.5 Hz, 1 H), 5.36 (d, J = 17.3 Hz, 1 H), 5.40 (d, J = 10.3 Hz, 1 H), 5.91 (ddd, J = 8.7 Hz, 2 H), 7.26 (d, J = 8.7 Hz, 2 H).

 ^{13}C NMR (125 MHz, CDCl₃): δ = 25.4, 27.2, 55.2, 61.2, 70.1, 77.7, 78.0, 78.8, 108.7, 113.8 (2 C), 120.3, 129.5, 129.6 (2 C), 134.8, 159.3.

HRMS–FAB: m/z [M + H]⁺ calcd for C₁₇H₂₅O₅: 309.1702; found: 309.1700.

(2*S*,3*S*,4*R*)-2,3-*O*-Isopropylidenedioxy-4-(4-methoxybenzyloxy)hex-5-enal (16)

DMSO (2.52 mL, 36 mmol) was added dropwise to a stirred soln of oxalyl chloride (2.33 mL, 27 mmol) in CH₂Cl₂ (80 mL) at -78 °C under argon. After 20 min, a soln of **15** (2.74 g, 8.9 mmol) in CH₂Cl₂ (40 mL) was added at -78 °C, and stirring was continued for 30 min at the same temperature. After addition of *i*-Pr₂NEt (12.0 mL, 69 mmol) at -78 °C, the mixture was gradually warmed to 0 °C over 1 h. The reaction was quenched with H₂O (20 mL) at 0 °C, and the mixture was extracted with Et₂O (2 × 80 mL). The combined extracts were washed successively with 3% aq HCl (2 × 50 mL), sat. aq NaHCO₃ (2 × 40 mL), and brine (50 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane–EtOAc, 5:1).

Yield: 2.59 g (95%); colorless oil; $[\alpha]_D^{20}$ –75.2 (*c* 1.00, CHCl₃).

IR (neat): 831, 1033, 1072, 1250, 1381, 1516, 1614, 1724, 2868, 2988, 3076 $\rm cm^{-1}.$

¹H NMR (500 MHz, CDCl₃): $\delta = 1.37$ (s, 3 H), 1.57 (s, 3 H), 3.78 (dd, J = 2.4, 8.2 Hz, 1 H), 3.80 (s, 3 H), 4.01 (d, J = 10.9 Hz, 1 H), 4.37 (dd, J = 2.0, 8.2 Hz, 1 H), 4.39 (d, J = 10.9 Hz, 1 H), 4.49 (dd, J = 2.4, 8.1 Hz, 1 H), 5.35 (d, J = 17.3 Hz, 1 H), 5.38 (d, J = 10.3 Hz, 1 H), 5.94 (ddd, J = 8.1, 10.3, 17.3 Hz, 1 H), 6.85 (d, J = 8.7 Hz, 2 H), 7.22 (d, J = 8.7 Hz, 2 H), 9.56 (d, J = 2.0 Hz, 1 H).

 ^{13}C NMR (125 MHz, CDCl₃): δ = 25.1, 26.5, 55.2, 69.5, 77.6, 80.5, 82.9, 111.2, 113.6 (2 C), 119.9, 129.2 (2 C), 129.9, 134.3, 159.1, 201.3.

HRMS–FAB: m/z [M + H]⁺ calcd for C₁₇H₂₃O₅: 307.1546; found: 307.1559.

Methyl (2*S*,3*S*,4*R*)-2,3-*O*-Isopropylidenedioxy-4-(4-methoxybenzyloxy)hex-5-enoate (5)

NaClO₂ (4.52 g, 50 mmol) was added in small portions to a stirred soln of **16** (3.04 g, 9.9 mmol) and NaH₂PO₄ (7.76 g, 50 mmol) in DMSO–H₂O (4:1, 100 mL) at r.t. After 1 h, the reaction was quenched with 15% aq Na₂S₂O₃ (30 mL) at 0 °C, and the mixture was extracted with EtOAc (3×70 mL). The organic layer was washed with brine (2×50 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded the corresponding carboxylic acid (2.80 g), which was directly, without further purification, used for the next reaction.

A 0.5 M soln of CH_2N_2 in Et_2O (24.0 mL, 12 mmol) was added dropwise to a stirred soln of the crude carboxylic acid (2.80 g) in MeOH (20 mL) at 0 °C, and stirring was continued for 30 min at r.t. The reaction was quenched with AcOH (2.0 mL) at 0 °C, and the resulting mixture was diluted with Et_2O (100 mL). The organic layer was washed with sat. aq NaHCO₃ (3 × 30 mL) and brine (30 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane–EtOAc, 3:1); this gave **5** as a colorless oil.

Yield: 2.61 g (78%, over 2 steps); $[\alpha]_D^{20}$ –34.2 (*c* 1.01, CHCl₃).

IR (neat): 823, 1076, 1248, 1381, 1514, 1614, 1730, 1761, 2839, 2988, 3076 $\rm cm^{-1}.$

¹H NMR (500 MHz, CDCl₃): $\delta = 1.39$ (s, 3 H), 1.60 (s, 3 H), 3.53 (s, 3 H), 3.79 (s, 3 H), 4.01 (dd, J = 4.0, 8.0 Hz, 1 H), 4.20 (d, J = 11.5 Hz, 1 H), 4.43 (dd, J = 4.0, 7.3 Hz, 1 H), 4.51 (d, J = 11.5 Hz, 1 H), 4.67 (d, J = 7.3 Hz, 1 H), 5.34 (d, J = 17.3 Hz, 1 H), 5.36 (d, J = 10.4 Hz, 1 H), 5.90 (ddd, J = 8.0, 10.4, 17.3 Hz, 1 H), 6.85 (d, J = 8.6 Hz, 2 H), 7.23 (d, J = 8.6 Hz, 2 H).

 ^{13}C NMR (125 MHz, CDCl₃): δ = 25.5, 26.6, 51.8, 55.2, 69.7, 75.3, 78.4, 80.4, 110.8, 113.5 (2 C), 119.7, 128.5 (2 C), 130.4, 134.8, 158.8, 170.2.

HRMS–FAB: m/z [M + H]⁺ calcd for C₁₈H₂₅O₆: 337.1651; found: 337.1668.

Methyl (2*S*,3*S*,4*R*)-2-{[(4*R*)-3-*tert*-Butoxycarbonyl-2,2-dimethyloxazolidin-4-yl]hydroxymethyl}-2,3-*O*-isopropylidenedioxy-4-(4-methoxybenzyloxy)hex-5-enoate (18)

A 1.0 M soln of NaHMDS in THF (2.96 mL, 3.0 mmol) was added dropwise to a stirred soln of **5** (796 mg, 2.4 mmol) in anhyd THF (25 mL) at 0 °C under argon. After 20 min, a soln of the Garner aldehyde (**6**; 598 mg, 2.6 mmol) in anhyd THF (10 mL) was added at -78 °C, and the resulting soln was stirred for 4 h at the same temperature. The reaction was quenched with sat. aq NH₄Cl (20 mL) at -78 °C, and the mixture was extracted with Et₂O (2 × 50 mL). The combined extracts were washed with sat. aq NaHCO₃ (2 × 30 mL) and brine (30 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane–EtOAc, 8:1 to 5:1); this gave **18** as a white solid; yield: 930 mg (69%). (Note that the presence of rotamers in the *tert*-butyl carbamate group resulted in extensive line broadening, and, in some instances, doubling of signals in the ¹H NMR and ¹³C NMR spectra.)

Colorless needles (recrystallization, hexane–CH₂Cl₂, 4:1); mp 86.5–88.0 °C; $[\alpha]_D^{20}$ –13.1 (*c* 1.08, CHCl₃).

IR (KBr): 772, 843, 1067, 1251, 1370, 1399, 1518, 1614, 1688, 1724, 2939, 2982, 3090, 3489 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 1.46 (br s, 3 H), 1.49 (br s, 3 H), 1.50 (br s, 9 H), 1.62 (s, 6 H), 2.52 (br s, 0.5 H), 2.93 (br s, 0.5 H), 3.46 (s, 3 H), 3.78 (s, 3 H), 3.85 (dd, *J* = 6.5, 9.6 Hz, 1 H), 3.90–4.10 (m, 2 H), 4.14 (d, *J* = 11.4 Hz, 1 H), 4.23–4.38 (m, 2 H), 4.50 (d, *J* = 11.4 Hz, 1 H), 4.59 (br s, 1 H), 5.31–5.40 (m, 2 H), 5.96 (ddd, *J* = 8.0, 10.3, 17.6 Hz, 1 H), 6.83 (d, *J* = 8.6 Hz, 2 H), 7.21 (d, *J* = 8.6 Hz, 2 H).

 ^{13}C NMR (125 MHz, CDCl₃): δ = 27.0 (4 C), 28.5 (3 C), 52.0, 55.2, 58.9, 63.7, 69.7, 71.8, 77.3, 80.5, 83.7, 86.2, 93.5, 111.7, 113.5 (2 C), 119.2, 128.4 (2 C), 130.6, 135.5, 152.3, 158.8, 171.3.

HRMS–FAB: m/z [M + H]⁺ calcd for C₂₉H₄₄NO₁₀: 566.2965; found: 566.2938.

Methyl (2*S*,3*S*,4*R*)-2-{[(4*S*)-3-*tert*-Butoxycarbonyl-2,2-dimethyloxazolidin-4-yl]methyl}-2,3-*O*-isopropylidenedioxy-4-(4-methoxybenzyloxy)hex-5-enoate (19)

A 1.0 M soln of NaHMDS in THF (4.00 mL, 4.0 mmol) was added dropwise to a stirred soln of **18** (924 mg, 1.6 mmol) in anhyd THF (35 mL) at 0 °C under argon. After 20 min, CS₂ (0.90 mL, 15 mmol) was added dropwise to the above soln at 0 °C, and stirring was continued for 20 min at the same temperature. MeI (1.00 mL, 16 mmol) was then added dropwise to the stirred soln at 0 °C, and then the resulting soln was allowed to warm to r.t. over 30 min. The reaction was quenched with sat. aq NH₄Cl (10 mL) at 0 °C, and the mixture was extracted with Et₂O (3 × 40 mL). The organic layer was washed with sat. aq NaHCO₃ (2 × 30 mL) and brine (30 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was immediately passed through silica gel (hexane–EtOAc, 2:1); this gave the corresponding methyl xanthate (1.35 g) as a colorless oil, which was directly used for the next reaction without further purification.

n-Bu₃SnH (1.20 mL, 4.5 mmol) and AIBN (66.0 mg, 0.40 mmol) were added successively to a soln of the crude methyl xanthate (1.35 g) in toluene (40 mL). For the deaeration of the reaction mixture, it was frozen in liquid N_2 , and the reaction vessel was evacuated in vacuo for 30 min before being filled with dry argon. The mixture was heated at reflux for 1 h under argon. After cooling, the reaction mixture was purified by column chromatography (hexane–EtOAc, 10:1 to 3:1); this gave **19** as a colorless oil. (Note that the presence of rota-

mers in the *tert*-butyl carbamate group resulted in extensive line broadening, and, in some instances, doubling of signals in the ¹H NMR and ¹³C NMR spectra.)

Yield: 475 mg [53%, or 92% based on recovery of starting material **18** (389 mg, 42%)]; $[\alpha]_{D}^{20}$ –10.4 (*c* 0.72, CHCl₃).

IR (neat): 770, 852, 1088, 1249, 1389, 1516, 1615, 1698, 1732, 2937, 2982, 3080 $\rm cm^{-1}.$

¹H NMR (500 MHz, CDCl₃): δ = 1.45 (s, 6 H), 1.47 (s, 9 H), 1.56 (br s, 3 H), 1.59 (s, 3 H), 1.82 (br s, 1 H), 2.45 (br s, 0.5 H), 2.65 (br s, 0.5 H), 3.51 (s, 3 H), 3.78 (s, 3 H), 3.79 (br, 1 H), 3.85 (br, 1 H), 4.03 (d, *J* = 2.8 Hz, 1 H), 4.08 (br, 1 H), 4.19 (d, *J* = 11.4 Hz, 1 H), 4.25 (br, 1 H), 4.48 (br, 1 H), 5.31–5.41 (m, 2 H), 5.95 (ddd, *J* = 7.8, 10.3, 17.7 Hz, 1 H), 6.83 (d, *J* = 8.6 Hz, 2 H), 7.22 (d, *J* = 8.6 Hz, 2 H).

 ^{13}C NMR (125 MHz, CDCl₃): δ = 27.0 (2 C), 27.7 (3 C), 28.5 (2 C), 41.4, 52.0, 54.8, 55.2, 67.1, 69.8, 78.0, 79.9, 84.0, 87.8, 93.1, 111.4, 113.5 (2 C), 119.4, 128.4 (2 C), 130.5, 134.9, 151.6, 158.8, 171.6.

HRMS–FAB: $m/z [M + H]^+$ calcd for C₂₉H₄₄NO₉: 550.3016; found: 550.3039.

(2*S*,3*S*,4*R*)-2-{[(4*S*)-3-*tert*-Butoxycarbonyl-2,2-dimethyloxazolidin-4-yl]methyl}-2,3-*O*-isopropylidenedioxy-4-(4-methoxybenzyloxy)hex-5-enal (20)

A 1.0 M soln of DIBAL-H in toluene (0.11 mL, 0.11 mmol) was added dropwise to a stirred soln of **19** (54.0 mg, 0.10 mmol) in anhyd CH₂Cl₂ (2 mL) at -100 °C under argon. After 1 h, the reaction was quenched with MeOH (1 mL) at -100 °C. A 15% aq soln of Rochelle salt (5 mL) was added to the mixture at 0 °C, and stirring was continued for 1 h at r.t. The resulting mixture was extracted with Et₂O (3 × 20 mL). The combined extracts were washed with brine (2 × 20 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane–EtOAc, 5:1); this gave **20** as a colorless oil. (Note that the presence of rotamers in the *tert*-butyl carbamate group resulted in extensive line broadening, and, in some instances, doubling of signals in the ¹H NMR and ¹³C NMR spectra.)

Yield: 45.0 mg (88%); $[\alpha]_D^{20}$ –30.5 (*c* 0.63, CHCl₃).

IR (neat): 770, 849, 1092, 1250, 1389, 1516, 1614, 1698, 1730, 2935, 2982, 3070 $\rm cm^{-1}.$

¹H NMR (500 MHz, CDCl₃): δ = 1.47 (s, 6 H), 1.49 (s, 9 H), 1.55 (s, 3 H), 1.57 (br s, 3 H), 1.75 (br, 1 H), 2.41 (br s, 0.5 H), 2.65 (br s, 0.5 H), 3.75 (br, 1 H), 3.79 (s, 3 H), 3.82–3.90 (br, 2 H), 4.02 (d, *J* = 2.9 Hz, 1 H), 4.05–4.20 (br, 2 H), 4.44 (br, 1 H), 5.33–5.43 (m, 2 H), 5.91 (ddd, *J* = 8.0, 10.3, 17.3 Hz, 1 H), 6.84 (d, *J* = 8.6 Hz, 2 H), 7.22 (d, *J* = 8.6 Hz, 2 H), 9.52 (br s, 1 H).

 ^{13}C NMR (125 MHz, CDCl₃): δ = 27.6 (4 C), 28.5 (3 C), 39.2, 54.3, 55.2, 67.7, 69.9, 77.2, 80.1, 87.3, 89.0, 93.3, 111.7, 113.7 (2 C), 119.9, 129.6 (3 C), 134.4, 151.7 159.2, 200.6.

HRMS–FAB: $m/z [M + H]^+$ calcd for $C_{28}H_{42}NO_8$: 520.2910; found: 520.2883.

$(3R,4R,5S,6R)-4-\{[(4S)-3-(tert-Butoxycarbonyl)-2,2-dimethyloxazolidin-4-yl]methyl\}-4,5-O-isopropylidene-6-(4-methoxybenzyloxy)octa-1,7-diene-3,4,5-triol (4)$

A 0.97 M soln of $H_2C=CH_2MgBr$ in THF (0.50 mL, 0.49 mmol) was added dropwise to a stirred soln of **20** (58.0 mg, 0.11 mmol) in anhyd THF (2.5 mL) at 0 °C under argon. After 1 h, the reaction was quenched with sat. aq NH₄Cl (5 mL) at 0 °C, and the mixture was extracted with Et₂O (3 × 20 mL). The combined extracts were washed with sat. aq NaHCO₃ (2 × 20 mL) and brine (20 mL), and dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane–EtOAc, 6:1); this gave **4** as a colorless oil. (Note that the presence

of rotamers in the *tert*-butyl carbamate group resulted in extensive line broadening, and, in some instances, doubling of signals in the ¹H NMR and ¹³C NMR spectra.)

Yield: 57 mg (93%); $[\alpha]_D^{20}$ +17.5 (*c* 1.21, CHCl₃).

IR (neat): 770, 855, 1107, 1248, 1393, 1514, 1615, 1670, 1700, 2935, 2982, 3090, 3423 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): $\delta = 1.37$ (s, 6 H), 1.40 (s, 3 H), 1.46 (s, 9 H), 1.51 (s, 3 H), 1.58 (m, 0.7 H), 1.73 (br, 0.3 H), 2.18 (br, 0.3 H), 2.29 (br d, 0.7 H, J = 14.5 Hz), 3.10 (br, 0.3 H), 3.73 (d, J = 6.0 Hz, 1 H), 3.79 (s, 3 H), 3.80–4.00 (m, 1.7 H), 4.00–4.20 (br, 1 H), 4.25–4.38 (br, 2 H), 4.44 (br, 1 H), 4.47 (br d, J = 11.4 Hz, 1 H), 4.60 (br d, J = 11.4 Hz, 1 H), 5.21 (dt, J = 10.8, 2.0 Hz, 1 H), 5.29–5.51 (m, 3 H), 5.98 (m, 0.7 H), 6.15 (m, 0.6 H), 6.38 (m, 0.7 H), 6.85 (d, J = 7.7 Hz, 2 H), 7.28 (d, J = 7.7 Hz, 2 H).

 ^{13}C NMR (125 MHz, CDCl₃): δ = 24.3, 26.6, 27.6, 28.0, 28.3 (3 C), 39.7, 53.8, 55.2, 68.7, 70.2, 72.0, 80.5, 84.1, 84.5, 87.1, 92.7, 107.9, 113.7 (2 C), 114.7, 119.2, 129.4 (2 C), 130.5, 135.6, 139.0, 152.4, 159.1.

HRMS–FAB: $m/z [M + H]^+$ calcd for C₃₀H₄₆NO₈: 548.3223; found: 548.3203.

(1*S*,2*R*,3*R*,6*R*)-2-{[(4*S*)-3-tert-Butoxycarbonyl-2,2-dimethyloxazolidin-4-yl]methyl}-1,2-*O*-isopropylidene-6-(4-methoxybenzyloxy)cyclohex-4-ene-1,2,3-triol (21)

[RuCl₂(=CHPh)(PCy₃)₂] (24.0 mg, 29 μ mol) was added to a soln of **4** (160 mg, 0.29 mmol) in anhyd CH₂Cl₂ (200 mL). For the deaeration of the reaction mixture, it was frozen in liquid N₂, and the reaction vessel was evacuated in vacuo for 30 min before being filled with dry argon. The mixture was then heated at reflux for 12 h. After cooling, the reaction mixture was stirred for 3 h at r.t. under air. Evaporation of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane–EtOAc, 8:1); this gave **21** as a colorless oil. (Note that the presence of rotamers in the *tert*-butyl carbamate group resulted in extensive line broadening, and, in some instances, doubling of signals in the ¹H NMR and ¹³C NMR spectra.)

Yield: 146 mg (96%); $[\alpha]_D^{20}$ -48.9 (*c* 1.20, CHCl₃).

IR (neat): 772, 850, 1078, 1252, 1391, 1514, 1612, 1696, 2936, 2982, 3040, 3524 $\rm cm^{-1}.$

¹H NMR (500 MHz, CDCl₃): δ = 1.33 (s, 3 H), 1.35 (s, 6 H), 1.44 (s, 3 H), 1.47 (s, 6 H), 1.60 (s, 3 H), 1.95 (dd, *J* = 11.3, 13.5 Hz, 1 H), 2.08 (dd, *J* = 10.6, 11.9 Hz, 1 H), 2.83 (d, *J* = 9.9 Hz, 1 H), 3.79 (s, 3 H), 3.91 (dd, *J* = 6.1, 8.3 Hz, 1 H), 4.02–4.30 (m, 4 H), 4.35 (m, 1 H), 4.43 (d, *J* = 11.3 Hz, 1 H), 4.52 (d, *J* = 11.3 Hz, 1 H), 5.96 (ddd, *J* = 1.8, 5.1, 9.9 Hz, 1 H), 6.04 (br d, *J* = 9.9 Hz, 1 H), 6.85 (d, *J* = 8.6 Hz, 2 H), 7.22 (d, *J* = 8.6 Hz, 2 H).

 ^{13}C NMR (125 MHz, CDCl₃): δ = 23.2, 26.9, 27.0, 27.3, 28.3 (3 C), 45.3, 54.0, 55.3, 67.8, 67.9, 70.6, 70.7, 79.6, 81.6, 82.4, 93.1, 109.4, 113.8 (2 C), 126.1, 129.5 (2 C), 130.3, 137.2, 151.5, 159.4.

HRMS–FAB: $m/z [M + H]^+$ calcd for $C_{28}H_{42}NO_8$: 520.2910; found: 520.2887.

(1*S*,2*S*,3*R*,6*R*)-1-{[(4*S*)-3-*tert*-Butoxycarbonyl-2,2-dimethyloxazolidin-4-yl]methyl}-6-(*tert*-butyldimethylsiloxy)-1,2-*O*-isopropylidene-3-(4-methoxybenzyloxy)cyclohex-4-ene-1,2-diol (22)

TBSOTf (0.20 μ L, 0.86 mmol) was added dropwise to a stirred soln of **21** (221 mg, 0.43 mmol) in anhyd CH₂Cl₂ (12 mL) containing 2,6-lutidine (0.14 μ L, 1.2 mmol) at 0 °C under argon. After 30 min at r.t., the reaction was quenched with H₂O (5 mL) at 0 °C, and the mixture was extracted with EtOAc (2 × 30 mL). The organic layer was washed with brine (2 × 20 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane–EtOAc, 10:1); this gave **22** as a colorless oil. (Note that the presence of rotamers in the *tert*butyl carbamate group resulted in extensive line broadening, and, in some instances, doubling of signals in the ¹H NMR and ¹³C NMR spectra.)

Yield: 250 mg (93%); $[\alpha]_D^{20}$ -42.2 (*c* 0.88, CHCl₃).

IR (neat): 775, 835, 1094, 1250, 1391, 1514, 1615, 1692, 2932, 2982, 3045 $\rm cm^{-1}.$

¹H NMR (500 MHz, DMSO- d_6): $\delta = 0.04$ (s, 6 H), 0.83 (s, 9 H), 1.36 (s, 3 H), 1.38 (s, 15 H), 1.45 (s, 3 H), 1.78–1.90 (br, 2 H), 3.74 (s, 3 H), 3.80–4.45 (m, 6 H), 4.49 (d, J = 11.4 Hz, 1 H), 4.54 (d, J = 11.4 Hz, 1 H), 5.93 (br, 1 H), 6.05 (ddd, J = 1.6, 4.7, 9.7 Hz, 1 H), 6.97 (d, J = 8.6 Hz, 2 H), 7.25 (d, J = 8.6 Hz, 2 H).

¹³C NMR (125 MHz, DMSO- d_6): δ = -4.4, -4.2, 17.7, 25.7 (6 C), 27.0 (2 C), 28.8 (2 C), 43.2, 53.7, 54.9, 66.9, 67.7, 69.8, 74.5, 78.9, 82.7, 83.9, 92.2, 109.0, 113.5 (2 C), 129.2 (2 C), 130.0, 130.6, 132.4, 150.7, 158.6.

HRMS–FAB: m/z [M + H]⁺ calcd for C₃₄H₅₆NO₈Si: 634.3775; found: 634. 3781.

(1*S*,2*S*,3*R*,6*R*)-1-[(2*S*)-2-(*tert*-Butoxycarbonylamino)-3-hydroxypropyl]-6-(*tert*-butyldimethylsiloxy)-1,2-*O*-isopropylidene-3-(4-methoxybenzyloxy)cyclohex-4-ene-1,2-diol (2)

A soln of **22** (212 mg, 0.34 mmol) and PPTS (50.0 mg, 0.20 mmol) in EtOH (8 mL) was heated at 60 °C for 3 h. After cooling, the reaction mixture was diluted with Et_2O (50 mL). The organic layer was washed with 1 M NaOH (15 mL) and brine (15 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane-EtOAc, 6:1); this gave **2** as a white solid; yield: 113 mg [57%, or 84% based on recovered **22** (68 mg, 32%)].

White needles (recrystallization, hexane–CHCl₃, 5:1); mp 112.5–113.5 °C; $[\alpha]_D^{20}$ –61.5 (*c* 1.03, CHCl₃).

IR (KBr): 777, 842, 1080, 1250, 1370, 1518, 1535, 1616, 1672, 2934, 2957, 3045, 3290, 3449, 3557 $\rm cm^{-1}.$

¹H NMR (500 MHz, CDCl₃): $\delta = 0.03$ (s, 3 H), 0.05 (s, 3 H), 0.84 (s, 9 H), 1.42 (s, 9 H), 1.44 (s, 3 H), 1.49 (s, 3 H), 1.68 (dd, J = 8.0, 14.8 Hz, 1 H), 1.87 (dd, J = 6.0, 14.8 Hz, 1 H), 2.87 (br, 1 H), 3.63 (m, 1 H), 3.72–3.80 (m, 2 H), 3.80 (s, 3 H), 3.96 (d, J = 4.3 Hz, 1 H), 4.23 (d, J = 5.6 Hz, 1 H), 4.48 (m, 1 H), 4.57 (d, J = 11.3 Hz, 1 H), 4.67 (d, J = 11.3 Hz, 1 H), 4.82 (br, 1 H), 5.92 (dd, J = 2.2, 9.7 Hz, 1 H), 6.08 (m, 1 H), 6.87 (d, J = 8.6 Hz, 2 H), 7.29 (d, J = 8.6 Hz, 2 H).

 13 C NMR (125 MHz, CDCl₃): δ = –4.6, –4.1, 18.1, 26.0 (3 C), 27.2, 27.3, 28.4 (3 C), 42.1, 49.3, 55.3, 65.6, 67.3, 71.2, 77.3, 79.6, 83.5, 85.7, 110.3, 113.8 (2 C), 129.6 (2 C), 130.3, 131.3, 132.8, 155.5, 159.2.

HRMS–FAB: m/z [M + H]⁺ calcd for C₃₁H₅₂NO₈Si: 594.3462; found: 594.3435.

(4S)-5-(Benzyloxy)-4-methylpent-2-yne (24)

CBr₄ (1.92 g, 5.8 mmol) was added to a stirred soln of (*R*)-**23**²⁰ (344 mg, 1.9 mmol) and PPh₃ (3.04 g, 12 mmol) in anhyd CH₂Cl₂ (40 mL) at 0 °C. After 10 min, the mixture was quenched with sat. aq NaHCO₃ (15 mL), and extracted with CH₂Cl₂ (2×50 mL). The combined extracts were washed with brine (2×30 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane–EtOAc, 1:0 to 10:1); this gave the corresponding dibromoalkene (645 mg) as a colorless oil. A 1.58 M soln of *n*-BuLi in hexane (2.46 mL, 3.9 mmol) was added dropwise to a stirred soln of the dibromoalkene (645 mg, 1.9 mmol) in anhyd THF (20 mL) at –78 °C under argon. After 1 h, the reaction mixture was allowed to warm to

FEATURE ARTICLE

0 °C, and MeI (0.14 mL, 2.3 mmol) was added. The resulting soln was further stirred for 1 h at 0 °C. The reaction was quenched with sat. aq NH₄Cl (10 mL) at 0 °C, and the mixture was extracted with Et₂O (2 × 40 mL). The combined extracts were washed with sat. aq NaHCO₃ (2 × 20 mL) and brine (20 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane–EtOAc, 50:1).

Yield: 342 mg (94%, 2 steps); colorless oil; $[\alpha]_D^{20}$ –3.0 (c 1.59, CHCl₃).

IR (neat): 610, 698, 735, 1028, 1097, 1204, 1360, 1454, 1497, 1605, 2859, 2971, 3030 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 1.17 (d, *J* = 6.9 Hz, 3 H), 1.80 (d, *J* = 2.4 Hz, 3 H), 2.70 (m, 1 H), 3.32 (dd, *J* = 7.5, 9.0 Hz, 1 H), 3.48 (dd, *J* = 6.1, 9.0 Hz, 1 H), 4.56 (dd, *J* = 12.2, 16.2 Hz, 2 H), 7.27–7.37 (m, 5 H).

¹³C NMR (125 MHz, CDCl₃): δ = 3.5, 18.1, 26.7, 72.9, 74.3, 76.4, 81.1, 127.5, 127.6 (2 C), 128.3 (2 C), 138.3.

(2E,4S)-5-(Benzyloxy)-2-iodo-4-methylpent-2-ene (9)

A soln of **24** (101 mg, 0.54 mmol) and Cp₂ZrHCl (348 mg, 1.4 mmol) in anhyd benzene (3 mL) was heated at 40 °C for 30 min. After cooling of the mixture, a soln of I₂ (274 mg, 1.1 mmol) in anhyd benzene (2 mL) was added at r.t. After 30 min, the reaction was quenched with sat. Na₂S₂O₃ (2 mL) at 0 °C, and the mixture was extracted with CH₂Cl₂ (3 × 20 mL). The combined extracts were washed successively with 3% aq HCl (15 mL), sat. aq NaHCO₃ (15 mL), and brine (15 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane–EtOAc, 100:1); this gave **9**, containing an inseparable regioisomer, as a pale yellow oil.

Yield: 118 mg (70%, 13% regioisomer); $[a]_D^{20}$ –12.3 (*c* 1.25, CHCl₃).

IR (neat): 610, 698, 735, 853, 1049, 1098, 1204, 1375, 1453, 1495, 1638, 2855, 2926, 2963, 3030 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 1.00 (d, *J* = 6.8 Hz, 3 H), 2.40 (s, 3 H), 2.77 (m, 1 H), 3.29 (dd, *J* = 3.4, 6.7 Hz, 2 H), 4.52 (s, 2 H), 6.00 (dd, *J* = 1.5, 9.5 Hz, 1 H), 7.25–7.37 (m, 5 H).

¹³C NMR (125 MHz, CDCl₃): δ = 17.0, 27.9, 36.1, 73.0, 74.2, 94.5, 127.5, 127.6, 127.6, 128.3, 138.4, 143.8.

HRMS–FAB: m/z [M + H]⁺ calcd for C₁₃H₁₇IO: 317.0402; found: 317.0422.

(2*S*,4*R*)-5-(*tert*-Butyldiphenylsiloxy)-2,4-dimethylpentan-1-ol (26)

An O₃ stream was bubbled through a soln of **25**²³ (369 mg, 1.0 mmol) in CH₂Cl₂–MeOH (3:1, 20 mL) at –78 °C until the color of the soln had changed to blue. The mixture was purged with argon, and then NaBH₄ (304 mg, 8.1 mmol) in EtOH–H₂O (1:1, 15 mL) was added at –78 °C. The mixture was allowed to warm to r.t. over 1 h. The reaction was quenched with 3% aq HCl (10 mL) at 0 °C, and the mixture was extracted with CH₂Cl₂ (2 × 40 mL). The combined extracts were washed with sat. aq NaHCO₃ (2 × 20 mL) and brine (20 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane–EtOAc, 1:0 to 20:1).

Yield: 315 mg (84%); colorless oil; $[\alpha]_{D}^{20}$ +1.74 (*c* 1.45, CHCl₃).

IR (neat): 505, 613, 702, 740, 1362, 1389, 1427, 1472, 1589, 1892, 1960, 2859, 2930, 2957, 3052, 3071, 3355 $\rm cm^{-1}.$

¹H NMR (500 MHz, $CDCl_3$): $\delta = 0.88$ (d, J = 6.7 Hz, 3 H), 0.93 (m, 1 H), 0.95 (d, J = 6.7 Hz, 3 H), 1.05 (s, 9 H), 1.26 (br, 1 H), 1.46 (m, 1 H), 1.64 (m, 1 H), 1.74 (m, 1 H), 3.35 (dd, J = 6.7, 10.5 Hz, 1 H), 3.41 (dd, J = 6.3, 9.9 Hz, 1 H), 3.46 (dd, J = 5.1, 10.6 Hz, 1 H), 3.52 (dd, J = 5.4, 9.9 Hz, 1 H), 7.35–7.45 (m, 6 H), 7.63–7.69 (m, 4 H).

Synthesis 2007, No. 4, 622–637 $\,$ © Thieme Stuttgart \cdot New York

¹³C NMR (125 MHz, CDCl₃): δ = 17.4, 17.9, 19.3, 26.8 (3 C), 33.12, 33.13, 37.1, 68.3, 68.7, 127.58 (2 C), 127.59 (2 C), 129.5 (2 C), 133.94, 133.95, 135.62 (2 C), 135.63 (2 C).

HRMS–FAB: $m/z [M + H]^+$ calcd for C₂₃H₃₅O₂Si: 371.2406; found: 371.2432.

(2*R*,4*S*)-1-(*tert*-Butyldiphenylsiloxy)-5-iodo-2,4-dimethylpentane (10)

MsCl (0.13 mL, 1.7 mmol) was added dropwise to a stirred soln of **26** (417 mg, 1.1 mmol) in anhyd CH_2Cl_2 (6 mL) containing Et_3N (0.31 mL, 2.2 mmol) at r.t. After 1 h, the reaction was quenched with sat. aq NaHCO₃ (5 mL) at 0 °C, and the mixture was extracted with CH_2Cl_2 (2 × 25 mL). The combined extracts were washed successively with 3% aq HCl (15 mL), sat. aq NaHCO₃ (15 mL), and brine (15 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded the corresponding mesylate (515 mg) as a colorless oil, which was used for the next reaction without further purification.

A mixture of the crude mesylate (515 mg) and NaI (1.01 g, 6.7 mmol) in acetone (10 mL) was heated at reflux for 6 h. After cooling, the mixture was diluted with H₂O (5 mL), and extracted with Et₂O (3×30 mL). The combined extracts were washed with sat. aq Na₂S₂O₃ (20 mL) and brine (20 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane–EtOAc, 20:1).

Yield: 513 mg (95%, 2 steps); colorless oil; $[\alpha]_{D}^{20}$ +7.3 (*c* 1.42, CHCl₃).

IR (neat): 503, 615, 702, 739, 824, 1111, 1194, 1262, 1389, 1427, 1460, 1589, 2859, 2930, 2959, 3071 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 0.93 (d, *J* = 6.3 Hz, 3 H), 0.94 (d, *J* = 6.7 Hz, 3 H), 1.00 (m, 1 H), 1.06 (s, 9 H), 1.47 (m, 2 H), 1.70 (m, 1 H), 3.07 (dd, *J* = 5.9, 9.6 Hz, 1 H), 3.23 (dd, *J* = 3.6, 9.5 Hz, 1 H), 3.43 (dd, *J* = 6.1, 9.9 Hz, 1 H), 3.49 (dd, *J* = 5.5, 9.9 Hz, 1 H), 7.35–7.45 (m, 6 H), 7.66 (dd, *J* = 0.9, 6.5 Hz, 4 H).

 ^{13}C NMR (125 MHz, CDCl₃): δ = 17.3, 18.0, 19.3, 21.4, 26.9 (3 C), 31.8, 33.1, 40.4, 68.8, 127.6 (4 C), 129.6 (2 C), 133.9 (2 C), 135.6 (4 C).

HRMS–FAB: $m/z [M + H]^+$ calcd for C₂₃H₃₄IOSi: 481.1424; found: 481.1407.

(2*S*,3*E*,6*S*,8*R*)-1-(Benzyloxy)-9-(*tert*-butyldiphenylsiloxy)-2,4,6,8-tetramethylnon-3-ene (28)

A 1.0 M soln of ZnCl₂ in Et₂O (0.58 mL, 0.58 mmol) was added to a stirred soln of 10 (279 mg, 0.58 mmol) in anhyd Et₂O (2 mL) at r.t. For the deaeration of the reaction mixture, it was frozen in liquid N₂, and the reaction vessel was evacuated in vacuo for 30 min before being filled with dry argon (freeze-vacuum deaeration technique). A 1.41 M soln of t-BuLi in pentane (1.24 mL, 1.8 mmol) was added to a stirred soln of the above mixture at -78 °C over 10 min under argon. The resulting soln was gradually warmed to -40 °C and stirred for 30 min at the same temperature. The mixture was allowed to warm to r.t., and stirring was continued for 1 h. A soln of 9 (160 mg, 0.51 mmol, including 13% of an inseparable 3-iodo regioisomer) in anhyd THF (2 mL) and Pd(PPh₃)₄ (58.0 mg, 51 µmol) in anhyd THF (2 mL) [degassed by freeze-vacuum deaeration technique for 30 min] were added successively to the above cloudy suspension of alkylzinc reagent, and the resulting mixture was stirred for 13 h at r.t. while shielded from light. The reaction was quenched with H₂O (10 mL) at 0 °C, and the mixture was extracted with Et₂O $(3 \times 20 \text{ mL})$. The organic layer was washed with sat. aq NaHCO₃ (20 mL) and brine (20 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane-CH₂Cl₂, 4:1).

Yield: 188 mg (81%); colorless oil; $[\alpha]_D^{20}$ +11.8 (*c* 1.00, CHCl₃).

IR (neat): 503, 569, 613, 700, 739, 1109, 1262, 1387, 1427, 1456, 1589, 1740, 2857, 2928, 2959, 3071 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): $\delta = 0.74$ (d, J = 6.1 Hz, 3 H), 0.92 (d, J = 6.7 Hz, 3 H), 0.97 (d, J = 6.7 Hz, 3 H), 1.05 (s, 9 H), 1.20–1.35 (m, 2 H), 1.56 (d, J = 1.2 Hz, 3 H), 1.58–1.65 (m, 2 H), 1.75 (m, 1 H), 2.00 (m, 1 H), 2.71 (m, 1 H), 3.22 (dd, J = 7.6, 9.1 Hz, 1 H), 3.32 (dd, J = 6.1, 9.1 Hz, 1 H), 3.40 (dd, J = 6.6, 9.8 Hz, 1 H), 3.50 (dd, J = 5.2, 9.8 Hz, 1 H), 4.52 (d, J = 12.1 Hz, 1 H), 4.47 (d, J = 12.1 Hz, 1 H), 4.90 (d, J = 9.0 Hz, 1 H), 7.26–7.45 (m, 11 H), 7.67 (dd, J = 1.4, 7.9 Hz, 4 H).

¹³C NMR (125 MHz, CDCl₃): δ = 16.2, 17.8, 18.0, 19.3, 19.8, 26.9 (3 C), 28.1, 33.0, 33.2, 41.4, 47.7, 69.0, 72.9, 75.5, 127.4, 127.5 (2 C), 127.6 (4 C), 128.3 (2 C), 128.9, 129.5 (2 C), 134.12, 134.13, 134.5, 135.62 (2 C), 135.64 (2 C), 138.8.

HRMS–FAB: $m/z [M + H]^+$ calcd for $C_{36}H_{51}O_2Si$: 543.3658; found: 543.3640.

(2*S*,3*E*,6*S*,8*R*)-9-(*tert*-Butyldiphenylsiloxy)-2,4,6,8-tetramethylnon-3-en-1-ol (29)

A 0.26 M soln of LiDBB in THF (4.0 mL, 1.1 mmol) [prepared from Li (8.7 mg, 1.3 mmol) and DTBB (346 mg, 1.3 mmol)] was added dropwise to a stirred soln of **28** (40.0 mg, 70 μ mol) in anhyd THF (1.5 mL) at -78 °C. After 5 min, the reaction was quenched with brine (5 mL) at 0 °C, and the resulting mixture was extracted with EtOAc (2 × 20 mL). The combined extracts were washed with brine (2 × 10 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane–EtOAc, 10:1).

Yield: 30.0 mg (90%); pale yellow oil; $[\alpha]_D^{20}$ –7.73 (*c* 1.00, CHCl₃).

IR (neat): 505, 615, 702, 741, 824, 939, 1032, 1111, 1188, 1260, 1387, 1427, 1460, 1589, 2859, 2928, 2957, 3362 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): $\delta = 0.76$ (d, J = 6.0 Hz, 3 H), 0.92 (d, J = 6.7 Hz, 3 H), 0.94 (d, J = 6.7 Hz, 3 H), 1.05 (s, 9 H), 1.32 (m, 2 H), 1.53 (s, 1 H), 1.58 (d, J = 1.2 Hz, 3 H), 1.60–1.68 (m, 2 H), 1.77 (m, 1 H), 2.04 (dd, J = 9.7, 17.3 Hz, 1 H), 2.60 (m, 1 H), 3.25–3.55 (m, 4 H), 4.85 (d, J = 9.4 Hz, 1 H), 7.35–7.45 (m, 6 H), 7.67 (dd, J = 1.4, 7.9 Hz, 4 H).

¹³C NMR (125 MHz, CDCl₃): δ = 16.4, 17.1, 17.8, 19.3, 19.8, 26.9 (3 C), 28.1, 33.2, 35.5, 41.5, 47.8, 68.0, 69.0, 127.6 (4 C), 128.5, 129.5 (2 C), 134.09, 134.10, 135.6 (4 C), 136.8.

HRMS–FAB: $m/z [M + H]^+$ calcd for $C_{29}H_{45}O_2Si$: 453.3189; found: 453.3189.

(3*R*,4*E*,7*S*,9*R*)-10-(*tert*-Butyldiphenylsiloxy)-3,5,7,9-tetramethyldec-4-ene (30)

TsCl (63.0 mg, 0.33 mmol) was added to a stirred soln of **29** (51.0 mg, 0.11 mmol) in anhyd CH₂Cl₂ (0.9 mL) containing Et₃N (0.14 mL, 0.99 mmol) and DMAP (1.3 mg, 11 µmol) at r.t. After 3 h, the reaction mixture was diluted with CH₂Cl₂ (30 mL), and the organic layer was washed successively with 3% aq HCl (2×10 mL), sat. aq NaHCO₃ (2×10 mL), and brine (10 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded the corresponding tosylate **8** (80 mg) as a pale yellow oil, which was used for the next reaction without further purification.

A 1.2 M soln of MeLi in Et₂O (5.50 mL, 6.6 mmol) was added dropwise to a stirred suspension of CuI (628 mg, 3.3 mmol) in anhyd Et₂O (3 mL) at -10 °C under argon. After 30 min, a soln of the crude tosylate **8** (80 mg) in anhyd Et₂O (5 mL) was added to the above mixture at -40 °C. The resulting mixture was allowed to warm to r.t., and stirring was continued for 13 h. The reaction was quenched with 25% NH₄OH (3 mL) and sat. aq NH₄Cl (3 mL) at 0 °C, and then the resulting mixture was stirred under air until the color of the soln had changed to bright blue. The mixture was extracted with Et₂O (2 × 20 mL), and the combined extracts were washed with brine $(2 \times 10 \text{ mL})$, and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane).

Yield: 49.2 mg (97%, 2 steps); pale yellow oil; $[\alpha]_{D}^{20}$ –5.22 (*c* 1.18, CHCl₃).

IR (neat): 503, 615, 702, 738, 822, 1111, 1262, 1385, 1427, 1460, 2859, 2928, 2959 $\rm cm^{-1}.$

¹H NMR (500 MHz, CDCl₃): $\delta = 0.75$ (d, J = 6.1 Hz, 3 H), 0.81 (t, J = 7.4 Hz, 3 H), 0.89 (d, J = 6.7 Hz, 3 H), 0.94 (d, J = 6.7 Hz, 3 H), 1.05 (s, 9 H), 1.13–1.23 (m, 2 H), 1.27–1.39 (m, 2 H), 1.52 (d, J = 1.3 Hz, 3 H), 1.56–1.65 (m, 2 H), 1.75 (m, 1 H), 1.97 (dd, J = 9.3, 16.8 Hz, 1 H), 2.22 (m, 1 H), 3.40 (dd, J = 6.6, 9.8 Hz, 1 H), 3.56 (dd, J = 5.1, 9.8 Hz, 1 H), 4.83 (d, J = 9.3 Hz, 1 H), 7.34–7.44 (m, 6 H), 7.66 (dd, J = 1.3, 7.7 Hz, 4 H).

 ^{13}C NMR (125 MHz, CDCl₃): δ = 12.0, 16.2, 17.9, 19.3, 20.0, 21.0, 26.9 (3 C), 28.2, 30.5, 33.2, 34.1, 41.3, 47.9, 69.0, 127.6 (4 C), 129.46, 129.47, 132.3, 132.9, 134.14, 134.16, 135.63 (2 C), 135.65 (2 C).

HRMS–FAB: $m/z [M + H]^+$ calcd for C₃₀H₄₇OSi: 451.3396; found: 451.3412.

(2R,4S,6E,8R)-2,4,6,8-Tetramethyldec-6-en-1-ol (31)

A 1.0 M soln of TBAF in THF (0.61 mL, 0.61 mmol) was added to a stirred soln of **30** (91.7 mg, 0.20 mmol) in THF (1.5 mL) at r.t. After 24 h, the reaction mixture was diluted with Et_2O (30 mL), and the organic layer was washed with sat. aq NaHCO₃ (10 mL) and brine (10 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane–EtOAc, 20:1).

Yield: 39.5 mg (91%); colorless oil; $[\alpha]_D^{20}$ –9.23 (*c* 1.20, CHCl₃).

IR (neat): 511, 608, 702, 818, 880, 1036, 1111, 1260, 1377, 1458, 1547, 1669, 1725, 2276, 2870, 2922, 2957, 3353 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 0.82 (d, *J* = 7.5 Hz, 3 H), 0.84 (t, *J* = 7.4 Hz, 3 H), 0.91 (d, *J* = 6.7 Hz, 3 H), 0.94 (d, *J* = 6.7 Hz, 3 H), 1.15–1.25 (m, 2 H), 1.25–1.38 (m, 3 H), 1.56 (d, *J* = 1.4 Hz, 3 H), 1.64–1.70 (m, 2 H), 1.75 (m, 1 H), 2.00 (dd, *J* = 9.4, 17.1 Hz, 1 H), 2.24 (m, 1 H), 3.37 (dd, *J* = 6.9, 10.4 Hz, 1 H), 3.53 (dd, *J* = 5.0, 10.5 Hz, 1 H), 4.86 (d, *J* = 9.2 Hz, 1 H).

¹³C NMR (125 MHz, CDCl₃): δ = 12.0, 16.2, 17.5, 20.2, 21.0, 28.2, 30.5, 33.2, 34.1, 40.9, 47.7, 68.3, 132.2, 133.1.

HRMS–FAB: m/z [M]⁺ calcd for C₁₄H₂₈O: 212.2140; found: 212.2131.

Methyl (2*E*,4*E*,6*E*,8*R*,10*S*,12*E*,14*R*)-8,10,12,14-Tetramethylhexadeca-2,4,6,12-tetraenoate (34)

A mixture of **31** (39.5 mg, 0.19 mmol), 3-Å MS (33 mg), and NMO (76.0 mg, 0.65 mmol) in CH_2Cl_2 (2.8 mL) was stirred for 10 min at r.t. *n*-Pr₄NRuO₄ (13.0 mg, 37 µmol) was added to the above mixture at r.t. After 10 min, the mixture was filtered through a short pad of silica gel, and the filtrate was concentrated in vacuo (water bath < 25°C); this gave aldehyde **32** (26.1 mg) as a colorless oil, which was used immediately for the next reaction.

A soln of 33^{29} (76.0 mg, 0.32 mmol) in anhyd THF (0.7 mL) was added dropwise to a stirred soln of LDA in anhyd THF [prepared from *i*-Pr₂NH (50.0 µL, 0.32 mmol) and 1.58 M *n*-BuLi in *n*-hexane (0.20 mL, 0.32 mmol) in THF (0.34 mL)] at -78 °C. After 30 min, a soln of the crude aldehyde **32** (26.1 mg) in anhyd THF (0.7 mL) was added slowly to the above soln, and the resulting mixture was stirred for 30 min at -78 °C and for 30 min at -30 °C. The reaction was quenched with sat. aq NH₄Cl (5 mL) at -30 °C, and the resulting mixture was extracted with Et₂O (2 × 30 mL). The combined extracts were washed with brine (2 × 15 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue,

FEATURE ARTICLE

which was purified by column chromatography (hexane–EtOAc, 100:1); this gave a mixture of **34** and its 6'Z-isomer (49.2 mg, 82%; ratio 73:27, by 500-MHz ¹H NMR). A pure sample of **34** was isolated from this mixture by HPLC [DAICEL CHIRALPAK AD-H, i.d. 10×250 mm; hexane–*i*-PrOH, 200:1, 1.0 mL·min⁻¹; UV detection: 254 nm absorbance; $t_{\rm R}$ (**34**): 12.5 min, $t_{\rm R}$ (6'Z-isomer): 10.8 min].

Yield: 27.2 mg (46%, 2 steps); colorless oil; $[a]_D^{20}$ –1.57 (*c* 1.45, CHCl₃).

IR (neat): 505, 613, 702, 740, 1362, 1389, 1427, 1472, 1589, 1892, 1960, 2859, 2930, 2957, 3052, 3071, 3355 $\rm cm^{-1}.$

¹H NMR (500 MHz, CDCl₃): $\delta = 0.80$ (d, J = 6.5 Hz, 3 H), 0.84 (d, J = 7.4 Hz, 3 H), 0.90 (d, J = 6.7 Hz, 3 H), 1.00 (d, J = 6.7 Hz, 3 H), 1.02 (m, 1 H), 1.19 (m, 1 H), 1.27–1.39 (m, 2 H), 1.52 (d, J = 1.2 Hz, 3 H), 1.56 (m, 1 H), 1.76 (dd, J = 7.3, 13.2 Hz, 1 H), 1.88 (dd, J = 6.8, 13.1 Hz, 1 H), 2.23 (m, 1 H), 2.34 (m, 1 H), 3.74 (s, 3 H), 4.85 (d, J = 9.4 Hz, 1 H), 5.73 (dd, J = 8.5, 15.2 Hz, 1 H), 5.84 (d, J = 11.3, 14.9 Hz, 1 H), 6.52 (dd, J = 10.7, 14.9 Hz, 1 H), 7.31 (dd, J = 11.3, 15.2 Hz, 1 H).

 ^{13}C NMR (125 MHz, CDCl₃): δ = 12.1, 16.2, 19.5, 21.1, 21.4, 28.4, 30.6, 34.1, 35.0, 44.0, 48.3, 51.5, 119.5, 127.8, 128.2, 132.1, 133.1, 141.5, 145.1, 146.5, 167.7.

HRMS–FAB: m/z [M + H]⁺ calcd for C₂₁H₃₄O₂: 319.2637; found: 319.2623.

(2*E*,4*E*,6*E*,8*R*,10*S*,12*E*,14*R*)-8,10,12,14-Tetramethylhexadeca-2,4,6,12-tetraenoyl Chloride (3)

A soln of **34** (26.2 mg, 82 µmol) in 2 M aq KOH–THF–MeOH (3:3:1, 2 mL) was stirred for 12 h at r.t. The reaction mixture was acidified with 1 M HCl (2 mL) at 0 °C and extracted with Et₂O (3×15 mL). The combined organic extracts were dried (MgSO₄). Concentration of the soln in vacuo afforded carboxylic acid **35** (26.0 mg), which was used for the following reaction without further purification.

Oxalyl chloride (21.0 μ L, 0.24 mmol) was added to a stirred soln of the crude carboxylic acid **35** (26.0 mg) in CH₂Cl₂ (2 mL) containing DMF (6.0 μ L, 82 μ mol) at r.t. After 1 h, the solvent was removed in vacuo; this afforded acid chloride **3** (26.0 mg) as a colorless oil, which was used immediately, without further purification, for the next reaction.

(2*E*,4*E*,6*E*,8*R*,10*S*,12*E*,14*R*)-*N*-{(1*S*)-2-[(3*aS*,4*R*,7*R*,7*aS*)-4-(*tert*-Butyldimethylsiloxy)-7-hydroxy-2,2-dimethyl-7,7a-dihydro-1,3-benzodioxol-3a(4*H*)-yl]-1-(hydroxymethyl)ethyl}-8,10,12,14-tetramethylhexadeca-2,4,6,12-tetraenamide (37)

TMSOTf (0.26 mL, 1.4 mmol) was added dropwise to a stirred soln of **2** (32.0 mg, 54 μ mol) in anhyd CH₂Cl₂ (2 mL) containing 2,6-lutidine (0.28 mL, 2.4 mmol) at 0 °C. The soln was stirred for 3 h at r.t. MeOH (2 mL) was added, and stirring was continued for 1 h at r.t. The mixture was diluted with H₂O (5 mL), and extracted with Et₂O (5 × 30 mL). The combined extracts were washed with sat. aq NaHCO₃ (30 mL) and brine (30 mL), and then dried (Na₂SO₄). Concentration of the soln in vacuo afforded amino alcohol **36** (80 mg, containing a small amount of 2,6-lutidine), which was used for the next reaction without further purification.

A soln of crude acid chloride **3** (26.0 mg, ca. 80 μ mol) in anhyd CH₂Cl₂ (2 mL) was added dropwise to a stirred soln of the crude amino alcohol **36** (80.0 mg, containing a small amount of 2,6-lutidine) in anhyd CH₂Cl₂ (2 mL) containing Et₃N (80 μ L, 0.58 mmol) at r.t. After 1 h, 60% aq AcOH (1.0 mL) was added, and the soln was stirred for 6 h at r.t. The mixture was diluted with H₂O (15 mL) and then extracted with Et₂O (2 × 50 mL). The combined extracts were

washed with sat. aq NaHCO₃ (30 mL) and brine (30 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue of **37**, which was purified by column chromatography (hexane–EtOAc, 2:1 to 1:1).

Yield: 26.0 mg (73%); colorless oil; $[\alpha]_D^{20}$ –33.7 (*c* 0.97, CHCl₃).

IR (neat): 777, 837, 1062, 1251, 1375, 1460, 1543, 1609, 1649, 2928, 2957, 3040, 3285 $\rm cm^{-1}.$

¹H NMR (500 MHz, CDCl₃): $\delta = 0.07$ (s, 6 H), 0.80 (d, J = 6.5 Hz, 3 H), 0.84 (t, J = 7.4 Hz, 3 H), 0.88 (s, 9 H), 0.90 (d, J = 6.7 Hz, 3 H), 0.99 (d, J = 6.6 Hz, 3 H), 1.03 (m, 1 H), 1.20 (dt, J = 13.4, 7.7 Hz, 1 H), 1.27–1.36 (m, 2 H), 1.44 (s, 3 H), 1.48 (s, 3 H), 1.52 (s, 3 H), 1.56 (m, 1 H), 1.76 (dd, J = 7.4, 15.0 Hz, 1 H), 1.81–1.91 (m, 2 H), 1.93 (dd, J = 6.3, 15.0 Hz, 1 H), 2.23 (m, 1 H), 2.33 (m, 1 H), 2.41 (br, 1 H), 3.06 (br, 1 H), 3.68 (br d, J = 9.0 Hz, 1 H), 3.85 (dd, J = 4.0, 11.4 Hz, 1 H), 3.90 (d, J = 4.4 Hz, 1 H), 4.17 (m, 1 H), 4.31 (d, J = 5.2 Hz, 1 H), 4.72 (m, 1 H), 4.84 (br d, J = 9.3 Hz, 1 H), 5.70 (dd, J = 8.4, 15.2 Hz, 1 H), 5.77 (d, J = 14.9 Hz, 1 H), 5.87–5.94 (m, 2 H), 6.05–6.13 (m, 2 H), 6.18 (dd, J = 11.3, 14.8 Hz, 1 H), 6.49 (dd, J = 10.7. 14.8 Hz, 1 H), 7.23 (dd, J = 11.3, 14.8 Hz, 1 H).

 13 C NMR (125 MHz, CDCl₃): δ = -4.5, -4.1, 12.1, 16.2, 18.2, 19.5, 21.1, 21.4, 26.0 (3 C), 27.5 (2 C), 28.3, 30.5, 34.1, 35.0, 41.9, 44.0, 48.3, 48.6, 65.5, 67.4, 70.7, 83.9, 86.9, 110.7, 122.1, 127.7, 128.3, 131.9, 132.1, 133.1, 133.7, 140.6, 141.8, 145.7, 166.1.

HRMS–FAB: m/z [M + H]⁺ calcd for $C_{38}H_{66}NO_6Si$: 660.4659; found: 660.4642.

$(2E,4E,6E,8R,10S,12E,14R)\text{-}N-\{(1S)\text{-}1-(Acetoxymethyl)\text{-}2-\\[(3aS,4R,7R,7aS)\text{-}4-(tert\text{-}butyldimethylsiloxy)\text{-}7-hydroxy\text{-}2,2-dimethyl\text{-}7,7a\text{-}dihydro\text{-}1,3-benzodioxol\text{-}3a(4H)\text{-}yl]ethyl]\text{-}8,10,12,14\text{-}tetramethylhexadeca\text{-}2,4,6,12\text{-}tetraenamide (38)}$

A soln of Ac₂O (5.4 μ L, 52 μ mol) in CH₂Cl₂ (0.2 mL) was added to a stirred soln of **37** (34.0 mg, 52 μ mol) in CH₂Cl₂ (1.6 mL) containing py (20 μ L, 0.24 mmol) and DMAP (0.4 mg, 3.2 μ mol) at r.t. After 20 min, the mixture was diluted with H₂O (5 mL), and extracted with Et₂O (3 × 20 mL). The combined extracts were washed with sat. aq NaHCO₃ (2 × 15 mL) and brine (15 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane–EtOAc, 5:1).

Yield: 26.0 mg [72%, or 81% based on recovered **37** (4.0 mg, 12%)]; colorless oil; $[\alpha]_D^{20}$ –26.5 (*c* 0.96, CHCl₃).

IR (neat): 777, 837, 1061, 1250, 1371, 1460, 1539, 1610, 1649, 1744, 2928, 2957, 3055, 3281 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): $\delta = 0.09$ (s, 6 H), 0.80 (d, J = 6.5 Hz, 3 H), 0.84 (t, J = 7.4 Hz, 3 H), 0.88 (s, 9 H), 0.90 (d, J = 6.7 Hz, 3 H), 0.99 (d, J = 6.6 Hz, 3 H), 1.02 (m, 1 H), 1.19 (dt, J = 13.4, 7.8 Hz, 1 H), 1.27–1.35 (m, 2 H), 1.38 (s, 3 H), 1.44 (s, 3 H), 1.52 (d, J = 1.2 Hz, 3 H), 1.56 (m, 1 H), 1.73–1.81 (m, 2 H), 1.85–1.91 (m, 2 H), 2.07 (s, 3 H), 2.23 (m, 1 H), 2.33 (m, 1 H), 2.41 (m, 1 H), 3.87 (d, J = 4.6 Hz, 1 H), 4.21 (d, J = 3.8, 11.3 Hz, 1 H), 4.33 (dd, J = 6.0, 11.3 Hz, 1 H), 4.41–4.49 (m, 2 H), 4.68 (br s, 1 H), 4.84 (d, J = 9.4 Hz, 1 H), 5.65–5.80 (m, 2 H), 5.75 (d, J = 14.9 Hz, 1 H), 5.87 (dd, J = 2.7, 9.6 Hz, 1 H), 6.04–6.12 (m, 2 H), 6.18 (dd, J = 11.3, 14.8 Hz, 1 H), 6.49 (dd, J = 10.7, 14.8 Hz, 1 H), 7.23 (dd, J = 11.3, 14.8 Hz, 1 H).

¹³C NMR (125 MHz, CDCl₃): δ = -4.5, -4.1, 12.1, 16.2, 18.2, 19.5, 20.9, 21.1, 21.4, 26.0 (3 C), 27.5, 27.6, 28.3, 30.5, 34.1, 35.0, 42.6, 44.0, 46.1, 48.3, 66.6, 67.3, 70.7, 84.0, 87.1, 110.6, 122.0, 127.7, 128.3, 132.1, 132.6, 133.1, 133.1, 140.7, 141.9, 145.8, 165.6, 171.4.

HRMS–FAB: m/z [M + H]⁺ calcd for C₄₀H₆₈NO₇Si: 702.4765; found: 702.4748.

(2E,4E,6E,8R,10S,12E,14R)-N-{(1S)-1-(Acetoxymethyl)-2-[(3aS,4R,7R,7aS)-4-(tert-butyldimethylsiloxy)-7-mesyloxy-2,2dimethyl-7,7a-dihydro-1,3-benzodioxol-3a(4H)-yl]ethyl}-8,10,12,14-tetramethylhexadeca-2,4,6,12-tetraenamide (39)

MsCl (8.0 µL, 0.1 mmol) was added to a stirred soln of 38 (31.0 mg, 38 µmol) in CH₂Cl₂ (2 mL) containing Et₃N (82.0 µL, 0.60 mmol) at r.t. After 1 h, the reaction was quenched with H₂O (2 mL), and the mixture was extracted with Et₂O (3×20 mL). The combined extracts were washed with 1 M HCl (2×10 mL), sat. aq NaHCO₃ $(2 \times 10 \text{ mL})$, and brine (10 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane-EtOAc, 6:1).

Yield: 32.0 mg (93%); colorless oil; $[\alpha]_D^{20}$ –31.2 (*c* 1.07, CHCl₃).

IR (neat): 777, 841, 1078, 1177, 1250, 1364, 1460, 1539, 1611, 1649, 1744, 2928, 2957, 3045, 3271 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): $\delta = 0.09$ (s, 3 H), 0.10 (s, 3 H), 0.80 (d, J = 6.5 Hz, 3 H), 0.84 (t, J = 7.4 Hz, 3 H), 0.89 (s, 9 H), 0.90 (d, *J* = 6.7 Hz, 3 H), 1.00 (d, *J* = 6.6 Hz, 3 H), 1.02 (m, 1 H), 1.19 (dt, J = 13.4, 7.8 Hz, 1 H), 1.27–1.35 (m, 2 H), 1.38 (s, 3 H), 1.47 (s, 3 H), 1.52 (d, J = 1.0 Hz, 3 H), 1.56 (m, 1 H), 1.73–1.81 (m, 2 H), 1.81-1.91 (m, 2 H), 2.08 (s, 3 H), 2.23 (m, 1 H), 2.33 (m, 1 H), 3.10 (s, 3 H), 3.99 (d, *J* = 4.9 Hz, 1 H), 4.22 (dd, *J* = 3.7, 11.3 Hz, 1 H), 4.34 (dd, J = 5.8, 11.3 Hz, 1 H), 4.43 (m, 1 H), 4.50 (d, J = 5.4 Hz, 1 H), 4.84 (d, J = 9.3 Hz, 1 H), 5.52 (m, 1 H), 5.65–5.75 (m, 2 H), 5.75 (d, J = 15.1 Hz, 1 H), 5.93 (dd, J = 2.4, 9.7 Hz, 1 H), 6.08 (dd, *J* = 10.7, 15.1 Hz, 1 H), 6.17 (dd, *J* = 11.3, 14.8 Hz, 1 H), 6.25 (m, 1 H), 6.49 (dd, J = 10.7. 14.8 Hz, 1 H), 7.21 (dd, J = 11.3, 14.8 Hz, 1 H).

¹³C NMR (125 MHz, CDCl₃): $\delta = -4.6, -4.2, 12.1, 16.2, 18.2, 19.5,$ 20.9, 21.1, 21.4, 26.0 (3 C), 27.4, 27.6, 28.3, 30.6, 34.1, 35.0, 38.4, 42.5, 44.0, 46.1, 48.3, 66.4, 66.8, 80.8, 83.8, 84.1, 111.5, 121.9, 127.6, 128.3, 129.2, 132.1, 133.1, 134.7, 140.7, 141.9, 145.8, 165.5, 171.3.

HRMS-FAB: m/z [M + H]⁺ calcd for C₄₁H₇₀NO₉SSi: 780.4541; found: 780.4541.

[(2E,4E,6E,8R,10S,12E,14R)-N-{(1S)-1-(Acetoxymethyl)-2-[(3aS,7R,7aS)-7-mesyloxy-2,2-dimethyl-4-oxo-7,7a-dihydro-1,3-benzodioxol-3a(4H)-yl]ethyl}-8,10,12,14-tetramethylhexadeca-2,4,6,12-tetraenamide (40)

A 1 M soln of TBAF in THF (30.0 µL, 30 µmol) was added to a stirred soln of 39 (24.0 mg, 30 µmol) in THF (1.5 mL) at r.t. After 30 min, additional 1 M TBAF in THF (20.0 µL, 20 µmol) was added. After 20 min, the mixture was diluted with Et₂O (30 mL), and the organic layer was washed with 1 M NaOH (8 mL) and brine (8 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded the corresponding alcohol (22.0 mg) as a colorless oil, which was used for the next reaction without further purification.

DMP (48.0 mg, 0.11 mmol) was added to a stirred soln of the crude alcohol (22.0 mg) in CH₂Cl₂ (2 mL) at r.t. After 20 min, the reaction was quenched with 15% aq Na2S2O3 (2 mL) at 0 °C, and the mixture was extracted with Et_2O (3 × 20 mL). The combined extracts were washed with 1 M NaOH (10 mL) and brine (10 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane-EtOAc, 6:1 to 2:1).

Yield: 20.0 mg (98%, 2 steps); colorless oil; $[\alpha]_{D}^{20}$ –21.8 (*c* 0.98, CHCl₃).

IR (neat): 794, 858, 1071, 1177, 1237, 1368, 1457, 1534, 1611, 1653, 1696, 1741, 2926, 2959, 3378 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): $\delta = 0.80$ (d, J = 6.5 Hz, 3 H), 0.84 (t, J = 7.4 Hz, 3 H), 0.90 (d, J = 6.7 Hz, 3 H), 1.00 (d, J = 6.7 Hz, 3 H), 1.02 (m, 1 H), 1.19 (dt, J = 13.4, 7.8 Hz, 1 H), 1.26 (s, 3 H), 1.27-1.35 (m, 2 H), 1.33 (s, 3 H), 1.52 (d, J = 1.0 Hz, 3 H), 1.56 (m, 1 H),

1.76 (dd, J = 7.4, 13.1 Hz, 1 H), 1.85–1.94 (m, 2 H), 2.06 (s, 3 H), 2.20 (dd, J = 9.2, 15.6 Hz, 1 H), 2.23 (m, 1 H), 2.33 (m, 1 H), 3.22 (s, 3 H), 4.10 (dd, J = 4.8, 11.3 Hz, 1 H), 4.23 (dd, J = 5.1, 11.3 Hz, 1 H), 4.42–4.50 (m, 2 H), 4.84 (d, J = 9.5 Hz, 1 H), 5.52 (dd, J = 1.7, 3.8 Hz, 1 H), 5.71 (dd, J = 8.4, 15.2 Hz, 1 H), 5.75 (d, J = 15.0 Hz, 1 H), 5.84 (d, J = 7.6 Hz, 1 H), 6.08 (dd, J = 10.8, 15.0 Hz, 1 H), 6.18 (dd, J = 11.3, 14.9 Hz, 1 H), 6.26 (d, J = 10.1 Hz, 1 H), 6.48 (dd, *J* = 10.7, 14.8 Hz, 1 H), 6.83 (ddd, *J* = 1.9, 4.8, 10.1 Hz, 1 H), 7.21 (dd, J = 11.3, 14.9 Hz, 1 H).

¹³C NMR (125 MHz, CDCl₃): δ = 12.1, 16.2, 19.5, 20.8, 21.1, 21.4, 26.7, 27.1, 28.4, 29.7, 30.5, 34.1, 35.0, 39.0, 44.0, 45.4, 48.3, 66.1, 70.6, 77.7, 81.1, 110.0, 122.0, 127.6, 128.2, 130.5, 132.1, 133.1, 138.9, 140.8, 141.9, 145.9, 165.7, 171.0, 197.2.

HRMS-FAB: m/z [M + H]⁺ calcd for C₃₅H₅₄NO₉S: 664.3519; found: 664.3544.

(2E,4E,6E,8R,10S,12E,14R)-N-[(1S)-2-Acetoxy-1-{[(1S,2S,3S)-2,3-epoxy-1-hydroxy-6-oxocyclohex-4-enyl]methyl]ethyl]-8,10,12,14-tetramethylhexadeca-2,4,6,12-tetraenamide (41)

A soln of 40 (6.6 mg, 6.9 µmol) and Cl₃CCO₂H (66.0 mg, 0.42 mmol) in CH₂Cl₂ (0.36 mL) containing H₂O (18 µL) was heated at reflux for 3 h. The mixture was basified with 2 M NaOH (0.22 mL) at 0 °C, and the resulting mixture was stirred for 10 min at r.t. The mixture was diluted with H₂O (8 mL), and extracted with Et₂O $(3 \times 30 \text{ mL})$. The combined extracts were washed with brine $(2 \times 20 \text{ mL})$ mL) and dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (hexane-EtOAc, 2:1 to 1:3).

Yield: 2.4 mg [45%, or 82% based on recovered 40 (3.0 mg, 45%)]; colorless oil; $[\alpha]_{D}^{20}$ +46.9 (*c* 0.12, CHCl₃).

IR (neat): 736, 837, 1006, 1238, 1373, 1456, 1539, 1609, 1651, 1682, 1732, 2924, 2959, 3353 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): $\delta = 0.80$ (d, J = 6.5 Hz, 3 H), 0.84 (t, J = 7.4 Hz, 3 H), 0.90 (d, J = 6.7 Hz, 3 H), 0.99 (d, J = 6.7 Hz, 3 H), 1.02 (m, 1 H), 1.19 (m, 1 H), 1.27–1.35 (m, 2 H), 1.52 (d, J = 1.3 Hz, 3 H), 1.76 (dd, J = 7.3, 13.0 Hz, 1 H), 1.87 (dd, J = 7.1, 13.0 Hz, 1 H), 1.96–2.10 (m, 3 H), 2.07 (s, 3 H), 2.23 (m, 1 H), 2.33 (m, 1 H), 3.58 (dt, J = 1.6, 3.9 Hz, 1 H), 3.72 (d, J = 3.8 Hz, 1 H), 4.02 (m, 1)H), 4.14 (m, 1 H), 4.22–4.32 (m, 2 H), 4.83 (d, J = 9.4 Hz, 1 H), 5.71 (dd, J = 8.4, 15.4 Hz, 1 H), 5.73 (d, J = 15.0 Hz, 1 H), 5.86 (d, J = 7.1 Hz, 1 H), 6.08 (dd, J = 10.8, 15.1 Hz, 1 H), 6.17 (dd, J = 11.3, 15.0 Hz, 1 H), 6.23 (dd, J = 1.6, 9.9 Hz, 1 H), 6.49 (dd, *J* = 10.6. 14.8 Hz, 1 H), 7.13 (dd, *J* = 3.9, 9.9 Hz, 1 H), 7.23 (dd, J = 11.4, 14.9 Hz, 1 H).

¹³C NMR (125 MHz, CDCl₃): δ = 12.1, 16.2, 19.5, 20.8, 21.1, 21.4,28.3, 30.5, 34.1, 35.0, 37.6, 44.0, 45.6, 48.0, 48.3, 55.8, 65.9, 76.6, 121.7, 127.1, 128.3, 130.1, 132.1, 133.1, 140.9, 142.1, 144.7, 145.9, 166.1, 171.1, 197.6.

HRMS-FAB: *m*/*z* [M + H]⁺ calcd for C₃₁H₄₆NO₆: 528.3325; found: 528.3333.

(2E,4E,6E,8R,10S,12E,14R)-N-[(1S)-1-{[(1S,2S,3S)-2,3-Epoxy-1-hydroxy-6-oxocyclohex-4-enyl]methyl}-2-hydroxyethyl]-8,10,12,14-tetramethylhexadeca-2,4,6,12-tetraenamide [(+)scyphostatin; 1]

A mixture of 41 (3.2 mg, 6.0 µmol) and lipase PS (16 mg) in acetone $(0.16\,mL)$ containing phosphate buffer (pH 7) $(0.16\,mL)$ was stirred for 10 h at r.t. The reaction mixture was diluted with Et₂O (50 mL), and the resulting mixture was filtered through a small pad of Celite. The filtrate was washed with sat. aq NaHCO₃ (10 mL) and brine (10 mL), and then dried (MgSO₄). Concentration of the soln in vacuo afforded a residue, which was purified by column chromatography (acetone-EtOAc, 1:4). The ¹H NMR and ¹³C NMR, IR, and HRMS-FAB spectra (see below) are compatible with those of natural (+)-scyphostatin (1).

Yield: 1.8 mg (60%); colorless amorphous powder; $[a]_{D}^{25}$ +61.0 (*c* 0.08, MeOH) [lit.¹ $[a]_{D}^{25}$ +66.4 (*c* 0.09, MeOH)].

IR (neat): 802, 1039, 1261, 1377, 1460, 1543, 1609, 1651, 1703, 1738, 2924, 2957, 3346 cm⁻¹.

¹H NMR (500 MHz, CD₃OD) δ = 0.83 (d, *J* = 6.5 Hz, 3 H), 0.86 (t, *J* = 7.4 Hz, 3 H), 0.91 (d, *J* = 6.9 Hz, 3 H), 1.00 (d, *J* = 6.7 Hz, 3 H), 1.03 (m, 1 H), 1.19 (m, 1 H), 1.33–1.38 (m, 2 H), 1.59 (d, *J* = 1.3 Hz, 3 H), 1.59 (m, 1 H), 1.79 (m, 1 H), 1.85–1.91 (m, 2 H), 2.08 (dd, *J* = 3.4, 14.7 Hz, 1 H), 2.27 (m, 1 H), 2.35 (m, 1 H), 3.46 (dd, *J* = 5.7, 10.9 Hz, 1 H), 3.52 (dd, *J* = 5.1, 10.9 Hz, 1 H), 3.59 (dt, *J* = 1.6, 3.9 Hz, 1 H), 3.67 (d, *J* = 3.9 Hz, 1 H), 4.06 (m, 1 H), 4.85 (m, 1 H), 5.70 (dd, *J* = 8.7, 15.1 Hz, 1 H), 5.90 (d, *J* = 15.0 Hz, 1 H), 6.07 (dd, *J* = 1.6, 9.9 Hz, 1 H), 6.15 (dd, *J* = 10.7, 15.1 Hz, 1 H), 6.25 (dd, *J* = 11.1, 14.9 Hz, 1 H), 6.53 (dd, *J* = 10.6, 14.7 Hz, 1 H), 7.12–7.17 (m, 2 H).

 ^{13}C NMR (125 MHz, CD₃OD) δ = 13.0, 16.9, 20.4, 22.0, 22.4, 30.0, 32.2, 35.9, 36.8, 40.3, 45.7, 48.5, 49.8, 50.1, 58.7, 66.1, 78.0, 124.2, 129.9, 130.4, 132.5, 134.0, 134.7, 141.9, 142.9, 146.3, 146.7, 169.1, 200.1.

HRMS–FAB: $m/z [M + H]^+$ calcd for C₂₉H₄₄NO₅: 486.3220; found: 486.3194.

Acknowledgment

We greatly thank Dr. H. Kogen, Dr. T. Ogita, and Dr. T. Akiyama (Sankyo Co., Ltd.) for fruitful discussions and the generous gift of natural (+)-scyphostatin (1). We also thank Dr. N. Sugimoto and Dr. N. Kawahara (National Institute of Health Sciences) for HRMS– FAB measurements and assistance with NMR spectroscopy experiments. This work was supported in part by Grants-in-Aid for Scientific Research on Priority Areas (17035073) and for High Technology Research Program from the Ministry of Education, Culture, Sports, Science, and Technology of Japan.

References

- Tanaka, M.; Nara, F.; Suzuki-Konagai, K.; Hosoya, T.; Ogita, T. J. Am. Chem. Soc. 1997, 119, 7871.
- (2) (a) Nara, F.; Tanaka, M.; Hosoya, T.; Suzuki-Konagai, K.; Ogita, T. J. Antibiot. 1999, 52, 525. (b) Nara, F.; Tanaka, M.; Masuda-Inoue, S.; Yamasato, Y.; Doi-Yoshioka, H.; Suzuki-Konagai, K.; Kumakura, S.; Ogita, T. J. Antibiot. 1999, 52, 531.
- (3) (a) Uchida, R.; Tomoda, H.; Arai, M.; Omura, S. J. Antibiot.
 2001, 54, 882. (b) Tanaka, M.; Nara, F.; Yamasato, Y.; Ono, Y.; Ogita, T. J. Antibiot. 1999, 52, 827. (c) Tanaka, M.; Nara, F.; Yamasato, Y.; Masuda-Inoue, S.; Doi-Yoshioka, H.; Kumakura, S.; Enokita, R.; Ogita, T. J. Antibiot. 1999, 52, 670. (d) Uchida, R.; Tomoda, H.; Dong, Y.; Omura, S. J. Antibiot. 1999, 52, 572.
- (4) (a) Claus, R. A.; Wuestholz, A.; Mueller, S.; Bockmeyer, C. L.; Riedel, N. H.; Kinscherf, R.; Deigner, H.-P. *ChemBioChem* 2005, *6*, 726. (b) Taguchi, M.; Goda, K.; Sugimoto, K.; Akama, T.; Yamamoto, K.; Suzuki, T.; Tomishima, Y.; Nishiguchi, M.; Arai, K.; Takahashi, K.; Kobori, T. *Bioorg. Med. Chem. Lett.* 2003, *13*, 3681. (c) Taguchi, M.; Sugimoto, K.; Goda, K.; Akama, T.; Yamamoto, K.; Suzuki, T.; Tomishima, Y.; Nishiguchi, M.; Arai, K.; Takahashi, K.; Kobori, T. *Bioorg. Med. Chem. Lett.* 2003, *13*, 1963. (d) Yokomatsu, T.; Murano, T.; Akiyama, T.; Koizumi, J.; Shibuya, S.; Tsuji, Y.; Soeda, S.; Shimeno, H. *Bioorg. Med. Chem. Lett.* 2003, *13*, 229. (e) Pitsinos, E. N.; Wascholowski, V.; Karaliota, S.; Rigou, C.;

Couladouros, E. A.; Giannis, A. ChemBioChem 2003, 4, 1223. (f) Lindsey, C. C.; Gómez-Díza, C.; Villalba, J. M.; Pettus, T. R. R. Tetrahedron 2002, 58, 4559. (g) Hakogi, T.; Monden, Y.; Taichi, M.; Iwama, S.; Fujii, S.; Ikeda, K.; Katsumura, S. J. Org. Chem. 2002, 67, 4839.
(h) Yokomatsu, T.; Takechi, H.; Akiyama, T.; Shibuya, S.; Kominato, T.; Soeda, S.; Shimeno, H. Bioorg. Med. Chem. Lett. 2001, 11, 1277. (i) Arenz, C.; Gartner, M.; Wascholowski, V.; Giannis, A. Bioorg. Med. Chem. 2001, 9, 2901. (j) Arenz, C.; Thutewohl, M.; Block, O.; Altenbach, H.-J.; Waldmann, H.; Giannis, A. ChemBioChem 2001, 2, 141. (k) Arenz, C.; Giannis, A. Eur. J. Org. Chem. 2001, 137. (l) Hakogi, T.; Monden, Y.; Iwama, S.; Katsumura, S. Org. Lett. 2000, 2, 2627. (m) Arenz, C.; Giannis, A. Angew. Chem. Int. Ed. 2000, 39, 1440.

- (5) For reviews, see: (a) Wascholowski, V.; Giannis, A. Drug News Perspect. 2001, 14, 581. (b) Hannun, Y. A.; Luberto, C.; Argraves, K. M. Biochemistry 2001, 40, 4893.
 (c) Kolter, T.; Sandhoff, K. Angew. Chem. Int. Ed. 1999, 38, 1532. (d) Hannun, Y. A. In Sphingolipid-Mediated Signal Transduction; Hannun, Y. A., Ed.; Springer: New York, 1997, 1.
- (6) Chatterjee, S. Arterioscler. Thromb. Vasc. Biol. **1998**, 18, 1523.
- (7) Amtmann, E.; Zoeller, M. *Biochem. Pharmacol.* **2005**, *69*, 1141.
- (8) (a) Lepine, S.; Lakatos, B.; Courageot, M.-P.; Le Stunff, H.; Sulpice, J.-C.; Giraud, F. J. Immunol. 2004, 173, 3783.
 (b) Numakawa, T.; Nakayama, H.; Suzuki, S.; Kubo, T.; Nara, F.; Numakawa, Y.; Yokomaku, D.; Araki, T.; Ishimoto, T.; Ogura, A.; Taguchi, T. J. Biol. Chem. 2003, 278, 41259. (c) Shin, H.-M.; Han, T.-H. Mol. Immunol. 1999, 36, 197.
- (9) Saito, S.; Tanaka, N.; Fujimoto, K.; Kogen, H. Org. Lett. 2000, 2, 505.
- (10) Hoye, T. R.; Tennakoon, M. A. Org. Lett. 2000, 2, 1481.
- (11) (a) Pitsinos, E. N.; Cruz, A. Org. Lett. 2005, 7, 2245. (b) Kenworthy, M. N.; Taylor, R. J. K. Org. Biomol. Chem. 2005, 3, 603. (c) Takagi, R.; Tojo, K.; Iwata, M.; Ohkata, K. Org. Biomol. Chem. 2005, 3, 2031. (d) Kenworthy, M. N.; McAllister, G. D.; Taylor, R. J. K. Tetrahedron Lett. 2004, 45, 6661. (e) Tan, Z.; Negishi, E.-i. Angew. Chem. Int. Ed. 2004, 43, 2911. (f) McAllister, G. D.; Taylor, R. J. K. Tetrahedron Lett. 2004, 45, 2551. (g) Takagi, R.; Tsuyumine, S.; Nishitani, H.; Miyanaga, W.; Ohkata, K. Aust. J. Chem. 2004, 57, 439. (h) Miyanaga, W.; Takagi, R.; Ohkata, K. Heterocycles 2004, 64, 75. (i) Murray, L. M.; O'Brien, P.; Taylor, R. J. K. Org. Lett. 2003, 5, 1943. (j) Eipert, M.; Maichle-Mössmer, C. M.; Maier, M. E. Tetrahedron 2003, 59, 7949. (k) Fujioka, H.; Kotoku, N.; Sawama, Y.; Nagatomi, Y.; Kita, Y. Tetrahedron Lett. 2002, 43, 4825. (l) Takagi, R.; Miyanaga, W.; Tamura, K.; Ohkata, K. Chem. Commun. 2002, 2096. (m) Runcie, K. A.; Taylor, R. J. K. Org. Lett. 2001, 3, 3237. (n) Gurjar, M. K.; Hotha, S. Heterocycles 2000, 53, 1885.
- (12) (a) Katoh, T.; Izuhara, T.; Yokota, W.; Inoue, M.; Watanabe, K.; Nobeyama, A.; Suzuki, T. *Tetrahedron* **2006**, *62*, 1590.
 (b) Izuhara, T.; Katoh, T. *Org. Lett.* **2001**, *3*, 1653.
 (c) Izuhara, T.; Katoh, T. *Tetrahedron Lett.* **2000**, *41*, 7651.
- (13) Izuhara, T.; Yokota, W.; Inoue, M.; Katoh, T. *Heterocycles* 2002, 56, 553.
- (14) Inoue, M.; Yokota, W.; Murugesh, M. G.; Izuhara, T.; Katoh, T. Angew. Chem. Int. Ed. 2004, 43, 4207.
- (15) (a) Dondoni, A.; Perrone, D. Synthesis 1997, 527.
 (b) Dondoni, A.; Perrone, D. Org. Synth. 1997, 77, 64.
 (c) Garner, P.; Park, J. M. J. Org. Chem. 1987, 52, 2361.
- (16) Ballou, C. E. J. Am. Chem. Soc. 1957, 79, 165.

- (17) Barton, D. H. R.; McCombie, S. W. J. Chem. Soc., Perkin Trans. 1 1975, 1574.
- (18) For recent reviews, see: (a) Nicolaou, K. C.; Bulger, P. G.; Sarlah, D. *Angew. Chem. Int. Ed.* **2005**, *44*, 4490.
 (b) Trnka, T. M.; Grubbs, R. H. *Acc. Chem. Res.* **2001**, *34*, 18.
- (19) Schwab, P.; France, M. B.; Ziller, J. W.; Grubbs, R. H. *Angew. Chem., Int. Ed. Engl.* **1995**, *34*, 2039.
- (20) Walsh, T. F.; Toupence, R. B.; Ujjainwalla, F.; Young, J. R.; Goulet, M. T. *Tetrahedron* **2001**, *57*, 5233.
- (21) Corey, E. J.; Fuchs, P. L. Tetrahedron Lett. 1972, 3769.
- (22) Hart, D. W.; Blackburn, T. F.; Schwartz, J. J. Am. Chem. Soc. **1975**, 97, 679.
- (23) Roush, W. R.; Palkowitz, A. D.; Ando, K. J. Am. Chem. Soc. 1990, 112, 6348.
- (24) Negishi, E.; Valente, L. F.; Kobayashi, M. J. Am. Chem. Soc. 1980, 102, 3298.

- (25) For a recent review, see: Handbook of Organopalladium Chemistry for Organic Synthesis, Vol. 1; Negishi, E., Ed.; Wiley-Interscience: New York, 2002, 597.
- (26) Smith, A. B. III; Beauchamp, T. J.; LaMarche, M. J.; Kaufman, M. D.; Qiu, Y.; Arimoto, H.; Jones, D. R.; Kobayashi, K. J. Am. Chem. Soc. **2000**, *122*, 8654.
- (27) Shimshock, S. J.; Waltermire, R. E.; DeShong, P. J. Am. Chem. Soc. 1991, 113, 8791.
- (28) Griffith, W. P.; Ley, S. V.; Whitcombe, G. P.; White, A. D. J. Chem. Soc., Chem. Commun. 1987, 1625.
- (29) Kinoshita, M.; Takami, H.; Taniguchi, M.; Tamai, T. Bull. Chem. Soc. Jpn. 1987, 60, 2151.
- (30) Sakaitani, M.; Ohfune, Y. J. Org. Chem. 1990, 55, 870.
- (31) (a) Ireland, R. E.; Liu, L. J. Org. Chem. 1993, 58, 2899.
 (b) Dess, D. B.; Martin, J. C. J. Am. Chem. Soc. 1991, 113, 7277. (c) Dess, D. B.; Martin, J. C. J. Org. Chem. 1983, 48, 4155.