

Two Approaches toward the Formal Total Synthesis of Oseltamivir Phosphate (Tamiflu): Catalytic Enantioselective Three-Component Reaction Strategy and L-Glutamic Acid Strategy

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Supporting Information

ABSTRACT: Two independent formal total syntheses of oseltamivir phosphate were successfully achieved: the first utilized a copper-catalyzed asymmetric three-component reaction strategy, and the second utilized L-glutamic acid γ ester as a chiral source to install the correct stereochemistry. Both strategies used Dieckmann condensation to construct a six-membered ring core, after which manipulation of the functional groups and protecting groups accessed Corey's intermediate for the synthesis of oseltamivir phosphate. While the first synthesis was accomplished via four purification steps in 25.7% overall yield, albeit with moderate optical purity (76% ee), the second strategy achieved the synthesis via six purification steps in 19.8% overall yield with perfect enantiocontrol.

■ INTRODUCTION

Despite long-standing preventive efforts, influenza continues to be one of the most serious threats to public health. Strains of the influenza virus easily mutate to escape host defense systems (such as the case of the H1N1 subtype in 2009), 1,2 and avian influenza strains sometimes newly acquire infectivity between humans resulting in very high mortality rate. Although immunization is a reliable and established method to prevent human from being infected by the seasonal viruses, the preparation of vaccines against novel strains usually lags behind their detection, allowing for flu outbreaks. Possibilities that highly lethal strains, such as the H5N1 subtype discovered in 1997,³ will acquire infectivity between humans are of particular concern because of the lack of immunologic warheads. An alternative and powerful means to combat influenza is the use of antivirals, such as viral neuraminidase inhibitors.⁴

Neuraminidase is a viral enzyme that catalyzes the cleavage of the sialic acid residue from glycoproteins to liberate proliferated viruses from infected cells. Six years after the report of the firstin-class neuraminidase inhibitor, zanamivir (currently marketed

as Relenza),⁵ the best-in-class compound, oseltamivir phosphate (Tamiflu), developed by Gilead Science and Roche, was approved as a clinical medicine against influenza A and B viruses by the FDA in 1999. Notably, Tamiflu has demonstrated positive effects against the H5N1 and H1N1 subtypes and is in clinical use. A huge global stockpile of oseltamivir phosphate is needed, however, to prepare for an outbreak of influenza virus, especially newly discovered hazardous strains that can cause serious mortality.

The process of manufacturing Tamiflu used by Roche involves a chemical synthesis beginning with (-)-shikimic acid, which can be isolated from the seeds of Chinese star anise (plant of *Illicium* family). Because the supply of shikimic acid from the plant is susceptible to climate fluctuations, a variety of complementary methods have been developed to secure shikimic acid reserves.8

Step-economic synthetic routes employing inexpensive starting materials as well as less costly reaction and purification conditions would be valuable alternative solutions to the supply problem. Since the first reports of asymmetric total syntheses of oseltamivir phosphate in 2006, 9,10 a myriad of synthetic studies have been published. 11,12 Unfortunately, none of these synthetic methods have been developed to the commercial scale due to the high total cost (too many steps, low conversion rates, and costly reagents), impracticality (harsh conditions,

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malodorous reagents, demands for special equipment, high dilutions, and excessive wastes), and safety issues (flammable or explosive reagents and intermediates). In our continuing efforts toward the development of more practical preparative routes, our group reported several catalytic asymmetric syntheses of oseltamivir phosphate using a yttrium-catalyzed asymmetric ring-opening reaction of *meso*-aziridines with TMSN₃ or HOMO-raising barium-catalyzed asymmetric Diels—Alder reaction as key transformations to manipulate the stereochemistry. Despite the improved step-economy, the most recent synthetic route ^{13d} still requires a potentially explosive azide intermediate, which hinders its technical transfer to industry.

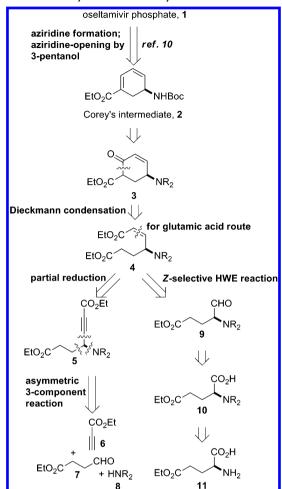
Herein we disclose a formal total synthesis of oseltamivir phosphate by two novel strategies: one using a catalytic asymmetric reaction and the other using enantiopure starting material to install the requisite stereochemistry. Both syntheses were accomplished via the Dieckmann condensation to form the six-membered ring skeleton, after which the identical protocols led to patent-free Corey's synthetic intermediate of oseltamivir phosphate, 2. 10 The original method reported by Corey takes seven steps to access to 2 and uses the starting material of low boiling point. For other approaches to synthesize Corey's intermediate, Kann et al. produced 2 through a diene-iron carbonyl intermediate and reported its optical resolution, 11a and Okamura et al. synthesized Corey's intermediate 2 using a Diels-Alder reaction of hydroxy pyridone and ethyl acrylate in a racemic manner. 11e Trost et al. also produced an intermediate that is different from 2 only in the protective group on the nitrogen atom (phthaloyl group) in an elegant manner using an enantioselective palladiumcatalyzed allylic amination reaction as a key transformation.^{11f} However, this method includes a step that needs microwave irradiation. Our approaches provide two alternative strategies to access Corey's intermediate in practical manners in terms of total yield and stereocontrol.

RESULTS AND DISCUSSION

The two novel approaches to our formal total synthesis of oseltamivir are summarized in Scheme 1. Retrosynthetically, the conjugated diene system of 2 precedes a cyclohexene-derived β ketoester 3 that can be disconnected to 4 by Dieckmann condensation. The route then branches off to one of two pathways, a catalytic asymmetric synthesis and a chiral pool method. The catalytic asymmetric synthesis takes advantage of a copper-catalyzed enantioselective three-component reaction to give optically active propargylic amines. As reported by Knochel et al., 14 the reaction proceeds via several events in a stepwise manner. The first step is in situ generation of an enamine from aldehyde 7 and secondary amine 8. Then, the copper(I) catalyst activates the enamine as a Lewis acid, accompanied by concomitant tautomerization into the iminium form, which is followed by copper(I) acetylide formation en route to its nucleophilic addition to the iminium resulting in propargylic amine 5. Subsequent partial reduction of the triple bond affords 4.

The chiral pool method utilizes the preinstalled chirality of L-glutamic acid as the origin of the stereochemistry of the target compound **2**. An α -amino aldehyde **9** is readily prepared from a commercially available L-glutamic acid γ -ester, which is followed by a Z-selective Horner–Wadsworth–Emmons (HWE) reaction to afford **4**.

Scheme 1. Retrosynthesis of Corey's intermediate 2



According to Knochel's report, the influence of the substituents of each substrate on the enantioselectivity of the three-component reaction can be summarized as follows: (1) trimethylsilylacetylene is the most preferable, aryl-substituted acetylene decreases the selectivity (never exceeding 90% ee), (2) bisbenzylamine gives excellent selectivity, but simple diallylamine substantially lowers the enantioselectivity, (3) aliphatic aldehydes provide better selectivity compared to aryl congeners, and α -alkyl-branched aliphatic aldehydes are the most preferable. The choice of the secondary amine is important not only because it is a determinant of the selectivity; the substituents should act as a protecting group of the primary amine and must be removed without damaging other functionalities upon exchange with a Boc group. Because removal of the benzyl group was expected to be problematic in the presence of a double bond, which always exists on the intermediates later than 4 bis(4-methoxybenzyl)amine was examined in a preliminary study. Unfortunately, oxidative deprotection did not work. We then decided to use diallylamine, which is removable via allylic substitution using Pd(0) as a catalyst. In addition, the absence of examples of propiolate ester such as 6 in the original reports led us to optimize reaction conditions (Table 1).

Initially, a reaction with 6, 7, and 8a was conducted under the standard conditions reported by Knochel with slight modification: with the catalyst system comprising 5 mol % of CuBr and 5.5 mol % of (R)-Quinap, 15 the reactions were

Table 1. Screening for Chiral Ligand in the Catalytic Asymmetric Three-Component Reaction

entry	ligand	R	X	Y	yield (%) ^e	ee (%) ^f
1	(R)-Quinap	Н	5.5	4	70	7
$2^{a,c}$	L1	Н	20	12	43	3
3	L2	Н	5.5	3	73	1
4	(R)-Monophos	Н	11	12	88	7
5	(R)-MOP	Н	11	12	6	3
6	(R)-Segphos	Н	5.5	12	21	4
7	(R,R)-i-Pr-DuPhos	Н	5.5	14	17	7
8	(S,S)-Ph-BPE	Н	5.5	14	25	2
9	(R,R)-Ph-Box	Н	5.5	3	88	2^g
10	(R,R)-Ph-Pybox	Н	5.5	12	89	43 ^g
11	(S,S)-4-MeO-C ₆ H ₄ - Pybox	Н	5.5	14	91	33
12	(R,R) -4-F- C_6H_4 -Pybox	Н	5.5	12	87	46 ^g
13	(R,R)-2-naphthyl-Pybox	Н	5.5	14	94	42^g
14 ^a	(S,S)-Ph-Pybox	Н	11	12	83	38
15 ^b	(S,S)-Ph-Pybox	Н	11	12	76	47
$16^{b,c}$	(S,S)-Ph-Pybox	Н	20	24	78	59
$17^{b,c}$	(S,S)-Ph-Pybox	Ph	20	24	91	64
$18^{b,d}$	(S,S)-Ph-Pybox	Ph	20	24	84	76
	× × ×	Ph PPh ₂	L2 : Ph₂P Pr	NH ₂ L Ph		

 a10 mol % of CuBr was used. b10 mol % of CuBr $_2$ was used. cThe reaction was performed at 0 °C. dThe reaction was performed at -15 °C. eNMR yield using 2-methoxynaphthalene as an internal standard. fDetermined by HPLC. $^g(R)$ -Isomer predominated.

performed in toluene in the presence of MS 4Å for 3 h at room temperature. The product was obtained in good yield (70%), but the enantioselectivity was as low as 7% ee. This outcome forced us to refine the catalyst system in an ad hoc manner to fit it to our particular system. Toward this end, we performed a broad screening for chiral ligands. Another P,N-ligand structurally closely related to Quinap reported by Carreira et al. 16 (L1) also afforded similarly unsatisfactory selectivity (entry 2), whereas a different type of P,N-ligand, L2, afforded an almost racemic adduct, albeit with clean conversion (entry 3, 73% yield, 1% ee). Next, testing of a variety of chiral monodentate and bidentate phosphorus-based ligands (phosphine- and phosphoramidite-based) revealed very poor enantioselectivity (entries 4-8, 6-88% yield, 2-7% ee). We then turned our attention to the use of Box ligands, which are well recognized as privileged chiral ligands containing two oxazoline rings and are commonly used in Lewis acidic coppercatalyzed asymmetric reactions. While a simple (R,R)-Ph-Box gave rise to the almost racemic product (entry 9), the tridentate variant (R,R)-Ph-Pybox, originally developed by Nishiyama and

co-workers,¹⁷ affected the asymmetric induction (entry 10, 89% yield, the antipode of 43% ee). Further tuning of the electronic and steric characteristics of the aryl groups located at oxazoline moieties had almost no beneficial effects on the enantiose-lectivity (entries 11–13). Notably, a catalyst system comprising Cu(II) and Ph-Pybox ligand (10 mol % of CuBr₂ and 20 mol % of (*S*,*S*)-Ph-Pybox) was slightly more effective compared with the Cu(I) system (entries 14 and 15). Indeed, an improved enantioselectivity up to 59% ee was observed at lower temperature at 0 °C, although an increased loading of the ligand and an extended reaction time (24 h) were required to fulfill reasonable conversions (entry 16, 78%). Encouraged by this result, the effects of a secondary amine were then examined.

Knochel reported that the use of a secondary amine with additional steric bulk, bis(2-phenylallyl)amine **8b**, improved enantioselectivity in many cases. This embellished allyl functionality can be uneventfully removed by Pd(0)-mediated allylic substitution. Accordingly, **8b** was tested under the conditions optimized for our system, as discussed above, resulting in better chemical yield and stereoselectivity (entry 17, 91% yield, 64% ee). Eventually, 84% yield with 76% ee was attained at a reaction temperature of $-15\,^{\circ}$ C, as demonstrated in entry 18. Further modifications of the conditions, however, failed to improve the enantioselectivity.

The overall process according to the catalytic asymmetric three-component reaction strategy is summarized in Scheme 2. With the optically active propargylamine products in hand, the succeeding partial reduction of the triple bond was examined. When adduct 5a was subjected to the conditions with poisoned Pd-mediated hydrogenation, such as Lindlar's catalyst, reduction of the allyl group afforded Z-olefin 4a as an inseparable mixture of products. The highest chemoselectivity was realized by the siloxane-based reduction method reported by Trost ((Me₂HSi)₂O, AcOH, 2.5 mol % of Pd₂(dba)₃·CHCl₃, and 10 mol % of $P(o\text{-tol})_3$), ¹⁹ but the reaction continued to suffer from a substantial undesired reduction of allyl groups. The same reaction conditions as above with the phenyl-substituted substrate 5b, however, escaped the overreduction to give Zolefin 4b in 84% yield without any trace of reduction of the 2phenylallyl groups, presumably because of the increased steric bulk.

The stage was set for the second key transformation of the synthesis, Dieckmann condensation. Preliminarily, several bases to promote this reaction were screened with 4a as the substrate. Use of 3 equiv of LHMDS at −40 °C led to substantially better conversion (ca. 90% yield) compared to the cases using KOt-Bu (31%) and NaH (no reaction). The transformation completed in 30 min at a higher temperature (-10 °C) without affecting the reaction course to give cyclization product 3a (see Scheme 3 also). The same protocol was effective for the synthesis via 4b to afford 3b, which was subjected to a 1,2reduction of the enone moiety employing NaBH₄ to give 13. The two 2-phenylallyl groups of 13 were removed by allylic substitution in the presence of a catalytic amount of Pd(PPh₃)₄ and N,N-dimethylbarbituric acid as a nucleophile, 20 which was followed by introduction of the Boc group to give 14 as a mixture of diastereomers in 58% yield over four steps. Further efforts to shorten the reaction sequence by introducing the Boc group prior to Dieckmann condensation failed: anion species derived from 4 by deprotonation of carbamate NH attacked at the carbonyl group of ester to give rise to a dead-end byproduct. At the end of the synthesis, mesylation of the

Scheme 2. Formal Total Synthesis of Oseltamivir Phosphate via a Catalytic Asymmetric Three-Component Reaction^a

"Conditions: (a) See Table 1 (runs 16–18), CuBr₂ (10 mol %), (*S*,*S*)-Ph-Pybox (20 mol %), 6 (2 equiv), 7 (1 equiv), 8a or 8b (2 equiv), MS 4Å, toluene, –15 or 0 °C, 24 h; (b) Pd₂(dba)₃·CHCl₃ (2.5 mol %), P(*o*-tol)₃ (10 mol %), (Me₂HSi)₂O (1 equiv), AcOH (1 equiv), toluene, 45 or 40 °C, 19 or 24 h, yield 36% for 4a, 84% for 4b; (c) LHMDS (3 equiv), THF, –40 °C, 1 h; NaBH₄ (2 equiv), MeOH, –20 °C, 30 min; Pd(PPh₃)₄ (10 mol %), *N*,*N*-dimethylbarbituric acid (6 equiv), CH₂Cl₂, rt, 1 h; Boc₂O (5 equiv), NaHCO₃ aq, CH₃CN, rt, 1 h, yield 55% for 4 steps; (d) MsCl (1.1 equiv), Et₃N (2 equiv), CH₂Cl₂, 4 °C, 10 min; DBU (3.1 equiv), CH₂Cl₂, rt, 1 h, 87% for 2 steps.

hydroxyl group and subsequent β -elimination afforded the target compound of this study, Corey's intermediate **2**, in good yield (87%).

Despite the overall fair efficiency of the synthetic route described above, enhancing the enantioselectivity of the asymmetric three-component reaction would be a formidable task. Therefore, we decided to switch our strategy to the chiral pool method, sharing the same synthetic intermediate 4 with the first synthetic route (Scheme 1). Historically, many asymmetric total syntheses have been accomplished using readily available chiral compounds, especially abundant natural products, as their origin of chirality. Of those, natural amino acids are inexpensive, inexhaustible chiral pools. Among the natural amino acids, L-glutamic acid is one of the least expensive starting materials for asymmetric syntheses. Previously, Saicic and co-workers achieved the synthesis of oseltamivir phosphate by taking advantage of L-glutamic acid as a chiral source, but the synthesis also made use of the Evans oxazolidinone-based diastereoselective aldol reaction to furnish the requisite

Scheme 3. Formal Total Synthesis of Oseltamivir Phosphate via the Glutamic Acid Route a

"Conditions: (a) 60% HClO₄ (1.1 equiv), t-BuOAc (excess), rt, 2 d, 77%; (b) allyl bromide (6 equiv), NaHCO₃ (4 equiv), EtOH, reflux, 24 h, yield 84%; (c) HCO₂H (excess), 80 °C, 1 h; ClCO₂Et (3 equiv), Et₃N (5 equiv), THF, -10 °C, 15 min, then NaBH₄ (6 equiv), MeOH, 0 °C, 30 min, yield 80% for 2 steps; (d) SO₃·pyridine (4 equiv), Et₃N (3.6 equiv), DMSO, 0 °C, 2 h; (CF₃CH₂O)₂P(=O)CH₂CO₂Et (1.1 equiv), 18-crown-6 (5 equiv), KHMDS (1.2 equiv), THF, -78 °C, 2 h, yield 80% for 2 steps; (e) LHMDS (3 equiv), THF, -40 °C, 1 h; NaBH₄ (2 equiv), MeOH, -20 °C, 30 min; Pd(PPh₃)₄ (10 mol %), N,N-dimethylbarbituric acid (6 equiv), CH₂Cl₂, rt, 1 h; Boc₂O (5 equiv), CH₃CN, rt, 1 h, yield 55% for 4 steps; (f) MsCl (1.1 equiv), Et₃N (2 equiv), CH₂Cl₂, 4 °C, 10 min; DBU (3 equiv), rt, 1 h, 87% for 2 steps.

stereochemistry. ^{11z} In our approach, the α -carbon of L-glutamic acid is the sole determinant of the stereochemistry of oseltamivir phosphate, which made the overall process more concise and efficient.

Synthesis along this line commenced with commercially available L-glutamic acid γ -ethyl ester 11 (>99% ee) as shown in Scheme 3. The α -carboxyl group of 11 was masked as *tert*-butyl ester 15 under acidic conditions with HClO₄ in the presence of t-BuOAc (77%).²¹ Then, the allyl groups were uneventfully introduced to the amino group of 15 to give a fully protected Lglutamic acid derivative 16 in 84% yield. Subsequent treatment of HCO₂H with 16 unveiled the α -carboxyl group, which was converted to the corresponding primary alcohol 17 via the formation of mixed anhydride followed by reduction with NaBH₄ in 80% yield over two steps.^{22'} After thorough investigation of the conditions for the subsequent oxidation of the hydroxyl group (e.g., DMP, Swern, TEMPO), Parikh-Doering oxidation (SO₃·pyridine, Et₃N, DMSO) was found to produce the best conversion. Because the aldehyde product 18 was labile upon purification, the crude material was directly used in the subsequent Z-selective HWE reaction.

Toward this end, two well-known Z-selective HWE reactions were tested. While Ando's protocol²³ with $(o\text{-tol})_2P(=O)CH_2CO_2Et$, NaI, and DBU gave the Z-olefin in 62% yield, accompanied by a substantial amount of the E-isomer (Z/E ratio; 80/20), the procedure reported by Still and Gennari²⁴ using $(CF_3CH_2O)_2P(=O)CH_2CO_2Et$, KHMDS, and 18-crown-6 achieved good reactivity (80% over 2 steps) and excellent Z/E selectivity (98/2) to give 4a. Subsequently, essentially the same reaction scheme developed for the first strategy led to the desired Corey's intermediate 2. In fact, the four-step procedure to 20 in 55% yield was followed by the two-step transformation into 2 in 87% yield as shown in Scheme 2. The optical purity of the final product 2 obtained by this method was 99% ee, confirming that no racemization occurred during the synthesis.

CONCLUSION

In summary, we developed two independent synthetic methods to access the synthetic precursor (2) of oseltamivir phosphate reported by Corey. In the first strategy, we took advantage of the copper-catalyzed asymmetric three-component reaction of terminal alkyne, aldehyde, and secondary amine to afford the propargylic amine product from which a substrate for Dieckmann condensation, Z-olefin 4b, was derived. The second strategy used the chirality of a readily available, optically pure Lglutamic acid derivative as the origin of the stereochemistry of target compound 2 and accessed the substrate of Dieckmann condensation 4a via a Z-selective HWE reaction of 18. The first synthesis was accomplished via five purification steps in 25.7% overall yield albeit with moderate optical purity (76% ee), and the second synthesis was achieved via six purification steps in 19.8% overall yield in optical pure form. Studies to further refine the glutamic acid route to make it more cost-effective by reducing the purification steps and tuning the reaction conditions are underway.

■ EXPERIMENTAL SECTION

All reactions were performed in oven-dried round-bottom flasks and test tubes with a Teflon-coated magnetic stirring bar under Ar atmosphere unless otherwise noted. Air- and moisture-sensitive liquids were transferred via a gastight syringe and a stainless steel needle. All workup and purification procedures were carried out with reagent grade solvents under ambient atmosphere. Flash chromatography was performed using silica gel 60 (230-400 mesh). Infrared (IR) spectra were recorded on a Fourier transform IR spectrophotometer. NMR was recorded on a 400 MHz spectrometer. For ¹H NMR (400 MHz), chemical shifts of protons are reported in parts per million downfield from tetramethylsilane and are referenced to residual protium in the NMR solvent (CDCl₃, δ 7.26 ppm). For ¹³C NMR (100 MHz), chemical shifts are reported in the scale relative to NMR solvent (CDCl₃, δ 77.0 ppm) as an internal reference. NMR data are reported as follows: chemical shifts, multiplicity (s: singlet, d: doublet, dd: doublet of doublets, t: triplet, q: quartet, m: multiplet), coupling constant (Hz), and integration. Optical rotation was measured using a 2-mL cell with a 1.0-dm path length on a polarimeter. High-resolution mass spectra (ESI-Orbitrap) were measured on an ESIMS instrument equipped with an Orbitrap detector. HPLC analysis was conducted on an HPLC system equipped with chiral stationary-phase columns (0.46 $cm \times 25 cm$).

(*S*)-Diethyl 4-(Diallylamino)hept-2-ynedioate (5a, Table 1, entry 16). To a tube containing CuBr₂ (4.5 mg, 20 μ mol, 10 mol %) and (*S*,*S*)-Ph-Pybox (14.8 mg, 40 μ mol, 20 mol %) was added toluene (0.8 mL) with stirring at room temperature. After 30 min, MS 4Å (120 mg), ethyl propiolate 6 (40.5 μ L, 0.400 mmol, 2 equiv), aldehyde 7^{25} (25.0 μ L, 0.200 mmol, 1 equiv), and diallylamine 8a (49.2 μ L, 0.400

mmol, 2 equiv) were successively added at 0 $^{\circ}\text{C}\textsc{,}$ and the mixture was stirred for 24 h at the same temperature. The reaction mixture was quenched with 1.0 M aqueous HCl. The resulting biphasic mixture was extracted with EtOAc, and the combined organic extract was washed with brine and dried over Na₂SO₄. Evaporation of volatiles under reduced pressure gave the crude mixture, which was used to determine ee (HPLC conditions are described below) and NMR yield (10 mg of 2-methoxynaphthalene was used as an internal standard). To collect physicochemical data, a crude material was purified with silica gel column chromatography (n-hexane/AcOEt = 8/1 to 6/1) to afford 6aas a pale yellow oil. ¹H NMR (CDCl₃, 400 MHz) δ 5.73 (m, 2H), 5.21 (d, J = 17.0 Hz, 2H), 5.11 (d, J = 10.5 Hz, 2H), 4.21 (q, J = 7.1 Hz,2H), 4.11 (q, J = 7.0 Hz, 2H), 3.71 (dd, J = 7.8 Hz, 7.1 Ĥz, 1H), 3.28 (m, 2H), 2.87 (dd, J = 14.0 Hz, 7.8 Hz, 2H), 2.42 (m, 2H), 1.99 (m, 2H), 1.30 (t, J = 7.1 Hz, 3H), 1.24 (t, J = 7.0 Hz, 3H); 13 C NMR (CDCl₃, 100 MHz) δ 172.9, 153.6, 135.9, 117.7, 85.9, 77.6, 62.0, 60.5, 53.7, 51.3, 30.9, 27.9, 14.3, 14.0; IR (neat, cm⁻¹) 3081, 2981, 2819, 2225, 1740, 1712; ESI-HRMS calcd for $C_{17}H_{25}NO_4Na$ [M + Na]⁺ 330.1676, found 330.1674; $[\alpha]^{22}_{D} = -40.1$ (34% ee, a sample obtained under different conditions, c 0.97, CHCl₃); HPLC (n-hexane/2propanol = 50/1, CHIRALPAK IC, 0.5 mL/min, 254 nm) t_R = 17.8 min (minor), 19.6 min (major).

(S)-Diethyl 4-(Bis(2-phenylallyl)amino)hept-2-ynedioate (5b, Table 1, entry 18). Substrates 6 (40.5 μ L, 0.400 mmol, 2 equiv), 7 (25.0 µL, 0.200 mmol, 1 equiv), and bis(2-phenylallyl)amine 8b (94.1 μ L, 0.400 mmol, 2 equiv) were used in the presence of catalyst comprising CuBr₂ (4.5 mg, 20 μ mol, 10 mol %) and (S,S)-Ph-Pybox (14.8 mg, 40 μ mol, 20 mol %). After stirring for 24 h at -15 °C, the reaction mixture was quenched with 1.0 M aqueous HCl. The resulting biphasic mixture was extracted with EtOAc, and the combined organic extract was washed with brine and dried over Na2SO4. Evaporation of volatiles under reduced pressure gave the crude mixture, which was used to determine NMR yield and ee (HPLC conditions are described below), then was purified by silica gel column chromatography (nhexane/AcOEt = 12/1 to 8/1) to give 5b (86.4 mg, 94%, 76% ee) as a pale yellow oil. 1 H NMR (CDCl₃, 400 MHz) δ 7.28–7.11 (m, 10H), 5.43 (s, 2H), 5.27 (s, 2H), 4.24 (q, J = 7.1 Hz, 2H), 4.07-3.90 (m, 2H), 3.86 (d, J = 13.8 Hz, 2H), 3.72-3.60 (m, 1H), 3.18 (d, J = 13.8 Hz, 2H), 2.00-1.82 (m, 3H), 1.82-1.68 (m, 1H), 1.32 (dd, J = 7.1Hz, 7.1 Hz, 3H), 1.16 (dd, J = 7.1 Hz, 7.1 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 172.8, 153.5, 144.8, 139.3, 128.0, 127.5, 126.6, 116.5, 85.3, 78.0, 62.0, 60.2, 55.4, 50.6, 30.2, 27.5, 14.1, 14.0; IR (neat, cm⁻¹) 2222, 1733, 1711, 1244, 1182, 908, 779, 752, 696, 407; ESI-HRMS calcd for $C_{29}H_{34}NO_4$ [M + H]⁺ 460.2482, found 460.2477; $[\alpha]_D^{29}$ = -38.7 (73% ee, a sample obtained under different conditions, c 1.02, CHCl₃); HPLC (n-hexane/2-propanol = 50/1, CHIRALPAK IC, 0.5 mL/min, 254 nm) t_R = 22.3 min (minor), 24.5 min (major).

(5)-Diethyl 4-(Diallylamino)-(Z)-hept-2-enedioate (4a). To $Pd_2(dba)_3$ ·CHCl $_3$ (105 mg, 0.102 mmol, 2.5 mol %) and $P(o\text{-tol})_3$ (124 mg, 0.406 mmol, 10 mol %) dissolved in toluene (10.3 mL) were added (Me $_2$ HSi) $_2$ O (0.718 mL, 4.06 mmol), AcOH (0.232 mL, 4.06 mmol), and toluene solution (10 mL) of 5a (1.25 g, 4.06 mmol) successively at room temperature. The mixture was stirred at 45 °C for 19 h, then allowed to reach room temperature, and quenched with saturated aqueous NaHCO $_3$. The aqueous layer was extracted with EtOAc, and the organic layer was washed with brine, dried over Na $_2$ SO $_4$, and concentrated *in vacuo*. The resulting residue was purified by silica gel column chromatography (n-hexane/EtOAc = 9/1 to 7/1) to afford 4a as a colorless oil (450 mg, 36%). Spectral data are shown later (as a product of the glutamic acid route). NMR spectra are included in Supporting Information.

(\$)-Diethyl 4-(Bis(2-phenylallyl)amino)-(Z)-hept-2-enedioate (4b). To Pd₂(dba)₃·CHCl₃ (243 mg, 0.235 mmol, 2.5 mol %) and P(o-tol)₃ (286 mg, 0.940 mmol, 10 mol %) dissolved in toluene (23.5 mL) were added (Me₂HSi)₂O (1.66 mL, 9.40 mmol), AcOH (0.537 mL, 9.40 mmol), and toluene solution (23.5 mL) of 5b (4.31 g, 9.39 mmol) successively at room temperature. The mixture was stirred at 40 °C for 24 h, then allowed to reach to room temperature, and quenched with saturated aqueous NaHCO₃. The aqueous layer was extracted with EtOAc, and the organic layers were washed with brine,

dried over Na₂SO₄, and concentrated *in vacuo*. The resulting residue was purified by silica gel column chromatography (n-hexane/EtOAc = 12/1 to 8/1) to afford 4b as a colorless oil (3.65 g, 84%). ¹H NMR (CDCl₃, 400 MHz) δ 7.29–7.23 (m, 4H), 7.21–7.16 (m, 6H), 6.27 (dd, J = 11.4 Hz, 11.4 Hz, 1H), 5.94 (d, J = 11.4 Hz, 1H), 5.32 (s, 2H), 5.17 (s, 2H), 4.46–4.32 (m, 1H), 4.14–4.04 (m, 2H), 4.03 (q, J = 7.2 Hz, 1H), 3.83 (d, J = 14.1 Hz, 2H), 3.19 (d, J = 14.1 Hz, 2H), 2.08–1.96 (m, 1H), 1.94–1.72 (m, 2H), 1.71–1.58 (m, 1H), 1.21 (dd, J = 7.2 Hz, 7.2 Hz, 3H), 1.19 (dd, J = 7.2 Hz, 7.2 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 173.8, 165.6, 146.0, 145.7, 140.1, 127.9, 127.3, 126.6, 122.7, 115.2, 60.1, 60.0, 55.2, 54.4, 30.8, 27.2, 14.2; IR (neat, cm⁻¹) 2980, 2820, 1721, 1631, 1416, 1186, 1029, 906, 779, 698; ESI-HRMS calcd for C₂₉H₃₆NO₄ [M + H]⁺ 462.2639, found 462.2632; $[\alpha]^{27}_{D}$ = +82.0 (73% ee, c 1.04, CHCl₃).

(2*R*,5*S*)-Ethyl 5-(*tert*-Butoxycarbonylamino)-2-hydroxycyclo-hex-3-enecarboxylate (14). To a stirred solution of diester (4b) (3.61 g, 7.83 mmol) in THF (15.7 mL) was slowly added a 1.0 M THF solution of LHMDS (23.5 mL, 23.5 mmol) at -40 °C. The resultant solution was stirred at the same temperature for 1 h, diluted with EtOAc, and quenched with saturated aqueous NH₄Cl. The biphasic mixture was extracted with EtOAc, and combined organic extracts were washed with brine and dried over Na₂SO₄. After evaporation of the organic solvent under reduced pressure, the crude product including 3b (keto-enol mixture) was obtained as a pale yellow oil, which was used for the next reaction without further purification.

To a stirred solution of the crude material including **3b** (maximum 7.83 mmol) in MeOH (39.2 mL) was added NaBH₄ (592 mg, 15.7 mmol) at -20 °C. The resultant solution was stirred at the same temperature for 30 min and then quenched with saturated aqueous NH₄Cl (slowly added). After warming to room temperature, MeOH was removed under the reduced pressure. The aqueous layer was extracted with EtOAc, and the combined organic layers were washed with brine and dried over Na₂SO₄. After evaporation of the volatiles under reduced pressure, the crude product including **13** was obtained as pale yellow oil, which was used for the next step without further purification.

To a stirred solution of crude ester (13, maximum 7.83 mmol) in CH₂Cl₂ (39.2 mL) were successively added Pd(PPh₃)₄ (905 mg, 0.783 mmol, 10 mol %) and N,N-dimethylbarbituric acid (7.34 g, 47.0 mmol) at room temperature. The resulting solution was stirred at the same temperature for 1 h, and CH2Cl2 was removed under reduced pressure. To this mixture were successively added a 1.0 M solution of Boc₂O in CH₃CN (39.2 mL, 39.2 mmol) and saturated aqueous NaHCO₃ (39.2 mL) at room temperature. The resulting solution was vigorously stirred at the same temperature for 1 h, diluted with EtOAc, and quenched with saturated aqueous NaHCO3. The biphasic mixture was extracted with EtOAc, and combined organic extracts were washed with brine and dried over Na2SO4. After evaporation of volatiles under reduced pressure, the crude mixture was purified by silica gel column chromatography (n-hexane/AcOEt = 1/1) to give the desired product (14) (1.23 g, diastereomeric mixture, 55% yield over 4 steps) as a pale yellow solid. The diastereomers could be partially separated upon careful silica gel column chromatography. Usually, the mixture of the isomers was used in the next step without separation.

(1*R*,2*R*,5*S*)-Ethyl 5-(*tert*-Butoxycarbonylamino)-2-hydroxycyclohex-3-enecarboxylate (14, less polar isomer). ¹H NMR (CDCl₃, 400 MHz) δ 5.77–5.71 (m, 1H), 5.65–5.59 (m, 1H), 4.53–4.47 (m, 1H), 4.47–4.40 (m, 1H), 4.35 (brs, 1H), 4.24–4.12 (m, 2H), 2.62–2.52 (m, 1H), 2.51–2.38 (m, 1H), 1.43 (s, 9H), 1.27 (dd, *J* = 4.8 Hz, 4.8 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 173.8, 155.2, 131.6, 130.5, 79.6, 67.7, 61.0, 48.0, 47.3, 31.7, 28.3, 14.1; IR (neat, cm⁻¹) 3424, 3361, 2976, 1718, 1682, 1523, 1181, 1053; ESI-HRMS calcd for C₁₄H₂₃NNaO₅ [M + Na]⁺ 308.1468, found 308.1474; [α]²⁸_D = -53.4 (70% ee, *c* 1.04, CHCl₃).

(15,2*R*,5*S*)-Ethyl 5-(*tert*-Butoxycarbonylamino)-2-hydroxycyclohex-3-enecarboxylate (14, more polar isomer). ¹H NMR (CDCl₃, 400 MHz) δ 6.00–5.91 (m, 1H), 5.86–5.76 (m, 1H), 4.54–4.42 (m, 1H), 4.40 (dd, *J* = 4.1 Hz, 4.1 Hz, 1H), 4.32–4.22 (m, 1H), 4.19 (q, *J* = 7.1 Hz, 2H), 2.80–2.67 (m, 1H), 2.30–2.15 (m, 1H),

1.99–1.87 (m, 1H), 1.43 (s, 9H), 1.27 (dd, J = 7.1 Hz, 7.1 Hz, 3H); 13 C NMR (CDCl₃, 100 MHz) δ 174.0, 155.0, 131.0, 129.9, 79.6, 63.6, 60.9, 44.4, 41.6, 28.3, 27.0, 14.1; IR (neat, cm⁻¹) 3361, 2979, 2934, 1690, 1517, 1367, 1248, 1170, 1012, 755; ESI-HRMS calcd for C₁₄H₂₃NNaO₅ [M + Na]⁺ 308.1468, found 308.1473; $[\alpha]^{28}_{D}$ = -133.3 (70% ee, c 1.02, CHCl₃).

(S)-Ethyl 5-((tert-Butoxycarbonyl)amino)cyclohexa-1,3-dienecarboxylate (2). To a stirred solution of the residue were successively added 14 (172 mg, 0.603 mmol) in CH₂Cl₂ (3 mL), MsCl (51.3 μ L, 0.663 mmol), and Et₃N (167 μ L, 1.21 mmol) at 4 °C, and the mixture was stirred for 10 min at the same temperature. DBU (279 μ L, 1.87 mmol) was added to the mixture, and the mixture was further stirred for 1 h at room temperature. The mixture was diluted with CH2Cl2 and washed with H2O, and the aqueous layer was backextracted with CH2Cl2. The combined organic layers were washed with 1 N HCl, saturated aqueous NaHCO3, and brine successively and dried over Na2SO4. The mixture was concentrated to give crude material that was purified with silica gel column chromatography (nhexane/Et₂O = 3/1 to 2/1) to afford 2 (140 mg, 0.523 mmol, 87% yield) as a pale yellow viscous oil. ESI-HRMS calcd for C₁₄H₂₁NO₄Na [M + Na]+ 290.1363, found 290.1361. Physicochemical data was identical to those reported. 10 Other spectral data are shown later (as a product of the glutamic acid route). NMR spectra obtained by this sample are included in Supporting Information.

(S)-1-tert-Butyl 5-Ethyl 2-Aminopentanedioate (15). L-Glutamic acid γ -ethyl ester (11, 10.0 g, 57.1 mmol) was dissolved in aqueous 60% HClO₄ (6.9 mL, 62.8 mmol) with stirring in an ice bath. t-BuOAc (250 mL) was added, and the stirring was continued until a homogeneous solution was obtained. The mixture was maintained at room temperature for 2 days, then 0.1 N HCl (350 mL) was added to the mixture, and Et₂O was added. The aqueous phase was separated from the organic phase, and the aqueous phase was adjusted to pH = 9with aqueous Na₂CO₃. The aqueous phase was extracted with EtOAc, and the organic layers were combined, washed with brine, dried over anhydrous Na₂SO₄, and concentrated to afford the desired ester 15 as a colorless oil (10.14 g, 77%). The crude product was NMR pure and was used directly without purification in the next step. ¹H NMR $(CDCl_3, 400 \text{ MHz}) \delta 4.07 (q, J = 7.2 \text{ Hz}, 2H), 3.28 (dd, J = 8.2 \text{ Hz},$ 5.3 Hz, 1H), 2.38 (dd, J = 7.6 Hz, 7.6 Hz, 2H), 2.02–1.92 (m, 1H), 1.80-1.68 (m, 1H), 1.51 (s, 2H), 1.40 (s, 9H), 1.19 (dd, J = 7.1 Hz, 7.1 Hz, 3H); 13 C NMR (CDCl₃, 100 MHz) δ 174.8, 173.2, 81.1, 60.3, 54.2, 30.6, 29.8, 27.9, 14.1; IR (neat, cm⁻¹) 3385, 2979, 2935, 1731; ESI-HRMS calcd for $C_{11}H_{22}NO_4$ [M + H]⁺ 232.1543, found 232.1546; $[\alpha]^{25}_{D} = 12.4$ (c 1.04, CHCl₃).

(S)-1-tert-Butyl 5-Ethyl-2-(diallylamino)pentanedioate (16). Diester 15 (9.4 g, 40.6 mmol) was dissolved in EtOH (100 mL), and powdered NaHCO₃ (13.65 g, 0.163 mol) and allyl bromide (21.1 mL, 0.244 mol) were added at room temperature. The mixture was stirred under reflux for 20 h, then cooled to room temperature, filtered through a Celite pad using EtOAc, and concentrated to give crude material, which was purified by silica gel column chromatography (nhexane/EtOAc = 32/1) to afford 16 as a colorless oil (10.59 g, 84%). ¹H NMR (CDCl₃, 400 MHz) δ 5.78–5.65 (m, 2H), 5.15 (d, J = 17.2 Hz, 2H), 5.07 (d, J = 10.1 Hz, 2H), 4.10 (q, J = 7.1 Hz, 2H), 3.38-3.28 (m, 3H), 3.01 (dd, J = 14.4 Hz, 8.0 Hz, 2H), 2.45-2.26 (m, 2H),2.00-1.80 (m, 2H), 1.45 (s, 9H), 1.23 (dd, J = 7.1 Hz, 7.1 Hz, 3H; 13 C NMR (CDCl₃, 100 MHz) δ 173.5, 171.9, 136.7, 116.9, 80.9, 61.1, 60.2, 53.3, 30.9, 28.3, 24.6, 14.2; IR (neat, cm⁻¹): 3078, 2978, 2819, 2934, 2841, 1734; ESI-HRMS calcd for $C_{17}H_{29}NO_4Na [M + Na]^+$ 334.1989, found 334.1987; $[\alpha]^{25}_{D} = -55.7$ (c 1.1, CHCl₃).

(5)-Ethyl 4-(Diallylamino)-5-hydroxypentanoate (17). Diester 16 (1.4 g, 4.5 mmol) was dissolved in HCO₂H (15 mL) and stirred at 80 °C for 1 h. The reaction mixture was cooled to room temperature, and HCO₂H was removed by azeotropic distillation with toluene (3 times) and CHCl₃ (3 times) under reduced pressure to afford the crude mixture. To the resultant residue in THF (30 mL) at -10 °C was added Et₃N (3.1 mL, 22.5 mmol), followed by ethyl chloroformate (1.28 mL, 13.5 mmol). After 15 min, NaBH₄ (1.02 g, 27 mmol) was added in one portion. MeOH (30 mL) was then added dropwise to the mixture at 0 °C. The solution was stirred for an additional 30 min

at the same temperature and then quenched with saturated aqueous NH₄Cl. The organic solvents were evaporated under reduced pressure, and the product was extracted with EtOAc, washed with brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure to afford crude material, which was purified by silica gel column chromatography (n-hexane/EtOAc = 1/4) to afford 17 as a colorless oil (869 mg, 80%). ¹H NMR (CDCl₃, 400 MHz) δ 5.80–5.67 (m, 2H), 5.21–5.05 (m, 4H), 4.10 (q, J = 7.2 Hz, 2H), 3.47 (dd, J = 10.6, 5.2 Hz, 1H), 3.34–3.21 (m, 3H), 2.96 (dd, J = 14.2 Hz, 7.8 Hz, 2H), 2.90–2.78 (m, 1H), 2.32–2.17 (m, 2H), 1.95–1.83 (m, 1H), 1.48–1.35 (m, 1H), 1.23 (dd, J = 7.1 Hz, 7.1 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 173.1, 136.4, 117.4, 60.6, 60.5, 59.2, 52.1, 31.5, 21.0, 14.2; IR (neat, cm⁻¹): 3440, 2978, 2934, 2876, 1733, 1642; ESI-HRMS calcd for C₁₃H₂₄NO₃ [M + H]⁺ 242.1751, found 242.1752; [α]²⁵_D = 23.1 (c 1.0, CHCl₃).

(5)-Ethyl 4-(Diallylamino)-5-oxopentanoate (18). Et₃N (0.21 mL, 1.49 mmol) was added to 17 (100 mg, 0.41 mmol) at 0 °C. Then, SO₃-pyridine (264 mg, 1.66 mmol) in DMSO (1.5 mL) was slowly added at 0 °C. After the addition completed, the mixture was stirred at the same temperature for 2 h. The reaction mixture was then quenched by the addition of cold water (1 mL) and extracted with ether. The combined organic layers were washed with 5% citric acid and brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure to afford crude 18 as a colorless oil, which was used for the succeeding reaction without further purification. ¹H NMR (CDCl₃, 400 MHz) δ 9.66 (s, 1H), 5.75–5.65 (m, 2H), 5.16–5.05 (m, 4H), 4.06 (q, J = 7.1 Hz, 2H), 3.26–3.11 (m, 5H), 2.44–2.37 (m, 1H), 2.31–2.23 (m, 1H), 1.97–1.88 (m, 1H), 1.81–1.72 (m, 1H), 1.19 (t, J = 7.1 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 203.6, 173.2, 136.0, 117.8, 66.9, 60.4, 53.7, 31.0, 19.9, 14.2.

(S)-Diethyl 4-(Diallylamino)-(Z)-hept-2-enedioate (4a). A solution of $(CF_3CH_2O)_2P(=O)CH_2CO_2Et$ (151 mg, 0.455 mmol) and 18-crown-6 (547 mg, 2.07 mmol) in THF (4 mL) was cooled to -78 °C and treated with KHMDS (0.5 M in toluene, 0.95 mL, 0.476 mmol). The mixture was stirred at -78 °C for 45 min, and then 18 (maximum 0.414 mmol) in THF (3 mL) was slowly added at -78 °C. After the addition was completed, the mixture was stirred at −78 °C for 2 h. Then, the reaction mixture was quenched with saturated aqueous NH₄Cl. After the mixture was warmed to room temperature, the solvent was evaporated under reduced pressure and extracted with EtOAc. The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure to afford crude material as a colorless oil (Z/E = 98/2), which was purified by silica gel column chromatography (n-hexane/EtOAc = 11.5/1) to afford Z-olefin 4a as a colorless oil (103 mg, 80% over 2 steps). 1 H NMR (CDCl₃, 400 MHz) δ 6.12 (dd, J = 11.4 Hz, 10.3 Hz, 1H), 5.89 (d, J = 11.4 Hz, 1H), 5.80-5.70 (m, 2H), 5.13-5.04 (m, 4H), 4.45 (m, 1H), 4.15-4.07 (m, 4H), 3.26 (m, 2H), 2.91 (dd, J = 14.4 Hz, 7.4 Hz, 2H), 2.46 (m, 1H), 2.30 (m, 1H), 1.93 (m, 1H), 1.73 (m, 1H), 1.28–1.21 (m, 6H); 13 C NMR (CDCl₃, 100 MHz) δ 173.7, 165.8, 147.4, 136.5, 122.1, 116.7, 60.2, 60.1, 56.0, 52.5, 31.1, 27.3, 14.2; IR (neat, cm⁻¹) 2980, 2936, 1734, 1732; ESI-HRMS calcd for $C_{17}H_{28}NO_4 [M + H]^+ 310.2013$, found 310.2015; $[\alpha]^{25}D = 138.5$ (c 0.91, CHCl₃).

(55)-Ethyl 5-((tert-Butoxycarbonyl)amino)-2-hydroxycyclo-hex-3-enecarboxylate (14). To a stirred solution of 4a (340 mg, 1.10 mmol) in THF (5.49 mL) was slowly added a 1.0 M THF solution of LHMDS (3.30 mL, 3.30 mmol) at -40 °C, and the mixture was stirred for 30 min at the same temperature. EtOAc and saturated aqueousNH₄Cl were added to quench the reaction, and the aqueous layer was extracted with EtOAc. The combined organic layers were washed with brine, dried over Na₂SO₄, filtered, and concentrated to give crude 19 (keto—enol mixture) as a pale yellow oil, which was used for the next reaction without further purification.

To a stirred solution of crude 19 (maximum 1.10 mmol) in MeOH (5.49 mL) was added NaBH₄ (83.2 mg, 2.20 mmol) at -20 °C, and the mixture was stirred for 30 min at the same temperature. After the addition of saturated aqueous NH₄Cl, MeOH was removed under reduced pressure. EtOAc was added, and the resulting aqueous layer was extracted with EtOAc. The combined organic layers were washed

with brine, dried over Na_2SO_4 , filtered, and concentrated to give crude **20**, which was partially purified by silica gel column chromatography (n-hexane/EtOAc = 2/1) to afford a sample including **20** (180 mg, 0.680 mmol at maximum, diastereomeric mixture) as a pale yellow oil.

To a stirred solution of partially purified 20 (180 mg, maximum 0.680 mmol) in CH₂Cl₂ (3.39 mL) were successively added $Pd(PPh_3)_4$ (78.4 mg, 67.8 μ mol, 10 mol %) and N,N-dimethylbarbituric acid (636 mg, 4.07 mmol) at room temperature, and the mixture was stirred for 1 h. After removing CH2Cl2 under the reduced pressure, a 0.82 M solution of Boc₂O in CH₃CN (4.13 mL, 3.39 mmol) and saturated aqueous NaHCO3 were successively added at room temperature, and the mixture was vigorously stirred for 3 h at the same temperature. The mixture was diluted with EtOAc and saturated aqueous NaHCO3, and the aqueous layer was extracted with EtOAc. The combined organic layers were washed with brine, dried over Na2SO4, filtered, and concentrated to give a crude mixture, which was purified with silica gel column chromatography (n-hexane/AcOEt = 1/1) to afford 14 (175 mg, 0.613 mmol, diastereomeric mixture, 56% yield for 4 steps) as a pale yellow solid. NMR and IR data were identical to those described above. ESI-HRMS calcd for $C_{14}H_{23}NO_5Na [M + Na]^+ 308.1468$, found 308.1468. NMR spectra obtained by this sample (diastereomeric mixture) are included in Supporting Information.

(5)-Ethyl 5-((*tert*-Butoxycarbonyl)amino)cyclohexa-1,3-dienecarboxylate (2). The same protocol described above was applied to 14 synthesized via the glutamic acid route. Physicochemical data were identical to those reported ¹⁰ as follows. ¹H NMR (CDCl₃, 400 MHz) δ 7.03 (d, J = 3.9 Hz, 1H), 6.18–6.09 (m, 2H), 4.61 (m, 1H), 4.42 (m, 1H), 4.20 (q, J = 7.1 Hz, 2H), 2.76–2.61 (m, 2H), 1.42 (s, 9H), 1.29 (t, J = 7.1 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 166.8, 154.9, 132.7, 131.7, 127.0, 124.8, 79.5, 60.6, 43.5, 28.8, 28.4, 14.3; IR (neat, cm⁻¹) 3352, 2978, 1705; ESI-HRMS calcd for C₁₄H₂₁NO₄Na [M + Na]⁺ 290.1363, found 290.1361; $\left[\alpha\right]^{25}_{D} = -205.5$ (98.9% ee, c 1.1, CHCl₃), lit. $\left[\alpha\right]^{20}_{D} = -217$ (>99% ee, c 1.1, CHCl₃); ^{14a} HPLC (n-hexane/2-propanol = 50/1, CHIRALPAK AD-H, 1.0 mL/min, 254 nm) t_R = 16.4 min (minor), 19.3 min (major).

ASSOCIATED CONTENT

S Supporting Information

¹H and ¹³C NMR spectra for all new compounds and HPLC data. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The authors declare no competing financial interest.

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■ REFERENCES

- (1) Neumann, G.; Chen, H.; Gao, G. F.; Shu, Y.; Kawaoka, Y. Cell Res. 2010, 20, 51.
- (2) Oseltamivir-resistant strains have been already discovered: van der Vries, E.; Stelma, F. F.; Boucher, C. A. B. *N. Engl. J. Med.* **2010**, 363, 1381.
- (3) (a) Moscona, A. N. Engl. J. Med. 2005, 353, 1363. (b) Boltz, D. A.; Aldridge, J. R., Jr.; Webster, R. G.; Govorkova, E. A. Drugs 2010, 70, 1349.

- (4) (a) Dawood, F. S.; Jain, S.; Finelli, L.; Shaw, M. W.; Lindstrom, S.; Garten, R. J.; Gubareva, L. V.; Xu, X.; Bridges, C. B.; Uyeki, T. M. N. Engl. J. Med. 2009, 360, 2605. (b) Garten, R. J.; Davis, C. T.; Russell, C. A.; Shu, B.; Lindstrom, S.; Balish, A.; Sessions, W. M.; Xu, X.; Skepner, E.; Deyde, V.; Okomo-Adhiambo, M.; Gubareva, L.; Barnes, J.; Smith, C. B.; Emery, S. L.; Hillman, M. J.; Rivailler, P.; Smagala, J.; de Graaf, M.; Burke, D. F.; Fouchier, R. A. M.; Pappas, C.; Alpuche-Aranda, C. M.; Lopez-Gatell, H.; Olivera, H.; Lopez, I.; Myers, C. A.; Faix, D.; Blair, P. J.; Yu, C.; Keene, K. M.; Dotson, P. D., Jr.; Boxrud, D.; Sambol, A. R.; Abid, S. H.; George, K. S.; Bannerman, T.; Moore, A. L.; Stringer, D. J.; Blevins, P.; Demmler-Harrison, G. J.; Ginsberg, M.; Kriner, P.; Waterman, S.; Smole, S.; Guevara, H. F.; Belongia, E. A.; Clark, P. A.; aBeatrice, S. T.; Donis, R.; Katz, J.; Finelli, L.; Bridges, C. B.; Shaw, M.; Jernigan, D. B.; Uyeki, T. M.; Smith, D. J.; Klimov, A. I.; Cox, N. J. Science 2009, 325, 197. (c) Smith, G. J. D.; Vijaykrishna, D.; Bahl, J.; Lycett, S. J.; Worobey, M.; Pybus, O. G.; Ma, S. K.; Cheung, C. L.; Raghwani, J.; Bhatt, S.; Peiris, J. S. M.; Guan, Y.; Rambaut, A. Nature 2009, 459, 1122.
- (5) (a) von Itzstein, M.; Wu, W. Y.; Kok, G. B.; Pegg, M. S.; Dyason, J. C.; Jin, B.; Phan, T. V.; Smythe, M. L.; White, H. F.; Oliver, S. W.; Colman, P. M.; Varghese, J. N.; Ryan, D. M.; Woods, J. M.; Bethell, R. C.; Hotham, V. J.; Cameron, J. M.; Penn, C. R. Nature 1993, 363, 418. (b) von Itzstein, M. Nat. Rev. Drug Discovery 2007, 6, 967.
- (6) (a) Kim, C. U.; Lew, W.; Williams, M. A.; Liu, H. T.; Zhang, L. J.; Swaminathan, S.; Bischofberger, N.; Chen, M. S.; Mendel, D. B.; Tai, C. Y.; Laver, W. G.; Stevens, R. C. J. Am. Chem. Soc. 1997, 119, 681. (b) Kim, C. U.; Lew, W.; Williams, M. A.; Wu, H. W.; Zhang, L. J.; Chen, X. W.; Escarpe, P. A.; Mendel, D. B.; Laver, W. G.; Stevens, R. C. J. Med. Chem. 1998, 41, 2451.
- (7) (a) Karpf, M.; Trussardi, R. J. Org. Chem. 2001, 66, 2044. (b) Rohloff, J. C.; Kent, K. M.; Postich, M. J.; Becker, M. W.; Chapman, H. H.; Kelly, D. E.; Lew, W.; Louie, M. S.; McGee, L. R.; Prisbe, E. J.; Schultze, L. M.; Yu, R. H.; Zhang, L. J. J. Org. Chem. 1998, 63, 4545. (c) Abrecht, S.; Harrington, P.; Iding, H.; Karpf, M.; Trussardi, R.; Wirz, B.; Zutter, U. Chimia 2004, 58, 621.
- (8) (a) Draths, K. M.; Knop, D. R.; Frost, J. W. J. Am. Chem. Soc. 1999, 121, 1603. (b) Ghosh, S.; Chisti, Y.; Banerjee, U. C. Biotechnol. Adv. 2012, 30, 1425 and references therein. See also: (c) Farina, V.; Brown, J. D. Angew. Chem., Int. Ed. 2006, 45, 7330.
- (9) Fukuta, Y.; Mita, T.; Fukuda, N.; Kanai, M.; Shibasaki, M. *J. Am. Chem. Soc.* **2006**, *128*, 6312.
- (10) Yeung, Y.-Y.; Hong, S.; Corey, E. J. J. Am. Chem. Soc. 2006, 128, 6310.
- (11) (a) Bromfield, K. M.; Graden, H.; Hagberg, D. P.; Olsson, T.; Kann, N. Chem. Commun. 2007, 3183. (b) Satoh, N.; Akiba, T.; Yokoshima, S.; Fukuyama, T. Angew. Chem., Int. Ed. 2007, 46, 5734. (c) Satoh, N.; Akiba, T.; Yokoshima, S.; Fukuyama, T. Tetrahedron 2009, 65, 3239. (d) Shie, J.-J.; Fang, J.-M.; Wang, S.-Y.; Tsai, K.-C.; Cheng, Y.-S. E.; Yang, A.-S.; Hsiao, S.-C.; Su, C.-Y.; Wong, C.-H. J. Am. Chem. Soc. 2007, 129, 11892. (e) Kipassa, N. T.; Okamura, H.; Kina, K.; Hamada, T.; Iwagawa, T. Org. Lett. 2008, 10, 815-816. (f) Trost, B. M.; Zhang, T. Angew. Chem., Int. Ed. 2008, 47, 3759. (g) Zutter, U.; Iding, H.; Spurr, P.; Wirz, B. J. Org. Chem. 2008, 73, 4895. (h) Shie, J.-J.; Fang, J.-M.; Wong, C.-H. Angew. Chem., Int. Ed. 2008, 47, 5788. (i) Matveenko, M.; Willis, A. C.; Banwell, M. G. Tetrahedron Lett. 2008, 49, 7018. (j) Ishikawa, H.; Suzuki, T.; Hayashi, Y. Angew. Chem., Int. Ed. 2009, 48, 1304. (k) Ishikawa, H.; Suzuki, T.; Orita, H.; Uchimaru, T.; Hayashi, Y. Chem.—Eur. J. 2010, 16, 12616. (1) Zhu, S.; Yu, S.; Wang, Y.; Ma, D. Angew. Chem., Int. Ed. 2010, 49, 4656. (m) Sullivan, B.; Carrera, I.; Drouin, M.; Hudlicky, T. Angew. Chem., Int. Ed. 2009, 48, 4229. (n) Werner, L.; Machara, A.; Hudlicky, T. Adv. Synth. Catal. 2010, 352, 195. (o) Mandai, T.; Oshitari, T. Synthesis 2009, 783. (p) Oshitari, T.; Mandai, T. Synlett 2009, 787. (q) Sun, H.; Lin, Y.-J.; Wu, Y.-L.; Wu, Y. Synlett 2009, 2473. (r) Osato, H.; Jones, I. L.; Chen, A.; Chai, C. L. L. Org. Lett. 2010, 12, 60. (s) Wichienukul, P.; Akkarasamiyo, S.; Kongkathip, N.; Kongkathip, B. Tetrahedron Lett. 2010, 51, 3208. (t) Ma, J.; Zhao, Y.; Ng, S.; Zhang, J.; Zeng, J.; Than, A.; Chen, P.; Liu, X.-W. Chem.—Eur. J. 2010, 16, 4533. (u) Weng, J.; Li, Y.-B.; Wang, R.-B.; Li, F.-Q.; Liu, C.; Chan, A. S. C.;

- Lu, G. J. Org. Chem. 2010, 75, 3125. (v) Kamimura, A.; Nakano, T. J. Org. Chem. 2010, 75, 3133. (w) Ko, J. S.; Keum, J. E.; Ko, S. Y. J. Org. Chem. 2010, 75, 7006. (x) Ishikawa, H.; Bondzic, B. P.; Hayashi, Y. Eur. J. Org. Chem. 2011, 6020. (y) Trost, B. M.; Zhang, T. Chem.—Eur. J. 2011, 17, 3630. (z) Trajkovic, M.; Ferjancic, Z.; Saicic, R. N. Org. Biomol. Chem. 2011, 9, 6927. (aa) Werner, L.; Machara, A.; Sullivan, B.; Carrera, I.; Moser, M.; Adams, D. R.; Hudlicky, T.; Andraos, J. J. Org. Chem. 2011, 76, 10050. (ab) Oh, H.-S.; Kang, H.-Y. J. Org. Chem. 2012, 77, 8792. (ac) Rehak, J.; Hut'ka, M.; Latika, A.; Brath, H.; Almassy, A.; Hajzer, V.; Durmis, J.; Toma, Š.; Šebesta, R. Synthesis 2012, 44, 2424. For alternative approaches using (–)-shikimic acid as starting materials, see: (ad) Karpf, M.; Trussardi, R. Angew. Chem., Int. Ed. 2009, 48, 5760–5762. (ae) Nie, L.-D.; Shi, X.-X. Tetrahedron Asymmetry 2009, 20, 124–129. (af) Nie, L.-D.; Shi, X.-X.; Ko, K. H.; Lu, W.-D. J. Org. Chem. 2009, 74, 3970–3973.
- (12) For reviews, see: (a) Shibasaki, M.; Kanai, M. Eur. J. Org. Chem. 2008, 1839. (b) Farina, V.; Brown, J. D. Angew. Chem., Int. Ed. 2006, 45, 7330–7334. (c) Magano, J. Chem. Rev. 2009, 109, 4398. (d) Andraos, J. Org. Process. Res. Dev. 2009, 13, 161. (e) Magano, J. Tetrahedron 2011, 67, 7875.
- (13) (a) Mita, T.; Fukuda, N.; Roca, F. X.; Kanai, M.; Shibasaki, M. Org. Lett. 2007, 9, 259. (b) Yamatsugu, K.; Kamijo, S.; Suto, Y.; Kanai, M.; Shibasaki, M. Tetrahedron Lett. 2007, 46, 1403. (c) Yamatsugu, K.; Yin, L.; Kamijo, S.; Kimura, Y.; Kanai, M.; Shibasaki, M. Angew. Chem., Int. Ed. 2009, 48, 1070. (d) Yamatsugu, K.; Kanai, M.; Shibasaki, M. Tetrahedron 2009, 65, 6017.
- (14) (a) Gommermann, N.; Koradin, X.; Polborn, K.; Knochel, P. *Angew. Chem., Int. Ed.* **2003**, 42, 5763. (b) Gommermann, N.; Knochel, P. *Chem.—Eur. J.* **2006**, 12, 4380. (c) Gommermann, N.; Knochel, P. *Tetrahedron* **2005**, 61, 11418.
- (15) Valk, J. M.; Claridge, T. D. W.; Brown, J. M.; Hibbs, D.; Hursthouse, M. B. *Tetrahedron: Asymmetry* **1995**, *6*, 2597.
- (16) Knopfel, T. F.; Aschwanden, P.; Ichikawa, T.; Watanabe, T.; Carreira, E. M. Angew. Chem., Int. Ed. 2004, 43, 5971.
- (17) (a) Nishiyama, H.; Sakaguchi, H.; Nakamura, T.; Horihata, M.; Kondo, M.; Itoh, K. *Organometallics* **1989**, *8*, 846. (b) Desimoni, G.; Faita, G.; Quadrelli, P. *Chem. Rev.* **2003**, *103*, 3119.
- (18) Gommermann, N.; Knochel, P. Chem. Commun. 2005, 4175.
- (19) Trost, B. M.; Braslau, R. Tetrahedron Lett. 1989, 30, 4657.
- (20) Garrohelion, F.; Merzouk, A.; Guibe, F. J. Org. Chem. 1993, 58, 6109.
- (21) Hu, J.; Miller, M. J. J. Am. Chem. Soc. 1997, 119, 3462.
- (22) Ishizumi, K.; Koga, K.; Yamada, S.-i. Chem. Pharm. Bull. 1968, 16, 492.
- (23) (a) Ando, K. J. Org. Chem. 1999, 64, 8406. (b) Ando, K.; Oishi, T.; Hirama, M.; Ohno, H.; Ibuka, T. J. Org. Chem. 2000, 65, 4745.
- (24) Still, W. C.; Gennari, C. Tetrahedron Lett. 1983, 24, 4405.
- (25) Izquierdo, J.; Rodriguez, S.; Gonzalez, F. V. Org. Lett. 2011, 13, 3856.