

Design of HIV-1 Protease Inhibitors with Pyrrolidinones and Oxazolidinones as Novel P1'-Ligands To Enhance Backbone-Binding Interactions with Protease: Synthesis, Biological Evaluation, and Protein–Ligand X-ray Studies[∞]

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Structure-based design, synthesis, and biological evaluation of a series of novel HIV-1 protease inhibitors are described. In an effort to enhance interactions with protease backbone atoms, we have incorporated stereochemically defined methyl-2-pyrrolidinone and methyl oxazolidinone as the P1'-ligands. These ligands are designed to interact with Gly-27' carbonyl and Arg-8 side chain in the S1'-subsite of the HIV protease. We have investigated the potential of these ligands in combination with our previously developed bis-tetrahydrofuran (bis-THF) and cyclopentanyltetrahydrofuran (Cp-THF) as the P2-ligands. Inhibitor **19b** with a (*R*)-aminomethyl-2-pyrrolidinone and a Cp-THF was shown to be the most potent compound. This inhibitor maintained near full potency against multi-PI-resistant clinical HIV-1 variants. A high resolution protein–ligand X-ray crystal structure of **19b**-bound HIV-1 protease revealed that the P1'-pyrrolidinone heterocycle and the P2-Cp-ligand are involved in several critical interactions with the backbone atoms in the S1' and S2 subsites of HIV-1 protease.

Introduction

Advances in the treatment of HIV/AIDS with HIV-1 protease inhibitors in combination with reverse transcriptase inhibitors have been widely documented.¹ The combination therapy, also known as highly active antiretroviral therapy (HAART), blocks critical viral replication at two different stages of the replication cycle.² The HAART regimens have resulted in dramatic reduction of blood plasma viral load levels, increased CD4⁺ lymphocyte counts, and improved life expectancy and significantly reduced HIV/AIDS-related mortality in the developed world.³ Despite these important advances, effective long-term suppression of HIV infection with HAART regimens is a complex issue in medicine for a number of reasons. These include drug side effects, poor penetration into protected HIV reservoir sites, poor oral bioavailability, and interactions between drugs.⁴ Perhaps one of the most daunting problems in future management of HIV is the emergence of drug-resistant HIV-1 variants and the transmission of these viral strains.^{5,6} Thus, development of antiretroviral therapy with broad-spectrum activity and minimal drug side effects is critical for an effective management of current and future HIV/AIDS treatment. We recently reported the design and development of a number of exceedingly potent nonpeptidic HIV-1 protease inhibitors (PIs) **1–3** (Figure 1).^{7–9} One of those PIs is darunavir (**1**, TMC-

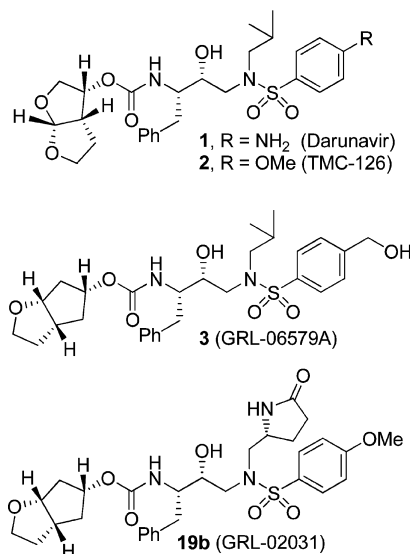


Figure 1. Structures of inhibitors **1–3** and **19b**.

114), which was approved by the FDA in 2006 for the treatment of HIV/AIDS patients who are harboring drug-resistant HIV and do not respond to other therapies.¹⁰ More recently, darunavir has received full approval for all HIV/AIDS patients.¹¹

To combat drug resistance, our structure-based design strategies are to maximize the protease active-site interactions with the inhibitor and particularly to promote extensive hydrogen bonding with the protein backbone atoms.¹² It is evident that active site backbone conformation of mutant proteases is only minimally distorted compared to that of the wild-type HIV-1 protease.^{13,14} Therefore, the “backbone binding” strategy may be important to combat drug resistance.¹² Using high resolution protein–ligand X-ray structures of **1**- and **3**-bound HIV-1

[∞] The PDB accession code for **19b**-bound HIV-1 protease X-ray structure is 3H5B.

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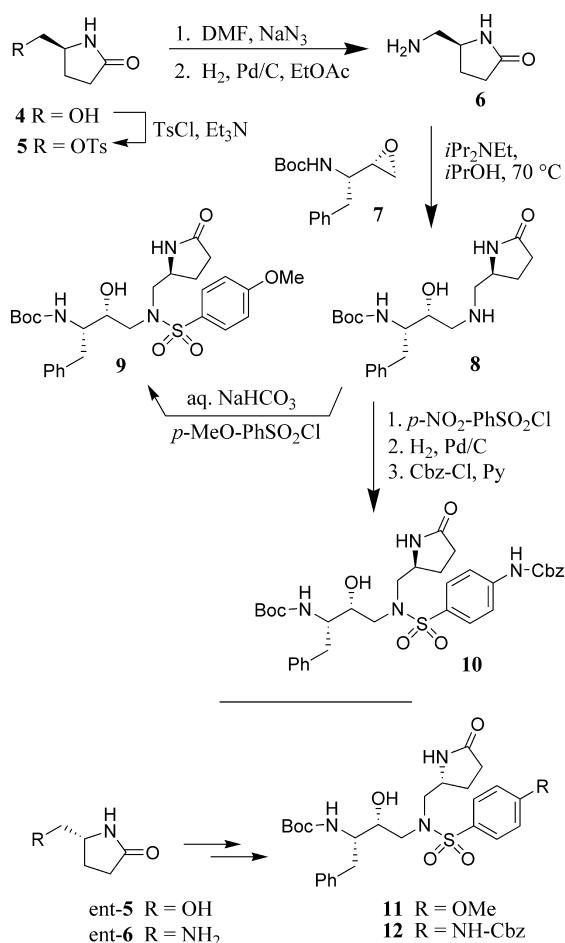
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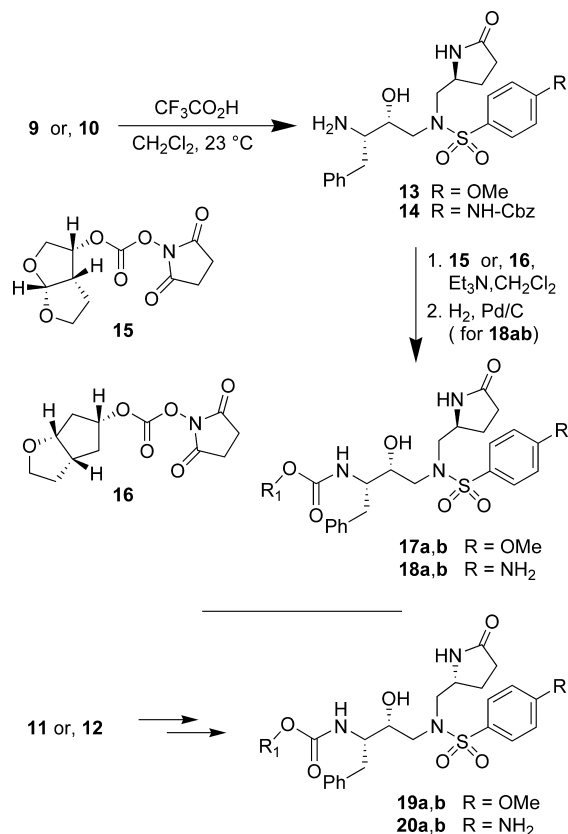
Abbreviations: HIV, human immunodeficiency virus; bis-THF, bis-tetrahydrofuran; Cp-THF, cyclopentanyltetrahydrofuran; PI, protease inhibitor; HAART, highly active antiretroviral therapy; APV, amprenavir; DRV, darunavir; SQV, saquinavir; IDV, indinavir; LPV, lopinavir; RTV, ritonavir.

Scheme 1. Synthesis of Lactam Containing Sulfonamide Isosteres

protease, we have shown that these PIs were engaged in extensive hydrogen bonding interactions with the backbone atoms throughout the active site cavity from the S2 to S2' regions.^{9,15} To further enhance "backbone binding" interactions, we became interested in designing an appropriately functionalized P1'-ligand that could interact with the backbone atoms, particularly with the Gly-27' and Arg-8 in the S1'-subsite. This enhancement of "backbone binding" interaction may lead to inhibitors with improved drug-resistance profiles. Herein, we report the design, synthesis, and biological evaluation of a series of potent HIV-1 protease inhibitors that incorporated structure-based designed stereochemically defined lactam and oxazolidinone derivatives as the P1'-ligands in combination with the bis-THF or Cp-THF as the P2-ligands. Inhibitor **19b** incorporating a (*R*)-5-aminomethyl-2-pyrrolidinone as the P1'-ligand and Cp-THF as the P2-ligand is the most potent PI in the series. Interestingly, this PI has retained full potency against a range of multidrug-resistant HIV-1 variants. The protein-ligand X-ray structure of **19b**-bound HIV-1 protease revealed important molecular insight into the ligand-binding site interactions.

Chemistry

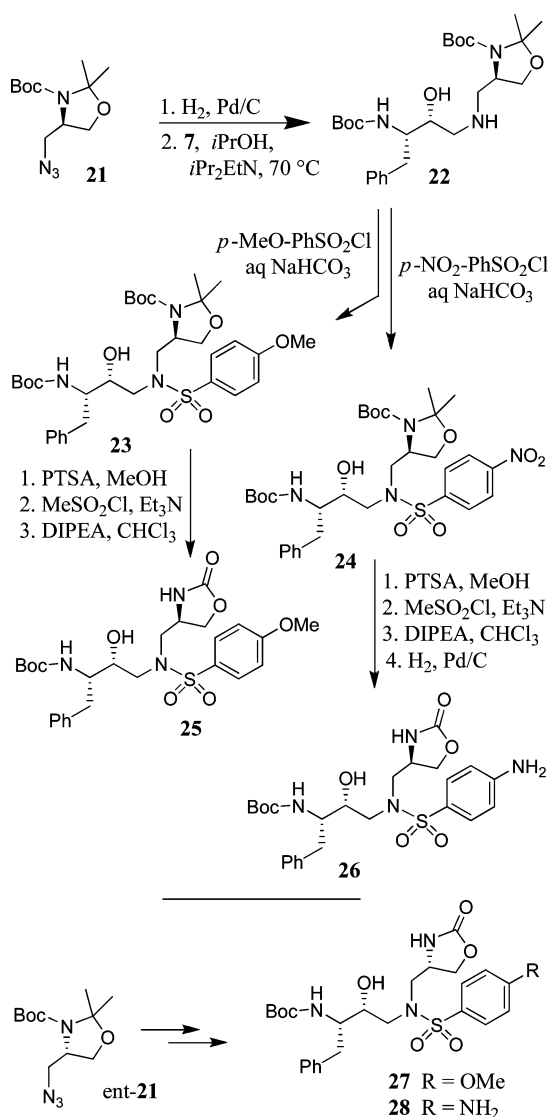
The optically active synthesis of the requisite 5-aminomethyl-2-pyrrolidinone for P1'-ligands and their conversion to respective sulfonamide isostere are shown in Scheme 1. Commercially available 5-(*S*)-hydroxymethyl-2-pyrrolidinone **4** was reacted with tosyl chloride and triethylamine to provide tosylate **5**. Displacement of the tosylate with sodium azide in DMF at 55 °C for 9 h provided the azide derivative in 92% yield over two

Scheme 2. Synthesis of Lactam Containing PIs

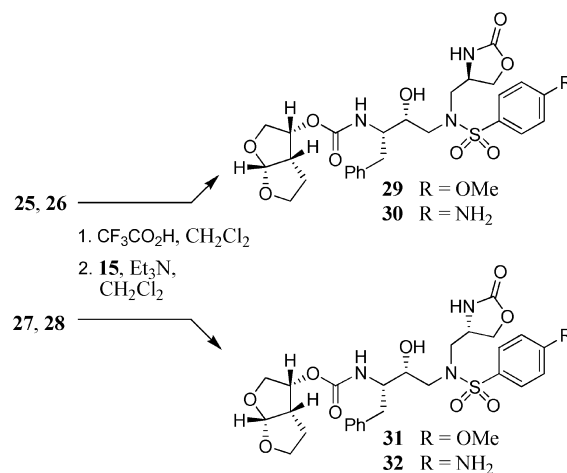
steps. Catalytic hydrogenation of the azide over 10% Pd-C in ethyl acetate afforded optically active amine **6** in quantitative yield. 5-(*R*)-Hydroxymethyl-2-pyrrolidinone (*ent*-**5**) was similarly converted to optically active amine *ent*-**6** in comparable yield. Amine **6** was reacted with commercially available epoxide **7** in the presence of *i*-Pr₂NEt (DIPEA) in 2-propanol at 70 °C for 36 h to provide epoxide-opened product **8** in 85% yield.¹⁶ Amine **8** was converted to *p*-methoxybenzenesulfonamide derivative **9** by reaction with *p*-methoxybenzenesulfonyl chloride in the presence of aqueous NaHCO₃ in quantitative yield. Treatment of amine **8** with *p*-nitrobenzenesulfonyl chloride afforded the corresponding nitrosulfonamide. Catalytic hydrogenation over 10% Pd-C gave the corresponding aniline derivative, which was reacted with benzyl chloroformate in the presence of pyridine to furnish Cbz-derivative **10** in 63% yield for three steps. Enantiomeric amine (*ent*-**6**) was converted to the respective methoxy and Cbz-derived **11** and **12** by analogous procedures.

The synthesis of various PIs incorporating methylpyrrolidinones as the P1'-ligand is shown in Scheme 2. Exposure of Boc-derivatives **9** and **10** to 30% CF₃CO₂H in CH₂Cl₂ at 23 °C for 40 min resulted in the respective amines **13** and **14**. Alkoxy-carbonylation of amine **13** with activated mixed carbonates **15**¹⁶ and **16**⁹ in the presence of Et₃N in CH₂Cl₂ furnished inhibitors **17a** and **17b** in 98% and 87% yields, respectively.¹⁷ Alkoxy-carbonylation of amine **14** with activated carbonates **15** and **16** afforded the corresponding Cbz-protected urethanes. Removal of the Cbz-group by catalytic hydrogenation over 10% Pd-C in ethyl acetate provided inhibitor **18a** and **18b** in 58% and 62% yields, respectively. Sulfonamide derivatives **11** and **12** containing enantiomeric P1'-ligands were converted to inhibitors **19a,b** and **20a,b** by following analogous procedures.

The synthesis of sulfonamide isosteres incorporating methyl-oxazolidinone as the P1'-ligand is shown in Scheme 3. Optically

Scheme 3. Synthesis of Sulfonamide Isosteres with P₁'-Oxazolidinone

active dimethyloxazolidines **21** and *ent-21* were prepared by following the procedure described by Dondini and co-workers.¹⁸ Reduction of these azides by catalytic hydrogenation in methanol afforded the respective amine. Reaction of **21**-derived amine with epoxide **7** in the presence of $i\text{-Pr}_2\text{NEt}$ in 2-propanol afforded amine **22** in 41% yield. Reaction of amine **22** with *p*-methoxybenzenesulfonyl chloride or *p*-nitrobenzenesulfonyl chloride as described previously afforded sulfonamide derivatives **23** and **24** in 80% and 92% yields, respectively. The isopropylidene functionality in **23** and **24** was converted to the corresponding oxazolidinone derivative in a three-step sequence involving (1) treatment of **23** by a catalytic amount of *p*-toluenesulfonic acid (PTSA) in methanol, resulting in the removal of the isopropylidene group, (2) reaction of the resulting Boc-amino alcohol with mesyl chloride in the presence of triethylamine to provide the corresponding mesylate, and (3) treatment of the resulting mesylate with $i\text{-Pr}_2\text{NEt}$ in chloroform at reflux. This has provided oxazolidinone **25** in 45% yield over three steps. The nitrosulfonamide derivative **24** was similarly converted to the corresponding oxazolidinone. Catalytic hydrogenation of the resulting nitro derivative with 10% Pd-C in methanol provided aniline derivative **26** in 37% overall yield

Scheme 4. Synthesis of Oxazolidinone-Derived PIs

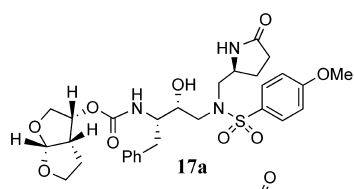
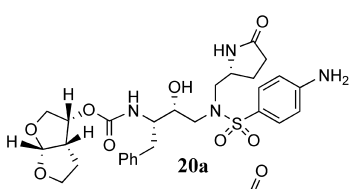
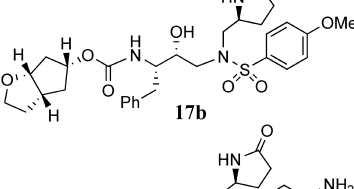
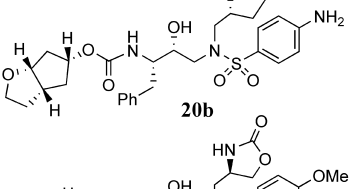
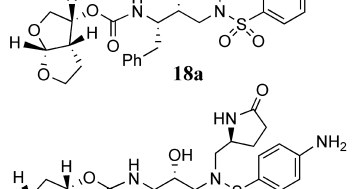
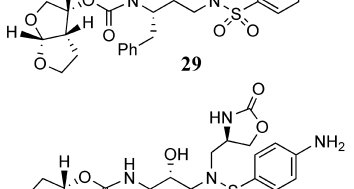
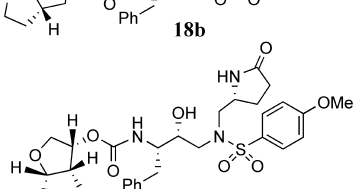
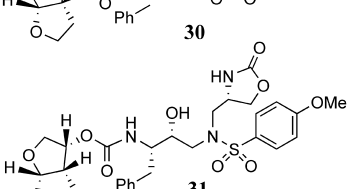
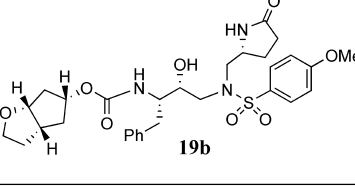
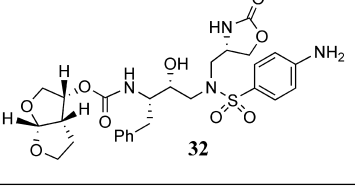
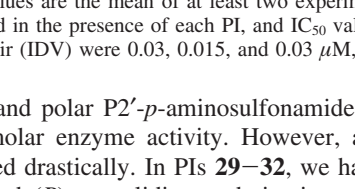
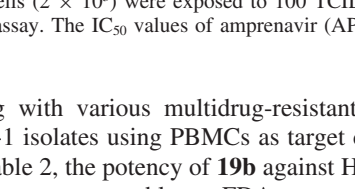
over four steps. Enantiomeric azide *ent-21* was converted to oxazolidinone derivatives **27** and **28** by following analogous procedures.

The synthesis of inhibitors containing oxazolidinone as P₁'-ligand and bis-THF as the P₂-ligand is shown in Scheme 4. Treatment of oxazolidinones **25**–**28** with 30% $\text{CF}_3\text{CO}_2\text{H}$ in CH_2Cl_2 at 23°C afforded the corresponding amines. Reaction of the resulting amines with activated mixed carbonate **15** in the presence of Et_3N in CH_2Cl_2 afforded the target inhibitors **29**–**32** in excellent yields (80–90%). The structures of these inhibitors are shown in Table 1.

Results and Discussion

Our examination of the X-ray structure of 1-bound HIV-1 protease and its respective modeling initially suggested that a methyl-2-pyrrolidinone may interact well with residues in the S₁'-site.¹⁵ As shown in Table 1, our first set of inhibitors contain a (*R*)-hydroxyethylamine sulfonamide isostere with either the bis-THF or Cp-THF as the P₂-ligand and *p*-methoxysulfonamide or *p*-aminosulfonamide as the P₂'-ligand. The enzyme inhibitory potency of these PIs was evaluated according to the procedure reported by Toth and Marshall.¹⁹ Inhibitor **17a** with (*S*)-methyl-2-pyrrolidinone displayed an enzyme K_i of 1 nM. Inhibitor **17b** with a Cp-THF showed a 3-fold improvement of potency. Antiviral activity of these inhibitors was determined in MT-2 human T-lymphoid cells exposed to HIV-1_{LAI}. Interestingly, both inhibitors have shown dramatic reduction in antiviral activity. Inhibitors **17a** and **17b** have shown IC₅₀ values of 0.48 and 0.23 μM , respectively. However, these inhibitors are significantly less potent compared to inhibitors with an isobutyl group as the P₁'-ligand.^{7c,9} Incorporation of *p*-aminosulfonamide (PIs **18a** and **18b**) as the P₂'-ligand led to a drop in enzyme inhibitory as well as antiviral potency. Inhibitor **19a** containing (*R*)-methyl-2-pyrrolidinone as the P₁'-ligand has shown 10-fold enhancement of enzyme K_i over the (*S*)-isomer **17a**. It showed a slight improvement in antiviral activity compared to inhibitor **17a**. Inhibitor **19b** with (*R*)-methyl-2-pyrrolidinone as the P₁'-ligand and Cp-THF as the P₂-ligand resulted in the most potent inhibitor in the series. It has shown an enzymatic K_i of 99 pM and a 10-fold improvement (IC₅₀ = 0.026 μM) in antiviral activity relative to epimeric (*S*)-pyrrolidinone derivative **17b**, suggesting an important role for the P₁'-ring stereochemistry. Indeed, an X-ray structure of **19b**-bound HIV-1 protease revealed that the pyrrolidinone carbonyl and the NH functionalities were positioned to hydrogen-bond with residues in the S₁'-site. Interestingly, the combination of P₁'-methylpyrrolidi-

Table 1. Enzymatic Inhibitory Activity of Lactam and Oxazolidinone Containing Inhibitors^b

Entry	Inhibitor	K _i (nM)	IC ₅₀ (nM) ^a	Entry	Inhibitor	K _i (nM)	IC ₅₀ (nM) ^a
1.		0.85±0.02	0.48±0.05	7.		0.85±0.2	>1
2.		0.31±0.03	0.23±0.08	8.		0.31±0.03	0.60±0.24
3.		0.28±0.03	>1	9.		0.28±0.03	0.48±0.17
4.		1.27±0.15	>1	10.		0.31±0.03	>1
5.		0.12±0.003	0.25±0.11	11.		0.035±0.01	0.31±0.21
6.		0.099±0.003	0.026±0.002	12.		0.24±0.03	>1

^a Values are the mean of at least two experiments. ^b Human T-lymphoid (MT-2) cells (2×10^3) were exposed to 100 TCID₅₀ values of HIV-1_{LA1} and cultured in the presence of each PI, and IC₅₀ values were determined using the MTT assay. The IC₅₀ values of amprenavir (APV), saquinavir (SQV), and indinavir (IDV) were 0.03, 0.015, and 0.03 μ M, respectively.

none and polar P2'-*p*-aminosulfonamide led to PIs with subnanomolar enzyme activity. However, antiviral activity was reduced drastically. In PIs **29–32**, we have incorporated both (*S*)- and (*R*)-oxazolidinone derivatives as substitutes for the respective pyrrolidinone isomers. As can be seen, oxazolidinone derivatives **29–32** have shown subnanomolar enzyme inhibitory potency. Inhibitors with *p*-methoxysulfonamide as the P2'-ligand displayed comparable antiviral activity relative to pyrrolidinone derivatives. Consistent with stereochemical preference, the (*R*)-oxazolidinone with *p*-methoxysulfonamide has shown better enzyme K_i values. However, the antiviral activity of these compounds is very similar. In general, both pyrrolidinone and oxazolidinone functionalities appear to be nicely accommodated in the S1'-site.

While inhibitor **31** is very potent in enzyme inhibitory assay, the significant reduction of antiviral potency is possibly due to poor cellular permeability of this polar functionality. Inhibitor **19b** appeared to be most potent among the series of inhibitors examined. It exhibited comparable antiviral activity with the FDA approved PIs amprenavir, saquinavir, and indinavir in the same assay.

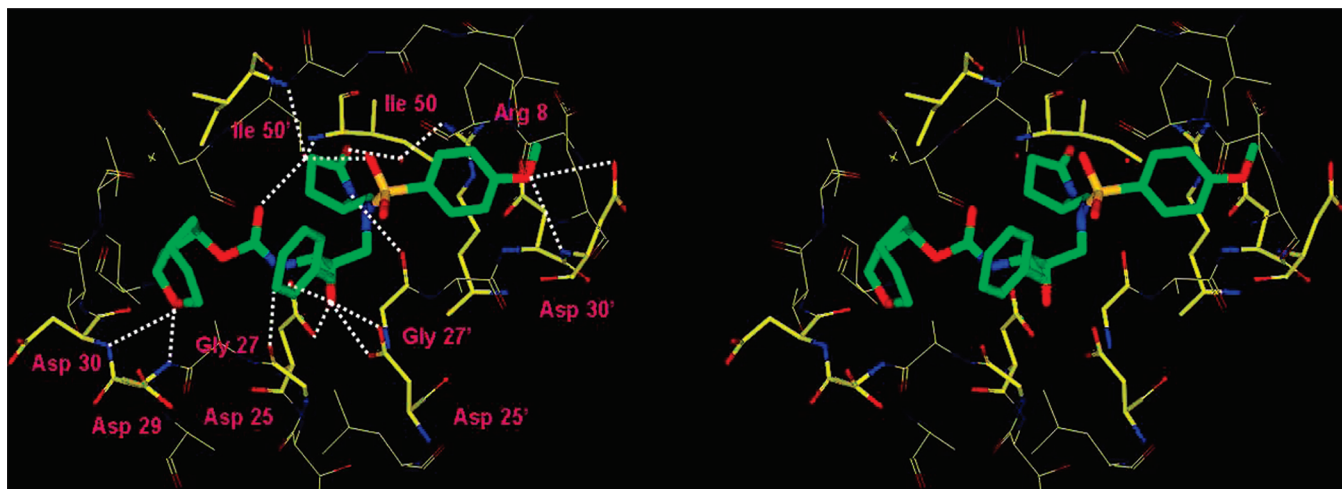
Inhibitor **19b** was subsequently examined for its activity against a clinical wild-type X₄-HIV-1 isolate (HIV-1_{ERS104pre})

along with various multidrug-resistant clinical X₄- and R₅-HIV-1 isolates using PBMCs as target cells.^{8b} As can be seen in Table 2, the potency of **19b** against HIV-1_{ER104pre} (IC₅₀ = 28 nM) was comparable to FDA approved PIs indinavir, amprenavir, and lopinavir with IC₅₀ values of 28, 25, and 30 nM, respectively. Darunavir, on the other hand, is nearly 10-fold more potent (IC₅₀ = 3.6 nM) than **19b** and the above-mentioned PIs. Interestingly, of all the PIs tested, indinavir was least able to suppress the replication of the multidrug-resistant clinical isolate examined (HIV-1_{MDR/MM}, HIV-1_{MDR/TM}, HIV-1_{MDR/C}, and HIV-1_{MDR/G}) with IC₅₀ values greater than 1 μ M. Both amprenavir and lopinavir displayed 10-fold or greater reduction in potency except against HIV-1_{MDR/G}, where lopinavir showed a 5-fold reduction in potency. A more detailed virologic study using inhibitor **19b** will be published elsewhere.²⁰ Darunavir has maintained impressive activity against all the multidrug-resistant variants. Inhibitor **19b**, while less potent than darunavir, maintained near full potency against multidrug-resistant clinical isolates examined. This impressive drug-resistance property of **19b** is possibly due to its extensive interactions, particularly its ability to make extensive hydrogen bonding throughout the active site of the protease's backbone. Furthermore, inhibitor **19b** blocked the infection and replication

Table 2. Anti-HIV Activity of **19b** against Selected Clinical Isolates Highly Resistant to Multiple Protease Inhibitors^a

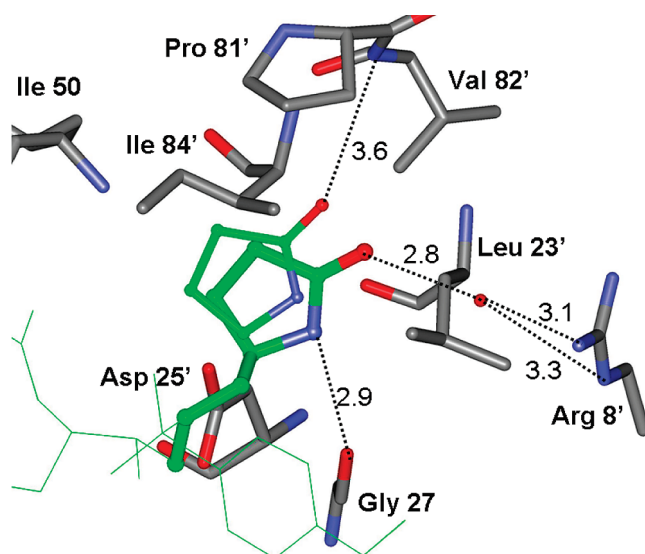
virus	phenotype	EC ₅₀ (μM)				
		IDV	APV	LPV	DRV	19b
HIV-1 _{ERS104pre} (wild-type)	X4	0.028 ± 0.005	0.025 ± 0.006	0.03 ± 0.001	0.0036 ± 0.0002	0.028 ± 0.004
HIV-1 _{TM} (MDR)	X4	>1 (>36)	0.25 ± 0.02 (10)	0.73 ± 0.53 (24)	0.0036 ± 0.0002 (1)	0.029 ± 0.004 (1)
HIV-1 _{MM} (MDR)	R5	>1 (>36)	0.32 ± 0.03 (13)	0.72 ± 0.31 (24)	0.019 ± 0.009 (5)	0.042 ± 0.002 (2)
HIV-1 _C (MDR)	X4	>1 (>36)	0.35 ± 0.03 (14)	0.32 ± 0.01 (11)	0.015 ± 0.001 (4)	0.023 ± 0.007 (1)
HIV-1 _G (MDR)	X4	0.29 ± 0.07 (10)	0.33 ± 0.16 (13)	0.14 ± 0.01 (5)	0.014 ± 0.006 (4)	0.027 ± 0.001 (1)

^a Amino acid substitutions identified in the protease-encoding regions of HIV-1_{ERS104pre}, HIV-1_{TM}, HIV-1_{MM}, HIV-1_C, and HIV-1_G compared to the consensus B sequence cited from the Los Alamos data base include L63P, L10I/K14R/R41K/M46L/I54V/L63P/A71V/V82A/L90M/I93L, L10I/K43T/M46L/I54V/L63P/A71V/V82A/L90M/Q92K, L10I/I15V/K20R/L24I/M36I/M46L/I54V/I62V/L63P/K70Q/V82A/L89M, and L10I/V11I/T12E/I15V/L19I/R41K/M46L/L63P/A71T/V82A/L90M, respectively. The EC₅₀ values were determined by employing PHA-PBM as target cells and the inhibition of p24 *Gag* protein production as an end point. All values were determined in duplicate or triplicate, and those shown are derived from the results of three independent experiments. Numbers in parentheses represent fold changes of EC₅₀ values against each isolate compared to EC₅₀ values against HIV-1_{ERS104pre}. MDR: multidrug-resistant.

**Figure 2.** Stereoview of the major conformation of the X-ray structure of inhibitor **19b**-bound HIV-1 protease.

of each of the HIV-1_{NL4-3} variants exposed to and selected by up to 5 μM saquinavir, amprenavir, indinavir, nelfinavir, or ritonavir and a 1 μM lopinavir or atazanavir with EC₅₀ values ranging from 0.036 to 0.14 μM.²⁰

X-ray Crystallography. The binding mode of inhibitor **19b** was determined from the X-ray crystal structure of its complex with wild-type HIV-1 protease. The crystal structure was solved and refined at 1.29 Å resolution with an *R* factor of 14.1%. In this high resolution structure, the inhibitor was bound to the HIV-1 protease active site in two orientations with the relative occupancy of 0.8/0.2. The protease dimer comprises residues 1–99 and 1'–99' of the two subunits, and the inhibitor binding site is formed by both subunits. The P1'-pyrrolidine ring also showed two alternative conformations with equal occupancy and related by about 18° rotation around the C12–C13 bond. A stereoview of the major conformation is shown in Figure 2 (only one conformation is shown for P1'). As shown, extensive interactions from P2 to P2' were observed between the inhibitor and the protease active site, most notably favorable polar interactions including hydrogen bonds, weaker C–H···O and C–H···π interactions. The isostere hydroxyl group forms asymmetric hydrogen bonds to the carboxylate oxygen atoms of the catalytic Asp25 and Asp25' with distances of 2.4–3.3 Å. Also, four direct hydrogen bonds are formed between the oxygens or nitrogens of the inhibitor atoms and the protease backbone atoms. These include cyclic ether oxygen of the P2-Cp-THF and the Asp-29 NH, the urethane NH with the carbonyl oxygen of Gly-27, P2'-methoxy oxygen and Asp-30' NH. One conformation of the P1'-pyrrolidinone formed a hydrogen bond between the NH and the carbonyl oxygen of Gly-27' and a water-mediated hydrogen bond between the P1'-pyrrolidinone

**Figure 3.** Protease interactions with the two alternate conformations of the inhibitor's pyrrolidine ring. The inhibitor is in green with thick bonds for the major and thin bonds for the minor conformations of the pyrrolidine ring. Hydrogen bonds are shown as dotted lines. Distances between donor and acceptor atoms are shown in Å.

carbonyl and the side chain of Arg-8. The other conformation of the P1' group formed hydrophobic and C–H···O interactions with Pro-81' and Val-82', as shown in Figure 3. Also, there exists a tetracoordinated water-mediated interaction where the amides of Ile50 and Ile50' donate hydrogen bonds, and the inhibitor's urethane carbonyl and one of the sulfonamide oxygen

accept hydrogen bonds from the water molecule. These interactions are conserved in a majority of other HIV-1 protease complexes with inhibitors²¹ or substrate analogues.²²

The weaker polar interactions such as C—H...O and water— π interactions can be analyzed accurately in this high resolution structure.^{23,24} These interactions are important for inhibitor-protease binding and must be considered in the design of inhibitors. The C—H...O interactions of the inhibitor with the carbonyl oxygens of Gly-48, Gly-48', and Gly-27' mimic the conserved hydrogen bonds observed with peptide analogue structures.^{21,22} A conserved water— π interaction is observed between the P2' aromatic ring of the inhibitor and the amide of Asp29', which is similar to the interaction with darunavir and other structure-based designed PIs from our laboratories.^{25,26} The bigger polar P1' group of the 2-pyrrolidinone ring in inhibitor **19b** instead of the isobutyl group in PIs **1** and **2** introduces a new direct hydrogen bond with the backbone of HIV-1 protease and one new water-mediated hydrogen bond between the inhibitor and the side chain residue of the protease. The two alternative conformations of the P1' group with occupancy of 0.5/0.5 provide more flexible binding within the S1' subsite, which is likely to enhance the inhibition of resistant proteases.

As mentioned earlier, inhibitor **19b** maintained near full potency against multidrug-resistant clinical isolates examined. On the other hand, **19b** is less potent than darunavir possibly due to the bigger and less optimum size of the P1'-ligand. The design strategy of incorporating new polar interactions with conserved backbone regions of the protease warrants further investigation in light of the current molecular insight into these ligand-binding site interactions.

Conclusion

We have designed a number of HIV-1 protease inhibitors with methyl-2-pyrrolidinone and methyloxazolidinone as the P1'-ligand to enhance hydrogen bonding with the protein backbone atoms in the S1'-subsite. The ligands were synthesized in enantiomerically pure forms, and a series of inhibitors were prepared and evaluated in combination with P2-bis-THF and P2-Cp-THF ligands. In general, these inhibitors exhibited enzyme inhibitory activity lower than the corresponding inhibitors with a P1'-isobutyl group. Our SAR studies indicated the importance of ligand stereochemistry and also preference for the P2-Cp-THF ligand. Interestingly, the polar P1'-ligand influenced their cellular properties. The inhibitors exhibited reduction in antiviral activity possibly due to changes in the molecule's overall lipophilicity. Our investigation resulted in the identification of inhibitor **19b** which has displayed similar antiviral potency as the other FDA approved inhibitors such as indinavir, lopinavir, and amprenavir. Inhibitor **19b**, however, is nearly 10-fold less potent than darunavir. Of particular importance, **19b** has maintained full potency against the examined panel of multidrug-resistant HIV-1 variants. A high resolution X-ray structure of **19b**-bound HIV-1 protease revealed a new hydrogen bonding of the P1'-pyrrolidinone NH with the backbone carbonyl of Gly27'. Also, there is a water mediated hydrogen bond with the pyrrolidinone carboxyl and Arg8' side chain. Furthermore, the P1'-pyrrolidinone showed two alternative conformations that filled the S1' subsite. These new interactions and the conformational flexibility most likely contributed to its impressive properties against multidrug-

resistant clinical variants. Further investigations including optimization of ligand-binding properties are in progress.

Experimental Section

General. All moisture sensitive reactions were carried out under nitrogen or argon atmosphere. Anhydrous solvents were obtained as follows: THF, diethyl ether, and benzene, distilled from sodium and benzophenone; dichloromethane, pyridine, triethylamine, and diisopropylethylamine, distilled from CaH₂. All other solvents were HPLC grade. Silica gel column chromatography was performed with Whatman 240–400 mesh silica gel under low pressure. TLC was carried out with E. Merck silica gel 60-F-254 plates. ¹H and ¹³C NMR spectra were recorded on Varian Mercury 300 and Bruker Avance 400 and 500 spectrometers. Optical rotations were measured using a Perkin-Elmer 341 polarimeter.

(S)-5-(Aminomethyl)-2-pyrrolidinone 6. To a stirred solution of (S)-5-(hydroxymethyl)-2-pyrrolidinone **4** (300 mg, 2.61 mmol) and *p*-toluenesulfonyl chloride (646 mg, 3.34 mmol) in CH₂Cl₂ (6 mL) at 0 °C was added DMAP (64 mg, 0.52 mmol) and Et₃N (472 μ L, 3.34 mmol). The resulting mixture was allowed to warm to 23 °C and stirred for 12 h. The reaction was then quenched with 7 mL of water, and the aqueous layer was extracted with CH₂Cl₂. The combined organic extracts were washed with 1 N HCl and dried over anhydrous Na₂SO₄. Removal of solvent under reduced pressure followed by flash chromatography purification (100% EtOAc as the eluent) yielded (S)-toluenesulfonate **5** (0.7 g, 93%) as a yellowish solid. *R*_f = 0.50 (5% MeOH in CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.75–1.80 (m, 1H), 2.19–2.35 (m, 3H), 2.44 (s, 3H), 3.85–3.92 (m, 2H), 4.00–4.03 (m, 1H), 6.49 (s, 1H), 7.35 (d, 2H, *J* = 8.0 Hz), 7.77 (d, 2H, *J* = 8.1 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 21.6, 22.7, 29.2, 52.5, 71.9, 121.9, 130.0, 132.3, 145.3, 178.2.

To a stirred solution of the above tosylate (638 mg, 2.37 mmol) in DMF (20 mL) was added NaN₃ (462 mg, 2.37 mmol). The resulting solution was stirred at 55 °C for 9 h. Removal of solvent under reduced pressure followed by flash chromatography purification (6% MeOH in CHCl₃ as the eluent) provided the (S)-azidopyrrolidinone (330 mg, 99%) as a yellow oil. *R*_f = 0.50 (10% MeOH in CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.76–1.84 (m, 1H), 2.18–2.44 (m, 3H), 3.28 (dd, 1H, *J* = 6.5, 12.2 Hz), 3.43 (dd, 1H, *J* = 4.6, 12.3 Hz), 3.77–3.83 (m, 1H), 7.38 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 23.9, 29.7, 53.6, 55.8, 178.7.

To a solution of the above azide (125 mg, 0.89 mmol) in EtOAc (10 mL) was added Pd/C (15 mg). The mixture was stirred at 23 °C under a hydrogen filled balloon for 4 h, then filtered over Celite, and the filter cake was washed with EtOAc and MeOH. Removal of solvent under reduced pressure followed by flash chromatography purification (5% MeOH in CHCl₃ as the eluent) afforded the corresponding (S)-amine **6** (105 mg, quantitative) as a yellow oil. *R*_f = 0.05 (20% MeOH in CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.41 (brs 2H), 1.56–1.65 (m, 1H), 2.01–2.12 (m, 1H), 2.19–2.24 (m, 2H), 2.52 (dd, 1H, *J* = 7.5, 12.8 Hz), 2.69 (dd, 1H, *J* = 4.3, 12.9 Hz), 3.50–3.57 (m, 1H), 7.90 (brs, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 24.0, 30.2, 47.3, 57.1, 179.0.

tert-Butyl-(2R,3R)-3-hydroxy-4-[(S)-5-oxopyrrolidin-2-yl]-methylamino]-1-phenylbutan-2-yl-carbamate 8. To a solution of amine **6** (107 mg, 0.94 mmol) in *i*-PrOH (5 mL) were added *tert*-butyl-[S-(R,R)]-(1-oxiran-2-yl)-2-phenylethyl carbamate **7** (62 mg, 0.23 mmol) and DIPEA (204 μ L, 1.2 mmol). The resultant mixture was stirred at 65 °C for 18 h and then concentrated under reduced pressure. Flash chromatography purification (15% MeOH in CHCl₃ as the eluent) yielded title compound **8** (76 mg, 85%). *R*_f = 0.47 (25% MeOH in CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.30 (s, 9H), 1.62–1.71 (m, 1H), 2.13–2.18 (m, 1H), 2.30–2.32 (m, 2H), 2.52 (d, 1H, *J* = 8.86 Hz), 2.64–2.73 (m, 4H), 2.96 (d, 1H, *J* = 9.8 Hz), 3.54 (s, 1H), 3.72–3.75 (m, 4H), 4.99 (brs, 1H), 7.15–7.26 (m, 5H), 8.02 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 24.6, 28.3, 30.2, 36.3, 51.7, 54.0, 54.4, 55.3, 71.5, 79.2, 126.2, 128.2, 129.4, 138.1, 155.9, 178.9; LRMS-ESI (*m/z*) [*M* + Na]⁺ 400.

tert-Butyl-(2R,3R)-3-hydroxy-4-(4-methoxy-N-(((S)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-ylcarbamate 9. To a stirred solution of amine **8** (22 mg, 0.06 mmol) in CH₂Cl₂ (3 mL) and aqueous saturated NaHCO₃ (3 mL) was added 4-methoxybenzenesulfonyl chloride (35.6 mg, 0.17 mmol). This reaction mixture was stirred for 12 h followed by extraction of the aqueous layer with CH₂Cl₂; the combined organic extracts were dried over anhydrous Na₂SO₄. Removal of solvent under reduced pressure followed by flash chromatography purification (6% MeOH in CHCl₃ as the eluent) provided compound **9** (31 mg, quantitative). *R_f* = 0.40 (10% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.28 (s, 9H), 1.54–1.62 (m, 1H), 2.14–2.21 (m, 1H), 2.32–2.35 (m, 2H), 2.68–2.75 (m, 2H), 2.72 (s, 3H), 2.81–2.88 (m, 2H), 2.97–3.03 (m, 3H), 3.64–3.72 (m, 1H), 4.01–4.05 (m, 1H), 5.06 (d, 1H, *J* = 8.9 Hz), 6.93 (d, 2H, *J* = 8.6 Hz), 7.16–7.19 (m, 3H), 7.27–7.28 (m, 3H), 7.61 (d, 2H, *J* = 8.5 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 23.9, 28.1, 29.6, 29.9, 36.1, 53.4, 53.9, 54.5, 55.5, 56.0, 71.9, 79.5, 114.3, 126.3, 126.3, 129.3, 129.5, 137.7, 155.9, 163.1, 178.4; LRMS-ESI (*m/z*) [*M* + Na]⁺ 570.

tert-Butyl-(2R,3R)-4-(4-Cbz-amino-N-(((S)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-ylcarbamate 10. To a stirred solution of amine **8** (93.6 mg, 0.25 mmol) in CH₂Cl₂ (10 mL) and aqueous saturated NaHCO₃ (10 mL) was added 4-nitrobenzenesulfonyl chloride (60 mg, 0.27 mmol). This reaction mixture was stirred for 7 h followed by extraction of the aqueous layer with CH₂Cl₂; the combined organic extracts were dried over anhydrous Na₂SO₄. Removal of solvent under reduced pressure followed by flash chromatography purification (dry transfer, 8% MeOH in CHCl₃ as the eluent) provided (*S*)-nitrosulfonamide (112 mg, 80%) as a yellowish solid. *R_f* = 0.56 (10% MeOH in CHCl₃).

The above nitrosulfone (103 mg, 0.18 mmol) was dissolved in EtOAc (20 mL), and Pd/C (11 mg) was added. The mixture was stirred under a hydrogen filled balloon for 8 h at 23 °C. It was then filtered over Celite, and the filter cake was washed with EtOAc and MeOH. Removal of solvent under reduced pressure followed by flash chromatography purification (2.5% MeOH in CHCl₃ as the eluent) afforded the corresponding (*S*)-amine (77 mg, 79%) as a white solid. *R_f* = 0.26 (5% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.35 (s, 9H), 1.59–1.57 (m, 1H), 2.15–2.23 (m, 1H), 2.29–2.42 (m, 2H), 2.82–2.87 (m, 3H), 3.02 (dd, 1H, *J* = 4.75, 14.0), 3.13 (dd, 1H, *J* = 10, 13.2 Hz), 3.30 (dd, 2H, *J* = 1.8, 14.4 Hz), 3.73–3.82 (m, 1H), 3.90–3.95 (m, 1H), 3.99 (d, 1H, *J* = 6.2 Hz), 4.72 (d, 1H, *J* = 8.2 Hz), 6.72 (d, 2H, *J* = 7.9 Hz), 7.20–7.32 (m, 5H), 7.37 (s, 1H), 7.57 (d, 2H, *J* = 8.2 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 24.3, 28.6, 30.0, 36.5, 53.6, 54.4, 54.9, 56.6, 72.6, 79.8, 114.4, 125.0, 126.7, 128.7, 129.9, 130.0, 138.3, 151.5.

To a stirred solution of the above amine (33.1 mg, 0.06 mmol) in CH₂Cl₂ (3 mL) was added pyridine (30 μL, 0.37 mmol) and benzyl chloroformate (20 μL, 0.137 mmol). This reaction mixture was stirred for 3 h, then quenched with 5 drops of benzylamine followed by removal of solvent under reduced pressure. Column chromatography over silica gel (2.5% MeOH in CHCl₃ as the eluent) provided (*S*)-Cbz-amine **10** (41 mg, 99%) as a white solid. *R_f* = 0.37 (5% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.56 (brs, 1H), 2.12–2.18 (m, 1H), 2.26–2.41 (m, 2H), 2.71–2.78 (m, 2H), 2.79–2.89 (m, 1H), 3.02 (dd, 1H, *J* = 8.2, 18.0 Hz), 3.19–3.26 (m, 1H), 3.38 (d, 1H, *J* = 14.4 Hz), 3.77 (brs, 1H), 3.94–3.99 (m, 2H), 4.73 (d, 1H, *J* = 8.5 Hz), 5.21 (s, 2H), 7.19–7.22 (m, 3H), 7.25–7.29 (m, 3H), 7.31–7.39 (m, 4H), 7.57 (d, 2H, *J* = 7.6 Hz), 7.69 (d, 2H, *J* = 7.7 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 24.1, 28.7, 30.1, 36.6, 53.3, 54.3, 54.8, 56.8, 67.7, 72.6, 80.0, 118.6, 126.8, 128.7, 128.8, 128.9, 129.0, 129.4, 129.9, 131.6, 136.0, 138.1, 143.0, 153.3, 156.3, 178.9; LRMS-ESI (*m/z*) [*M* + Na]⁺ 689.

tert-Butyl-(2R,3R)-3-hydroxy-4-(4-methoxy-N-(((R)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-ylcarbamate 11. To a stirred solution of (*R*)-5-(hydroxymethyl)-2-pyrrolidinone *ent*-**5** (500 mg, 4.34 mmol) and *p*-toluenesulfonyl chloride (1.08 g, 5.6 mmol) in CH₂Cl₂ (10 mL) at 0 °C were added DMAP (106 mg, 0.87 mmol) and Et₃N (780 μL, 5.6 mmol). The

resulting mixture was allowed to warm to 23 °C and stirred for 12 h. The reaction was then quenched with 10 mL of water, and the aqueous layer was extracted with CH₂Cl₂. The combined organic extracts were washed with 1 N HCl and dried over anhydrous Na₂SO₄. Removal of solvent under reduced pressure followed by flash chromatography (2.5% MeOH in CHCl₃ as the eluent) yielded the (*R*)-toluenesulfonate (1.8 g, 93%) as a yellowish solid. *R_f* = 0.50 (5% MeOH in CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.70–1.79 (m, 1H), 2.13–2.35 (m, 3H), 2.42 (s, 3H), 3.84–3.89 (m, 2H), 3.96–3.04 (m, 1H), 6.76 (s, 1H), 7.33 (d, 2H, *J* = 8.04 Hz), 7.76 (d, 2H, *J* = 8.2 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 21.6, 22.7, 29.3, 52.6, 71.9, 127.9, 130.0, 132.3, 145.3, 178.1.

To a stirred solution of the above toluenesulfonate (1.08 g, 4.03 mmol) in DMF (30 mL) was added NaN₃ (1.31 g, 20.2 mmol). The resulting solution was stirred at 55 °C for 12 h. Solvent was then removed under reduced pressure followed by flash chromatography purification (6% MeOH in CHCl₃ as the eluent), providing the (*R*)-azidopyrrolidinone (558 mg, 99%) as a yellow oil. *R_f* = 0.44 (5% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.72–1.79 (m, 1H), 2.13–2.37 (m, 3H), 3.22 (dd, 1H, *J* = 6.3, 12.3 Hz), 3.37 (dd, 1H, *J* = 4.7, 12.3 Hz), 3.72–3.78 (m, 1H), 7.69 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 24.3, 30.2, 54.1, 56.1, 179.3.

To a solution of the above azide (528 mg, 3.77 mmol) in EtOAc (35 mL) was added Pd/C (40 mg). The mixture was stirred at 23 °C under a hydrogen filled balloon for 4 h, then filtered over Celite, and the filter cake was washed with EtOAc and MeOH. Removal of solvent under reduced pressure followed by flash chromatography purification (5% MeOH in CHCl₃ as the eluent) afforded the (*R*)-amine *ent*-**6** (257 mg, 95%) as a yellow oil. *R_f* = 0.05 (20% MeOH in CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.59–1.69 (m, 1H), 2.06–2.15 (m, 1H), 2.23–2.30 (m, 4H), 2.57 (dd, 1H, *J* = 7.6, 12.8 Hz), 2.74 (dd, 1H, *J* = 4.1, 12.9 Hz), 3.56–3.65 (m, 1H), 7.80 (brs, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 24.1, 30.2, 47.1, 56.8, 179.0.

To a solution of amine *ent*-**6** (430 mg, 3.76 mmol) in *i*-PrOH (20 mL) were added *tert*-butyl-[*S*-(*R,R*)]-(–)-(1-oxiranyl-2-phenylethyl)carbamate **7** (250 mg, 0.94 mmol) and *i*-Pr₂EtN (1.5 mL, 8.6 mmol). The resultant mixture was stirred at 65 °C for 36 h and then concentrated under reduced pressure. Flash chromatography purification (10% MeOH in CHCl₃ as the eluent) yielded the (*R*)-hydroxylamine (**8R**) (300 mg, 84%). *R_f* = 0.33 (20% MeOH in CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.33 (s, 9H), 1.64–1.73 (m, 1H), 2.13–2.22 (m, 1H), 2.28–2.34 (m, 2H), 2.54 (dd, 1H, *J* = 9.0, 11.8 Hz), 2.65 (dd, 1H, *J* = 7.2, 13.2 Hz), 2.73–2.85 (m, 4H), 2.94 (dd, 1H, *J* = 4.4, 14.0 Hz), 3.43 (s, 1H), 3.51–3.60 (m, 1H), 3.70–3.76 (m, 1H), 3.79–3.84 (m, 1H), 5.02 (d, 1H, *J* = 8.9 Hz), 7.16–7.27 (m, 5H), 7.94 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 24.6, 28.3, 30.2, 36.3, 52.1, 54.2, 54.6, 55.4, 71.6, 79.2, 126.2, 128.3, 129.4, 138.1, 155.9, 178.9; LRMS-ESI (*m/z*) [*M* + Na]⁺ 400.

To a stirred solution of above (*R*)-hydroxylamine (**8R**) (40 mg, 0.105 mmol) in CH₂Cl₂ (4 mL) and aqueous saturated NaHCO₃ (4 mL) was added 4-methoxybenzenesulfonyl chloride (66 mg, 0.318 mmol). This reaction mixture was stirred for 12 h followed by extraction of the aqueous layer with CH₂Cl₂; the combined organic extracts were dried over anhydrous Na₂SO₄. Removal of solvent under reduced pressure followed by flash chromatography purification (4% MeOH in CHCl₃ as the eluent) provided compound **11** (54 mg, 93%). *R_f* = 0.40 (10% MeOH in CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.32 (s, 9H), 1.64–1.68 (m, 1H), 2.17–2.21 (m, 2H), 2.34–2.40 (m, 2H), 2.76–2.84 (m, 1H), 2.91–3.06 (m, 3H), 3.16–3.29 (m, 2H), 3.75–3.80 (m, 1H), 3.84 (s, 3H), 3.96–4.02 (m, 2H), 4.99 (d, 1H, *J* = 8.7 Hz), 6.95 (d, 2H, *J* = 8.8 Hz), 7.16–7.28 (m, 5H), 7.68 (d, 2H, *J* = 8.8 Hz), 7.93 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 24.4, 28.2, 29.9, 35.7, 35.4, 54.7, 55.2, 55.6, 55.8, 57.8, 73.9, 79.5, 114.4, 126.2, 128.3, 129.3, 129.5, 138.0, 155.9, 163.1, 178.6; LRMS-ESI (*m/z*) [*M* + Na]⁺ 670.

tert-Butyl-(2R,3R)-4-(4-Cbz-amino-N-(((R)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-yl-carbamate 12. To a stirred solution of (R)-hydroxylamine (8R) (116 mg, 0.3 mmol) in CH₂Cl₂ (10 mL) and aqueous saturated NaHCO₃ (10 mL) was added 4-nitrobenzenesulfonyl chloride (74 mg, 0.33 mmol). This reaction mixture was stirred for 12 h followed by extraction of the aqueous layer with CH₂Cl₂; the combined organic extracts were dried over anhydrous Na₂SO₄. Removal of solvent under reduced pressure followed by flash chromatography purification (dry transfer, 5% MeOH in CHCl₃ as the eluent) provided the (R)-nitrosulfonamide (164 mg, 96%) as a yellowish solid. *R_f* = 0.56 (10% MeOH in CHCl₃).

The above nitrosulfonamide (154 mg, 0.27 mmol) was redissolved in EtOAc (25 mL) and treated with Pd/C (16 mg) under argon. Argon was then replaced with a hydrogen filled balloon, and the mixture was allowed to stir for 12 h at 23 °C. It was then filtered over Celite, and the filter cake was washed with EtOAc and MeOH. Removal of solvent under reduced pressure followed by flash chromatography purification (6% MeOH in CHCl₃ as the eluent) afforded the corresponding (R)-aniline (123 mg, 83%) as an amorphous solid. *R_f* = 0.45 (10% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.31 (s, 9H), 1.58–1.64 (m, 1H), 2.15–2.21 (m, 1H), 2.29 (t, 2H, *J* = 8.2 Hz), 2.73–2.86 (m, 3H), 2.99 (dd, 1H, *J* = 4.4, 13.9 Hz), 3.23 (d, 1H, *J* = 13.8 Hz), 3.30 (d, 1H, *J* = 14.8 Hz), 3.74 (brs, 1H), 3.92 (brs, 1H), 3.99 (d, 1H, *J* = 5.7), 4.31 (s, 1H), 5.01 (d, 1H, *J* = 9.1 Hz), 6.63 (d, 2H, *J* = 8.5 Hz), 7.16–7.21 (m, 3H), 7.24–7.27 (m, 2H), 7.48 (d, 2H, *J* = 8.5 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 24.3, 28.1, 29.8, 35.5, 54.6, 55.0, 55.5, 57.7, 73.8, 79.5, 113.9, 125.0, 126.2, 128.2, 129.3, 129.4, 137.8, 151.0, 155.9, 178.2; LRMS-ESI (*m/z*) [*M* + Na]⁺ 555.

To a stirred solution of the above (R)-aniline (101 mg, 0.19 mmol) in CH₂Cl₂ (15 mL) was added pyridine (34 μL, 0.41 mmol) and benzyl chloroformate (60 μL, 0.41 mmol). This reaction mixture was stirred for 1.5 h, then quenched with 3 drops of benzylamine, followed by removal of solvent under reduced pressure. Column chromatography over silica gel (6% MeOH in CHCl₃ as the eluent) provided the (R)-Cbz-amine 12 (120 mg, 95%) as a white solid. *R_f* = 0.35 (10% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.29 (s, 9H), 1.55–1.65 (m, 1H), 2.15–2.23 (m, 1H), 2.26–2.31 (m, 2H), 2.60–2.75 (m, 3H), 2.97 (dd, 1H, *J* = 8.2, 18.1 Hz), 3.29 (d, 1H, *J* = 17.7 Hz), 3.36 (dd, 1H, *J* = 2.4, 14.9 Hz), 3.64 (s, 1H), 3.88–3.92 (m, 1H), 3.98–4.02 (m, 1H), 5.12 (d, 1H, *J* = 9.0 Hz), 5.17 (s, 2H), 7.14–7.19 (m, 3H), 7.21–7.28 (m, 3H), 7.30–7.37 (m, 4H), 7.54 (d, 2H, *J* = 8.6 Hz), 7.63 (d, 2H, *J* = 8.8); ¹³C NMR (125 MHz, CDCl₃) δ 24.8, 28.6, 30.2, 36.0, 55.1, 55.8, 55.9, 58.2, 67.6, 74.4, 80.0, 118.6, 126.7, 128.6, 128.7, 128.8, 129.0, 129.0, 129.7, 130.3, 131.2, 136.1, 138.2, 143.4, 153.6, 178.9; LRMS-ESI (*m/z*) [*M* + Na]⁺ 689.

(3R,3aS,6aR)-Hexahydrofuro[2,3-*b*]furan-3-yl(2S,3R)-3-hydroxy-4-(4-methoxy-N-(((S)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-yl-carbamate 17a. A solution of compound 9 (10 mg, 0.02 mmol) in 30% trifluoroacetic acid (in CH₂Cl₂, 3 mL) was stirred 23 °C for 40 min, then concentrated under reduced pressure to give the crude amine 13S. This residue was redissolved in CH₂Cl₂ (3 mL), treated with Et₃N (20 μL, 0.13 mmol), followed by carbonate 15 (5.5 mg, 0.02 mmol) and stirred at 23 °C for 12 h. The reaction mixture was then concentrated under reduced pressure, and the residue was purified by flash chromatography (2% MeOH in CHCl₃ as the eluent) to give inhibitor 17a (11.3 mg, 98%) as a white solid. *R_f* = 0.48 (10% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.48 (dd, 1H, *J* = 5.5, 13.2 Hz), 1.58–1.68 (m, 2H), 2.17–2.26 (m, 1H), 2.34–2.49 (m, 2H), 2.76 (dd, 1H, *J* = 9.8, 14.0 Hz), 2.85–2.95 (m, 3H), 3.10–3.16 (m, 2H), 3.22 (dd, 1H, (dd, 1H, *J* = 9.9, 13.7 Hz), 3.67–3.74 (m, 2H), 3.82–3.85 (dt, 1H, *J* = 1.8, 8.4 Hz), 3.87 (s, 3H), 3.94 (dd, 1H, *J* = 6.2, 9.6 Hz), 3.96–4.01 (m, 1H), 4.04–4.08 (m, 1H), 5.0 (q, 1H, *J* = 6.1, 7.9 Hz), 5.64 (d, 1H, *J* = 5.2 Hz), 6.98 (d, 2H, *J* = 8.9 Hz), 7.19–7.29 (m, 3H), 7.26–7.29 (m, 2H), 7.58 (brs, 1H), 7.69 (d, 2H, *J* = 8.8 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 23.9, 25.7, 29.5, 35.9, 45.2, 53.4, 53.9, 55.1, 55.6, 56.4, 69.5, 70.8, 72.3, 73.3, 109.2, 114.4, 126.4, 128.3, 128.6, 129.2, 129.4, 137.6, 155.4,

163.2, 178.5. LRMS-ESI (*m/z*) [*M* + H]⁺ 604.2; HRMS-ESI (*m/z*) [*M* + H]⁺ calcd for C₂₉H₃₈N₃O₉S 604.2329, found 604.2332.

(3aS,5R,6aR)-Hexahydro-2H-cyclopenta[*b*]furan-5-yl-(2S,3R)-3-hydroxy-4-(4-methoxy-N-(((S)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-yl-carbamate 17b. A solution of compound 9 (11.8 mg, 0.02 mmol) in 30% trifluoroacetic acid (in CH₂Cl₂, 1.5 mL) was stirred at 23 °C for 40 min, then concentrated under reduced pressure to give the crude amine 13S. This crude residue was redissolved in CH₂Cl₂ (1.5 mL), treated with Et₃N (63 μL, 0.45 mmol), followed by carbonate 16 (6.4 mg, 0.02 mmol), and stirred at 23 °C for 6 h. The reaction mixture was then concentrated under reduced pressure and the residue was purified by flash chromatography (1% MeOH in CHCl₃ as the eluent) to give inhibitor 17b (11.5 mg, 87%) as a white solid. *R_f* = 0.35 (5% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.45 (d, 1H, *J* = 14.3 Hz), 1.55–1.59 (m, 1H), 1.88 (d, 1H, *J* = 15.1 Hz), 1.95–2.06 (m, 3H), 2.17–2.24 (m, 1H), 2.33–2.48 (m, 2H), 2.60–2.67 (m, 1H), 2.78 (dd, 1H, *J* = 9.1, 14.1 Hz), 2.88–2.97 (m, 2H), 3.09 (dd, 1H, *J* = 4.3, 14.1 Hz), 3.12–3.18 (m, 2H), 3.64–3.68 (m, 1H), 3.82–3.85 (m, 2H), 3.86 (s, 3H), 3.89–3.95 (m, 1H), 3.99–4.05 (m, 1H), 4.39–4.42 (m, 1H), 4.69 (d, 1H, *J* = 4.1 Hz), 4.87–4.90 (m, 1H), 4.91 (d, 1H, *J* = 8.9 Hz), 6.98 (d, 2H, *J* = 8.9 Hz), 7.20–7.23 (m, 3H), 7.27–7.30 (m, 2H), 7.42 (s, 1H), 7.70 (d, 2H, *J* = 8.9 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 24.0, 29.8, 33.8, 35.9, 38.3, 39.4, 41.5, 53.2, 53.3, 53.9, 54.8, 55.5, 56.3, 67.6, 72.2, 83.7, 114.3, 126.4, 128.4, 128.7, 129.3, 129.5, 137.5, 156.1, 163.1, 178.3. LRMS-ESI (*m/z*) [*M* + Na]⁺ 624.0; HRMS-ESI (*m/z*) [*M* + Na]⁺ calcd for C₃₀H₃₉N₃NaO₈S 624.2356, found 624.2352.

(3R,3aS,6aR)-Hexahydrofuro[2,3-*b*]furan-3-yl(2S,3R)-4-(4-amino-N-(((S)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-yl-carbamate 18a. The Cbz-protected amine 10 (31 mg, 0.04 mmol) was treated with 30% trifluoroacetic acid in CH₂Cl₂ (6 mL) and stirred at 23 °C for 40 min, then concentrated under reduced pressure to give the crude amine 14S. This residue was redissolved in CH₂Cl₂ (6 mL), charged with Et₃N (64 μL, 0.46 mmol), followed by carbonate 15 (14 mg, 0.05 mmol), and stirred at 23 °C for 12 h. Reaction was quenched with 3 drops of benzylamine and concentrated under reduced pressure. Flash chromatography purification (4% MeOH in CHCl₃ as the eluent) provided the Cbz-protected inhibitor (23 mg, 86%) as a white solid. *R_f* = 0.46 (10% MeOH in CHCl₃).

To the above Cbz-protected inhibitor (13.3 mg, 0.018 mmol), in EtOAc (6 mL) under argon, was added Pd/C (3 mg). The mixture was stirred at 23 °C under a hydrogen filled balloon for 3 h, then filtered over Celite, and the filter cake was washed with EtOAc and MeOH. Removal of solvent under reduced pressure followed by flash chromatography purification (3% MeOH in CHCl₃ as the eluent) provided the title inhibitor 18a (7.4 mg, 68%) as a white solid. *R_f* = 0.19 (5% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.44 (d, 1H, *J* = 5.4 Hz), 1.59–1.69 (m, 2H), 2.16–2.25 (m, 1H), 2.36 (t, 2H, *J* = 7.9 Hz), 2.68 (dd, 1H, *J* = 9.8, 13.9 Hz), 2.85–2.95 (m, 3H), 3.03–3.09 (m, 2H), 3.13 (dd, 1H, *J* = 4.4, 14.1 Hz), 3.67–3.72 (m, 2H), 3.79–3.89 (m, 3H), 3.93 (dd, 1H, *J* = 6.0, 9.7 Hz), 4.01–4.06 (m, 1H), 4.96 (q, 1H, *J* = 5.9, 7.9 Hz), 5.63 (d, 1H, *J* = 5.1 Hz), 6.72 (d, 2H, *J* = 8.2 Hz), 7.17–7.21 (m, 3H), 7.24–7.28 (m, 2H), 7.50 (d, 2H, *J* = 8.4 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 24.3, 26.2, 30.0, 36.6, 45.8, 53.6, 54.2, 55.6, 56.6, 70.0, 71.5, 72.7, 73.6, 109.7, 114.6, 124.9, 126.8, 128.8, 129.7, 129.9, 130.3, 151.4, 155.9, 178.3. LRMS-ESI (*m/z*) [*M* + H]⁺ 589.2; HRMS-ESI (*m/z*) [*M* + H]⁺ calcd for C₂₈H₃₇N₄O₈S 589.2332, found 589.2336.

(3aS,5R,6aR)-Hexahydro-2H-cyclopenta[*b*]furan-5-yl-(2S,3R)-4-(4-amino-N-(((S)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-yl-carbamate 18b. The Cbz-protected amine 10 (29.6 mg, 0.04 mmol) was treated with 30% trifluoroacetic acid (in CH₂Cl₂, 6 mL) and stirred at 23 °C for 40 min, then concentrated under reduced pressure to give the crude amine 14S. The residue was redissolved in CH₂Cl₂ (6 mL), charged with Et₃N (31 μL, 0.22 mmol), followed by carbonate 16 (13.1 mg, 0.05 mmol), and stirred at 23 °C for 4 h. Reaction was

quenched with 2 drops of benzylamine and concentrated under reduced pressure. Flash chromatography purification (4% MeOH in CHCl₃ as the eluent) provided the Cbz-protected inhibitor (26.1 mg, 82%) as a white solid. R_f = 0.49 (10% MeOH in CHCl₃).

To a solution of the above protected inhibitor (17 mg, 0.02 mmol), in EtOAc (5 mL) under argon, was added Pd/C (3 mg). The mixture was stirred at 23 °C under a H₂ filled balloon for 5 h, then filtered over Celite, and the filter cake was washed with EtOAc and MeOH. Removal of solvent under reduced pressure, followed by flash chromatography purification (5% MeOH in CHCl₃ as the eluent) provided inhibitor **18b** (14 mg, 75%) as a white solid. R_f = 0.27 (10% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.41–1.45 (m, 1H), 1.56–1.67 (m, 3H), 1.85 (d, 1H, J = 13.4 Hz), 1.96–2.04 (m, 3H), 2.14–2.21 (m, 1H), 2.31–2.42 (m, 2H), 2.61 (brs, 1H), 2.74–2.85 (m, 3H), 3.09 (dd, 1H, J = 4.4, 14.4 Hz), 3.15–3.20 (m, 1H), 3.25 (d, 1H, J = 14.3 Hz), 3.66–3.70 (q, 1H, J = 7.1, 7.4 Hz), 3.83–3.88 (m, 2H), 3.91–3.96 (m, 1H), 3.96–4.25 (m, 1H), 4.40 (t, 1H, J = 5.9 Hz), 4.87 (brs, 1H), 4.93 (d, 1H, J = 8.9 Hz), 6.69 (d, 2H), 7.18–7.22 (m, 3H), 7.28–7.30 (m, 2H), 7.55 (d, 2H, J = 8.4 Hz), 7.63 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 24.3, 30.1, 34.1, 36.7, 38.7, 39.8, 41.9, 53.5, 54.3, 55.3, 56.8, 68.0, 72.8, 84.1, 114.6, 125.3, 126.8, 128.8, 129.9, 130.1, 138.1, 151.4, 156.6, 178.9. LRMS-ESI (m/z) [M + H]⁺ 586.9; HRMS-ESI (m/z) [M + H]⁺ calcd for C₂₉H₃₉N₄O₇S 587.2539, found 587.2539.

(3R,3aS,6aR)-Hexahydrofuro[2,3-*b*]furan-3-yl-(2S,3R)-3-hydroxy-4-(4-methoxy-*N*-(((*R*)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-ylcarbamate 19a. A solution of compound **11** (20 mg, 0.03 mmol) in 30% trifluoroacetic acid (in CH₂Cl₂, 3 mL) was stirred 23 °C for 40 min, then concentrated under reduced pressure. The residue was redissolved in CH₂Cl₂ (3 mL), treated with Et₃N (51 μL, 0.36 mmol), followed by carbonate **15** (11 mg, 0.04 mmol), and stirred at 23 °C for 12 h. The reaction mixture was then concentrated under reduced pressure and the residue was purified by flash chromatography (2% MeOH in CHCl₃ as the eluent) to give inhibitor **19a** (21 mg, 92%) as a white solid. R_f = 0.41 (10% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.44 (dd, 1H, J = 5.6, 13.2 Hz), 1.57–1.67 (m, 2H), 1.81–1.91 (m, 1H), 2.19–2.27 (m, 1H), 2.33–2.41 (m, 2H), 2.73 (dd, 1H, J = 10.35, 13.9 Hz), 2.83–2.91 (m, 2H), 2.95 (dd, 1H, J = 8.9, 14.9 Hz), 3.11 (dd, 1H, J = 4.2, 14.0 Hz), 3.26–3.30 (ddd, 2H, J = 2.6, 7.0, 14.3 Hz), 3.64–3.70 (m, 1H), 3.74 (dd, 1H, J = 5.5, 9.8 Hz), 3.76–3.81 (dt, 1H, J = 1.62, 8.0 Hz), 3.87 (s, 3H), 3.89–3.93 (q, 1H, J = 4.0, 5.6 Hz), 4.03–4.06 (m, 2H), 4.98–5.02 (q, 1H, J = 5.65, 7.9 Hz), 5.62 (d, 1H, J = 5.4 Hz), 6.98 (d, 2H, J = 8.9 Hz), 7.17–7.27 (m, 5H), 7.70 (d, 2H, J = 8.8 Hz), 7.89 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 24.2, 25.7, 29.8, 35.4, 45.4, 55.0, 55.3, 55.5, 55.7, 58.1, 69.5, 71.0, 73.3, 74.0, 109.2, 114.4, 126.3, 128.3, 128.8, 129.2, 129.4, 137.8, 155.5, 163.2, 178.5. LRMS-ESI (m/z) [M + Na]⁺ 626.3; HRMS-ESI (m/z) [M + Na]⁺ calcd for C₂₉H₃₇N₃ Na O₉S 626.2148, found 626.2156.

(3aS,5R,6aR)-Hexahydro-2H-cyclopenta[*b*]furan-5-yl-(2S,3R)-3-hydroxy-4-(4-methoxy-*N*-(((*R*)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-ylcarbamate 19b. A solution of compound **11** (21 mg, 0.04 mmol) in 30% trifluoroacetic acid (in CH₂Cl₂, 3 mL) was stirred at 23 °C for 40 min, then concentrated under reduced pressure. The residue was redissolved in CH₂Cl₂ (3 mL), treated with Et₃N (27 μL, 0.19 mmol), followed by carbonate **16** (12 mg, 0.04 mmol), and stirred at 23 °C for 12 h. The reaction mixture was then concentrated under reduced pressure and the residue was purified by flash chromatography (1% MeOH in CHCl₃ as the eluent) to give inhibitor **19b** (21 mg, 93%) as a white solid. R_f = 0.31 (5% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.48 (d, 1H, J = 14.1 Hz), 1.58–1.62 (m, 1H), 1.83–2.04 (m, 5H), 2.19–2.25 (m, 1H), 2.31–2.41 (m, 2H), 2.59–2.67 (m, 1H), 2.73 (dd, 1H, J = 9.0, 13.9 Hz), 3.03 (dd, 1H, J = 7.0, 15.0 Hz), 3.08–3.16 (m, 2H), 3.19 (d, 1H, J = 14.9 Hz), 3.57–3.63 (m, 1H), 3.83–3.86 (m, 3H), 3.86 (s, 3H), 3.94–3.99 (m, 1H), 4.78 (d, 1H, J = 13.7 Hz), 4.90 (s, 1H), 5.36 (d, 1H, J = 8.1 Hz), 6.97 (d, 2H, J = 8.6 Hz), 7.18–7.28 (m, 5H), 7.46 (d, 1H, J = 18.4 Hz), 7.70 (d, 2H, J = 8.8 Hz); ¹³C NMR (125 MHz, CDCl₃) δ

22.5, 30.0, 31.4, 33.9, 35.8, 38.2, 39.5, 41.7, 54.6, 54.9, 55.5, 56.0, 57.7, 67.7, 74.0, 83.9, 114.4, 126.2, 128.2, 128.9, 129.3, 129.4, 137.9, 156.1, 163.1, 178.5. LRMS-ESI (m/z) [M + H]⁺ 601.7; HRMS-ESI (m/z) [M + H]⁺ calcd for C₃₀H₄₀N₃O₈S 602.2536, found 602.2536.

(3R,3aS,6aR)-Hexahydrofuro[2,3-*b*]furan-3-yl-(2S,3R)-4-(4-amino-*N*-(((*R*)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-ylcarbamate 20a. A solution of the (*R*)-aniline **12** (15 mg, 0.03 mmol) in 30% trifluoroacetic acid (in CH₂Cl₂, 3 mL) was stirred at 23 °C for 40 min, then concentrated under reduced pressure. The residue was redissolved in CH₂Cl₂ (3 mL), treated with Et₃N (40 μL, 0.28 mmol), followed by carbonate **15** (9.2 mg, 0.03 mmol), and stirred at 23 °C for 6 h. The reaction mixture was then concentrated under reduced pressure and the residue was purified by flash chromatography (4% MeOH in CHCl₃ as the eluent) to give inhibitor **20a** (12.5 mg, 75%) as a white solid. R_f = 0.26 (10% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.35 (dd, 1H, J = 5.7 Hz), 1.50–1.63 (m, 2H), 2.15–2.23 (m, 1H), 2.27–2.31 (m, 2H), 2.56–2.72 (m, 3H), 2.82–2.87 (m, 2H), 3.07 (dd, 1H, J = 4.0, 14.0 Hz), 3.36 (q, 1H, J = 1.4, 3.5 Hz), 3.36 (q, 1H, J = 2.0, 3.4 Hz), 3.60–3.65 (m, 1H), 3.70 (dd, 1H, J = 5.4, 9.8 Hz), 3.75–3.79 (dt, 1H, J = 1.96, 8.3 Hz), 3.80–3.86 (m, 1H), 3.88 (dd, 1H, J = 5.9, 9.8 Hz), 3.90–3.94 (m, 1H), 3.99–4.02 (m, 1H), 4.92–4.97 (q, 1H, J = 5.7, 8.1 Hz), 5.58 (d, 1H, J = 5.8 Hz), 5.92 (d, 1H, J = 9.5 Hz), 6.63 (d, 2H, J = 8.7 Hz), 7.13–7.22 (m, 5H), 7.46 (d, 2H, J = 8.7 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 24.1, 25.7, 29.7, 35.3, 45.4, 55.0, 55.5, 55.6, 58.1, 69.5, 71.0, 73.1, 73.9, 109.2, 113.8, 124.2, 126.2, 128.2, 129.1, 129.3, 137.8, 151.4, 155.6, 178.6. LRMS-ESI (m/z) [M + Na]⁺ 611.4; HRMS-ESI (m/z) [M + Na]⁺ calcd for C₂₈H₃₆N₄NaO₈S 611.2152, found 611.2149.

(3aS,5R,6aR)-Hexahydro-2H-cyclopenta[*b*]furan-5-yl-(2S,3R)-4-(4-amino-*N*-(((*R*)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-ylcarbamate 20b. The Cbz-amine **12** (40 mg, 0.06 mmol) was treated with 30% trifluoroacetic acid (in CH₂Cl₂, 6 mL) and stirred at 23 °C for 40 min, then concentrated under reduced pressure. This residue was redissolved in CH₂Cl₂ (6 mL), charged with Et₃N (42 μL, 0.3 mmol), followed by carbonate **16** (19 mg, 0.07 mmol), and stirred at 23 °C for 12 h. Reaction was quenched with 3 drops of benzylamine and concentrated under reduced pressure. Flash chromatography purification (5% MeOH in CHCl₃ as the eluent) provided the Cbz-protected inhibitor (32.1 mg, 75%) as a white solid. R_f = 0.41 (10% MeOH in CHCl₃).

To the above Cbz-protected inhibitor (25.8 mg, 0.03 mmol), in EtOAc (5 mL) under argon, was added Pd/C (5 mg). The mixture was stirred at 23 °C under a hydrogen filled balloon for 3 h, then filtered over Celite, and the filter cake was washed with EtOAc and MeOH. Removal of solvent under reduced pressure followed by flash chromatography purification (7.5% MeOH in CHCl₃ as the eluent) provided the title inhibitor **20b** (16.2 mg, 77%) as a white solid. R_f = 0.51 (15% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.46 (m, 1H), 1.58–1.64 (m, 1H), 1.77–1.84 (m, 1H), 1.86–2.04 (m, 5H), 2.15–2.23 (m, 1H), 2.32–2.36 (m, 2H), 2.59–2.65 (m, 1H), 2.68 (dd, 1H, J = 9.2, 14.0 Hz), 2.95–3.04 (m, 2H), 3.11 (d, 2H, J = 13.7 Hz), 3.18 (d, 1H, J = 14.9 Hz), 3.57–3.62 (q, 1H, J = 6.9, 7.7 Hz), 3.82–3.87 (q, 2H, J = 6.5, 7.9 Hz), 3.92–3.97 (m, 1H), 4.30 (s, 2H), 4.37 (t, 1H, J = 5.7 Hz), 4.75 (s, 1H), 4.89 (s, 1H), 5.40 (d, 1H, J = 8.4 Hz), 6.66 (d, 2H, J = 8.6 Hz), 7.16–7.22 (m, 3H), 7.24–7.27 (m, 2H), 7.40 (s, 1H), 7.51 (d, 2H, J = 8.6 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 23.7, 30.0, 33.9, 35.9, 38.1, 39.5, 41.7, 54.6, 54.8, 55.9, 57.6, 67.7, 74.0, 76.6, 84.0, 114.1, 124.9, 126.2, 128.2, 129.3, 129.4, 137.9, 151.0, 156.2, 178.4. LRMS-ESI (m/z) [M + Na]⁺ 609.0; HRMS-ESI (m/z) [M + Na]⁺ calcd for C₂₉H₃₈N₄NaO₇S 609.2359, found 609.2362.

(*R*)-tert-Butyl-4-(((2R,3S)-3-(tert-butoxycarbonylamino)-2-hydroxy-4-phenylbutylamino)methyl)-2,2-dimethyloxazolidine-3-carboxylate 22. To a solution of the (*R*)-tert-butyl 4-(azidomethyl)-2,2-dimethyloxazolidine-3-carboxylate **21** (411 mg, 1.60 mmol) in MeOH (10 mL) was added Pd/C (40 mg). This mixture was stirred

at 23 °C under a hydrogen filled balloon for 2 h, then filtered over Celite, and the filter cake was washed with MeOH. Solvent was removed under reduced pressure, and the resultant residue was redissolved in *i*-PrOH, followed by addition of *tert*-butyl-[*S*-(*R,R*)]-(α)-(1-oxiranyl-2-phenylethyl)carbamate **7** (121 mg, 0.46 mmol). The mixture was allowed to stir at 65 °C for 24 h. Solvent was then removed under reduced pressure. Flash chromatography purification (2% MeOH in CHCl₃ as the eluent) yielded the (*R*)-amine **22** (93 mg, 41%) as a yellow oil. R_f = 0.32 (10% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.35 (s, 18H), 1.48 (s, 24H), 1.54 (s, 3H), 1.58 (s, 3H), 2.67–2.92 (m, 10), 2.95 (d, 1H, J = 4.1 Hz), 2.98 (d, 1H, J = 4.1 Hz), 3.47 (brs, 2H), 3.78 (brs, 3H), 3.89–4.02 (m, 5H), 4.61 (brs, 1H), 4.68 (brs, 1H), 7.18–7.22 (m, 6H), 7.27–7.30 (m, 4H); ¹³C NMR (125 MHz, CDCl₃) δ 22.9, 24.2, 26.6, 27.4, 28.1, 28.3, 36.4, 51.4, 56.9, 65.9, 70.7, 70.9, 79.3, 79.4, 79.6, 80.3, 93.4, 93.7, 126.2, 128.3, 129.4, 137.5, 137.7, 152.6, 155.8; LRMS-ESI (m/z) [M + Na]⁺ 516.

(*R*)-*tert*-Butyl-4-((*N*-((2*R*,3*S*)-3-(*tert*-butoxycarbonylamino)-2-hydroxy-4-phenylbutyl)-4-methoxyphenylsulfonamido)methyl)-2,2-dimethyloxazolidine-3-carboxylate **23.** To a stirred solution of amine **22** (37 mg, 0.07 mmol) in CH₂Cl₂ (4 mL) and aqueous saturated NaHCO₃ (4 mL) was added 4-methoxybenzenesulfonyl chloride (34 mg, 0.16 mmol). This reaction mixture was stirred for 12 h followed by extraction of the aqueous layer with CH₂Cl₂; the combined organic extracts were dried over anhydrous Na₂SO₄. Removal of solvent under reduced pressure followed by flash chromatography purification (30% EtOAc in hexane as the eluent) provided compound **23** (40 mg, 80%) as a white solid. R_f = 0.37 (40% EtOAc in hexane); ¹H NMR (500 MHz, CDCl₃) δ 1.34 (s, 9H), 1.48 (s, 9H), 1.60 (s, 6H), 2.84–2.89 (m, 1H), 2.95–3.06 (m, 2H), 3.15 (dd, 1H, J = 6.5, 13.0 Hz), 3.29 (d, 1H, J = 9.7 Hz), 3.41 (d, 1H, J = 15.1 Hz), 3.77 (brs, 2H), 3.85 (s, 3H), 3.90–3.94 (m, 1H), 3.99 (d, 1H, J = 9.0 Hz), 4.2 (brs, 1H), 4.73 (brs, 0.5H), 4.98 (brs, 0.5H), 6.95 (d, 2H, J = 8.0 Hz), 7.19–7.24 (m, 3H), 7.25–7.28 (m, 2H), 7.69 (d, 2H, J = 8.9 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 24.0, 27.4, 28.2, 28.3, 36.0, 53.3, 54.0, 55.4, 55.8, 56.3, 65.6, 72.4, 79.0, 80.9, 93.9, 114.2, 126.0, 128.1, 129.3, 129.6, 129.9, 137.8, 152.8, 155.4, 162.9; LRMS-ESI (m/z) [M + Na]⁺ 686.

(*R*)-*tert*-Butyl-4-((*N*-((2*R*,3*S*)-3-(*tert*-butoxycarbonylamino)-2-hydroxy-4-phenylbutyl)-4-nitrophenylsulfonamido)methyl)-2,2-dimethyloxazolidine-3-carboxylate **24.** To a stirred solution of amine **22** (41 mg, 0.08 mmol) in CH₂Cl₂ (4 mL) and aqueous saturated NaHCO₃ (4 mL) was added 4-nitrobenzenesulfonyl chloride (37 mg, 0.16 mmol). This reaction mixture was stirred for 12 h followed by extraction of the aqueous layer with CH₂Cl₂; the combined organic extracts were dried over anhydrous Na₂SO₄. Removal of solvent under reduced pressure followed by flash chromatography purification (30% EtOAc in hexane as the eluent) provided (*R*)-nitrosulfonamide **24** (51 mg, 92%) as a white solid. R_f = 0.52 (40% EtOAc in hexane); ¹H NMR (500 MHz, CDCl₃) δ 1.35 (s, 18H), 1.49 (s, 24H), 1.62 (s, 6H), 3.43 (bd, 2H, J = 9.6 Hz), 3.55 (d, 2H, J = 15.1 Hz), 3.75 (brs, 4H), 3.95 (s, 4H), 4.28 (brs, 2H), 4.63 (d, 2H, J = 7.8 Hz), 5.16 (brs, 2H), 7.19–7.23 (m, 6H), 7.25–7.31 (m, 4H), 7.92 (d, 4H, J = 8.8 Hz), 8.30 (d, 4H, J = 8.05 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 24.0, 27.3, 28.1, 28.2, 35.8, 53.7, 54.0, 55.5, 55.9, 65.6, 72.2, 79.4, 81.3, 94.1, 124.3, 126.3, 128.2, 128.4, 129.5, 137.4, 144.3, 149.9, 153.0, 155.5; LRMS-ESI (m/z) [M + Na]⁺ 701.

***tert*-Butyl-(2*S*,3*R*)-3-hydroxy-4-(4-methoxy-*N*-(((*R*)-2-oxooxazolidin-4-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-ylcarbamate **25**.** To (*R*)-toluenesulfonate **23** (32 mg, 0.05 mmol) in methanol (3 mL) was added *p*-toluenesulfonic acid monohydrate (2 mg), and the reaction mixture was allowed to stir at 23 °C. After 12 h the reaction reached maximum conversion and did not move any further. It was quenched with saturated aqueous NaHCO₃, extracted with EtOAc, washed with water and brine, and dried over Na₂SO₄. Flash chromatography purification (30–50% EtOAc in hexane) yielded 7 mg (21%) of starting material and 18 mg (58%) of the expected primary alcohol.

The above alcohol (17 mg, 0.0278 mmol), in CH₂Cl₂ (2 mL) and Et₃N (16 μ L, 0.11 mmol), was cooled to –10 °C and treated with methanesulfonyl chloride (2.5 μ L, 0.03 mmol). The reaction mixture was stirred at 0 °C for 2 h, followed by removal of solvent via rotavap and pump. The resulting residue was redissolved in chloroform (2 mL), treated with DIPEA (19 μ L, 0.11 mmol), refluxed for 8 h, and concentrated under reduced pressure. Flash chromatography purification (1% MeOH in CHCl₃ as the eluent) afforded (*R*)-oxazolidinone **25** (12 mg, 78%) as a white solid. R_f = 0.33 (5% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.32 (s, 9H), 2.77 (dd, 1H, J = 8.4, 13.9 Hz), 2.93–3.03 (m, 3H), 3.07 (d, 1H, J = 14.5 Hz), 3.13 (dd, 1H, J = 9.3, 14.1 Hz), 3.69–3.79 (m, 2H), 3.85 (s, 3H), 3.97 (dd, 1H, J = 5.2, 8.3 Hz), 4.23–4.29 (m, 1H), 4.43 (t, 1H, J = 8.7 Hz), 6.94 (d, 2H, J = 8.8 Hz), 7.18–7.24 (m, 3H), 7.26–7.30 (m, 2H), 7.64 (d, 2H, J = 8.6 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 28.1, 35.9, 52.0, 54.1, 54.6, 54.7, 55.5, 67.4, 71.9, 79.7, 114.4, 126.3, 128.4, 128.4, 129.3, 129.4, 137.5, 155.9, 159.4, 163.2; LRMS-ESI (m/z) [M + Na]⁺ 572.

***tert*-Butyl-(2*S*,3*R*)-4-(4-amino-*N*-(((*R*)-2-oxooxazolidin-4-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-ylcarbamate **26**.** To (*R*)-nitrosulfonamide **24** (42 mg, 0.06 mmol) in methanol (4 mL) was added *p*-toluenesulfonic acid monohydrate (2 mg), and the reaction mixture was allowed to stir at 23 °C. After 36 h the reaction appeared to have moved only 75% to completion, and formation of free amines was evident on TLC. At this point, it was quenched with saturated aqueous NaHCO₃, extracted with EtOAc, washed with water and brine, and dried over Na₂SO₄. Flash chromatography purification (30–50% EtOAc in hexane) yielded 7 mg (17%) of starting material and 24 mg (55%) of the expected primary alcohol.

The above alcohol (23 mg, 0.03 mmol), in CH₂Cl₂ (3 mL) and Et₃N (20 μ L, 0.14 mmol), was cooled to –10 °C and treated with methanesulfonyl chloride (53 μ L, 0.04 mmol). The reaction mixture was stirred at 0 °C for 2 h, followed by removal of solvent via rotavap and pump. The resultant residue was redissolved in chloroform (3 mL), treated with DIPEA (25 μ L, 0.14 mmol), and refluxed for 12 h; product crashed out of the chloroform and was found to be only slightly soluble in methanol. Solvent was removed under reduced pressure, and the crude material was treated with methanol (3 mL) and Pd/C (3 mg). The mixture was stirred at 23 °C under a H₂ filled balloon for 2 h, then filtered over Celite, and the filter cake was washed with MeOH. Removal of solvent, followed by flash chromatography purification (3% MeOH in CHCl₃ as the eluent) provided (*R*)-oxazolidinone **26** (13 mg, 68%) as a white solid. R_f = 0.19 (5% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.31 (s, 9H), 2.73 (dd, 1H, J = 8.7, 13.6 Hz), 2.88 (dd, 1H, J = 9.6, 14.5 Hz), 2.91–2.94 (m, 1H), 2.98 (dd, 1H, J = 4.2, 13.9 Hz), 3.10 (dd, 2H, J = 9.2, 14.0 Hz), 3.70–3.81 (m, 2H), 3.96 (dd, 1H, J = 5.2, 8.4 Hz), 4.21–4.24 (m, 1H), 4.40 (t, 1H, J = 8.7 Hz), 4.92 (d, 1H, J = 8.5 Hz), 6.63 (d, 2H, J = 8.7 Hz), 7.17–7.21 (m, 3H), 7.25–7.28 (m, 2H), 7.46 (d, 2H, J = 8.4 Hz); ¹³C NMR (125 MHz, CD₃OD) δ 28.1, 29.5, 35.9, 51.9, 52.0, 54.0, 54.7, 67.5, 72.1, 79.6, 113.9, 124.4, 126.3, 128.3, 129.3, 129.4, 137.6, 155.1, 156.0, 159.5; LRMS-ESI (m/z) [M + Na]⁺ 557.

***tert*-Butyl-(2*S*,3*R*)-3-hydroxy-4-(4-methoxy-*N*-(((*S*)-2-oxooxazolidin-4-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-ylcarbamate **27**.** To a solution of the (*S*)-*tert*-butyl 4-(azidomethyl)-2,2-dimethyloxazolidine-3-carboxylate *ent*-**21** (840 mg, 3.32 mmol) in MeOH (20 mL) was added Pd/C (80 mg). This mixture was stirred at 23 °C under a hydrogen filled balloon for 2 h, then filtered over Celite, and the filter cake was washed with MeOH. Solvent was removed under reduced pressure, and the resultant residue was redissolved in *i*-PrOH, followed by addition of *tert*-butyl-[*S*-(*R,R*)]-(α)-(1-oxiranyl-2-phenylethyl)carbamate **7** (218 mg, 0.83 mmol). The mixture was allowed to stir at 65 °C for 24 h. Solvent was then removed under reduced pressure. Flash chromatography purification (2% MeOH in CHCl₃ as the eluent) yielded the corresponding (*S*)-amine (**22S**) (187 mg, 46%) as an amorphous solid. R_f = 0.28 (10% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.35 (s, 18H), 1.48 (s, 24H), 1.54 (s, 3H), 1.58 (s, 3H), 2.64–2.91 (m, 10), 2.95–3.03 (m, 2H), 3.44 (brs, 1H), 3.50 (brs,

1H), 3.77 (brs, 3H), 3.87–4.00 (m, 5H), 4.61 (brs, 1H), 4.64 (brs, 1H), 7.18–7.22 (m, 6H), 7.27–7.30 (m, 4H); ¹³C NMR (125 MHz, CDCl₃) δ 22.9, 24.2, 26.6, 27.4, 28.1, 28.3, 36.4, 51.4, 56.7, 56.8, 65.9, 70.6, 79.3, 79.6, 80.3, 93.4, 93.7, 126.2, 126.3, 128.3, 129.3, 137.6, 137.7, 152.5, 155.8; LRMS-ESI (*m/z*) [*M* + Na]⁺ 576.

To a stirred solution of above *S*-amine (**22S**) (85 mg, 0.17 mmol) in CH₂Cl₂ (6 mL) and aqueous saturated NaHCO₃ (6 mL) was added 4-methoxybenzenesulfonyl chloride (71 mg, 0.34 mmol). This reaction mixture was stirred for 12 h followed by extraction of the aqueous layer with CH₂Cl₂; the combined organic extracts were dried over anhydrous Na₂SO₄. Removal of solvent under reduced pressure followed by flash chromatography purification (30% EtOAc in hexane as the eluent) provided (*S*)-toluenesulfonate (**23S**) (107 mg, 93%) as a white solid. *R*_f = 0.40 (40% EtOAc in hexane); ¹H NMR (500 MHz, CDCl₃) δ 1.34 (s, 18H), 1.46 (s, 24H), 1.58 (s, 6H), 2.82–2.87 (m, 2H), 2.96–3.13 (m, 4H), 3.18–3.24 (m, 3H), 3.32 (brs, 1H), 3.75 (brs, 1H), 3.80 (brs, 1H), 3.84 (s, 6H), 3.89–3.93 (4H), 4.07 (d, 2H, *J* = 9.2 Hz), 4.16–4.19 (m, 2H), 4.65 (brs, 1H), 4.87 (d, 1H, *J* = 7.31 Hz), 6.93–6.97 (m, 4H), 7.18–7.23 (6H), 7.26–7.28 (m, 4H), 7.64–7.71 (4H); ¹³C NMR (125 MHz, CDCl₃) δ 23.9, 27.3, 28.1, 28.2, 35.6, 52.4, 54.3, 55.2, 55.4, 56.8, 65.6, 71.8, 79.1, 80.9, 93.8, 114.2, 126.1, 128.2, 129.3, 129.4, 129.6, 137.9, 152.6, 155.6, 162.8; LRMS-ESI (*m/z*) [*M* + Na]⁺ 686.

To the above (*S*)-toluenesulfonate (**23S**) (96 mg, 0.14 mmol) in methanol (5 mL) was added *p*-toluenesulfonic acid monohydrate (4.5 mg, 0.02 mmol), and the reaction mixture was allowed to stir at 23 °C. After 8 h the reaction reached maximum conversion and did not move any further. It was quenched with saturated aqueous NaHCO₃, extracted with EtOAc, washed with water and brine, and dried over Na₂SO₄. Flash chromatography purification (30–50% EtOAc in hexane) yielded 13 mg (13%) of starting material and 59 mg (65%) of the expected primary alcohol.

The above alcohol (58 mg, 0.09 mmol), in CH₂Cl₂ (4 mL) and Et₃N (33 μL, 0.23 mmol), was cooled to –10 °C and treated with methanesulfonyl chloride (7.9 μL, 0.1 mmol). The reaction mixture was stirred at 0 °C for 2 h, followed by removal of solvent via rotavap and pump. The resultant residue was redissolved in chloroform (4 mL), treated with DIPEA (65 μL, 0.37 mmol), refluxed for 12 h, and concentrated under reduced pressure. Flash chromatography purification (2% MeOH in CHCl₃ as the eluent) afforded *S*-oxazolidinone **27** (44 mg, 85%) as a white solid. *R*_f = 0.27 (5% MeOH in CHCl₃); ¹H NMR (500 MHz, CD₃OD) δ 1.27 (s, 9H), 2.52 (dd, 1H, *J* = 10.9, 13.7 Hz), 2.67 (dd, 1H, *J* = 8.9, 14.9 Hz), 2.83 (dd, 1H, *J* = 6.1, 14.0 Hz), 3.10 (dd, 1H, *J* = 3.5, 13.8 Hz), 3.47–3.52 (m, 2H), 3.58–3.63 (m, 1H), 3.80–3.83 (m, 1H), 3.85 (s, 3H), 4.23–4.29 (m, 1H), 4.31 (dd, 1H, *J* = 4.8, 8.9 Hz), 4.47 (t, 1H, *J* = 8.6 Hz), 7.06 (d, 2H, *J* = 8.93 Hz), 7.12–7.16 (m, 1H), 7.20–7.24 (m, 4H), 7.75 (d, 2H, *J* = 8.9 Hz); ¹³C NMR (125 MHz, CD₃OD) δ 27.1, 35.6, 52.0, 54.3, 54.6, 54.7, 55.1, 68.0, 73.3, 78.5, 114.0, 125.5, 127.6, 128.9, 129.0, 129.3, 138.6, 156.5, 160.4, 163.3; LRMS-ESI (*m/z*) [*M* + Na]⁺ 572.

tert-Butyl-(2S,3R)-4-(4-amino-N-(((S)-2-oxooxazolidin-4-yl)-methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-ylcarbamate 28. To a stirred solution of *S*-amine (**22S**) (87 mg, 0.17 mmol) in CH₂Cl₂ (6 mL) and aqueous saturated NaHCO₃ (6 mL) was added 4-nitrobenzenesulfonyl chloride (78 mg, 0.35 mmol). This reaction mixture was stirred for 12 h followed by extraction of the aqueous layer with CH₂Cl₂; the combined organic extracts were dried over anhydrous Na₂SO₄. Removal of solvent under reduced pressure followed by flash chromatography purification (30% EtOAc in hexane as the eluent) provided *S*-nitrosulfonamide (**24S**) (102 mg, 85%) as a yellow solid. *R*_f = 0.54 (40% EtOAc in hexane); ¹H NMR (500 MHz, CDCl₃) δ 1.34 (s, 18H), 1.47 (s, 24H), 1.59 (s, 6H), 2.81 (dd, 2H, *J* = 8.0, 12.9 Hz), 3.03 (bd, 2H, *J* = 9.7 Hz), 3.16–3.36 (m, 8H), 3.82 (brs, 2H), 3.91 (dd, 4H, *J* = 5.1, 8.7 Hz), 4.00 (d, 2H, *J* = 6.2 Hz), 4.20 (q, 2H, *J* = 5.6, 5.7 Hz), 4.80 (brs, 1H), 4.86 (d, 1H, *J* = 7.2 Hz), 7.20–7.24 (m, 6H), 7.27–7.30 (m, 4H), 7.88 (d, 4H, *J* = 8.4 Hz), 8.29 (d, 4H, *J* = 8.4 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 23.9, 27.3, 28.1, 28.2, 35.8, 52.3, 54.5, 54.9, 56.6, 65.5, 71.6, 79.5, 81.1, 94.0, 124.2, 126.3,

128.4, 128.4, 128.5, 129.3, 137.7, 144.0, 149.9, 152.7, 155.8, 171.0; LRMS-ESI (*m/z*) [*M* + Na]⁺ 701.

To the above *S*-nitrosulfonamide (**24S**) (92 mg, 0.13 mmol) in methanol (5 mL) was added *p*-toluenesulfonic acid monohydrate (4 mg, 0.02 mmol), and the reaction mixture was allowed to stir at 23 °C. After 18 h the reaction appeared to have moved only 50% to completion, and formation of free amines was evident on TLC. At this point it was quenched with saturated aqueous NaHCO₃, extracted with EtOAc, washed with water and brine, and dried over Na₂SO₄. Flash chromatography purification (30–50% EtOAc in hexane) yielded 32 mg (35%) of starting material and 44 mg (51%) of the expected primary alcohol.

The above alcohol (43 mg, 0.06 mmol), in CH₂Cl₂ (3 mL) and Et₃N (24 μL, 0.17 mmol), was cooled to –10 °C and treated with methanesulfonyl chloride (5.5 μL, 0.07 mmol). The reaction mixture was stirred at 0 °C for 2 h, followed by removal of solvent via rotavap and pump. The resultant residue was redissolved in chloroform (2 mL), treated with DIPEA (29 μL, 0.17 mmol), and refluxed for 12 h; product crashed out of the chloroform and was found to be only slightly soluble in methanol. Solvent was removed under reduced pressure, and the crude material was treated with methanol (4 mL) and Pd/C (5 mg). The mixture was stirred at 23 °C under a hydrogen filled balloon for 3 h, then filtered over Celite, and the filter cake was washed with MeOH. Removal of solvent followed by flash chromatography purification (3% MeOH in CHCl₃ as the eluent) provided (*S*)-oxazolidinone **28** (21 mg, 58%) as a white solid. *R*_f = 0.21 (5% MeOH in CHCl₃); ¹H NMR (500 MHz, 1:1 CD₃OD/CDCl₃) δ 1.24 (s, 9H), 2.54 (d, 1H), 2.56 (dd, 1H, *J* = 8.7, 14.9 Hz), 2.70 (dd, 1H, *J* = 7.6, 14.3 Hz), 2.97 (dd, 1H, *J* = 4.0, 13.9 Hz), 3.32 (dd, 1H, *J* = 5.0, 14.3 Hz), 3.38 (dd, 1H, *J* = 3.1, 14.8 Hz), 3.64–3.70 (m, 1H), 3.78–3.83 (m, 1H), 4.05 (dd, 1H, *J* = 5.3, 8.8 Hz), 4.16–4.20 (m, 1H), 4.39 (t, 1H, *J* = 8.7 Hz), 6.59 (d, 2H, *J* = 8.7 Hz), 7.09–7.15 (m, 3H), 7.18–7.21 (m, 2H), 7.40 (d, 2H, *J* = 8.7 Hz); ¹³C NMR (125 MHz, 1:1 CD₃OD/CDCl₃) δ 31.8, 33.4, 39.4, 53.3, 56.9, 58.5, 58.8, 59.7, 71.8, 77.4, 83.3, 117.6, 127.4, 130.0, 132.0, 133.1, 133.2, 141.9, 156.0, 160.1, 164.0; LRMS-ESI (*m/z*) [*M* + Na]⁺ 557.

(3R,3aS,6aR)-Hexahydrofuro[2,3-*b*]furan-3-yl-(2S,3R)-3-hydroxy-4-(4-methoxy-N-(((R)-2-oxooxazolidin-4-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-ylcarbamate 29. A solution of *S*-aminosulfone **25** (10.9 mg, 0.02 mmol) in 30% trifluoroacetic acid (in CH₂Cl₂, 2 mL) was stirred at 23 °C for 40 min, then concentrated under reduced pressure. The residue was redissolved in CH₂Cl₂ (2 mL), treated with Et₃N (30 μL, 0.21 mmol), followed by carbonate **15** (6.3 mg, 0.02 mmol), and stirred at 23 °C for 12 h. The reaction mixture was then concentrated under reduced pressure and the residue was purified by flash chromatography (3% MeOH in CHCl₃ as the eluent) to give inhibitor **29** (10 mg, 85%) as a white solid. *R*_f = 0.54 (10% MeOH in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.39 (dd, 1H, *J* = 5.5, 13.1 Hz), 1.56–1.63 (m, 1H), 2.67 (dd, 1H, *J* = 10.0, 14.0 Hz), 2.85–2.90 (m, 1H), 2.95–3.01 (m, 2H), 3.10 (dd, 1H, *J* = 4.1, 14.1 Hz), 3.20 (d, 1H, *J* = 12.8 Hz), 3.28 (dd, 1H, *J* = 9.2, 14.1 Hz), 3.63–3.72 (m, 1H), 3.73 (dd, 1H, *J* = 5.2, 9.8 Hz), 3.79 (dt, 1H, *J* = 1.6, 8.2 Hz), 3.86 (s, 3H), 3.88–3.95 (m, 3H), 4.04 (dd, 1H, *J* = 5.0, 8.8 Hz), 4.25 (brs, 1H), 4.28–4.34 (m, 1H), 4.47 (t, 1H, *J* = 8.7 Hz), 4.99 (q, 1H, *J* = 5.6, 13.4 Hz), 5.38 (d, 1H, *J* = 9.8 Hz), 5.63 (d, 1H, *J* = 5.2 Hz), 6.87 (s, 1H), 6.97 (d, 2H, *J* = 8.9 Hz), 7.16–7.22 (m, 3H), 7.24–7.28 (m, 2H), 7.69 (d, 2H, *J* = 8.8 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 25.8, 35.9, 45.4, 51.9, 54.1, 55.0, 55.3, 55.6, 67.6, 69.6, 71.1, 72.5, 73.4, 109.2, 114.5, 126.4, 128.4, 128.4, 129.2, 129.4, 137.5, 155.6, 159.6, 163.3; LRMS-ESI (*m/z*) [*M* + Na]⁺ 628.2; HRMS-ESI (*m/z*) [*M* + Na]⁺ calcd for C₂₈H₃₅N₃O₁₀S 628.1941, found 628.1937.

(3R,3aS,6aR)-Hexahydrofuro[2,3-*b*]furan-3-yl-(2S,3R)-4-(4-amino-N-(((R)-2-oxooxazolidin-4-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-ylcarbamate 30. A solution of the free amine **26** (12 mg, 0.02 mmol) in 30% trifluoroacetic acid in CH₂Cl₂ (2 mL) was stirred at 23 °C for 40 min, then concentrated under reduced pressure. The residue was redissolved in CH₂Cl₂ (2 mL), treated with carbonate **15** (7.1 mg, 0.02 mmol) and Et₃N (40

μL , 0.28 mmol), and stirred at 23 °C for 12 h at 40 °C. The reaction mixture was then concentrated under reduced pressure and the residue was purified by flash chromatography (5% MeOH in CHCl_3 as the eluent) to give inhibitor **30** (11 mg, 85%) as a white solid. $R_f = 0.26$ (10% MeOH in CHCl_3); ^1H NMR (500 MHz, CDCl_3) δ 1.38 (dd, 1H, $J = 5.6$, 13.2 Hz), 1.54–1.63 (m, 1H), 2.61 (dd, 1H, $J = 9.5$, 13.9 Hz), 2.83–2.91 (m, 2H), 3.10 (d, 2H, $J = 13.3$ Hz), 3.18 (dd, 1H, $J = 9.6$, 13.9 Hz), 3.62–3.70 (m, 2H), 3.78–3.84 (m, 3H), 3.88 (dd, 1H, $J = 6.0$, 9.8 Hz), 3.97 (dd, 1H, $J = 5.0$, 8.9 Hz), 4.21–4.27 (m, 1H), 4.41 (t, 1H, $J = 8.7$ Hz), 4.93 (q, 1H, $J = 5.7$, 13.6 Hz), 5.60 (d, 1H, $J = 5.2$ Hz), 5.69 (d, 1H, $J = 8.8$ Hz), 6.64 (d, 2H, $J = 8.7$ Hz), 7.16–7.19 (m, 3H), 7.21–7.26 (m, 2H), 7.45 (d, 2H, $J = 8.7$ Hz); ^{13}C NMR (125 MHz, CDCl_3) δ 25.7, 29.5, 36.1, 45.3, 51.7, 54.1, 54.9, 55.1, 67.5, 69.5, 70.8, 71.0, 73.2, 109.2, 113.9, 123.9, 126.3, 128.3, 129.2, 129.4, 137.7, 151.4, 155.6, 159.6; LRMS-ESI (m/z) [$\text{M} + \text{Na}$] $^+$ 613.2; HRMS-ESI (m/z) [$\text{M} + \text{Na}$] $^+$ calcd for $\text{C}_{27}\text{H}_{34}\text{N}_4\text{O}_9\text{S}$ 613.1944, found 613.1939.

(3R,3aS,6aR)-Hexahydrofuro[2,3-b]furan-3-yl-(2S,3R)-3-hydroxy-4-(4-methoxy-*N*-(((S)-2-oxooxazolidin-4-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-ylcarbamate 31. A solution of *S*-aminosulfonamide **27** (16.5 mg, 0.03 mmol) in 30% trifluoroacetic acid (in CH_2Cl_2 , 3 mL) was stirred at 23 °C for 40 min, then concentrated under reduced pressure. The residue was redissolved in CH_2Cl_2 (3 mL), treated with Et_3N (41 μL , 0.29 mmol), followed by carbonate **15** (10 mg, 0.03 mmol), and stirred at 23 °C for 12 h. The reaction mixture was then concentrated under reduced pressure and the residue was purified by flash chromatography (3% MeOH in CHCl_3 as the eluent) to give inhibitor **31** (16 mg, 90%) as a white solid. $R_f = 0.5$ (10% MeOH in CHCl_3); ^1H NMR (500 MHz, CDCl_3) δ 1.32 (dd, 1H, $J = 5.4$, 13.2 Hz), 1.53–1.62 (m, 1H), 2.62 (dd, 1H, $J = 11.0$, 13.8 Hz), 2.71 (dd, 1H, $J = 7.25$, 14.6 Hz), 2.84–2.89 (m, 2H), 3.10 (dd, 1H, $J = 3.6$, 14.0 Hz), 3.61–3.67 (m, 1H), 3.74–3.82 (m, 2H), 3.86 (s, 3H), 3.87 (dd, 2H, $J = 5.8$, 10.1 Hz), 3.94–3.98 (m, 1H), 4.00–4.05 (m, 2H), 4.27–4.33 (m, 1H), 4.45 (t, 1H, $J = 8.8$ Hz), 4.99 (q, 1H, $J = 5.3$, 7.7 Hz), 5.60 (d, 1H, $J = 5.3$ Hz), 5.68 (d, 1H, $J = 9.2$ Hz), 6.98 (d, 2H, $J = 8.8$ Hz), 7.15–7.20 (m, 1H), 7.22–7.27 (m, 4H), 7.69 (d, 2H, $J = 8.8$ Hz); ^{13}C NMR (125 MHz, CDCl_3) δ 25.7, 43.4, 45.5, 53.2, 54.6, 54.9, 55.6, 56.5, 57.5, 69.6, 71.2, 73.3, 73.5, 109.3, 114.5, 126.3, 128.3, 129.2, 129.4, 137.8, 155.9, 159.61, 163.38; LRMS-ESI (m/z) [$\text{M} + \text{Na}$] $^+$ 628.2; HRMS-ESI (m/z) [$\text{M} + \text{Na}$] $^+$ calcd for $\text{C}_{28}\text{H}_{35}\text{N}_3\text{O}_{10}\text{S}$ 628.1941, found 628.1943.

(3R,3aS,6aR)-Hexahydrofuro[2,3-b]furan-3-yl-(2S,3R)-4-(4-amino-*N*-(((S)-2-oxooxazolidin-4-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-ylcarbamate 32. A solution of the free amine **29** (15 mg, 0.03 mmol) in 30% trifluoroacetic acid (in CH_2Cl_2 , 3 mL) was stirred at 23 °C for 40 min, then concentrated under reduced pressure. The residue was redissolved in CH_2Cl_2 (3 mL), charged with Et_3N (60 μL , 0.42 mmol), followed by carbonate **15** (8.9 mg, 0.03 mmol), and stirred at 23 °C for 12 h at 40 °C. The reaction mixture was then concentrated under reduced pressure and the residue was purified by flash chromatography (5% MeOH in CHCl_3 as the eluent) to give inhibitor **32** (13 mg, 80%) as a white solid. $R_f = 0.32$ (10% MeOH in CHCl_3); ^1H NMR (500 MHz, CDCl_3) δ 1.33 (dd, 1H, $J = 8.5$ Hz), 1.53–1.62 (m, 1H), 2.59 (dd, 1H, $J = 10.5$, 13.9 Hz), 2.66 (dd, 1H, $J = 7.4$, 14.7 Hz), 2.77 (dd, 1H, $J = 8.9$, 14.5 Hz), 2.82–2.89 (m, 1H), 3.10 (dd, 1H, $J = 3.4$, 14.0 Hz), 3.36 (dd, 1H, $J = 3.5$, 14.3 Hz), 3.44 (dd, 1H, $J = 4.5$, 14.6 Hz), 3.62–3.67 (m, 1H), 3.72 (dd, 1H, $J = 5.0$, 9.9 Hz), 3.77 (dt, 1H, $J = 1.8$, 8.2 Hz), 3.87 (dd, 1H, $J = 5.8$, 9.9 Hz), 3.92–3.98 (m, 2H), 3.99 (dd, 1H, $J = 5.0$, 9.0 Hz), 4.15–4.29 (m, 1H), 4.43 (t, 1H, $J = 8.8$ Hz), 4.97 (q, 1H, $J = 5.4$, 13.2 Hz), 5.60 (d, 1H, $J = 5.3$ Hz), 6.66 (d, 2H, $J = 8.7$ Hz), 7.15–7.18 (m, 1H), 7.20–7.25 (m, 4H), 7.49 (d, 2H, $J = 8.6$ Hz); ^{13}C NMR (125 MHz, CDCl_3) δ 25.7, 29.5, 34.7, 45.5, 53.3, 54.6, 55.0, 56.6, 67.5, 69.6, 71.1, 73.4, 109.3, 114.0, 124.2, 126.3, 128.3, 129.2, 129.4, 137.8, 151.2, 155.8, 159.6; LRMS-ESI (m/z) [$\text{M} + \text{Na}$] $^+$ 613.2; HRMS-ESI (m/z) [$\text{M} + \text{Na}$] $^+$ calcd for $\text{C}_{27}\text{H}_{34}\text{N}_4\text{O}_9\text{S}$ 613.1944, found 613.1938.

Determination of X-ray Structure of 19b-Bound HIV Protease (WT). The stabilized HIV-1 protease with the substitutions of Q7K, L33I, L63I, C67A, and C95A that reduce autoproteolysis and aggregation²⁷ was expressed and purified as described.²⁸ These mutations do not alter the inhibitor binding site, and the stabilized protease has kinetic parameters and stability indistinguishable from those of the unsubstituted enzyme.^{27,28} Inhibitor **19b** was dissolved in dimethyl sulfoxide (DMSO). Crystals were grown by the hanging drop vapor diffusion method using 1:5 molar ratio of protease (at 3.9 mg/mL) to inhibitor. The reservoir contained 0.1 M citrate phosphate buffer, pH 5.0, 0.35 M NaCl, and 4% DMSO. Crystals were mounted on a nylon loop and flash-frozen in liquid nitrogen with a cryoprotectant of 30% (v/v) glycerol. X-ray diffraction data were collected on the SER-CAT beamline of the Advanced Photon Source, Argonne National Laboratory. Diffraction data were processed using HKL2000 to 1.29 Å resolution.²⁸ Data were reduced in space group $P2_12_12_1$ with unit cell dimensions of $a = 58.11$ Å, $b = 86.42$ Å, $c = 45.97$ Å with one dimer in the asymmetric unit. The structure was solved by molecular replacement using the CCP4,^{29,30} using WT/GRL0255 complex (PDB 3DJK) as a starting model.²⁶ The structure was refined using SHELX97^{31,32} and refitted manually using the molecular graphics program COOT.³³ Alternative conformations were modeled for the protease residues where observed in the electron density maps. Anisotropic atomic displacement parameters (B -factors) were refined for all atoms including solvent molecules. Hydrogen atoms were automatically added by SHELXL in the last round of the refinement. The identity of ions and other solvent molecules from the crystallization conditions was deduced from the shape and peak height of the $2F_o - F_c$ and $F_o - F_c$ electron density, the hydrogen bond interactions, and interatomic distances. The solvent molecules were one sodium ion, two chloride ions, one glycerol molecule, and 207 water (including partial occupancy sites). The final R was 14.1% for the working set, and R_{free} was 18.2% for all data between 10 and 1.29 Å resolution. The rmsd values from ideal bonds and angle distances were 0.013 and 0.031 Å, respectively. The average B -factor was 15.9 and 22.7 Å² for protease main chain and side chain atoms, respectively, 14.1 Å² for inhibitor atoms, and 32.1 Å² for water atoms. The crystallographic coordinates and structure factors have been deposited in the Protein Databank (PDB) with access code 3H5B.^{34,35}

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Supporting Information Available: HPLC and HRMS data of inhibitors and crystallographic data. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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