

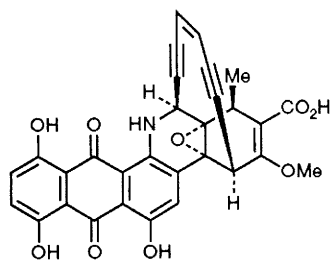
Synthesis and Biological Actions of Optically Active Enediynes Related to Dynemicin A

K. C. Nicolaou,* Y. P. Hong, W.-M. Dai, Z.-J. Zeng and W. Wrasidlo

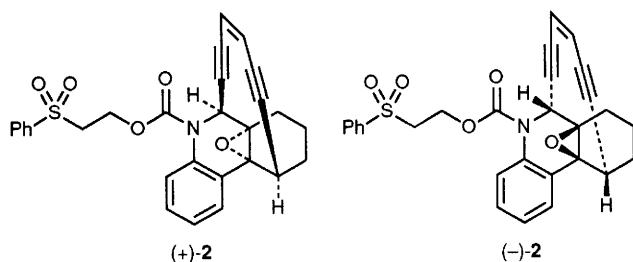
Department of Chemistry, 10666 N. Torrey Pines Road, The Scripps Research Institute, La Jolla, California 92037, USA
Department of Chemistry, 9500 Gilman Drive, University of California, San Diego, La Jolla, California 92093, USA

Enantiomerically pure enediynes (+)-**2** and (–)-**2** were synthesized using a chiral resolution method and were shown to exhibit different cytotoxicities against a number of tumour cells.

Dynemicin A **1**¹ is a recently discovered antitumour antibiotic of the enediyne class.² Its unique molecular architecture, potent antitumour activity and fascinating mechanism of action have stimulated extensive studies during the past few years.² Reports from these laboratories included disclosures on the synthesis and biology of a number of novel model systems of this natural product.^{3,4} Designed enediyne **2** equipped with a 2-(phenylsulfonyl)ethoxycarbonyl group represents a family of highly potent and selective cytotoxic agents as demonstrated in experiments with a variety of cell



1: dynemicin A



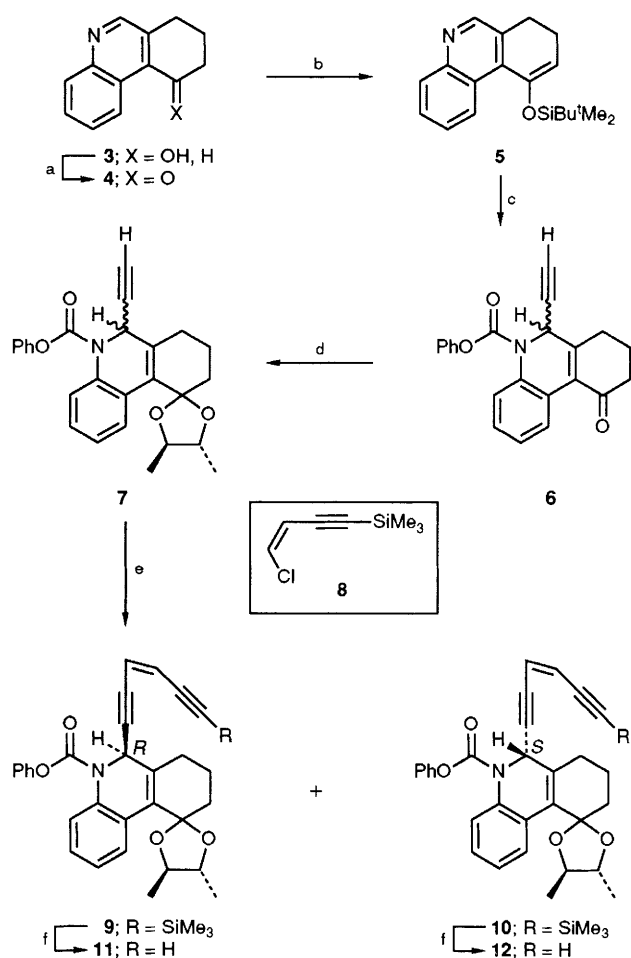
lines. Thus, racemic **2** exhibited DNA cleaving properties and an IC_{50} value of 10^{-11} mol dm^{-3} against Molt-4 leukaemia cells.^{3e} Owing to this significant observation it was deemed important to prepare pure enantiomers of **2** and test their biological action. In this communication we report the synthesis of enantiomers (+)-**2** and (–)-**2** and their biological action against a number of cell lines.

Since both enantiomers of **2** were desired for biological studies, a rapid access to both compounds through a resolution method was sought. Thus, the original synthesis of racemic **2**^{3a-e} was significantly modified as outlined in Scheme 1 to produce (+)-**2** and (–)-**2** in pure forms. Hydroxy quinoline **3** was oxidized to ketone **4**[†] using Jones reagent (98%) and then converted to enol silyl ether **5** in high yield. Sequential treatment of **5** with phenyl chloroformate and ethynylmagnesium bromide afforded, after acidic work-up, acetylenic compound **6** in 91% overall yield. Ketalization of **6** with (2*R*,3*R*)-butane-2,3-diol gave an inseparable mixture of diastereoisomers **7** (*ca.* 1 : 1 by ¹H NMR) which was coupled with vinyl chloride **8** under the influence of Pd⁰–Cu^I catalysis to afford a 1 : 1 mixture of enediynes **9** and **10** (63% total yield). Flash column chromatography (silica gel, 0.25% ethyl acetate in benzene) led to pure diastereoisomers **9** ($R_f = 0.22$ (silica gel, 0.25% ethyl acetate in benzene); $[\alpha]_D^{25} + 427$ (*c* 0.88, benzene)) and **10** ($R_f = 0.20$ (silica gel, 0.25% ethyl acetate in benzene); $[\alpha]_D^{25} - 397$ (*c* 0.95, benzene)) in 45 and 42% yield, respectively. Removal of the trimethylsilyl group from **9**

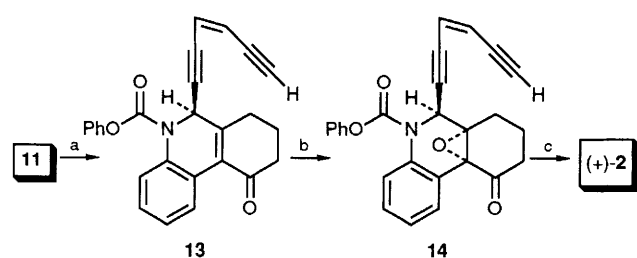
[†] All new compounds exhibited satisfactory spectral and analytical and/or exact mass data. Yields refer to chromatographically and spectroscopically homogeneous materials.

Table 1 Cytotoxicities of enediynes ($\pm$)-**2**, (+)-**2** and (–)-**2**

Cell type	Cell line	IC ₅₀ /mol dm ^{–3}		
		($\pm$)- 2	(+)- 2	(–)- 2
Melanoma	SK-Mel-28	6.3×10^{-6}	6.3×10^{-6}	6.3×10^{-6}
Pancreatic carcinoma	Capan-1	1.6×10^{-6}	3.9×10^{-7}	1.6×10^{-6}
Breast carcinoma	MCF-7/ADR ^a	1.6×10^{-6}	7.8×10^{-7}	1.6×10^{-6}
Promyelocytic leukemia	HL-60	3.9×10^{-6}	$>9.8 \times 10^{-8}$	7.8×10^{-7}
T-cell leukemia	Molt-4	1.0×10^{-11}	1.0×10^{-13}	1.0×10^{-7}

^a Adriamycin resistant cell line.

and **10** led to enediynes **11** and **12** in high yields. Assignment of absolute stereochemistry in this series was based on X-ray crystallographic analysis of **12**. Transformation of the diastereoisomeric compounds **11** and **12** into the targeted molecules (+)-**2** and (–)-**2** was carried out as illustrated in Scheme 2 for the synthesis of (+)-**2**. Thus, acid hydrolysis of



Scheme 2 Reagents and conditions: (a) 0.2 equiv. of *p*-MeC₆H₄SO₃H·H₂O, PhH–acetone–H₂O (100:1:1), reflux, 6 h, 85%; (b) 2.0 equiv. of *m*CPBA, aqueous NaHCO₃–CH₂Cl₂ (1:1), 25 °C, 1.5 h, **14**, 37% plus 13% of recovered **13**; (c) see ref. 3a–c

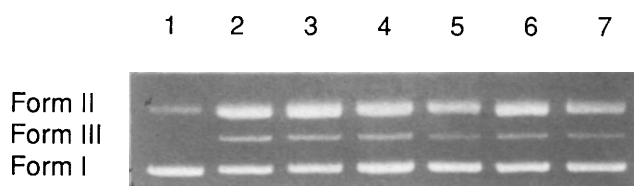


Fig. 1 Supercoiled DNA interaction with synthesized enediynes. Φ X174 DNA was incubated for 24 h at 37 °C in buffer (50 mmol dm^{–3} Tris–HCl, pH 8.5) with compounds ($\pm$)-**2**, (+)-**2**, and (–)-**2**. Lane 1: DNA control. Lanes 2–4: ($\pm$)-**2**, (+)-**2**, and (–)-**2** (1000 μ mol dm^{–3} each). Lanes 5–7: ($\pm$)-**2**, (+)-**2**, and (–)-**2** (100 μ mol dm^{–3} each). Key: Form I, supercoiled DNA; Form II, nicked DNA; Form III, linear DNA.

ketal **11** afforded enone **13** (85%) which was converted into epoxyketone **14** using chloroperbenzoic acid (*m*CPBA) under basic conditions (43% yield based on 87% conversion). From here on the sequence followed the reported pathway for the racemic series.^{3a–c} It is noteworthy that the enantiomer (+)-**2** had the same absolute stereochemistry⁵ and sign of optical rotation as dynemicin A {(+)-**2**: [α]_D²⁵ +586 (*c* 0.46, benzene); dynemicin A **1**: [α]_D²⁴ +270 (*c* 0.01, dimethylformamide)¹}. Enantiomer (–)-**2** {[α]_D²⁵ –562 (*c* 0.50, benzene)} was synthesized from **12** in a similar manner.

As shown in Fig. 1 compounds ($\pm$)-**2**, (+)-**2**, and (–)-**2** cleaved Φ X174 supercoiled DNA under basic conditions (pH 8.5) with comparable potencies (at 1000 and 100 μ mol dm^{–3} concentrations).⁶ More striking were the *in vitro* cytotoxicities of these compounds against tumour cell lines. As exhibited in Table 1 the (+)-enantiomer of **2** exhibited higher potencies against a number of cell lines, particularly the more sensitive Molt-4 leukaemia cells.

The described chemistry renders these enediynes available in their enantiomerically pure forms and provides support for the proposed⁵ absolute stereochemistry of dynemicin A. The biological observations point to rather selective interactions of these molecules with their target cells, particularly in the cases of the most sensitive cell types.

This work was financially supported by the National Institutes of Health (USA) and The Scripps Research Institute.

Received, 16th June 1992; Com. 2/03156F

References

- 1 M. Konishi, H. Ohkuma, K. Matsumoto, T. Tsuno, H. Kamei, T. Miyaki, T. Oki, H. Kawaguchi, G. D. VanDuyne and J. Clardy, *J. Antibiot.*, 1989, **42**, 1449; M. Konishi, H. Ohuma, T. Tsuno, T. Oki, G. D. VanDuyne and J. Clardy, *J. Am. Chem. Soc.*, 1990, **112**, 3715.
- 2 For a review, see: K. C. Nicolaou and W.-M. Dai, *Angew. Chem., Int. Ed. Engl.*, 1991, **30**, 1387.
- 3 (a) K. C. Nicolaou, C.-K. Hwang, A. L. Smith and S. V. Wendeborn, *J. Am. Chem. Soc.*, 1990, **112**, 7416; (b) K. C. Nicolaou, A. L. Smith, S. V. Wendeborn and C.-K. Hwang, *J. Am. Chem. Soc.*, 1991, **113**, 3106; (c) K. C. Nicolaou, W.-M. Dai, S. V. Wendeborn, A. L. Smith, Y. Torisawa, P. Maligres and C.-K. Hwang, *Angew. Chem., Int. Ed. Engl.*, 1991, **30**, 1302; (d) K. C. Nicolaou, Y.-P. Hong, Y. Torisawa, S.-C. Tsay and W.-M. Dai, *J. Am. Chem. Soc.*, 1991, **113**, 9878; (e) K. C. Nicolaou, W.-M. Dai, S.-C. Tsay, V. A. Estevez and W. Wrasidlo, *Science*, 1992, **256**, 1172; (f) K. C. Nicolaou, E. P. Schreiner and W. Stahl, *Angew. Chem., Int. Ed. Engl.*, 1991, **30**, 585; (g) K. C. Nicolaou, E. P. Schreiner, Y. Iwabuchi and T. Suzuki, *Angew. Chem., Int. Ed. Engl.*, 1992, **31**, 340.
- 4 For a number of other selected contributions to the synthesis of dynemicin A models, see: J. A. Porco, Jr., F. J. Schoenen, T. J. Stout, J. Clardy and S. L. Schreiber, *J. Am. Chem. Soc.*, 1990, **112**, 7410; P. A. Wender and C. K. Zercher, *J. Am. Chem. Soc.*, 1991, **113**, 2311; P. Magnus and S. M. Fortt, *J. Chem. Soc., Chem. Commun.*, 1991, 544; T. Nishikawa, A. Ino, M. Isobe and T. Goto, *Chem. Lett.*, 1991, 1271.
- 5 The absolute stereochemistry of dynemicin A was suggested based on a working model of its interaction with DNA, see: D. R. Landley, T. W. Doyle and D. L. Beveride, *J. Am. Chem. Soc.*, 1991, **113**, 4395; P. A. Wender, R. C. Kelly, S. Beckham and B. L. Miller, *Proc. Natl. Acad. Sci. USA*, 1991, **88**, 8835.
- 6 These results may arise from the lack of an extended aromatic ring skeleton in these compounds as compared with dynemicin A, which was proposed to intercalate into DNA prior to drug activation, see: Y. Sugiura, T. Shiraki, M. Konishi and T. Oki, *Proc. Natl. Acad. Sci. USA*, 1990, **87**, 3831.