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Studies towards the total synthesis of narbonolide: stereoselective preparation of the C1–C10 fragment $\stackrel{\leftrightarrow}{\sim}$

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Abstract

An efficient stereoselective synthesis of the Cl–Cl0 fragment of narbonolide is reported. The stereocentres at C2, 3, 4, 5, 8 and 9 in fragment 5 can be generated via an iterative asymmetric acyl-thiazolidinethione aldol reaction, whereas the stereocentre at C6 is installed by means of Myers alkylation.

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Ketolides¹ are a new class of macrolide antibiotics that have been shown to be active against a variety of bacteria including macrolide-resistant bacteria and mycobacteria. Ketolides differ from erythromycin A by harbouring a 3-keto group instead of a L-cladinose group. The 3-ketone modification has imparted this class of compounds with excellent activities against drug resistant bacterial infections especially the clinically important respiratory tract pathogen *Streptococcus pneumoniae*.²

The therapeutic promise shown by ketolides has led to a resurgence in macrolide antibiotic research in the pharmaceutical industry,³ with successful clinical candidates such as telithromycin^{4a} (1) from Aventis Pharma, and cethromycin (2) from Abbott Laboratories^{4b,c} (Fig. 1).

Narbonolide **3** is a 14-membered polyketide macrolactone biosynthesized by the pikromycin polyketide synthase (PKS) system of *Streptomyces venezuelae* ATCC 15439.⁵ Macrolide **3** consists of seven stereocentres including the sensitive chiral centre at C2. Its significant therapeutic potential, and structural similarity to the macrocytes in

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telithromycin and cethromycin have resulted in total syntheses from the groups of Masamune^{6a} and Fecik.^{6b,c} However, these syntheses have one or more complicating factors such as a low yield of the key macrocyclization reaction, inability to differentiate the C3 and C5 positions for chemoselective reactions and highly optimized protecting group strategies that decrease the synthetic efficiency.

In considering an alternate strategy for the synthesis of narbonolide and its structural analogues, we envisaged that

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the core structure of narbonolide **3** could be constructed via ring closing metathesis of the bis olefin **4**, which in turn could be made via esterification of the C1–C10 fragment **5** and C11–O14 fragment **6**. The stereocentres at C2, 3, 4, 5, 8 and 9 in fragment **5** can be generated via iterative asymmetric acyl-thiazolidinethione aldol reactions, whereas the stereocentre at C6 can be installed by means of Myers alkylation (Scheme 1). Herein, we report the stereoselective synthesis of the C1–C10 fragment of narbonolide.

Accordingly, our synthesis began with the aldolization of *N*-propionyl thiazolidinethione⁷ **7** with acrolein utilizing the Crimmins protocol^{8,9} (titanium mediated enolization using (–)-sparteine (1 equiv) and *N*-methyl-2-pyrrolidinone (1 equiv)) to yield the desired Evans *syn* aldol product **8**¹⁰ in 80% with excellent diastereoselectivity (20:1). Silylation of the aldol product **8** with TBSOTf afforded **9** in good yield. Reductive removal of the chiral auxiliary from thione 9 with LiBH₄ and subsequent iodination of the resulting alcohol 10 yielded iodide 11. Iodide 11 was then subjected to Myers alkylation¹¹conditions. Treatment of iodide 11 with (R,R)-pseudoephedrine propionate 12 at 40 °C proceeded cleanly to furnish amide 13 in 77% yield. Removal of the chiral auxiliary was accomplished with lithium amidotrihydroborate¹² to produce alcohol 14,¹³ which was further oxidized to aldehyde 15, in the presence of TEMPO and iodosobenzene diacetate (Scheme 2).

Aldolization of aldehvde 15 with N-propionvl thiazolidinethione 7 gave the non-Evans syn aldol product 16 via the Crimmins protocol (titanium mediated enolization using 1 equiv of diisopropylethylamine as base).^{8a} The reaction was readily scalable, providing reproducible results in terms of both yield and diastereoselectivity (20:1). Reductive removal of the thiazolidinethione of 16 with NaBH₄ produced diol 17. The 1,3-diol in 17 was then protected with 4-methoxybenzaldehyde dimethylacetal affording the corresponding acetal 18 which after reductive hydrolysis with DIBAL-H afforded primary alcohol 19 in good yield.¹⁴ The resulting alcohol was oxidized to the corresponding aldehyde 20 and subjected to aldol reaction with thiazolidinethione 7 to give non-Evans syn aldol adduct 21 in 83% yield and excellent diastereoselectivity $(\geq 96\%)$.⁸ The aldol product was silvlated to give 22, which on subsequent oxidative hydrolysis afforded C1-C10 fragment 5^{15} with all the required stereocentres (Scheme 3).

In summary, an efficient route for the synthesis of the C1-C10 fragment of narbonolide has been developed, which features aldol reactions to afford both Evans and non-Evans *syn* aldol products from thiazolidinethione 7, and Myers alkylation as key steps. Studies towards the total synthesis of narbonolide and its structural analogues for biological studies are currently underway in our laboratory.



Scheme 2. Reagents and conditions: (a) $TiCl_4$, (–)-sparteine, NMP, acrolein, CH_2Cl_2 , -78 °C to 0 °C, 80% (20:1); (b) TBSOTf, 2,6-lutidine, CH_2Cl_2 , 0 °C, 85%; (c) $LiBH_4$, Et_2O , H_2O , rt, 76%; (d) PPh₃, I_2 , imidazole, Et_2O : CH_3CN (4:1), rt, 79%; (e) **12**, ^{*i*}Pr₂NH, *n*-BuLi, THF, 77%; (f) ^{*i*}Pr₂NH, *n*-BuLi, BH₃NH₃, THF, 90%; (g) TEMPO, iodosobenzene diacetate, CH_2Cl_2 , rt, 95%.



Scheme 3. Reagents and conditions: (a) 7, TiCl₄, ${}^{i}Pr_{2}NEt$, CH₂Cl₂, 0 °C, 74% (20:1); (b) NaBH₄, EtOH, rt, 81%; (c) (OMe)₂CHC₆H₄OMe, CSA, CH₂Cl₂, rt, 92%; (d) DIBAL-H, CH₂Cl₂, 0 °C, 79%; (e) TEMPO, iodosobenzene diacetate, CH₂Cl₂, rt, 95%; (f) 7, TiCl₄, ${}^{i}Pr_{2}NEt$, CH₂Cl₂, 0 °C, 83% (24:1); (g) TBSOTf, ${}^{i}Pr_{2}NEt$, CH₂Cl₂, 0 °C to rt, 81%; (h) LiOH, 30% H₂O₂, THF:H₂O, 0 °C, 65%.

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References and notes

- (a) Agouridas, C.; Denis, A.; Auger, J.-M.; Benedetti, Y.; Bonnefoy, A.; Bretin, F.; Chantot, J.-F.; Dussarat, A.; Fromentin, C.; D'Ambrieres, S. G.; Lachaud, S.; Laurin, P.; Le Martret, O.; Loyau, V.; Tessot, N. J. Med. Chem. 1998, 41, 4080; (b) Zhanel, G. G.; Walters, M.; Noreddin, A.; Vercaigne, L. M.; Wierzbowski, A.; Embil, J. M.; Gin, A. S.; Douthwaite, S.; Hoban, D. J. Drugs 2002, 62, 12.
- Ma, Z.; Clark, R. F.; Brazzale, A.; Wang, S.; Rupp, M. J.; Li, L.; Griesgraber, G.; Zhang, S.; Yong, H.; Phan, L. T.; Nemoto, P. A.; Chu, D. T. W.; Plattner, J. J.; Zhang, X.; Zhong, P.; Cao, Z.; Nilius, A. M.; Shortridge, V. D.; Flamm, R.; Mitten, M.; Meulbroek, J.; Ewing, P.; Alder, J.; Or, Y. S. J. Med. Chem. 2001, 44, 4137.
- 3. Henninger, T. C. Expert Opin. Ther. Pat. 2003, 13, 787.
- (a) Denis, A.; Agouridas, C.; Auger, J.-M.; Benedetti, Y.; Bonnefoy, A.; Bretin, F.; Chantot, J.-F.; Dussarat, A.; Fromentin, C.; D'Ambrieres, S. G.; Lachaud, S.; Laurin, P.; Le Martret, O.; Loyau, V.; Tessot, N.; Pejac, J.-M.; Perron, S. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 3075; (b) Plata, D. J.; Leanna, M. R.; Rasmussen, M.; McLaughlin, M. A.; Condon, S. L.; Kerdesky, F. A. J.; King, S. A.; Peterson, M. J.; Stoner, E. J.; Wittenberger, S. J. *Cheminform* **2005**, *36*, 10; (c) Henninger, T. C.; Xu, X.; Abbanat, D.; Baum, E. Z.; Foleno, B. D.; Hilliard, J. J.; Bush, K.; Hlasta, D. J.; Macielag, M. J. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 4495.
- (a) Hori, T.; Maezawa, I.; Nagahama, N.; Suzuki, M. J. Chem. Soc., Chem. Commun. 1971, 304; (b) Maezawa, I.; Hori, T.; Kinumaki, A.; Suzuki, M. J. Antibiot. 1973, 26, 771; (c) Xue, Y.; Zhao, L.; Liu, H.-W.; Sherman, D. H. Proc. Natl. Acad. Sci. U.S.A. 1998, 95, 12111; (d) Xue, Y.; Sherman, D. H. Metall. Eng. 2001, 3, 15.
- (a) Kaiho, T.; Masamune, S.; Toyoda, T. J. Org. Chem. 1982, 47, 1612; (b) Venkatraman, L.; Aldrich, C. C.; Sherman, D. H.; Fecik, R.

A. J. Org. Chem. 2005, 70, 7267; (c) Venkatraman, L.; Salomon, C.
E.; Sherman, D. H.; Fecik, R. A. J. Org. Chem. 2006, 71, 9853.

- (a) Nagao, Y.; Yamada, S.; Kumagai, T.; Ochiai, M.; Fujita, E. J. *Chem. Soc., Chem. Commun.* **1985**, 1418; (b) Nagao, Y.; Hagiwara, Y.; Kumagai, T.; Ochiai, M.; Inoue, T.; Hashimoto, K.; Fujita, E. J. *Org. Chem.* **1986**, *51*, 2391; (c) Romero-Ortega, M.; Colby, D. A.; Olivo, H. F. *Tetrahedron Lett.* **2002**, *43*, 6439.
- (a) Crimmins, M. T.; Chaudhary, K. Org. Lett. 2000, 2, 775; (b) Crimmins, M. T.; King, B. W.; Tabet, E. A.; Chaudhary, K. J. Org. Chem. 2001, 66, 894; (c) Crimmins, M. T.; Caussanel, F. J. Am. Chem. Soc. 2006, 128, 3128.
- (a) Sai Baba, V.; Das, P.; Mukkanti, K.; Iqbal, J. *Tetrahedron Lett.* 2006, 47, 7927; (b) Nyayavadi, V.; Nanduri, S.; Vasu Dev, R.; Naidu, A.; Iqbal, J. *Tetrahedron Lett.* 2006, 47, 6667.
- (a) Crimmins, M. T.; Christie, H. S.; Chaudhary, K.; Long, A. J. Am. Chem. Soc. 2005, 127, 13810; (b) Crimmins, M. T.; Vanier, G. S. Org. Lett. 2006, 8, 2887.
- (a) Myers, A. G.; Yang, B. H.; Chen, H.; Gleason, J. L. J. Am. Chem. Soc. 1994, 116, 9361; (b) Myers, A. G.; Yang, B. H.; Chen, H.; McKinstry, L.; Kopecky, D. J.; Gleason, J. L. J. Am. Chem. Soc. 1997, 119, 6496.
- Myers, A. G.; Yang, B. H.; Kopecky, D. J. *Tetrahedron Lett.* 1996, 37, 3623.
- 13. Spectral data of compound 14: $[\alpha]_D^{25} + 16.4$ (*c* 0.50, CHCl₃); IR (neat): 3334, 2956, 2927, 1251, 1028, 837, 775 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): $\delta = 5.78$ (ddd, J = 17.0, 10.5, 6.5 Hz, 1H), 5.12 (dd, J = 17.0, 1.5 Hz, 1H), 5.08 (dd, J = 10.5, 1.5 Hz, 1H), 3.96–3.93 (m, 1H), 3.53 (dd, J = 10.5, 4.5 Hz, 1H), 3.39 (dd, J = 10.5, 6.5 Hz, 1H), 1.75–1.73 (m, 1H), 1.72–1.60 (m, 2H), 1.53–1.48 (m, 2H), 0.95 (d, J = 7.0 Hz, 3H), 0.89 (s, 9H), 0.87 (d, J = 7.0 Hz, 3H), 0.04 (s, 3H), 0.01 (s, 3H); ¹³C NMR (CDCl₃, 50 MHz): $\delta = 139.7$, 114.8, 77.5, 67.8, 37.0, 35.9, 33.1, 25.9, 18.2, 17.9, 15.7, -4.3, -4.9; ESI-MS: *m/z* (%) = 273 (100) [M+H]⁺; HRMS (ESI): [M+Na]⁺ calcd for C₁₅H₃₂O₂SiNa: 295.2071; found 295.2069.
- Anderson, J. C.; McDermott, B. P.; Griffin, E. J. *Tetrahedron* 2000, 56, 8747.
- 15. Spectral data of compound 5: $[\alpha]_D^{25}$ +1.6 (c 1.00, CHCl₃); IR (neat): 2954, 2929, 1707, 1514, 1249, 835, 775 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ = 7.26 (d, J = 8.5 Hz, 2H), 6.79 (d, J = 8.5 Hz, 2H), 5.74 (ddd, J = 17.0, 10.5, 6.5 Hz, 1H), 5.05 (dd, J = 17.0, 2.0 Hz, 1H), 5.00

(dd, J = 10.5, 1.2 Hz, 1H), 4.47 (d, J = 11.0 Hz, 1H), 4.36 (d, J = 11.0 Hz, 1H), 4.02–3.99 (m, 1H), 3.90–3.88 (m, 1H), 3.73 (s, 3H), 3.14 (dd, J = 5.5, 3.5 Hz, 1H), 2.67 (dd, J = 7.0, 3.5 Hz, 1H), 2.02–1.98 (m, 1H), 1.83–1.80 (m, 1H), 1.68–1.61 (m, 4H), 1.13 (d, J = 7.0 Hz, 3H), 0.97 (d, J = 6.5 Hz, 3H), 0.94 (d, J = 6.5 Hz, 3H), 0.90 (s, 3H), 0.905 (s, 9H), 0.88 (s, 9H), 0.02 (s, 6H), 0.001 (s, 6H); ¹³C

NMR (CDCl₃, 50 MHz): $\delta = 179.2$, 159.0, 140.1, 131.1, 129.0, 114.7, 113.6, 83.6, 76.4, 74.9, 73.1, 55.2, 43.0, 39.5, 37.5, 36.2, 33.6, 26.1, 25.9, 23.8, 20.8, 18.3, 16.9, 16.8, 11.2, 10.9, -4.0, -4.1, -4.2, -4.8; ESI-MS: m/z (%) = 659 (100) [M+Na]⁺, 637 (95) [M+H]⁺; HRMS (ESI): [M+H]⁺ calcd for C₃₅H₆₅O₆Si₂: 637. 0687; found 637. 0661.