

Bioorganic & Medicinal Chemistry Letters 13 (2003) 669-673

BIOORGANIC & MEDICINAL CHEMISTRY LETTERS

Synthesis and Anti-Influenza Evaluation of Orally Active Bicyclic Ether Derivatives Related to Zanamivir

Takeshi Masuda,^a Satoshi Shibuya,^a Masami Arai,^a Shuku Yoshida,^b Takanori Tomozawa,^b Akiko Ohno,^b Makoto Yamashita^b and Takeshi Honda^{a,*}

^aMedicinal Chemistry Research Laboratories, Sankyo Co., Ltd., 1-2-58 Hiromachi, Shinagawa-ku, Tokyo 140-8710, Japan ^bBiological Research Laboratories, Sankyo Co., Ltd., 1-2-58 Hiromachi, Shinagawa-ku, Tokyo 140-8710, Japan

Received 30 September 2002; accepted 18 November 2002

Abstract—We synthesized bicyclic ether sialidase inhibitors such as tetrahydro-furan-2-yl, tetrahydro-pyran-2-yl, and oxepan-2-yl derivatives related to zanamivir. These compounds substituted by diol at the C-3' and C-4' positions resulted in the retention of low nanomolar inhibitory activities against not only influenza A virus sialidase but also influenza A virus in cell culture. Compound **11a** in particular showed comparable efficacy in vivo relative to that of oseltamivir phosphate. © 2003 Elsevier Science Ltd. All rights reserved.

Sialidase is one of two glycoproteins expressed on the influenza virus surface and catalyzes the cleavage of sialic acid residues from glycoproteins, glycolipids and oligosaccharides. It has been shown that sialidase is required for release of newly produced virions from infected cells and that it facilitates the movement of the virus through the mucus of the respiratory tract.¹ The catalytic activity of sialidase is essential for influenza virus replication and infectivity. Accordingly, inhibitors of this enzyme are of interest as potential anti-influenza agents. Indeed, the effectiveness of sialidase inhibitors as anti-influenza agents has been demonstrated both in animal models and in human clinical trials by several research groups² and highlighted by recent approval for human use of zanamivir³ and oseltamivir phosphate⁴ (Fig. 1). Zanamivir is delivered by inhalation because of its low oral bioavailability whereas oseltamivir phosphate is administered orally. As part of our study of the structure activity relationships of 4-guanidino-7-substituted Neu5Ac2en derivatives, we reported that the appropriate alkyl ether derivatives at the C-7 position of 4-guanidino-Neu5Ac2en had more potent activity against influenza virus replication compared to zanamivir and that the multivalent sialidase inhibitor 3 (Fig. 1)

2 and the polyvalent sialidase inhibitor 3 were less effective than oseltamivir phosphate (data not shown) when delivered systemically to the mouse. There have been no reports of orally active derivatives related to zanamivir. Therefore, in order to develop an orally bioavailable sialidase inhibitor, we designed a newly synthesized sialidase inhibitor possessing cyclic ether moieties such as tetrahydro-furan-2-yl, tetrahydropyran-2-yl, and oxepan-2-yl groups in place of the glycerol side chain of zanamivir. We postulated that these compounds would possess modified physicochemical properties which could make them more suitable for systemic delivery.

carrying 4-guanidino-Neu5Ac2en analogues via a linker

of alkyl ether at the C-7 position was much more effective

than zanamivir in the mouse/influenza virus infection

model by intranasal administration.⁵ However, compound





⁰⁹⁶⁰⁻⁸⁹⁴X/03/\$ - see front matter \odot 2003 Elsevier Science Ltd. All rights reserved. doi:10.1016/S0960-894X(02)01039-9

^{*}Corresponding author. Tel.: +81-3-3492-3131; fax: +81-3-5436-8563; e-mail: thonda@shina.sankyo.co.jp

Herein we report the synthesis of tetrahydro-furan-2-yl, tetrahydro-pyran-2-yl and oxepan-2-yl derivatives related to zanamivir and their biological activities.

Chemistry

The synthesis of the tetrahydro-furan-2-yl, tetrahydropyran-2-yl, and oxepan-2-yl derivatives substituted by diol at the C-3' and C-4' positions is illustrated in Scheme 1. Compound 4^{5b} was alkylated with toluene-4sulfonic acid 2-(2,2-diethyl-[1,3]dioxolan-4-yl)-ethyl ester,⁶ allyl iodide, trifluoro-methanesulfonic acid 2,2difluoro-but-3-enyl ester,⁷ or 5-iodo-pent-1-ene in the presence of NaH in DMF to give the corresponding compound **5a**, **5b**, **5c**, or **5d**, in moderate yield. The



Scheme 1. Reagents and conditions: (a) alkylation reagents (2.0 equiv), NaH (2.0 equiv), DMF (50–70%); (b) (i) TBAF, THF (70–80%), (ii) Ac₂O, pyridine (90–100%), (iii) AcOH–H₂O (4:1) (70–80%), (iv) thiophosgene (1.2–2.4 equiv), dimethylaminopyridine (2.4–5.0 equiv), CH₂Cl₂ (80–90%); (c) P(OCH₃)₃, 120 °C (60–90%); (d) benzylidene-bis (tricyclo-hexylphosphine) dichlororuthenium (0.01 equiv), CH₂Cl₂ (50–100%); (e) OSO₄ (0.01 equiv), N-methylmorphorine N-oxide (1.2 equiv), acetone–H₂O (50–90%); (f) (i) Ac₂O, AcOH, cat. H₂SO₄ (10:10:1, v/v) (80–90%), (ii) NaN₃, Dowex 50W (H⁺), *t*-BuOH (70–80%), (iii) Lindlar cat., H₂. EtOH (70–80%), (iv) 1H-pyrazol-1-[*N*,*N'*-bis(*tert*-butoxycarbonyl)] carboxamidine, THF (95%); (g) (i) CF₃COOH–CH₂Cl₂ (ii) aq NaOH, (90–100%).

removal of the ketals with 80% acetic acid after protection of the 4-OH group by the acetyl group, followed by the formation of the thiocarbonates with thiophosgene and dimethylaminopyridine provided compounds 6a-6d, which were then reduced to give the corresponding di-terminal olefines 7a-7d. Subsequently, ring-closing metathesis (RCM) reactions with Grubbs' catalyst could be successively accomplished to afford the corresponding cyclic compounds 8a-8d. This is the first example of application of the RCM reaction for sialic acid modifications. Oxidation of compounds 8a-8d with OsO_4 and NMO provided the diols **9a–9d**, respectively, as single diastereomers. The (3'R, 4'R)-configurations of the diols of compounds 9a and 9b were determined by X-ray crystallographic analysis⁸ of compounds 11a and 11b, respectively.

The high diasteroselectivity of osmylation could be explained in terms of the most preferable conformation (A) of compound 8a based on molecular orbital calculation (3-21G* basis set) as shown in Figure 2. Thus, the oxygen of the tetrahydropyran moiety is orientated away from the oxygen of the dihydropyran moiety to avoid electrostatic repulsion and one side of the cyclic olefin is covered by the acetamide group. Therefore, osmylation occurred from the sterically more accessible face, the π -face opposite to that of the preexisting acetamide group. Compounds 9a-9d were converted to compounds 11a-11d under the same conditions as that described previously.⁵ The synthesis of the tetrahydrofuran-2-yl, tetrahydro-pyran-2-yl, and oxepan-2-yl derivatives substituted by diol at the C-4' and C-5' positions is illustrated in Scheme 2. Removal of the acetonide of compound 12, followed by treatment of compound 13 with sodium periodate gave the aldehyde 14. Subsequently, alkylation of compound 14 was performed with allyltrimethylsilane in the presence of TiCl₄ to provide the diastereomixture of the homoallyl alcohols (15a/ 15b = 2:1) which could be separated using a silica gel column. The (S)-configuration of the C-7 alcohol of compound 15b was determined by X-ray crystallographic analysis of 19c. Alkylation of the alcohols of compounds 15a and 15b with allyl trichloroacetimidate and TfOH, followed by ring closing metathesis reaction



Figure 2. The preferred conformation (A) of compound 8a calculated using Macromodel.

of compounds 16a and 16b provided compounds 17a and 17b, respectively, in good yield. Osmium oxidation of 17a and 17b each furnished mixtures of two *cis* diols (4'S, 5'R:4'R, 5'S=2:1), which were separated using a silica gel column regardless of the C-2' stereochemistry. Compounds 18a, 18b, 18c, and 18d were thus converted to the target compounds 19a, 19b, 19c, and 19d, respectively. The absolute configurations of the diols of compounds 19b and 19c were determined by their X-ray crystal-



Scheme 2. Reagents and conditions: (a) AcOH–H₂O (4:1) (70%); (b) NaIO₄, acetone–H₂O (90%); (c) allyltrimethylsilane (2.0 equiv), TiCl₄ (1.5 equiv) (77% as a mixture of *R* and *S* diastereomers); (d) allyl 2,2,2-trichloroacetimidate (10 equiv), TfOH (1.5 equiv), CH₂Cl₂–cyclohexane (20–25%); (e) benzylidene-bis(tricyclohexylphosphine)dichlororuthenium (0.01 equiv), CH₂Cl₂ (80%); (f) OsO₄ (0.01 equiv), *N* methylmorphorine *N*-oxide (1.2 equiv), acetone–H₂O (70–80% as a mixture of 4'S, 5' *R*-diol and 4'*R*, 5' S-diol); (g) (i) Ac₂O, AcOH, cat. H₂SO₄ (10:10:1, v/v) (80–90%), (ii) NaN₃, Dowex 50W (H⁺), *t*-BuOH (70–80%), (iii) Lindlar cat., H₂, EtOH (70–80%), (iv) 1*H*-pyrazol-1-[*N*,*N*'-bis(*tert*-butoxycarbonyl)] carboxamidine, THF (95%), (v) CF₃COOH–CH₂Cl₂, (vi) aq NaOH, (90–100%).



Scheme 3. Reagents and conditions: (a) Pd/C, H₂, EtOH (84%); (b) same conditions as those described in Scheme 1f and g.

lographic analysis.⁸ The olefin of compound **8a** was reduced with Pd/C under H_2 atmosphere to afford the tetrahydrofuran derivatives **20**, which yielded compound **21** as shown in Scheme 3.

Biological Activities

The influenza A virus sialidase inhibitory and plaque reduction activities⁹ of a range of the bicyclic sialidase inhibitors are summarized in Table 1. Tetrahydro-furan-2-yl, tetrahydro-pyran-2-yl, and oxepan-2-yl derivatives substituted by diol at the C-3' and C-4' positions, **11a**, **11b**, **11c**, and **11d**, exhibited comparable inhibitory

Table 1. Sialidase inhibitory and plaque reduction activities of bicyclic sialidase inhibitors related to zanamivir IC_{50} (ng/mL)^b



120					
Compd	R	Sialidase inhibitory assay	Plaque reduction assay		
		A/PR/8/34	A/Yamagata/32/89		
Zanamivir		0.6–12.0 (1.0) ^a	0.9–6.6 (1.0) ^a		
11a	HOW	10.3 (0.94)	0.6 (0.46)		
11b	но он	6.1 (1.22)	1.4 (0.82)		
11c		10.7 (0.88)	1.4 (0.53)		
11d	НО ОН	12.4 (1.12)	2.2 (1.47)		
21		240 (4.00)	>100		
19a	HO//, HO ^W	2.45 (22.2)	>100		
19b	HO	998 (90.7)	> 100		
19c	HO _{//} HO ^{//}	370 (30.8)	> 100		
19d	HO	3600 (300)	> 100		

^aSince IC_{50} values varied depending on the experiments, the relative potencies of the compounds to zanamivir are shown in the parentheses based on the IC_{50} values of zanamivir as a reference. IC_{50} values of zanamivir in enzyme inhibition and plaque reduction were 0.6-12.0 ng/mL and 0.9-6.6 ng/mL, respectively. ^bND, not determined.

activities against influenza A virus sialidase to that of zanamivir. Compound 11a showed slightly increased activity (2-fold) in a plaque reduction assay relative to zanamivir. However, removal of the diol at the C-3' and C-4' positions resulted in reduced inhibitory activity (21). Movement of the diol to the C-4' and C-5' positions from the C-3' and C-4' positions, respectively, showed a significant loss of enzyme inhibitory activity (11a vs 19a). Compounds 19a and 19c possessing the diol of the 4'S and 5'R configurations were relatively more potent than compounds 19b and 19d possessing the diol of the 4'R and 5'S configurations. This result suggests the position and the stereochemistry of the diol of the tetrahydro-pyran side chain must play an important role in binding affinity with a virus sialidase.

As shown in the model of the structure of compound 11a bound with influenza virus sialidase (Fig. 3), the carboxylate was held strongly by two arginine residues (Arg 371 and Arg 118). The guanidino group forms strong charge-charge type hydrogen bond interaction with Glu 119 and Asp 151. The diol interacts with the carboxylate of Glu 276 in a bidentate hydrogen bond donor-acceptor mode. This binding feature is not significantly different from that found in the zanamivir-sialidase complex.

Furthermore, the efficacy of orally administered compound **11a** was tested in the influenza virus infected mouse model on the basis of the survival rates for treated and infected mice relative to that for control mice. Compound **11a** was administered orally once daily for 5 days after infection. It was found that compound **11a** had similar efficacy relative to oseltamivir phosphate as shown in Table 2.

In summary, we prepared the tetrahydro-furan-2-yl, tetrahydro-pyran-2-yl, and oxepan-2-yl derivatives related to zanamivir using an RCM reaction. These bicyclic ether derivatives substituted by diol at the C-3' and C-4'



Figure 3. Model structure¹¹ of the complex of compound **11a** with influenza virus sialidase.

Table 2. The median survival time of compound $11a^{a}$ and oseltamivir phosphate^a and the results of the Log-rank test

Compd	Median survival time ^b (days)	p value ^c compared with	
	unie (duys)	Control	Oseltamivir phosphate
Control Oseltamivir phosphate 11a	7.5 10 10	<0.0001 <0.0001	0.1089

Mice were infected with influenza A/PR/8/34 (H1N1) virus. ^aCompound 11a and oseltamivir phosphate were administered orally at doses of 10 µmol/kg once daily for 5 days after infection. ^bThe median survival times were calculated by Kaplan–Meier curve. ^cStatistical significances of the median survival times were analyzed by the Log-rank test.

positions showed comparable sialidase inhibitory activities relative to that of zanamivir. Furthermore, compound 11a exhibited a similar oral efficacy in the mouse/ infection model to that of oseltamivir. This is the first example of an in vivo result for derivatives related to zanamivir.

Acknowledgements

We wish to thank Dr. Mutsuo Nakajima and Ms. Reiko Kametani for performing the sialidase assay, Dr. Shuichi Miyamoto and Atsushi Kasuya for molecular modeling studies, and Mr. Youji Furukawa for performing X-ray crystallographic analysis.

References and Notes

(a) Palese, P.; Compans, R. W. J. Gen. Virol. 1976, 33, 159.
(b) Palese, P.; Jobita, K.; Ueda, M.; Compans, R. W. Virology 1974, 61, 397.

2. (a) Babu, Y. S.; Chand, P.; Bantia, S.; Kotian, P.; Dehghani, A.; El-Kattan, Y.; Lin, T. H.; Hutchison, T. L.; Elliott, A. J.; Parker, C. D.; Ananth, S. L.; Horn, L. L.; Laver, G. W.; Montgomery, J. A. *J. Med. Chem.* **2000**, *43*, 3482. (b) Maring, C. J; Giranda, V. L.; Kempf, D. J.; Stoll, V. S.; Sun, M.; Zhao, C; Gu, Y. C.; Hanessian, S.; Wang, G. T. PCT Int. Appl., WO0128996. (c) Smith, P. W.; Sollis, S. L.; Howes, P. D.; Cherry, P. C.; Starkey, I. D.; Cobley, K. N.; Weston, H.; Scicinski, J.; Merritt, A.; Whittington, A.; Wyatt, P.; Taylor, N.; Green, D.; Bethell, R.; Madar, S.; Fenton, R. J.; Morley, P. J.; Pateman, T.; Beresford, A. *J. Med. Chem.* **1998**, *41*, 787.

3. (a) von Itzstein, M.; Wu, W.-Y.; Kok, G. B.; Pegg, M. S.; Dyason, J. C.; Jin, B.; Phan, T. V.; Smythe, M. L.; White, H. F.; Oliver, S. W.; Colman, P. M.; Varghese, J. N.; Ryan, D. M.; Woods, J. M.; Bethell, R. C.; Hotham, V. J.; Cameron, J. M.; Penn, C. R. *Nature* **1993**, *363*, 418. (b) Ryan, D. M.; Ticehurst, J.; Dempsey, M. H.; Penn, C. R. *Antimicrob. Agents Chemother.* **1994**, *38*, 2270.

4. (a) Kim, C. U.; Lew, W.; Williams, M. A.; Wu, H.; Zhang, L.; Chen, X.; Escarpe, P. A.; Mendel, D. B.; Laver, W. G.; Stevens, R. C. *J. Med. Chem.* **1998**, *41*, 2451. (b) Mendel, D. B.; Tai, C. Y.; Escarpe, P. A.; Li, W.; Sidwell, R. W.; Huffmann, J. H.; Sweet, C.; Jakeman, K. J.; Merson, J.; Lacy, S. A.; Lew, W.; Williams, M. A.; Zhang, L.; Chen, M. S.; Bischofberger, N.; Kim, C. U. *Antimicrob. Agents Chemother.* **1998**, *42*, 640.

5. (a) Honda, T.; Masuda, T.; Yoshida, S.; Arai, M.; Kobayashi, Y.; Yamashita, M. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 1921. (b) Honda, T.; Masuda, T.; Yoshida, S.; Arai, M.; Kaneko, S.; Yamashita, M. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 1925. (c) Honda, T.; Yoshida, S.; Arai, M.; Masuda, T.; Yamashita, M. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 1929.

6. Toluene-4-sulfonic acid 2-(2,2-diethyl-[1,3]dioxolan-4-yl)ethyl ester was prepared as follows: 1,2,4-butanetriol was reacted with acetone dimethyl acetal in the presence of TsOH, followed by treatment of TsCl and pyridine to give the target compound.

7. Trifluoro-methanesulfonic acid 2,2-difluoro-but-3-enyl ester was prepared as follows: 2,2-difluoro-3-butenoic acid ethyl ester was treated with LiAlH₄ to afford the 2,2-difluoro-3-butene-1-ol, which was reacted with $(TfO)_2O$ in the presence of pyridine to give the target compound.

8. ORTEP drawings of compounds 11a, 11b, 19b, and 19c:





Crystallographic data for the structures **11a**, **11b**, **19b**, and **19c** reported in this paper have been deposited with the Cambridge Crystallographic Data Center as supplementary publication nos. CCDC 188639, 188640, 188641, 188642, respectively. Copies of the data can be obtained, free of charge, on application to CCDC, 12 Union Road, Cambridge CB2 1EZ, UK (Fax: +44–1223-336033 or e-mail: deposit@ccdc.cam.ac.uk). 9. The methods for sialidase inhibitory and plaque reduction assays are detailed in a previous paper.^{5a}

10. The stereochemistry of the diol of **10d** was determined by ¹H NMR analysis. $J_{2',3'}=6.0$ Hz and $J_{3',4'}=2.5$ Hz indicated that the stereochemistry between H-2' and H-3' was a *trans* configuration and that between H-3' and H-4' was a *cis* configuration. Therefore, the configurations of the diol of **10d** were 3'*R* and 4'*R*.

11. A model of compound **11a** complexed with influenza virus sialidase was created based on the crystal structure of subtype N9 sialidase complexed with zanamivir (Varghese, J. N.; Epa, V. C.; Colman, P. M. *Protein Sci.* **1995**, *4*, 1081). The glycerol side chain of zanamivir was replaced by the 3,4-dihydroxy-tetrahydropyran-2-yl group of **11a**, and ligand conformation was optimized using the molecular mechanics program CHARMm (Accelrys Inc., San Diego, CA, USA).