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Novel Terminal Bipheny-Based Diapophytoene Desaturases (CrtN) Inhibitors as Anti-MRSA/VISR/LRSA Agents with Reduced hERG Activity

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Abstract

CrtN has been identified as an attractive and druggable target for treating pigmented S. aureus infections. More than 100 new compounds were synthesized that target the overwhelming the defects of the CrtN inhibitor **1**. Analogs **23a-b** demonstrated significant activity against pigmented *S. aureus* Newman and thirteen MRSA strains ($IC_{50} = 0.02 \sim 10.5$ nM), along with lower hERG inhibition ($IC_{50} > 30$ μ M, ~10-fold decrease in comparison with **1**). Furthermore, **23a-b** were confirmed to reduce the staphylococcal load in the kidney and heart in a mice model with normal treatment deeper than pretreatment ones, comparable even with vancomycin and linezolid. Remarkably, **23a** could strongly block the pigment biosynthesis of these nine multidrug-resistant MRSA strains, including excellent activity against LRSA strains and VISA strains *in vivo*, all which demonstrated that **23a** has huge potential against intractable MRSA, VISA and LRSA issues as a therapeutic drug.

Introduction

Staphylococcus aureus (S. aureus), a major human pathogen, causes serious skin and soft tissues infections,¹ respiratory disease and more serious illness like pneumonia, endocarditis and sepsis.² Infections caused by S. aureus has gained growing concern worldwide due to its capability for the rapid development of drug resistance induced by antibiotic misuse.³ Resistance to antibiotics represents one of the most pressing challenges to the infectious disease community.⁴⁻⁶ Increasing rates of infectious diseases caused by different methicillin-resistant Staphylococcus aureus (MRSA) strains have resulted in high infiltration of hospital and community settings.⁷⁻¹¹ Antibiotics that are approved for treatment of MRSA infections includevancomycin (VAN, glycopeptide antibiotic), linezolid (LZD, oxazolidinone antibiotic), daptomycin (lipopeptide antibiotic), dalbavancin (lipoglycopeptide antibiotic), oritavancin (glycopeptide antibiotic), ceftaroline (β -lactam antibiotic) and tedizolid (anoxazolidinone antibiotic). The appearance of VAN-intermediate resistance MRSA (VISA/MRSA)¹² and LZD-resistant MRSA (LRSA/MRSA) has especially left patients vulnerable. Recently, a report by the U.S. Centers for Disease Control and Prevention (CDC) stated that more than 2 million people have suffered MRSA infections, and 23,000 people have died directly due to these infections.¹³ Great importance has been attached by governments and organizations worldwide, including the ND4BB project initiated by the European Union¹⁴ and National Efforts by the United States.¹⁵ On 27 February 2017, the World Health Organization (WHO)

released its first list of the world's deadliest superbugs, a catalog of 12 families of bacteria. The treatment of MRSA (VISA or LRSA) is urgent at high priority tiers.¹⁶⁻²¹

Anti-virulence strategies aiming at "disarming" the pathogen rather than inhibiting growth with weak selective pressure for the development of antibiotic resistance are now gaining interest.²² Staphyloxanthin (STX) is an important virulence factor for pigmented S. aureus. Nonpigmented S. aureus is susceptible to be killed by reactive oxygen species. Hence, blocking STX biosynthesis is an underlying magnetically therapeutic target against all pigmented MRSA.²³ STX biosynthesis begins with the condensation of farnesyl diphosphate followed by a series of catalytic reactions of important enzymes (CrtM, CrtN, etc.). In 2008, Eric Oldfield and coworkers reported a CrtM inhibitor (BPH-652), which was introduced in our previous work,²⁴ and based on this work, several types of CrtM inhibitors have been identified by the same group, and BPH-652 was selected for these experiments because it had a good IC_{50} in pigment inhibition (110 nM).²⁵ The same group continued to develop a series of CrtM inhibitors (published in 2009) and the most active compounds are halogen-substituted phosphonosulfonates, with Ki values as low as 5 nM against the enzyme and IC_{50} values for STX inhibition in S. aureus as low as 11nM.²⁶ Furthermore, in 2012, Fuyang Lin and coworkers reported X-ray crystallographic structures of three inhibitors bond to CrtM, and their results provided structural clues for the mechanism and inhibition of the head-to-head prenyl transferases.²⁷

In our previous work, we demonstrated the enzyme CrtN to be an attractive and druggable target for fighting pigmented *S. aureus* infection²⁴ and a potent CrtN inhibitor **1** (5m²⁸, Figure 1), which could inhibit the STX biosynthesis of *S. aureus* Newman and three MRSA strains, sensitize four strains to immune clearance and effectively attenuate the virulence of three strains *in vivo*. However, prophylactic administration (24 h before infection) with high-dosage treatment (200 mg/kg) of **1**, along with the high hERG inhibition (IC₅₀ = 3.71 μ M), was impractical for clinical application.

Figure 1

In this study, we synthesized a series of CrtN inhibitors with new scaffolds, which not only demonstrated excellent activity against MRSA *in vitro* and *in vivo* at lower dosage, but also overcame the disadvantages of hERG inhibition. Linezolid and vancomycin, the last-resort antimicrobial agents, were introduced as positive control drugs into a murine model of *S. aureus* abscess formation to evaluate the effectiveness of our new analogues.

Design and Chemistry

In previous work, the varied scaffold and series of different substituents about benzofuran-derived CrtN inhibitors were not enough to investigate the relationship of structure and activity comprehensively. In this study, to further improve the affinity of the lead compound **1** and obtain novel structural scaffolds, chemical modifications were performed in four cycles. First, in region A (Figure 1), we first explored different

heteroaromatic rings and the related positions of substitutions (in region A) to generate novel scaffold compounds **2a-j**. In general, we observed that pigment inhibition varied greatly by the scaffold hopping, and compound **2b** revealed the strongest efficiency among these derivatives, which was enough for further modification study. Additionally, considering the structural diversity, compound **2j**, the second-best scaffold, was also identified as lead compound. Then, we replaced the *N*-methyl group of either **2b** or **2j** in region B (Figure 1) with various steric alkyl group (including hydrogen atom) and 6 analogues (**3a-f**) were designed (Table 1). Subsequently, we incorporated different substituted phenyls, furanyl, naphthalenyls, (cyclo)alkyls at the section C of either **2b** or **2j**, and seventy-four analogs (**4a-40a** and **4b-40b**) were further designed (Table 2-3). In addition, 10 analogs (**41a-45a** and **41b-45b**, Table 4) were prepared to evaluate whether the varies of linkers (allyl in region D) affected pigment inhibitory activity.

The synthesis routes of derivatives 2a-d and 5a-40a were outlined in Scheme 1. Intermediate 47 could be synthesized from commercial indole derivatives 46b via reductive amination, and further coupled with *trans*-4-fluorocinnamaldehyde by the reductive amination to afford the target analog 3a. Replacing the methyl with ethyl group or isopropyl group was to afford target analogs 3b-c. In parallel, 46a-b were suffered with methylamine under the reaction of reductive amination to yield intermediates 48a-b. Various substituted acraldehydes were reduced by sodium borohydride, and accompanied with the bromination reaction for obtaining the intermediates 49a-z and 50a-k. Finally, the nucleophilic reactions of compounds

51a-z and **52a-k** with **48a-b** yielded target analogs **2a-d** and **5a-40a**. Meanwhile, derivatives **2e-j**, **3d-f** and **5b-40b** were prepared as described in scheme 2 and synthesis of these derivatives corresponded closely to scheme 1

Scheme 1-2

Scheme 3 outlines the synthesis strategy for the synthesis of derivatives **41a-b**. Coupling of commercially obtained 4-iodo-1,1'-biphenyl and tert-butyldimethyl(prop-2-yn-1-yloxy)silane gave **56**, which was reduced by TBAF to give **57**, and further brominated by PBr₃ to give intermediate **58**. The coupling of intermediate **58** with either **48b** or **55f** yielded analogs **41a-b**.

Scheme 3

Derivatives **42a-b** were prepared as described in scheme 4. **49t** was reduced by sodium borohydride, followed by bromination to give intermediate **60**. The nucleophilic substitution of intermediate **60** with **48b** or **55f** yielded analogs **42a-b**

Scheme 4

Scheme 5 outlines the synthesis strategy for the synthesis of derivatives **43a-b**. By the Wittig-Horner olefination, **61** was yielded from [1,1'-biphenyl]-4-carbaldehyde, then through reduction and bromination, intermediate **63** was generated. The nucleophilic substitution of intermediate **63** with **48b** or **55f** yielded analogs **43a-b**.

Scheme 5

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Scheme 6 outlines the synthesis strategy used for the synthesis of derivatives **44a-b**. Through Wittig-Horner olefination, intermediate **64** was yielded from [1,1'-biphenyl]-4-carbaldehyde, then through reduction and bromination, intermediate **66** were generated. The nucleophilic substitution of intermediate **66** with **48b** or **55f** yielded analogs **44a-b**.

Scheme 6

As shown in scheme 7, by the Wittig-Horner olefination, intermediate **67** was yielded from (E)-3-([1,1'-biphenyl]-4-yl)acrylaldehyde, followed by reduction of the ethyl ester and bromination reaction to generate **69**. The nucleophilic substitution of compounds **69** with **48b** or **55f** yielded analogs **45a-b**.

Scheme 7

Results and Discussion

In total, 100 novel derivatives (2a-j, 3a-f, 4a-45a and 4b-45b) were designed and synthesized. Their chemical structures are shown in Tables 1-4. The details of the synthesis procedures and structural characterization are described in the *Supporting Information*. All derivatives were confirmed to have \geq 95% purity, and were identified with non-PAINS on the web at http://fafdrugs3.mti.univparis-diderot.fr/ recommended by editors from the ACS (American Chemical Society).

In vitro Pigment Inhibitory Activities against S. aureus Newman

These synthesized compounds were evaluated for pigment inhibitory activities against *S. aureus* Newman and the results are shown in Table 1-4. We replaced the *N*-methyl group with various steric alkyl groups (including hydrogen), and 6 analogs (**3a-f**) were designed (Table 1). As shown in Table 1, in the respect of the substituents with various alkyl groups at *N* atom (including hydrogen atom, **2b**, **2j**, **3a-f**), compounds with *N*-methyl group exhibited the best pigment inhibitory activity (**2b** *vs* **3a-c**; **2j** *vs* **3d-f**), indicating that the *N*-methyl group is the optimal substituent for pigment inhibition.

Table 1

As shown in Table 2-3, substitution of phenyl with various types of groups (\mathbb{R}^3) could significantly affect pigment inhibitory activity. Generally, (cyclo)alkyl or heteroaromatic or 1-naphthyl groups were detrimental to pigment inhibitory activity (**4a-9a** *vs* **2b**, **4b-9b** *vs* **2j**), while the introduction of 2-naphthyl group was favorable (**10a** *vs* **2b**), but inferior to analogue **2j**. Our previous studies found that the electron-withdrawing groups and electron-donating groups at the phenyl ring may have minor effects on activity.²⁸ To investigate the effect of substituted groups, more electron-withdrawing (fluoro, chloro, bromo, trifluoromethyl, difluoromethyl, nitro, phenyl, formate, cyan) and electron-donating groups (methyl, methoxyl, ethoxyl, *t*-butyl) were introduced at the phenyl ring. The results in Table 2-3 demonstrated that there was no relationship between activity and electronic effect. However, the substituted positions at the phenyl ring significantly affected the activity, and

substitution at the para position of the phenyl ring showed the best activity (**12a** *vs* **24a** *vs* **30a**, **12b** *vs* **24b** *vs* **30b**). Moreover, the substitution at the para-position with chloro, bromo, methyl, nitro, difluoromethyl, formate and phenyl (**12a**, **14a**, **15a**, **18a**, **19a**, **22a**, **23a** and **23b**) resulted in significant improvements in activity. Especially, analog **23a** (IC₅₀ = 2.0 ± 0.1 nM) and **23b** (IC₅₀ = 3.3 ± 0.4 nM) displayed better potency than other derivatives. Notably, the R³ group of both **23a** and **23b** was 4-phenylphenyl (biphenyl).

Table 2-3

As shown in Table 4, compounds **41a-45a** and **41b-45b** were assessed to determine whether the allyl linker had influence on the pigment inhibition potency. The results indicated that the change of the allyl linker moiety was detrimental to improve pigment inhibitory activity. When the allyl linker was replaced by propargyl (**41b**, $IC_{50} = 101.8 \pm 5.2 \text{ nM}$) or vinyl (**45b**, $IC_{50}=19.4 \pm 0.2 \text{ nM}$), the pigment inhibitory activity decreased. Additionally, other linkers listed in Table 4 were detrimental to the pigment inhibitory activity.

Table 4

Structure-Activity Relationship

Based on the structural features and pigment inhibitory activities data, the SARs are summarized in detail. (1) *N*-substituent variation exerts great influence on pigment inhibitory activity (**2b** *vs* **3a-c**, **2j** *vs* **3d-f**), and the *N*-methyl is optimal; (2) The impact of the electronic characteristics of substituent in the phenyl ring on the potency is
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weak, while the impact of the substituted position is critical. The substituent group on the para position can enhance the potency significantly(**12a** *vs* **24a** *vs* **30a**, **12b** *vs* **24b** *vs* **30b**); (3) Replacement the phenyl ring with furanyl or thiophene or (cyclo)alkyl is not tolerable to the potency (**2b** *vs* **7a/8a**, **2j** *vs* **7b/8b**); (4) the unsubstituted allyl linker is critical for high potency and the other linkers are on the contrary (**2b** *vs* **41a-45a**, **2j** *vs* **41-45b**).

Cytotoxicity of human embryonic kidney cell (HEK-293) and human hepatocellular carcinoma cell (HepG2)

Based on the *in vitro* pigment inhibitory activities, four compounds (**12a**, **14a** and **23a-b**), whose IC₅₀ values of pigment inhibitory activity of which are lower than 5nM, were selected to for cytotoxicity evaluation in HEK-293 and HepG2. As shown in Table 5, in comparison with amphotericin B, all compounds displayed far less cytotoxicity, especially, **23a** ($CC_{50} = 48.1 \mu$ M by HEK-293, and $CC_{50} = 105.8 \mu$ M by HepG2, respectively) were superior than amphotericin B ($CC_{50} = 4.2 \mu$ M by HEK-293, and $CC_{50} = 3.1 \mu$ M by HepG2, respectively).

Table 5

In vitro Pigment Inhibitory Activities against four MRSA strains

USA400 MW2 and USA300 LAC,²⁹⁻³⁰ two clones liable for the epidemic of community-acquired MRSA (CA-MRSA) infectious diseases in the United States; Mu50, a hospital-acquired MRSA (HA-MRSA) strain with VAN intermediate resistance MRSA (MRSA/VISA); NRS271, a linezolid resistance MRSA

(MRSA/LRSA);³¹⁻³² these four MRSA strains (Table S4, *supporting information*) were used to investigate pigment inhibition activity of **23a-b**. The results are shown in Table 6. We were delighted to find that the color of all strains (USA400 MW2, USA300 LAC, Mu50 and NRS271) faded by the incubation with our compounds **23a-b**, and the IC₅₀ value against MRSA were comparable with that against *S. aureus* Newman. Moreover, according to the results of the bacterial growth assay (Figure 2), different from conditional antibiotics, **23a-b** did not affect the growth of *S. aureus* strains and MRSAs, even given a high concentration (0.2 mM).

Table 6

Figure 2

In vitro Water Solubility, hERG Inhibition and CrtN Enzymatic Inhibitory Activities

Compounds with high affinity for the hERG ion channel could induce QT interval prolongation, which is frequently related to potentially lethal arrhythmias (Table S2, *Supporting Information*). As shown in Table 7, we tested hERG inhibition activity of **23a-b**, and surprisingly found their IC₅₀ values exceeded 30 μ M, which achieved our top priority to decrease the heart cardiotoxicity and gave us much confidence for further development. We next evaluated their capacities of inhibiting the enzymatic activity of CrtN *in vitro*. Using our previous protocol,²⁸ both compounds were found to significantly inhibitenzymatic activity of CrtN at nanomolar concentrations (Table 7), However, although **23a-b** represented excellent

pigment inhibitory activity and cytotoxicity and hERG inhibition, water solubility of **23a-b** decreased a lot than **1** (from 10.0 mg/mL to about 0.2 mg/mL), due to the lipophilic biphenyl.

Table 7

In vitro Metabolic Stability of 23a-b in Human Liver Microsome

Human liver microsome assay was utilized to preliminarily evaluate the effect of the stability of compounds in liver *in vitro*. As shown in Table 8, we took midazolam as control-compound as for its fast clearance rate in human liver microsome, and **23a-b** showed significant stability ($t_{1/2} = 50.7$ mins of **23a** and $t_{1/2} = 93.7$ mins of **23b**).

Table 8

In vitro Anti-fungal Activities of 23a-b.

Because leading compound **1** originated from the anti-fungal agent Naftifine,²⁶ we would like to investigate whether our compound reserved the capacity of antifungal. As shown in Table 9, three fungus strains and three first-line drugs were chosen to proceed with the *in vitro* assay. Compared with the three positive control groups, **23a-b** exhibited weak activities against all three dermatophytes, which verified our compounds losing theirs antifungal activities.

Tables 9

Effects of 23a-b on Sensitizing S. aureus to Immune Clearance.

Since pigment could serve as a protective antioxidant to confer the resistance to immune clearance, we speculated that non-pigment S. aureus could be vulnerable to be killed by the block of STX with the treatment of our CrtN inhibition. Herein, two assays were introduced to investigate the effect of 23a-b on sensitizing S. aureus to immune clearance in vitro, including hydrogen peroxide (H₂O₂) killing assay and human whole blood killing assay. To verify this argument experimentally, we first compared the susceptibility of mock-treatment with 23a-treated (1 μ M) S. aureus to H₂O₂ killing. As shown in Figure 3, non-pigmented S. aureus cells were more vulnerable to be killed by 1.5% H₂O₂ compared to the untreated S. aureus (mock) (survival, 3.0% vs 33.3%). In parallel, the survival of the known antioxidant nacetylcysteine (NAC)-treated S. aureus cells were far worse than the mock treatment as expected (51.3% vs 33.3%). Similarly, the survival percentage of the three MRSA strains greatly decreased (2.3% vs 28.7% in USA400 MW2; 2.3% vs 29.0% in USA300 LAC; 2.7% vs 24.3% in Mu50); correspondingly, the survival rates of the three NAC-treated MRSA strains worsened (54.3% vs 28.7% in USA400 MW2, 52.0% vs 29.0% in USA300 LAC, 53.0% vs 24.3% in Mu50). All results proved that the addition of H_2O_2 (with strong oxidation) exerted an impact on the MRSA strain survival, and the pigment definitely acted as the protective antioxidant. Subsequently, the other experiment was to compare the effect of analog-treated S. aureus with those of non-treated ones by human whole blood killing. The fresh human whole blood was added to 23a-treated (1 μ M) and untreated S. aureus, and the bacterial survival was measured. As shown in Figure 4, untreated S. aureus survived significantly better than

the **23a**-treated *S. aureus* (survival, 13.3% vs 1.3% in Newman, 20.0% vs 1.0% in USA400 MW2, 12.3% vs 0.7% in USA300 LAC, 9.6% vs 0.4% in Mu50. We then repeated the same experiment for **23b**, and the analysis was identical to that of **23a**. All collected results suggested that **23a-b** could render *S. aureus* more susceptible to immune clearance.

Figure 3-4

In vivo Effects of 23a-b on Attenuating the Virulence of *S. aureus* Newman and Two MRSA Strains.

Since 23a-b had promising activities *in vitro*, we next investigated *in vivo* efficacy in a systematic infection model. We subjected *S. aureus* Newman to a murine model of abscess formation via retro-orbital injection and measured the bacterial survival in the host organs. It was worth stressing that we first conducted the normal treatment procedure (giving the compounds 6 h after infection). As shown in Figure 5A-B, the mice were injected with 2.3×10^7 colony-forming units (CFU) of *S. aureus* Newman bacteria via retro-orbital injection. We provided the compounds with four different drug regiments—i.e., both 0.4 mg b.i.d. (giving the compounds twice a day)/4.5 d (180 mg/kg in total), and 0.1 mg b.i.d./4.5 d (45 mg/kg) in the pretreatment case (pretreatment with drugs or compounds 24 h in advance of the infection), along with both 0.4 mg b.i.d./3.5 d (140 mg/kg) and 0.1 mg b.i.d./3.5 d (35 mg/kg) in the normal treatment case (administered 6 h after the infection of strains), while LZD and VAN were set up as two positive control groups at the dosage of 0.4 mg b.i.d./4.5 d

(180 mg/kg) with pretreatment. The mice were sacrificed after 4.5 d, and we measured bacterial survival in kidneys and hearts. First, we analyzed 23a-b in terms of attenuating the virulence of Newman (Figure 5A-B). Compared to the untreated group, the staphylococcal loads of all 23a treated groups were significantly lower in kidneys and hearts (p < 0.01), while **23b**-treated groups were not as obvious. Surprisingly, both 23a-treatment and 23b-treatment with low dosage in the normal treatment group reduced bacterial load more than in the other three groups. For further investigation, we performed the same experiment on infection with Mu50 (VISA/MRSA) and NRS271 (LRSA/MRSA). The analysis of the data in Figure 5C-F revealed that compound-treatment groups had significantly decreased Mu50 and NRS271 staphylococcal loads in the kidneys and hearts, all of which corresponded to a greater than 95.0% decrease in surviving bacteria (significance P < 0.001), while all low-dosage compound-treatment groups were better than the high-dosage ones. Encouragingly, all low-dosage groups were not inferior to the positive-controlled ones; especially, 23a-treatment with low dosage (0.1 mg b.i.d./3.5 d) in the normal treatment case displayed best activity again among the groups.

Figure 5

Along with the encouraging outcome *in vivo* described above, we concluded **23a**-treatment with low dosage in the normal treatment group to be the group with the greatest potential. To confirm the most appropriate treatment dosage, we lowered the dosage to 0.05 mg b.i.d./3.5 d (17.5 mg/kg in total) and 0.01 mg b.i.d./3.5 d (3.5

mg/kg). As shown in Figure 6A-B, **23a** (17.5 mg/kg) significantly decreased the Mu50 *staphylococcal* loads in kidneys by 1.09 log₁₀ CFU (more than 91.8% decrease) and in hearts by 2.70 log₁₀ CFU (more than 99.8% decrease); however, it was not comparable to the 0.1 mg b.i.d./3.5 d (35 mg/kg) group. Generally, we confirmed **23a**-treatment with 0.1 mg b.i.d./3.5 d (35 mg/kg) dosage in normal treatment still as the best treatment. To further evaluate the efficacy of **23a** at affecting the outcome of *S. aureus* sepsis, we challenged animals with 2×10^8 CFU Newman bacteria. The untreated mice nearly died out (90% animal) within 4 days, with **23a** resulting in 80% animal survival. By the eighth day, more than 70% of the **23a**-treated mice were alive, demonstrating a slight advantage over the **1**-treated group (Figure 7). This investigation clearly proved that the *in vivo* **23a**-treatment weakened the virulence of *S. aureus* Newman.

Figure 6-7

In vitro Pigment Inhibitory Activities against additional 9 MRSA Strains.

In addition to four *S. aureus* strains above, 9 additional multidrug-resistant MRSA strains—NRS70, NRS100, NRS108, LRSA56, LRSA202,³³ LRSA205, HS663,³⁴ NF65Y and XN108³⁵ (Table S4, *supporting information*), were selected to further explore the effect of the lead compound **23a** against multiple resistant bacteria. First, the pigment inhibition of **23a** were conducted, and the results were shown in Table 10 indicated that **23a** could strongly block the pigment biosynthesis of these

nine MRSA strains (IC₅₀=0.02-10.5 nM), which meant the adaptability of our compound against MRSAs *in vitro*.

Table 10

In vivo Effects of 23a on Attenuating the Virulence of *S. aureus* LRSA102, LRSA56, NF65Y and XN108.

Following, based on *in vitro* pigment inhibitory activities (Table 10) and MIC values of the nine MRSA strains (Table S4, *Supporting Information*), we chose four representative MRSA strains, LRSA56 (LRSA), LRSA202 (LRSA), NF65Y (VISA) and XN108 (VISA) to further investigate *in vivo* effects of **23a** *in vivo* (0.1 mg b.i.d./3.5 d, in normal treatment, Figure 8).

As shown in Figure 8A, the mice were injected with 5.0×10^7 CFU of *S. aureus* LRSA56 bacteria via retro-orbital injection. Compared with untreated group, **23a**-treatment decreased the LRSA56 loads in kidneys by 0.77 log₁₀ CFU (more than 83.0% decrease) and in hearts by 0.73 log₁₀ CFU (more than 81.4% decrease). Compared with LZD-treated group, **23a**-treatment decreased the LRSA56 loads in kidneys by 0.34 log₁₀ CFU (more than 54.3% decrease) and in hearts by 0.55 log₁₀ CFU (more than 71.8% decrease), however, it showed no significant difference compared to the VAN-treatment group. As shown in Figure 7B, the analysis was just like Figure 7A, **23a** could be comparable to LZD, but worse than VAN.

As shown in Figure 7C-D, for VISA-resistance bacteria NF65Y and XN108, all the **23a**-treatment groups had significantly reduced *S. aureus* loads (p < 0.001), and

was comparable to the positive control groups, even though LAZ and VAN were given at higher doses. Especially, in Figure 7C, **23a**-treatment group showed extraordinary activity, exceeding VAN with significance p < 0.01. Consequently, we were quite confident that **23a** was the best candidate for further development.

Figure 8

In vivo Effects of 23a on Attenuating the Virulence of *S. aureus* Newman, Mu50 and NRS271 with Oral Administration.

Because all *in vivo* investigations above were through intraperitoneal injection, oral administration was utilized for more investigations about curative effects of **23a** (0.4 mg b.i.d./3.5 d, 0.2 mg b.i.d./3.5 d and 0.1 mg b.i.d./3.5 d, in normal treatment, Figure 9). We kept VAN and LZD as positive control groups (0.4 mg b.i.d./4.5 d, in pretreatment).

As shown in Figure 9A-B, the mice were injected with 3.1×10^7 CFU of *S. aureus* Newman bacteria via retro-orbital injection. In the kidneys, compared with untreated group, **23a**-treatment decreased the Newman loads by 1.00 log₁₀ CFU (90.0% decrease), but there was no significant difference in hearts. Compared with VAN-treated group, **23a**-treatment decreased the Newman loads in hearts by 0.38 log₁₀ CFU (more than 55.3% decrease with significance p < 0.1.). However, all **23a**-treatment groups were not better than positive-controlled groups. As for Figure 9C-F, the analysis was just like that of Figure 9A, the best dosage of the three **23a**-treatment groups was still 0.2 mg b.i.d./ 3.5 d, and not better than VAN and

LZD-treatment groups. Consequently, the most appropriate dose of 23a was 0.2 mg b.i.d./ 3.5 d with oral administration, which was not comparable to intraperitoneal injection.

Figure 9

In vitro Effects of 23a on CYP Enzymatic Inhibitory Activity.

The study on CYP drug metabolizing enzymatic could be used to predict the side effect of compounds in liver *in vitro*. The data in Table 11 showed the CYP enzymatic inhibition of **23a**, and the results indicated that **23a** exhibited no significant inhibition of CYP1A2, CYP2B6, CYP2C8, CYP2C9, CYP2C19, CYP2D6, CYP3A4M and CYP3A4T (IC₅₀ > 3 μ M), which preliminarily proved that our compound had no side effect in liver.

Table 11

In vivo Rat and Mice Pharmacokinetics (PK) Profile of 23a.

Because of its potency and attractive adaptability, compound **23a** was further evaluated by both rat (Table 12) and mice (Table 13) PK model. Initially, **23a** was characterized by low clearance (1750 mL/h/kg in rat and 1840 mL/h/kg in mice) and a terminal half-life after I.V. administration (2.46 h in rat and 2.05 h in mice), while the peak serum concentrations were observed at 2 h by P.O. administration. As shown in Table 12 and Table 13, the terminal half-life after P.O. administration was 1.29 h in rat and 3.35 h in mice. The oral bioavailability showed obvious difference between

species of **23a** which was 16.3% in rat and 83.3% in mice, which was high enough for a candidate drug. Furthermore, we supplemented PK study of intraperitoneal (I.P.) injection in rat as shown in Table12. Surprisingly, all the PK parameters were significantly higher than the two other methods of administrations, especially the terminal half-life reached 7.61 h, which made us confident that I.P. was the most appropriate method of administration for **23a**. *Table 12-13* Pharmacological Safety The favorable PK profile of **23a** along with its highly desirable inhibitory

potency against CrtN warranted its use in *in vitro* safety studies. The maximum tolerated dose of **23a** was determined for acute toxicity in rat. 30 rat were randomly divided into three groups and given single oral doses of 60 mg/kg, 250 mg/kg, or 1000 mg/kg of **23a** on the first day. The animals treated with **23a** did not exhibit any poisoning symptoms or mortality immediately or during the post-treatment period of 2 weeks. In addition, no abnormal behaviors or significant changes in the water/food consumption and body weight were observed during the period of the experiments. Therefore, **23a** was well tolerated up to a dose of 1000 mg/kg with no acute toxicity.

Resistance Development Study of 23a

As the drug resistance development was the central issue to design and evaluation of new antibiotics, we wondered whether these CrtN inhibitors could avoid the development of drug resistance under sequential H_2O_2 selection pressure.

Erythromycin, a macrolides antibiotics, was included as a positive control to compare with these CrtN inhibitors for their drug resistance development. In the initial susceptibility testing, we determined the MIC value of erythromycin and H_2O_2 were $0.125 \ \mu g/ml$ and 0.15% (v/v), respectively. Subsequent passaging for erythromycin was started with MIC/2 (0.0625 μ g/ml). And for H₂O₂ with CrtN inhibitors, we chose a concentration of 0.15‰ (v/v) H₂O₂ which imposed a selected pressure for bacteria growth and leading to a lagged time for bacteria to reach the exponential phase, and IC_{90} concentrations of **23a-b** for pigment production to initiate passaging. As it was shown in Figure.10, the MIC value of erythromycin against S.aureus Newman strain started to increase once beginning passaging, and the value had increased by a factor of over 16 after 12 passages. By contrast, the IC₅₀ value of the CrtN inhibitors for pigment production of 12th passage of S. aureus Newman (under IC₉₀ concentrations of 23a or 23b) was basically unchanged (1.8nM vs 2.0 nM for 23a, 2.6 nM vs 3.3 nM for **23b**, Table 12). Meanwhile, if CrtN was mutated under the sequential $H_2O_2 + CrtN$ inhibitors selection pressure, colored single colony would be appeared on the TSA plate containing these drugs. As shown in Fig. 11, when ~1000 CFUs of the 12th passage bacteria were spread onto TSA plate containing IC_{90} concentrations of **23a** or 23b, all the colonies still remained non-pigmented. In contrast, pigmented bacteria were observed from TSA plate without CrtN inhibitors. In total, 100 TSA plates containing IC₉₀ concentrations of 23a or 23b were included to determine the frequency of mutations. The frequencies of mutations at IC_{90} concentrations of CrtN

inhibitors for pigment production were less than ~ 1×10^{-5} for both **23a-b** after 12 sequential passaging.

Figure 10-11

Table 14

Finally, the MIC values of **23a** for the three other pathogens were investigated, including enterococcus faecium, *S. aureus* Newman, and pseudomonas aeruginosa, which all belong to the "ESKAPE" family. As shown in Table 15, the three values of MIC of **23a** were all above 500 μ g/mL, which proved our inhibitor do not affect the growth of these pathogens.

Table 15

Conclusion

In summary, based on the results of scaffold hopping, 3-substituted benzofuran and 7-substituted indole were key skeletons. Subsequently, with the variation of the *N*-substituents, allyl linkers and phenyl moiety, 100 new analogs were synthesized. To investigate the pigment inhibition of *S. aureus* Newman, the unambiguous SARs were obtained. The most valid four pigment inhibitors **12a**, **14a** and **23a-b** (pigment inhibition $IC_{50} < 5nM$) were chosen among the analogs to test their cytotoxicity for HEK-293 and HepG2 cells, in order to keep the diversity of main structure, we maintained **23a-b** for further study. **23a-b** had the capacity to block the pigment

biosynthesis of USA400 MW2, USA300 LAC, Mu50 and NRS271 at a comparable level to the Newman strain. Notably, the hERG inhibition activity of 23a (IC₅₀ = 34.8) μ M) and **23b** (IC₅₀ > 40 μ M) were largely improved compared to **1** (IC₅₀ = 3.71 μ M), ~ 10 fold decrease. We found that 23a-b had submicromolar activity in CrtN enzymatic inhibition assay without any bactericidal impact on S. aureus bacteria (up to 200 μ M) and proper human miarosome stability (50.7 mins of 23a and 93.7 mins of 23b for half-life time). Furthermore, 23a-b abandoned the antifungal activities (MIC > 8 $\mu g/mL$). According to the concept of antivirulence, **23a-b** (1 μ M)-sensitized S. aureus strains were killed by H_2O_2 and human whole blood. In the *in vivo* assay, **23a-b** were proven efficacious in the S. aureus Newman and multidrug resistant MRSA (Mu50 and NRS271) model, and 23a treatment with 0.1 mg b.i.d./3.5 d (35 mg/kg in total) in the normal treatment group was preferred. Next, all compound-treatment groups significantly decreased Mu50 (VISA/MRSA) loads in hearts and kidneys, which were compared with the efficacy of the positive groups, VAN and LZD. Regarding NRS271 (LRSA/MRSA), 23a-b strongly decreased the NRS271 loads in kidneys and hearts (> 99% decrease), which were also comparable with the positive control groups. Considering 23a had a slight advantage over 23b, we preferred 23a as the candidate drug and then proved it to be efficacious in S. aureus Newman sepsis model, with more than 70% survival after 8 d. Considering no better inhibitory activity under the lower dosage, the 0.1 mg b.i.d./3.5 d (35 mg/kg) case was guaranteed to be the most appropriate dosage. The pigment inhibitory activity of 23a against another nine MRSA strains was at the same level to the Newman strain, which demonstrated that 23a had a

broad spectrum bactericidal effect. Then two LRSA strains (LRSA56 and LRSA202) and two VISA strains (NF65Y and XN108) among the nine strains were tested in vivo, and 23a showed extraordinary activity again, comparable to LZD and VAN. Moreover, 23a showed no inhibitory activity on CYP enzymatic, and is characterized by an acceptable half-life, high volume of distribution and low clearance. 23a was a potential antibiotic and had 16.3% orally bioavailable in rat and 83.8% orally bioavailable in mice. When we tried I.P. as method of administration in rat, 23a showed extreme stability (7.61 h for half-life time). Lastly, we verified that 23a was not easy to induce the resistance development, and used **23b** as contrast. We developed 12 passages of S. aureus Newman under pressure of 23a-b and H₂O₂, and made sure that the frequencies of mutations at IC₉₀ concentrations of CrtN inhibitors for pigment production were $< \sim 1 \times 10^{-5}$. In total, 23a has the potential to be developed as therapeutic drugs, especially against intractable MRSA (VISA and LRSA) issues, by blocking virulence, and this class of antibiotics holds great promise in treatment of infections by difficult human pathogens.

Experimental Section

General Chemistry

Reagents and solvents were obtained from commercial suppliers at high quality and were used without further purification. TLC was performed on a HSGF 254 (150-200 μ m thickness; Yantai Huiyou Co., China). UV light and I₂ were used to monitor synthetic progress. Column chromatography was performed on silica gel (200-300 mesh), eluted with ethyl acetate and petroleum ether. NMR spectra data were obtained on a Bruker AMX-400 NMR using TMS as an internal standard. Chemical shifts were provided in parts per million. ¹H NMR data were reported from the aspect of multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet and br = broad), coupling constant (Hz) and integrated value. Low- and high-resolution mass spectral (MS) data were acquired with electron spray ionization (ESI) produced by a Finnigan MAT-95 and LCQ-DECA spectrometer. The purity of each compound (> 95%) was determined by HPLC on an Agilent 1100 with a quaternary pump and diode-array detector (DAD). The melting points of each compound were determined on an SGW X-4 melting point apparatus.

Preparation of Salts.

Taking **4a** as an example, to a solution of oily derivative **4a** (100 mg) in ethyl ether (10 mL) stirred at room temperature bubbled with hydrogen chloride gas for 1 min. After stirring for 15 min, the solvent was removed by rotary evaporation and the residue was suspended in ethyl acetate/petroleum ether (1:100, v/v, 10 mL) for an additional hour of agitation. The precipitate was filtrated and washed with ethyl acetate to obtain the final compound in the form of hydrochloride. All other final derivatives underwent through this process to yield an amorphous, solid form or oil. Spectroscopic data reported below are in their hydrochloride form.

(E)-N-(benzofuran-2-ylmethyl)-N-methyl-3-(4-(trifluoromethyl))phenyl)pro p-2-en-1-amine hydrochloride (2a).

A solution of intermediate 48a (X=O, 163.0 mg, 1 mmol), 51a (290.0 mg, 1.1 mmol) and K₂CO₃ (280.0 mg, 2 mmol) in DMF (10 mL) was stirred at room temperature overnight. The mixture was poured into water and extracted with EtOAc. The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, filtered and condensed. The residue was then purified via flash chromatography on silica gel, eluting with EtOAc/petroleum ether (1/5, v/v) to give the free base of **2a** as white solid. Yield: 44%. 2a was prepared by the general procedure given above as white solid, m.p. 161-162 °C. 1H NMR (400 MHz, MeOD) δ 7.72 – 7.63 (m, 5H), 7.60 – 7.54 (m, 1H), 7.45 – 7.36 (m, 1H), 7.35 – 7.27 (m, 1H), 7.20 (s, 1H), 7.02 (d, J = 15.8 Hz, 1H), 6.56 - 6.46 (m, 1H), 4.66 (s, 2H), 4.10 (dd, J = 14.3, 7.1 Hz, 2H), 2.95 (s. 3H); ¹³C NMR (101 MHz, MeOD) δ 155.67, 146.49, 139.13, 138.76, 127.49, 127.22, 127.22, 125.67, 125.50, 125.33, 125.29, 124.59, 123.34, 121.54, 119.72, 111.05, 110.86, 57.44, 51.29, 39.09. HRMS (ESI) m/z calcd for $C_{20}H_{19}F_3NO (M+H)^+$ 346.1419, found 346.1418.

(E)-N-(benzofuran-3-ylmethyl)-N-methyl-3-(4-(trifluoromethyl))phenyl)pro p-2-en-1-amine hydrochloride (2b).

Yield: 57%. **2b** was synthesized by the general procedure given above as white solid, m.p. 165-166 °C. ¹H-NMR (400 MHz, MeOD) δ 8.13 (s, 1H), 7.82 (d, J = 8.1 Hz, 1H), 7.73–7.64 (m, 5H), 7.60 (d, J = 7.8 Hz, 1H), 7.41 (dtd, J = 17.8, 7.3, 1.2 Hz, 2H), 7.01 (d, J = 15.9 Hz, 1H), 6.59–6.47 (m, 1H), 4.61 (d, J = 37.0 Hz, 2H), 4.24–4.07 (m, 1H), 4.00 (s, 1H), 2.92 (s, 3H). ¹³C NMR (101 MHz, MeOD) δ 157.89,

155.54, 147.94, 139.07, 138.78, 130.18, 127.26, 127.26, 126.49, 125.30, 124.15, 123.48, 120.12, 119.50, 119.27, 111.52, 109.86, 57.26, 47.87, 38.56. HRMS (ESI) m/z calcd for $C_{20}H_{19}F_{3}NO(M+H)^{+}$ 346.1419, found 346.1418.

(E)-N-(benzo[b]thiophen-2-ylmethyl)-N-methyl-3-(4-(trifluoromethyl))phen yl)prop-2-en-1-amine hydrochloride (2c).

Yield: 42%. **2c** was synthesized by the general procedure given above as white solid. m.p. 223-224 °C. ¹H-NMR (400 MHz, MeOD) δ 7.99 – 7.85 (m, 2H), 7.75 – 7.63 (m, 5H), 7.49 – 7.37 (m, 2H), 7.02 (d, J = 15.8 Hz, 1H), 6.53 (dt, J = 15.2, 7.4 Hz, 1H), 4.73 (s, 2H), 4.02 (s, 2H), 2.94 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 141.02, 139.18, 139.11, 138.93, 130.89, 129.50, 127.30, 127.30, 125.57, 125.51, 125.33, 125.30, 124.80, 124.12, 122.81, 122.08, 119.48, 57.18, 53.43, 38.51.HRMS (ESI) m/z calcd for C₂₀H₁₉F₃NS (M+H)⁺ 362.1190, found 360.1193.

(E)-N-(benzo[b]thiophen-3-ylmethyl)-N-methyl-3-(4-(trifluoromethyl))phen yl)prop-2-en-1-amine hydrochloride (2d).

Yield: 50%. **2d** was synthesized by the general procedure given above as white solid. m.p. 198-200 °C. 1H NMR (400 MHz, MeOD) δ 8.02 (s, 3H), 7.68 (s, 3H), 7.50 (d, J = 19.3 Hz, 2H), 6.99 (d, J = 16.0 Hz, 1H), 6.53 (s, 1H), 4.71 (d, J = 53.3 Hz, 2H), 4.09 (d, J = 40.8 Hz, 2H), 2.90 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 140.38, 139.08, 138.87, 137.86, 131.76, 127.27, 127.27, 125.50, 125.35, 125.31, 125.27, 124.98, 124.78, 124.48, 122.81, 121.27, 119.55, 57.55, 51.26, 38.81. HRMS (ESI) m/z calcd for C₂₀H₁₈F₃NS [M+H]⁺ 361.1112, found 361.1114.

(E)-N-((1H-indol-2-yl)methyl)-N-methyl-3-(4-(trifluoromethyl)phenyl)prop-2-en-1-amine hydrochloride (2e).

Yield: 25%. **2e** was synthesized by the general procedure given above as white oil. ¹H NMR (400 MHz, Acetone) δ 7.98 (s, 1H), 7.71 – 7.64 (m, 4H), 7.59 (d, J = 8.0 Hz, 1H), 7.42 (d, J = 8.3 Hz, 1H), 7.20 (t, J = 7.4 Hz, 1H), 7.07 (t, J = 7.5 Hz, 1H), 7.00 (d, J = 15.8 Hz, 1H), 6.55 – 6.45 (m, 1H), 4.62 (t, J = 11.2 Hz, 1H), 4.50 (d, J = 13.9 Hz, 1H), 4.11 (dd, J = 13.1, 7.0 Hz, 1H), 3.94 (dd, J = 13.4, 7.7 Hz, 1H), 2.89 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 139.12, 138.76, 137.21, 127.90, 127.25, 127.25, 125.78, 125.50, 125.32, 125.28, 124.51, 122.84, 120.46, 119.80, 119.63, 111.21, 106.12, 57.05, 52.11, 38.49. HRMS (EI) m/z calcd for C₂₀H₁₉F₃N₂ (M)⁺ 344.1500, found 344.1496.

(E)-N-((1H-indol-3-yl)methyl)-N-methyl-3-(4-(trifluoromethyl)phenyl)prop-2-en-1-amine hydrochloride (2f).

Yield: 78%. **2f** was synthesized by the general procedure given above as white solid. m.p. 165-168 °C. ¹H NMR (400 MHz, MeOD) δ 7.64 (d, J = 7.9 Hz, 1H), 7.62 – 7.54 (m, 4H), 7.35 (t, J = 7.5 Hz, 1H), 7.26 (s, 1H), 7.12 (dd, J = 11.0, 4.0 Hz, 1H), 7.05 (dd, J = 11.0, 3.9 Hz, 1H), 6.66 (d, J = 15.9 Hz, 1H), 6.48 (dt, J = 15.9, 6.8 Hz, 1H), 3.86 (s, 2H), 3.33 (d, J = 6.9 Hz, 2H); ¹³C NMR (101 MHz, MeOD) δ 139.09, 138.51, 136.68, 128.08, 127.19, 127.05, 127.05, 126.00, 125.37, 125.33, 124.57, 122.18, 120.09, 119.94, 117.59, 111.65, 102.64, 95.65, 56.57, 50.82, 38.09. HRMS (EI) m/z calcd for C₂₀H₁₉F₃N₂ (M)⁺ 344.1500, found 344.1499.

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(E)-N-((1H-indol-4-yl)methyl)-N-methyl-3-(4-(trifluoromethyl)phenyl)prop-2-en-1-amine hydrochloride (2g).

Yield: 79%. **2g** was synthesized by the general procedure given above as white solid. m.p. 155-157 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.24 (s, 1H), 7.54 (t, *J* = 8.8 Hz, 2H), 7.46 (d, *J* = 8.1 Hz, 2H), 7.33 (d, *J* = 7.9 Hz, 1H), 7.24 – 7.08 (m, 3H), 6.72 (s, 1H), 6.59 (d, *J* = 16.0 Hz, 1H), 6.50 – 6.37 (m, 1H), 3.85 (s, 2H), 3.28 (d, *J* = 6.4 Hz, 2H), 2.29 (d, *J* = 12.3 Hz, 3H); ¹³C NMR (101 MHz, MeOD) δ 139.11, 138.48, 135.18, 128.02, 127.11, 126.85, 126.85, 126.12, 125.39, 125.31, 124.67, 123.12, 122.56, 120.01, 119.84, 117.19, 111.25, 102.59, 95.59, 56.47, 50.72, 38.01. HRMS (EI) m/z calcd for C₂₀H₁₉F₃N₂ (M)⁺ 344.1500, found 344.1499.

(E)-N-((1H-indol-5-yl)methyl)-N-methyl-3-(4-(trifluoromethyl)phenyl)prop-2-en-1-amine hydrochloride (2h).

Yield: 88%. **2h** was synthesized by the general procedure given above as white solid. m.p. 129-131 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.22 (s, 1H), 7.63 – 7.51 (m, 3H), 7.46 (d, *J* = 8.1 Hz, 2H), 7.36 (d, *J* = 8.3 Hz, 1H), 7.20 (d, *J* = 8.0 Hz, 2H), 6.61 – 6.50 (m, 2H), 6.49 – 6.38 (m, 1H), 3.67 (s, 2H), 3.24 (d, *J* = 6.4 Hz, 2H), 2.28 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 138.95, 138.45, 137.11, 127.47, 127.15, 126.85, 125.78, 125.50, 125.33, 125.21, 124.89, 122.68, 120.35, 119.14, 119.53, 111.45, 106.52, 57.55, 53.45, 38.36. HRMS (EI) m/z calcd for C₂₀H₁₉F₃N₂ (M)⁺ 344.1500, found 344.1501.

(E)-N-((1H-indol-6-yl)methyl)-N-methyl-3-(4-(trifluoromethyl)phenyl)prop-2-en-1-amine hydrochloride (2i).

Yield: 65%. **2i** was synthesized by the general procedure given above as white solid. m.p. 134-137 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.16 (s, 1H), 7.60 (d, J = 8.1Hz, 1H), 7.55 (d, J = 8.1 Hz, 3H), 7.46 (d, J = 8.1 Hz, 2H), 7.39 (s, 1H), 7.20 (s, 1H), 7.10 (d, J = 8.0 Hz, 1H), 6.63 – 6.52 (m, 2H), 6.48 – 6.37 (m, 1H), 3.72 – 3.64 (m, 2H), 3.24 (d, J = 6.4 Hz, 2H), 2.29 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 139.04, 138.11, 136.98, 129.03, 127.09, 126.65, 126.65, 126.01, 125.58, 125.47, 124.87, 121.88, 120.02, 118.97, 117.81, 111.94, 101.54, 95.95, 56.67, 50.98, 38.31. HRMS (EI) m/z calcd for C₂₀H₁₉F₃N₂ (M)⁺ 344.1500, found 344.1498.

(E)-N-((1H-indol-7-yl)methyl)-N-methyl-3-(4-(trifluoromethyl)phenyl)prop-2-en-1-amine hydrochloride (2j).

Yield: 74%. **2j** was synthesized by the general procedure given above as white solid. m.p. 146-148 °C. ¹H NMR (400 MHz, MeOD) δ 7.98 (s, 1H), 7.71 – 7.64 (m, 4H), 7.59 (d, *J* = 8.0 Hz, 1H), 7.42 (d, *J* = 8.3 Hz, 1H), 7.20 (t, *J* = 7.4 Hz, 1H), 7.07 (t, *J* = 7.5 Hz, 1H), 7.00 (d, *J* = 12.8 Hz, 1H), 6.55 – 6.45 (m, 1H), 4.62 (t, *J* = 11.2 Hz, 1H), 4.50 (d, *J* = 13.9 Hz, 1H), 4.11 (dd, *J* = 13.1, 7.0 Hz, 1H), 3.94 (dd, *J* = 13.4, 7.7 Hz, 1H), 2.89 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 139.12, 138.76, 137.21, 127.90, 126.45, 126.45, 125.81, 125.47, 125.39, 125.11, 124.35, 122.74, 121.08, 119.81, 119.13, 111.23, 105.89, 57.01, 52.21, 38.39. HRMS (EI) m/z calcd for C₂₀H₁₉F₃N₂ (M)⁺ 344.1500, found 344.1501. (E)-N-((3a,7a-dihydrobenzofuran-3-yl)methyl)-3-(4-(trifluoromethyl)phenyl)prop-2-en-1-amine hydrochloride (3a).

To a solution of **47** (895.0 mg, 5 mmol) in methanol, **49a** (1320.0 mg, 5 mmol) and K₂CO₃ (140.0 mg, 10 mmol) were added in batches at room temperature. Thereafter the reaction mixture was stirred for 4 h and concentrated. The residue was poured into water and extracted with EtOAc. The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, filtered and condensed. The residue was then purified via flash chromatography on silica gel, eluting with EtOAc/petroleum ether (1/1, v/v) to give free base of **3a** as colorless oil. **3a** was prepared using general procedure of salification as colorless oil. Yield: 65%. ¹H-NMR (400 MHz, MeOD) δ 8.15 (s, 1H), 7.88 – 7.78 (m, 1H), 7.61 (s, 1H), 7.36 – 7.33 (m, 2H), 6.94 (dd, *J* = 7.5, 1.0 Hz, 1H), 6.89 (d, *J* = 8.1 Hz, 1H), 6.82 (d, *J* = 23.9 Hz, 2H), 5.65 (s, 1H), 4.62 (d, *J* = 37.6 Hz, 2H), 4.03 (d, *J* = 7.2 Hz, 2H); ¹³C NMR (101 MHz, MeOD) δ 160.71, 155.73, 148.11, 147.99, 130.40, 128.24, 128.17, 127.69, 126.32, 126.22, 124.29, 124.25, 123.04, 121.89, 113.19, 112.78, 110.78, 76.08, 51.45. HRMS (ESI) m/z calcd for C₁₉H₁₇F₃NO (M+H)⁺ 332.1262, found 332.1263.

(E)-N-((3a,7a-dihydrobenzofuran-3-yl)methyl)-N-ethyl-3-(4-(trifluoromethyl))phenyl)prop-2-en-1-amine (3b).

To a solution of the free base of **3a** (360.0 mg, 1.4 mmol) in DMF (10 mL) was added sodium hydride (52 mg, 1.4 mmol) in batches at 0 °C under N₂ atmosphere. The reaction mixture was stirred for 15 min and iodoethane (219.0 μ L, 2.7 mmol) was

added into the solution. The mixture was stirred at room temperature overnight, poured into water and extracted with EtOAc. The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, filtered and condensed. The residue was then purified via flash chromatography on silica gel, eluting with EtOAc/petroleum ether (1/5, v/v) to give the free base of **11b** as colorless oil. Yield: 50%..¹H-NMR (400 MHz, MeOD) δ 8.15 (s, 1H), 7.80 (d, *J* = 7.6 Hz, 1H), 7.59 (d, *J* = 6.4 Hz, 2H), 7.44 (ddd, *J* = 9.5, 5.9, 2.2 Hz, 3H), 7.41 – 7.33 (m, 2H), 6.98 (d, *J* = 15.8 Hz, 1H), 6.51 – 6.33 (m, 1H), 4.63 (s, 2H), 4.06 (dd, *J* = 7.0, 3.4 Hz, 2H), 3.37 (q, *J* = 7.3 Hz, 2H), 1.47 (t, *J* = 7.3 Hz, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.48 , 147.91, 140.15, 134.28, 128.57, 128.57, 127.29, 126.93, 126.46, 126.46, 125.28, 123.44, 123.11, 119.34, 119.31, 111.51, 109.83, 54.28, 48.31, 45.28, 8.22. HRMS (ESI) m/z calcd for C₂₁H₂₁NO (M+H)⁺ 360.1575, found 360. 1576.

(E)-N-((3a,7a-dihydrobenzofuran-3-yl)methyl)-N-isopropyl-3-(4-(trifluorom ethyl)phenyl)prop-2-en-1-amine (3c).

Yield: 50%. **3c** was synthesized using general procedure of salification as yellow oil. ¹H-NMR (400 MHz, MeOD) δ 8.03 (d, J = 5.7 Hz, 1H), 7.82 (d, J = 7.3 Hz, 1H), 7.77 (d, J = 7.2 Hz, 1H), 7.58 (dd, J = 7.9, 2.4 Hz, 1H), 7.37 (dddd, J = 33.9, 26.7, 12.6, 5.2 Hz, 4H), 7.20 (t, J = 7.9 Hz, 1H), 6.92 (td, J = 7.1, 4.0 Hz, 1H), 6.81 (d, J =8.1 Hz, 1H), 4.71 – 4.58 (m, 1H), 4.57 – 4.44 (m, 3H), 3.95 – 3.79 (m, 1H), 3.31 (d, J= 1.6 Hz, 6H); ¹³C NMR (101 MHz, MeOD) δ 155.57, 147.96, 139.62, 127.04, 127.04, 126.41, 125.86, 125.80, 125.74, 125.30, 123.47, 123.47, 123.08, 119.31, 114.63, 111.51, 109.86, 57.39, 47.54, 37.50, 37.50. HRMS (ESI) m/z calcd for $C_{22}H_{23}F_3NO(M+H)^+$ 374.1732, found 374.1733.

(E)-N-(benzofuran-3-ylmethyl)-N-methy-2-en-1-amine hydrochloride (4a).

(E)-1-bromobut-2-ene was bought from the chemical reagent companies Yield: 45%. **4a** was synthesized by the general procedure given above as white solid. m.p. 171-173 °C. ¹H-NMR (400 MHz, MeOD) δ 8.15 (s, 1H), 7.87 – 7.81 (m, 1H), 7.60 (dt, J = 10.1, 4.9 Hz, 1H), 7.46 – 7.37 (m, 2H), 6.96 – 6.83 (m, 1H), 6.19 – 6.08 (m, 1H), 4.60 (dd, J = 17.9, 9.2 Hz, 1H), 4.49 – 4.42 (m, 1H), 3.91 (dt, J = 23.7, 11.8 Hz, 1H), 3.74 (dd, J = 13.1, 7.9 Hz, 1H), 2.84 (s, 3H), 1.85 (dt, J = 5.1, 2.5 Hz, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.47, 147.93, 138.69, 126.46, 125.22, 123.42, 119.40, 119.00, 111.45, 109.90, 57.37, 48.17, 38.10, 16.87. HRMS (ESI) m/z calcd for C₁₄H₁₈NO (M+H)⁺ 284.2014, found 284.2008.

(E)-N-((1H-indol-7-yl)methyl)-N-methylbut-2-en-1-amine hydrochloride (4b).

Yield: 62%. **16a** was synthesized by the procedure of **4b** as white solid. m.p. 154-156 °C. ¹H NMR (400 MHz, MeOD) δ 7.67 (s, 1H), 7.36 (s, 1H), 7.19 (s, 1H), 7.11 (s, 1H), 6.56 (s, 1H), 6.02 (s, 1H), 5.62 (s, 1H), 4.43 (s, 2H), 3.71 – 6.38 (m, 2H), 2.65 (s, 3H), 1.71 – 1.61 (m, 3H); ¹³C NMR (126 MHz, MeOD) δ 138.25, 135.47, 129.26, 125.33, 124.17, 122.18, 119.07, 118.91, 112.20, 102.02, 57.51, 55.37, 37.78, 16.73. HRMS (EI) m/z calcd for C₁₄H₁₈N₂ (M)⁺ 214.1470, found 214.1471.

(E)-N-(benzofuran-3-ylmethyl)-3-cyclopentyl-N-methyprop-2-en-1-amine hydrochloride (5a).

Yield: 54%. **5a** was synthesized by the general procedure given above as white solid. m.p. 154-156 °C. ¹H-NMR (400 MHz, MeOD) δ 8.08 (s, 1H), 7.78 (d, *J* = 7.3 Hz, 1H), 7.60 (d, *J* = 7.9 Hz, 1H), 7.47 – 7.34 (m, 2H), 6.07 (dd, *J* = 15.4, 7.7 Hz, 1H), 5.62 (dt, *J* = 14.8, 7.3 Hz, 1H), 4.59 (d, *J* = 14.0 Hz, 1H), 4.42 (d, *J* = 13.9 Hz, 1H), 3.99 – 3.84 (m, 1H), 3.77 – 3.62 (m, 1H), 2.82 (s, 3H), 2.59 (dd, *J* = 15.9, 8.3 Hz, 1H), 1.85 (s, 2H), 1.77 – 1.58 (m, 4H), 1.39 (s, 2H); ¹³C NMR (101 MHz, MeOD) δ 155.44, 148.27, 147.84, 126.39, 125.28, 123.43, 119.27, 115.81, 111.50, 109.94, 57.47, 43.10, 38.13, 32.26, 32.26, 24.71, 24.71. HRMS (ESI) m/z calcd for C₁₈H₂₄NO (M+H)⁺ 270.1858, found 270.1857.

(E)-N-((1H-indol-7-yl)methyl)-3-cyclopentyl-N-methylprop-2-en-1-amine hydrochloride (5b).

Yield: 74%. **5b** was synthesized by the general procedure given above as white solid. m.p. 159-161 °C. ¹H NMR (400 MHz, MeOD) δ 7.66 (d, J = 7.8 Hz, 1H), 7.35 (d, J = 2.4 Hz, 1H), 7.19 (d, J = 7.2 Hz, 1H), 7.10 (t, J = 7.6 Hz, 1H), 6.55 (d, J = 3.1 Hz, 1H), 5.95 (dd, J = 12.8, 7.7 Hz, 1H), 5.59 (dt, J = 8.4, 1.8 Hz, 1H), 4.40 (d, J = 12.1 Hz, 2H), 3.65 (d, J = 7.2 Hz, 2H), 2.64 (s, 3H), 2.53 – 2.49 (m, 1H), 1.84 (d, J = 8.4 Hz, 2H), 1.75 – 1.56 (m, 4H), 1.40 – 1.31 (m, 2H); ¹³C NMR (126 MHz, MeOD) δ 147.78, 135.48, 129.27, 125.35, 124.21, 122.21, 119.02, 115.83, 112.14, 102.03,
57.58, 55.45, 42.95, 37.79, 32.15, 32.15, 24.59, 24.59. HRMS (EI) m/z calcd for $C_{18}H_{26}N_2$ (M)⁺ 268.1939, found 268.1940.

(E)-N-(benzofuran-3-ylmethyl)-3-cyclohexyl-N-methyprop-2-en-1-amine hydrochloride (6a).

Yield: 57%. **6a** was synthesized by the general procedure given above as yellow soil. ¹H-NMR (400 MHz, MeOD) δ 8.10 (s, 1H), 7.80 (d, J = 7.3 Hz, 1H), 7.58 (t, J =9.1 Hz, 1H), 7.47 – 7.30 (m, 2H), 6.19 – 5.97 (m, 1H), 5.71 – 5.53 (m, 1H), 4.52 (d, J= 46.0 Hz, 2H), 3.81 (d, J = 66.3 Hz, 2H), 2.80 (d, J = 14.3 Hz, 3H), 2.65 – 2.51 (m, 1H), 1.85 (dt, J = 11.2, 6.8 Hz, 2H), 1.75 – 1.50 (m, 5H), 1.44 – 1.23 (m, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.49, 149.31, 141.84, 125.28, 123.43, 119.27, 115.81, 112.54, 111.50, 109.94, 57.78, 55.55, 40.68, 37.86, 32.01, 32.01, 25.68, 25.50, 25.50. HRMS (ESI) m/z calcd for C₁₉H₂₆NO (M+H)⁺ 284.2014, found 284.2008.

(E)-N-((1H-indol-7-yl)methyl)-3-cyclohexyl-N-methylprop-2-en-1-amine hydrochloride (6b).

Yield: 72%. **6b** was synthesized by the general procedure given above as white solid. m.p. 165-167 °C. ¹H NMR (400 MHz, MeOD) δ 7.69 (d, J = 7.8 Hz, 1H), 7.37 (d, J = 3.1 Hz, 1H), 7.23 (d, J = 7.1 Hz, 1H), 7.12 (t, J = 7.6 Hz, 1H), 6.56 (d, J = 3.1 Hz, 1H), 5.99 (d, J = 2.4 Hz, 1H), 5.62 – 5.49 (m, 1H), 4.55 (s, 2H), 3.78 (s, 2H), 2.73 (s, 3H), 2.09 (d, J = 6.8 Hz, 1H), 1.85 – 1.45 (m, 5H), 1.35 – 1.25 (m, 2H), 1.24 – 1.05 (m, 3H); ¹³C NMR (101 MHz, MeOD) δ 148.94, 135.59, 129.37, 125.46, 124.35, 122.33, 119.12, 115.51, 112.20, 102.13, 57.78, 55.55, 40.68, 37.86, 32.01, 32.01,

25.68, 25.50, 25.50. HRMS (EI) m/z calcd for $C_{19}H_{26}N_2$ (M)⁺ 282.2096, found 282.2095.

(E)-N-(benzofuran-3-ylmethyl)-3-(furan-2-yl)-N-methylprop-2-en-1-amine hydrochloride (7a).

Yield: 35%. **7a** was synthesized by the general procedure given above as yellow oil. ¹H NMR (400 MHz, MeOD) δ 8.14 (d, J = 20.9 Hz, 1H), 7.81 (d, J = 7.2 Hz, 1H), 7.60 (d, J = 8.0 Hz, 1H), 7.54 (s, 1H), 7.40 (ddd, J = 15.0, 13.9, 6.8 Hz, 2H), 6.76 (dd, J = 25.9, 15.6 Hz, 1H), 6.60 – 6.44 (m, 2H), 6.28 – 6.12 (m, 1H), 4.66 (d, J = 13.9 Hz, 1H), 4.50 (d, J = 13.9 Hz, 1H), 4.10 (dt, J = 14.4, 7.2 Hz, 1H), 3.91 (dd, J = 13.1, 8.3 Hz, 1H), 2.93 – 2.76 (m, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.53, 150.91, 147.94, 143.54, 128.41, 125.28, 125.18, 123.46, 119.29, 111.50, 111.41, 111.07, 110.98, 109.81, 57.37, 48.14, 38.26. HRMS (ESI) m/z calcd for C₁₇H₁₈NO₂ (M+H)⁺ 268.1338, found 268.1334.

(E)-N-((1H-indol-7-yl)methyl)-3-(furan-2-yl)-N-methylprop-2-en-1-amine hydrochloride (7b).

Yield: 66%. **7b** was synthesized by the general procedure given above as white solid. m.p. 147-149 °C. ¹H NMR (500 MHz, MeOD) δ 7.71 (d, J = 7.8 Hz, 1H), 7.53 (s, 1H), 7.36 – 7.21 (m, 1H), 7.28 (t, J = 12.2 Hz, 1H), 7.19 – 7.11 (m, 1H), 6.75 – 6.66 (m, 1H), 6.58 (d, J = 3.1 Hz, 1H), 6.53 – 6.45 (m, 2H), 6.24 – 6.14 (m, 1H), 4.63 (s, 2H), 4.09 – 3.87 (m, 2H), 2.77 – 2.57 (m, 1H); ¹³C NMR (126 MHz, MeOD) δ 150.89, 143.33, 135.50, 129.31, 128.14, 125.39, 124.26, 122.30, 119.06, 113.94,

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112.00, 111.27, 110.71, 102.06, 57.47, 55.55, 37.86. HRMS (EI) m/z calcd for $C_{17}H_{18}N_2O(M)^+$ 266.1419, found 266.1418.

(E)-N-(benzofuran-3-ylmethyl)-N-methyl-3-(thiophen-2-yl)prop-2-en-1-amin e hydrochloride (8a).

Yield: 40%. **8a** was synthesized by the general procedure given above as yellow soil. ¹H-NMR (400 MHz, MeOD) δ 8.12 (s, 1H), 7.80 (d, J = 1.2 Hz, 1H), 7.61 – 7.58 (m, 1H), 7.44 (dd, J = 7.2, 1.2 Hz, 1H), 7.39 – 7.35 (m, 1H), 7.2 5 (t, J = 7.6 Hz, 2H), 7.16 (d, J = 7.5 Hz, 1H), 6.91 (d, J = 15.8 Hz, 1H), 6.41 – 6.31 (m, 1H), 4.66 (t, J =9.5 Hz, 1H), 4.54 – 4.46 (m, 1H), 4.11 (dt, J = 7.0, 5.7 Hz, 1H), 3.94 (dd, J = 13.1, 8.2 Hz, 1H), 2.90 (s, 3H); ¹³C NMR (126 MHz, MeOD) δ 155.42, 147.85, 140.92, 138.17 , 135.09, 129.40, 128.26, 127.21, 125.16, 123.81, 123.35, 119.24, 115.81, 111.39, 57.55, 48.12, 38.27. HRMS (ESI) m/z calcd for C₁₇H₁₈NOS (M+H)⁺ 360.1422, found 360.1421.

(E)-N-((1H-indol-7-yl)methyl)-N-methyl-3-(thiophen-2-yl)prop-2-en-1-amine hydrochloride (8b).

Yield: 43%. **8b** was synthesized by the general procedure given above as white solid. m.p. 144-147 °C. ¹H NMR (400 MHz, MeOD) δ 7.62 (d, J = 7.7 Hz, 1H), 7.35 – 7.30 (m, 2H), 7.16 (d, J = 7.1 Hz, 1H), 7.08 (d, J = 7.0 Hz, 2H), 7.02 – 6.92 (m, 1H), 6.91 (d, J = 4.2 Hz, 1H), 6.53 (d, J = 2.4 Hz, 1H), 6.19 – 6.07 (m, 1H), 4.29 (s, 2H), 3.66 (d, J = 7.2 Hz, 2H), 2.58 (d, J = 8.1 Hz, 3H); ¹³C NMR (101 MHz, MeOD) δ 140.66, 135.49, 130.70, 129.00, 127.20, 126.95, 125.20, 124.98, 123.32, 121.17,

119.38, 118.96, 115.56, 101.76, 58.18, 56.84, 39.11. HRMS (EI) m/z calcd for $C_{17}H_{18}N_2S$ (M)⁺ 282.1191, found 282.1189.

(E)-N-(benzofuran-3-ylmethyl)-N-methyl-3-(naphthalen-1-yl)prop-2-en-1-a mine hydrochloride (9a).

Yield: 46%. **9a** was synthesized by the general procedure given above as white solid. m.p. 149-151 °C. ¹H-NMR (400 MHz, MeOD) δ 8.16 (d, J = 8.7 Hz, 2H), 7.94 – 7.87 (m, 2H), 7.86 – 7.73 (m, 3H), 7.61 (d, J = 7.8 Hz, 1H), 7.59 – 7.34 (m, 5H), 6.49 – 6.34 (m, 1H), 4.67 (d, J = 36.8 Hz, 2H), 4.18 (d, J = 49.8 Hz, 2H), 2.97 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.54, 148.00, 138.05, 133.77, 132.67, 131.01, 129.06, 128.32, 126.456, 126.27, 125.76, 125.26, 124.07, 123.48, 122.98, 122.05, 119.37, 111.51, 109.89, 57.67, 48.18, 38.48. HRMS (ESI) m/z calcd for C₂₃H₂₂NO (M+H)⁺ 328.1701, found 328.1700.

(E)-N-((1H-indol-7-yl)methyl)-N-methyl-3-(naphthalen-1-yl)prop-2-en-1-am ine hydrochloride (9b).

Yield: 39%. **9b** was synthesized by the general procedure given above as white solid. m.p. 137-139 °C. ¹H NMR (400 MHz, MeOD) δ 8.15 (d, J = 8.1 Hz, 1H), 7.88 (dd, J = 8.2, 7.8 Hz, 2H), 7.72 – 7.63 (m, 3H), 7.60 – 7.46 (m, 4H), 7.36 (d, J = 3.1 Hz, 1H), 7.24 (d, J = 7.1 Hz, 1H), 7.12 (t, J = 7.6 Hz, 1H), 6.57 (d, J = 3.1 Hz, 1H), 6.47 – 6.36 (m, 1H), 4.43 (s, 2H), 3.88 (d, J = 7.1 Hz, 2H), 2.71 (s, 3H); ¹³C NMR (126 MHz, MeOD) δ 136.87, 135.43, 134.87, 133.62, 133.29, 130.90, 128.88, 128.30, 128.12, 125.92, 125.49, 125.12, 124.83, 123.73, 123.34, 123.12, 123.02, 121.14,

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118.88, 115.30, 58.32, 56.80, 39.18. HRMS (EI) m/z calcd for $C_{23}H_{22}N_2$ (M)⁺ 326.1783, found 326.1784.

(E)-N-(benzofuran-3-ylmethyl)-N-methyl-3-(naphthalen-2-yl)prop-2-en-1-a mine hydrochloride (10a).

Yield: 51%. **10a** was synthesized by the general procedure given above as white solid. m.p. 148-150 °C. ¹H-NMR (400 MHz, MeOD) δ 8.14 (s, 1H), 7.91 – 7.81 (m, 5H), 7.73 (dd, J = 8.7, 1.7 Hz, 1H), 7.63 – 7.59 (m, 1H), 7.53 – 7.48 (m, 2H), 7.41 (dtd, J = 18.2, 7.3, 1.3 Hz, 2H), 7.12 (d, J = 15.7 Hz, 1H), 6.60 – 6.44 (m, 1H); ¹³C NMR (126 MHz, MeOD) δ 155.43, 147.86, 140.73, 133.63, 133.40, 132.61, 128.09, 127.78, 127.44, 127.24, 126.35, 126.27, 126.17, 125.16, 123.36, 122.87, 119.27, 116.48, 111.39, 109.81, 57.58, 47.94, 38.34. HRMS (ESI) m/z calcd for C₂₃H₂₂NO (M+H)⁺ 328.1701, found 328.1700.

(E)-N-((1H-indol-7-yl)methyl)-N-methyl-3-(naphthalen-2-yl)prop-2-en-1-am ine hydrochloride (10b).

Yield: 55%. **10b** was synthesized by the general procedure given above as white solid. m.p. 173-174 °C. ¹H NMR (400 MHz, MeOD) δ 7.85 – 7.62 (m, 4H), 7.71 (t, *J* = 6.7 Hz, 2H), 7.53 – 7.45 (m, 2H), 7.39 (d, *J* = 3.2 Hz, 1H), 7.30 (d, *J* = 7.1 Hz, 1H), 7.16 (t, *J* = 7.6 Hz, 1H), 7.07 (d, *J* = 6.4 Hz, 1H), 6.58 (d, *J* = 3.2 Hz, 1H), 6.53 – 6.42 (m, 1H), 4.80 – 4.56 (m, 2H), 4.21 – 3.97 (m, 2H), 2.86 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 140.82, 135.62, 133.73, 133.49, 132.73, 129.46, 128.16, 127.87, 127.53, 127.34, 126.36, 126.27, 125.54, 124.42, 122.99, 122.48, 119.20, 116.39, 112.07,

102.22, 57.91, 55.69, 38.15. HRMS (EI) m/z calcd for $C_{23}H_{22}N_2$ (M)⁺ 326.1783, found 326.1786.

(E)-N-(benzofuran-3-ylmethyl)-N-methyl-3-phenylprop-2-en-1-amine hydrochloride (11a).

Yield: 60%. **11a** was synthesized by the general procedure given above as white solid. m.p. 167-169 °C. ¹H-NMR (400 MHz, MeOD) δ 8.12 (s, 1H), 7.80 (m, 1H), 7.60 (m, 1H), 7.48 (d, *J* = 23.1 Hz, 2H), 7.39 (m, 5H), 6.95 (d, *J* = 16.0 Hz, 1H), 6.40 (m, 1H), 4.68 (d, *J* = 13.7 Hz, 1H), 4.52 (d, *J* = 13.5 Hz, 1H), 4.12 (s, 1H), 3.95 (s, 1H), 2.88 (d, *J* = 17.6 Hz 3H); ¹³C NMR (101 MHz, MeOD) δ 155.52, 147.92, 140.80, 135.26, 128.78, 128.48, 128.48, 126.73, 126.73, 126.43, 125.26, 123.45, 119.33, 116.27, 111.49, 109.93, 57.61, 47.35, 38.39. HRMS (ESI) m/z calcd for C₁₉H₂₀NO (M+H)⁺ 278.1545, found 278.1545.

(E)-N-((1H-indol-7-yl)methyl)-N-methyl-3-phenylprop-2-en-1-amine hydrochloride (11b).

Yield: 76%. **11b** was synthesized by the general procedure given above as white solid. m.p. 180-181 °C. ¹H NMR (400 MHz, MeOD) δ 7.76 – 7.67 (m, 1H), 7.51 (d, *J* = 7.0 Hz, 2H), 7.40 – 7.25 (m, 5H), 7.16 (t, *J* = 7.6 Hz, 1H), 6.92 (d, *J* = 4.8 Hz, 1H), 6.57 (t, *J* = 6.4 Hz, 1H), 6.37 – 6.25 (m, 1H), 4.79 – 4.75 (m, 1H), 4.60 – 4.50 (m, 1H), 4.19 – 4.07 (m, 1H), 3.90 – 3.65 (m, 1H), 2.83 (s, 3H); ¹³C NMR (126 MHz, MeOD) δ 138.29, 135.42, 134.24, 134.12, 129.18, 128.39, 128.39, 128.05, 128.05, 125.19,

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125.18, 123.87, 121.85, 119.11, 118.25, 101.97, 57.75, 55.98, 38.40. HRMS (EI) m/z calcd for $C_{19}H_{20}N_2$ (M)⁺ 276.1626, found 276.1625.

(E)-N-(benzofuran-3-ylmethyl)-3-(4-chlorophenyl)-N-methylprop-2-en-1-am ine hydrochloride (12a).

Yield: 51%. **12a** was synthesized by the general procedure given above as white solid. m.p. 147-148 °C. ¹H-NMR (400 MHz, MeOD) δ 8.12 (s, 1H), 7.80 (d, J = 7.4 Hz, 1H), 7.60 (d, J = 8.0 Hz, 1H), 7.50 (d, J = 8.5 Hz, 2H), 7.46 – 7.34 (m, 4H), 6.92 (d, J = 16.3 Hz, 1H), 6.39 (dd, J = 15.2, 7.9 Hz, 1H), 4.59 (d, J = 36.9 Hz, 2H), 4.02 (d, J = 52.6 Hz, 2H), 2.90 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.48, 147.94, 139.33, 134.47, 128.61, 128.61, 128.22, 128.22, 126.41, 125.33 125.29, 123.47, 119.31, 117.20, 111.51, 109.81, 57.43, 47.21, 38.43. HRMS (ESI) m/z calcd for C₁₉H₁₉CINO (M+H)⁺ 312.1155, found 312.1154.

(E)-N-((1H-indol-7-yl)methyl)-3-(4-chlorophenyl)-N-methylprop-2-en-1-ami ne hydrochloride (12b).

Yield: 64%. **12b** was synthesized by the general procedure given above as white solid. m.p. 159-163 °C. ¹H NMR (400 MHz, MeOD) δ 7.66 (d, J = 7.6 Hz, 1H), 7.47 (d, J = 8.4 Hz, 2H), 7.42 – 7.35 (m, 3H), 7.22 (d, J = 7.0 Hz, 1H), 7.12 (t, J = 7.6 Hz, 1H), 6.79 (d, J = 2.8 Hz, 1H), 6.56 (d, J = 2.4 Hz, 1H), 6.44 – 6.33 (m, 1H), 4.41 (s, 2H), 3.79 (d, J = 6.7 Hz, 2H), 2.67 (s, 3H); ¹³C NMR (126 MHz, MeOD) δ 138.21, 135.45, 134.16, 134.09, 129.21, 128.42, 128.42, 128.01, 128.01, 125.24, 125.16,

123.96, 121.97, 119.00, 118.28, 101.96, 57.74, 55.99, 38.42. HRMS (EI) m/z calcd for $C_{19}H_{19}FN_2$ (M)⁺ 310.1237, found 310.1236.

(E)-N-(benzofuran-3-ylmethyl)-3-(4-fluorophenyl)-N-methylprop-2-en-1-am ine hydrochloride (13a).

Yield: 56%. **13a** was synthesized by the general procedure given above as white solid. m.p. 162-164 °C. ¹H-NMR (400 MHz, MeOD) δ 8.11 (s, 1H), 7.80 (d, J = 7.7 Hz, 1H), 7.64 – 7.51 (m, 3H), 7.40 (ddd, J = 15.1, 13.9, 6.8 Hz, 2H), 7.12 (t, J = 8.7 Hz, 2H), 6.92 (d, J = 15.8 Hz, 1H), 6.41 – 6.24 (m, 1H), 4.54 (s, 2H), 4.02 (d, J = 51.4 Hz, 2H), 2.90 (s, 3H). ¹³C NMR (101 MHz, MeOD) δ 155.53, 147.97, 139.51, 128.77, 128.77, 128.69, 126.37, 125.27, 123.46, 119.29, 116.17, 115.38, 115.17, 111.49, 109.88, 57.51, 47.88, 38.37. HRMS (ESI) m/z calcd for C₁₉H₁₉FNO (M+H)⁺ 296.1451, found 296.1450.

(E)-N-((1H-indol-7-yl)methyl)-3-(4-fluorophenyl)-N-methylprop-2-en-1-ami ne hydrochloride (13b).

Yield: 54%. **13b** was synthesized by the general procedure given above as white solid. m.p. 161-163 °C. ¹H NMR (400 MHz, MeOD) δ 7.68 (d, J = 7.9 Hz, 1H), 7.52 (d, J = 8.5Hz, 2H), 7.37 (d, J = 2.4 Hz, 1H), 7.23 (d, J = 4.6 Hz, 1H), 7.16 – 7.07 (m, 3H), 6.81 (d, J = 12.8 Hz, 1H), 6.57 (d, J = 3.1 Hz, 1H), 6.37 – 6.27 (m, 1H), 4.46 (s, 2H), 3.83 (d, J = 7.0 Hz, 2H), 2.70 (s, 3H); ¹³C NMR (126 MHz, MeOD) δ 138.96, 135.48, 131.73, 129.28, 128.61, 128.58, 125.35, 124.20, 122.20, 119.05, 118.15,

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116.45, 115.19, 115.02, 112.35, 102.03, 57.69, 55.70, 38.13. HRMS (EI) m/z calcd for C₁₉H₁₉FN₂ (M)⁺ 294.1532, found 294.1531.

(E)-N-(benzofuran-3-ylmethyl)-3-(4-bromophenyl)-N-methylprop-2-en-1-am ine hydrochloride (14a).

Yield: 43%. **14a** was synthesized by the general procedure given above as white solid. m.p. 164-165 °C. ¹H-NMR (400 MHz, MeOD) δ 8.12 (s, 1H), 7.81 (d, J = 7.4 Hz, 1H), 7.60 (d, J = 8.1 Hz, 1H), 7.54 (d, J = 8.4 Hz, 2H), 7.47 – 7.35 (m, 4H), 6.90 (d, J = 15.8 Hz, 1H), 6.51 – 6.31 (m, 1H), 4.58 (s, 2H), 4.01 (s, 2H), 2.89 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.50, 147.75, 139.49, 134.24, 131.64, 131.64, 128.45, 128.45, 126.39, 125.11, 123.47, 122.58, 119.29, 117.11, 111.30, 109.92, 57.43, 48.09, 38.39. HRMS (ESI) m/z calcd for C₁₉H₁₉BrNO (M+H)⁺ 356.0650, found 358.0636.

(E)-N-((1H-indol-7-yl)methyl)-3-(4-bromophenyl)-N-methylprop-2-en-1-ami ne hydrochloride (14b).

Yield: 51%. **14b** was synthesized by the general procedure given above as white solid. m.p. 131-134 °C. ¹H NMR (400 MHz, MeOD) δ 7.63 (d, J = 8.0 Hz, 1H), 7.51 (d, J = 8.4 Hz, 2H), 7.39 (d, J = 8.4 Hz, 2H), 7.35 (d, J = 3.1 Hz, 1H), 7.18 (d, J = 7.0Hz, 1H), 7.09 (t, J = 7.5 Hz, 1H), 6.72 (d, J = 6.8 Hz, 1H), 6.54 (d, J = 3.1 Hz, 1H), 6.46 – 6.35 (m, 1H), 4.29 (s, 2H), 3.67 (d, J = 6.9 Hz, 2H), 2.59 (s, 3H); ¹³C NMR (126 MHz, MeOD) δ 135.84, 135.38, 135.10, 131.33, 131.33, 128.79, 128.25, 128.04, 128.04, 124.73, 121.69, 121.56, 120.94, 118.83, 115.77, 58.22, 56.94, 39.31. HRMS (EI) m/z calcd for C₁₉H₁₉BrN₂ (M)⁺ 354.0732, found 354.0734.

(E)-N-(benzofuran-3-ylmethyl)-N-methyl-3-(p-tolyl)prop-2-en-1-amine hydrochloride (15a).

Yield: 46%. **15a** was synthesized by the general procedure given above as white solid. m.p. 164-165 °C. ¹H-NMR (400 MHz, MeOD) δ 8.13 (s, 1H), 7.83 (d, J = 7.2 Hz, 1H), 7.62 (dd, J = 15.1, 7.8 Hz, 2H), 7.40 (qd, J = 14.2, 6.8 Hz, 3H), 7.23 – 7.11 (m, 2H), 7.08 (d, J = 16.0 Hz, 1H), 6.59 – 6.38 (m, 1H), 4.61 (s, 2H), 4.09 (s, 2H), 2.91 (s, 3H), 2.38 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.53, 147.88, 140.78, 139.05, 132.50, 129.09, 129.09, 126.70, 126.70, 126.42, 125.26, 123.44, 119.33, 115.11, 111.49, 109.96, 57.75, 47.88, 38.38, 19.88. HRMS (ESI) m/z calcd for $C_{20}H_{22}NO (M+H)^+$ 292.1701, found 292.1702.

(E)-N-((1H-indol-7-yl)methyl)-N-methyl-3-(p-tolyl)prop-2-en-1-amine hydrochloride (15b).

Yield: 76%. **15b** was synthesized by the general procedure given above as white solid. m.p. 122-124 °C. ¹H NMR (400 MHz, MeOD) δ 7.69 (d, J = 7.8 Hz, 1H), 7.46 – 7.35 (m, 3H), 7.25 (d, J = 7.2 Hz, 1H), 7.19 (d, J = 7.9 Hz, 2H), 7.13 – 7.00 (m, 2H), 6.81 (d, J = 15.8 Hz, 1H), 6.57 (t, J = 5.5 Hz, 1H), 6.37 – 6.26 (m, 1H), 4.52 (s, 2H), 3.88 (d, J = 7.3 Hz, 2H), 2.73 (s, 3H), 2.35 (s, 3H); ¹³C NMR (126 MHz, MeOD) δ 140.50, 138.87, 135.50, 132.46, 129.30, 128.95, 128.95, 126.59, 126.59, 125.38, 124.25, 122.26, 119.06, 115.04, 112.15, 102.06, 57.85, 55.57, 37.97, 19.78. HRMS (EI) m/z calcd for C₂₀H₂₂N₂ (M)⁺ 290.1783, found 290.1784.

(E)-N-(benzofuran-3-ylmethyl)-3-(4-bromophenyl)-N-methylprop-2-en-1-am ine hydrochloride (16a).

Yield: 43%. **16a** was synthesized by the general procedure given above as yellow oil. ¹H-NMR (400 MHz, MeOD) δ 8.12 (s, 1H), 7.85 – 7.74 (m, 1H), 7.60 (d, J = 7.8Hz, 1H), 7.46 (d, J = 8.7 Hz, 2H), 7.42 – 7.35 (m, 2H), 6.98 – 6.91 (m, 2H), 6.88 (d, J = 15.7 Hz, 1H), 6.22 (dt, J = 15.5, 7.5 Hz, 1H), 4.66 (d, J = 13.9 Hz, 1H), 4.50 (d, J = 13.9 Hz, 1H), 4.10 (dd, J = 12.7, 7.2 Hz, 1H), 3.91 (dd, J = 13.0, 7.8 Hz, 1H), 3.81 (s, 3H), 2.89 (s, 3H). ¹³C NMR (101 MHz, MeOD) δ 160.61, 155.51, 147.93, 140.59, 128.18, 128.18, 127.87, 126.43, 125.25, 123.45, 119.36, 113.81, 113.81, 113.47, 111.48, 109.91, 57.84, 54.40, 48.13, 38.26. HRMS (ESI) m/z calcd for C₂₀H₂₁NO₂ (M+H)⁺ 307.1572, found 307.1575.

(E)-N-((1H-indol-7-yl)methyl)-3-(4-methoxyphenyl)-N-methylprop-2-en-1-a mine hydrochloride (16b).

Yield: 61%. **16b** was synthesized by the general procedure given above as white solid. m.p. 152-154 °C. ¹H NMR (500 MHz, MeOD) δ 7.68 (d, J = 7.9 Hz, 1H), 7.42 (d, J = 8.7 Hz, 2H), 7.36 (d, J = 3.1 Hz, 1H), 7.29 – 7.20 (m, 1H), 7.12 (t, J = 7.6 Hz, 1H), 6.91 (d, J = 4.6 Hz, 2H), 6.80 (d, J = 2.4 Hz, 1H), 6.56 (d, J = 6.4 Hz, 1H), 6.25 – 6.14 (m, 1H), 4.51 (s, 2H), 3.87 (d, J = 5.5 Hz, 2H), 3.80 (s, 3H), 2.72 (s, 3H); ¹³C NMR (126 MHz, MeOD) δ 160.42, 140.13, 135.50, 129.27, 128.04, 128.04, 127.88, 125.36, 124.26, 122.21, 119.06, 113.67, 113.67, 113.41, 112.25, 102.02, 57.94, 55.49, 54.28, 37.89. HRMS (EI) m/z calcd for C₂₀H₂₂N₂O (M)⁺ 306.1732, found 306.1731.

(E)-N-(benzofuran-3-ylmethyl)-3-(4-ethoxyphenyl)-N-methylprop-2-en-1-am ine hydrochloride (17a).

Yield: 41%. **17a** was synthesized by the general procedure given above as white solid. m.p. 153-154 °C. ¹H-NMR (400 MHz, MeOD) δ 8.12 (d, J = 5.9 Hz, 1H), 7.81 (d, J = 6.0 Hz, 1H), 7.60 (d, J = 8.1 Hz, 1H), 7.47 – 7.34 (m, 4H), 6.96 – 6.79 (m, 3H), 6.30 – 6.14 (m, 1H), 4.66 (d, J = 14.0 Hz, 1H), 4.50 (d, J = 13.8 Hz, 1H), 4.07 (ddd, J = 20.8, 13.6, 7.2 Hz, 3H), 3.91 (dd, J = 13.0, 8.0 Hz, 1H), 2.88 (s, 3H), 1.39 (t, J = 7.0 Hz, 3H); ¹³C NMR (101 MHz, MeOD) δ 159.90, 155.50, 147.96, 140.60, 128.18, 128.18, 127.76, 126.45, 125.25, 123.45, 119.40, 114.33, 113.82, 111.48, 109.91, 63.19, 57.85, 47.95, 38.25, 13.70. HRMS (ESI) m/z calcd for C₂₁H₂₄NO₂ (M+H)⁺ 322.1807, found 322.1808.

(E)-N-((1H-indol-7-yl)methyl)-3-(4-ethoxyphenyl)-N-methylprop-2-en-1-ami ne hydrochloride (17b).

Yield: 67%. **17b** was synthesized by the general procedure given above as white solid. m.p. 148-149 °C. ¹H NMR (400 MHz, MeOD) δ 7.70 (d, *J* = 7.8 Hz, 1H), 7.41 (d, *J* = 8.7 Hz, 2H), 7.36 (t, *J* = 6.6 Hz, 1H), 7.26 (d, *J* = 7.2 Hz, 1H), 7.13 (t, *J* = 7.6 Hz, 1H), 6.90 (t, *J* = 7.5 Hz, 2H), 6.83 – 6.81 (m, 1H), 6.57 (d, *J* = 3.2 Hz, 1H), 6.19 – 6.04 (m, 2H), 4.61 (s, 2H), 4.09 – 4.00 (m, 2H), 3.97 (d, *J* = 7.4 Hz, 2H), 2.81 (d, *J* = 27.0 Hz, 3H), 1.41 – 1.33 (m, 3H); ¹³C NMR (101 MHz, MeOD) δ 159.83, 140.27, 135.59, 129.38, 128.13, 128.13, 127.85, 125.46, 124.31, 122.30, 119.15, 114.29,

114.29, 113.64, 112.41, 102.13, 63.17, 58.09, 55.61, 38.02, 13.70. HRMS (EI) m/z calcd for $C_{21}H_{24}N_2O(M)^+$ 320.1889, found 320.1890.

(E)-N-(benzofuran-3-ylmethyl)-N-methyl-3-(4-nitrophenyl)prop-2-en-1-ami ne hydrochloride (18a).

Yield: 45%. **18a** was synthesized by the general procedure given above as yellow oil °C. ¹H NMR (400 MHz, MeOD) δ 8.25 (d, J = 8.5 Hz, 2H), 8.14 (s, 1H), 7.81 (t, J = 13.0 Hz, 1H), 7.75 (d, J = 8.8 Hz, 2H), 7.60 (d, J = 8.1 Hz, 1H), 7.41 (dq, J = 14.6, 7.2 Hz, 2H), 7.05 (d, J = 15.8 Hz, 1H), 6.63 (dd, J = 15.5, 7.8 Hz, 1H), 4.70 (d, J = 13.6 Hz, 1H), 4.57 (d, J = 14.0 Hz, 1H), 4.19 (dd, J = 13.2, 6.9 Hz, 1H), 4.06 – 3.94 (m, 1H), 2.92 (d, J = 7.7 Hz, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.51, 148.08, 147.82, 141.60, 138.11, 127.68, 127.68, 126.43, 125.28, 123.59, 123.59, 123.48, 121.35, 119.39, 111.50, 109.79, 57.12, 47.89, 38.62. HRMS (ESI) m/z calcd for C₁₉H₁₉N₂O₃ (M+H)⁺ 323.1396, found 323.1393.

(E)-N-((1H-indol-7-yl)methyl)-N-methyl-3-(4-nitrophenyl)prop-2-en-1-amin e hydrochloride (18b).

Yield: 73%. **18b** was synthesized by the general procedure given above as white solid. m.p. 133-136 °C. ¹H NMR (400 MHz, MeOD) δ 8.25 (d, J = 8.7 Hz, 2H), 7.75 – 7.65 (m, 3H), 7.39 (d, J = 2.4 Hz, 1H), 7.26 (d, J = 7.2 Hz, 1H), 7.14 (t, J = 7.6 Hz, 1H), 6.95 (d, J = 15.7 Hz, 1H), 6.65 – 6.54 (m, 2H), 4.53 (s, 2H), 3.94 (d, J = 5.5 Hz, 2H), 2.77 (s, 3H); ¹³C NMR (126 MHz, MeOD) δ 147.23, 142.30, 135.37, 134.98, 128.85, 127.40, 127.17, 127.17, 125.47, 124.82, 123.39, 123.39, 123.28, 121.15,

118.87, 115.33, 57.93, 56.99, 39.42. HRMS (EI) m/z calcd for $C_{19}H_{19}N_3O_2(M)^+$ 321.1477, found 321.1478.

(E)-N-(benzofuran-3-ylmethyl)-3-(4-(difluoromethyl)phenyl)-N-methylprop-2-en-1-amine hydrochloride (19a).

Yield: 55%. **19a** was synthesized by the general procedure given above as white solid. m.p. 145-146 °C. ¹H-NMR (400 MHz, MeOD) δ 8.11 (s, 1H), 7.81 (d, J = 7.5 Hz, 1H), 7.67 – 7.53 (m, 5H), 7.47 – 7.35 (m, 2H), 6.98 (d, J = 16.1 Hz, 1H), 6.84 (d, J = 56.0 Hz, 1H), 6.47 (dt, J = 15.1, 7.5 Hz, 1H), 4.55 (s, 2H), 3.99 (s, 2H), 2.91 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.57, 147.96, 139.62, 137.66, 127.04, 127.04, 126.41, 125.86, 125.80, 125.74, 125.30, 123.47, 119.31, 118.26, 114.63, 111.51, 109.86, 57.39, 48.14, 38.50. HRMS (ESI) m/z calcd for C₂₀H₂₀F₂NO (M+H)⁺ 328.1513, found 328.1512.

(E)-N-((1H-indol-7-yl)methyl)-3-(4-(difluoromethyl)phenyl)-N-methylprop-2 -en-1-amine hydrochloride (19b).

Yield: 64%. **19b** was synthesized by the general procedure given above as yellow oil. ¹H NMR (400 MHz, MeOD) δ 7.48 – 7.30 (m, 5H), 7.25 (d, J = 3.1 Hz, 1H), 7.03 (d, J = 6.9 Hz, 1H), 6.97 (t, J = 7.4 Hz, 1H), 6.86 (s, 1H), 6.72 (s, 1H), 6.60 (d, J = 16.1 Hz, 1H), 6.58 – 6.55 (m, 1H), 6.52 – 6.41 (m, 2H), 3.87 (s, 2H), 3.27 (d, J = 6.6 Hz, 2H), 2.28 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 139.53, 135.34, 131.99, 128.55, 128.42, 126.21, 126.21, 125.51, 125.45, 124.13, 121.62, 120.82, 119.28, 118.60,

117.24, 114.89, 101.15, 59.29, 58.82, 41.06. HRMS (EI) m/z calcd for $C_{20}H_{20}F_2N_2$ (M)⁺ 326.1595, found 326.1596.

(E)-4-(3-((benzofuran-3-ylmethyl)(methyl)amino)prop-1-en-1-yl)benzonitril e hydrochloride (20a).

Yield: 51%. **20a** was synthesized by the general procedure given above as yellow oil. ¹H-NMR (400 MHz, MeOD) δ 8.13 (d, J = 8.3 Hz, 1H), 7.83 (t, J = 7.1 Hz, 1H), 7.74 (d, J = 8.2 Hz, 3H), 7.69 (d, J = 8.3 Hz, 2H), 7.60 (d, J = 8.1 Hz, 1H), 7.47 – 7.35 (m, 3H), 6.99 (d, J = 15.7 Hz, 1H), 6.64 – 6.52 (m, 1H), 4.69 (d, J = 13.9 Hz, 1H), 4.56 (d, J = 14.0 Hz, 1H), 4.18 (dd, J = 13.3, 6.7 Hz, 1H), 4.00 (dd, J = 13.1, 8.1 Hz, 1H), 2.92 (s, 3H); ¹³C NMR (126 MHz, MeOD) δ 155.43, 147.86, 139.73, 138.46, 132.24, 132.24, 127.38, 127.38, 126.30, 125.19, 123.37, 120.51, 119.21, 117.95, 111.79, 111.41, 109.77, 57.08, 48.18, 38.54. HRMS (ESI) m/z calcd for C₂₀H₁₉NO (M+H)⁺ 303.1497, found 303.1496.

(E)-4-(3-(((1H-indol-7-yl)methyl)(methyl)amino)prop-1-en-1-yl)benzonitrile hydrochloride (20b).

Yield: 52%. **20b** was synthesized by the general procedure given above as white solid. m.p. 138-139 °C. ¹H NMR (400 MHz, MeOD) δ 7.67 (d, J = 8.4 Hz, 2H), 7.60 (s, 3H), 7.31 (s, 1H), 7.12 (s, 1H), 7.04 (s, 1H), 6.75 (d, J = 8.6 Hz, 1H), 6.53 (d, J = 4.6 Hz, 2H), 4.19 (s, 2H), 3.54 (d, J = 2.4 Hz, 2H), 2.52 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 140.07, 137.69, 135.55, 132.28, 132.28, 129.34, 127.41, 127.41, 125.38,

124.13, 122.17, 121.62, 119.13, 118.11, 112.96, 111.66, 102.10, 57.59, 56.21, 38.69. HRMS (EI) m/z calcd for $C_{20}H_{19}N_3$ (M)⁺ 301.1579, found 301.1581.

(E)-N-(benzofuran-3-ylmethyl)-3-(4-(tert-butyl)phenyl)-N-methylprop-2-en-1-amine hydrochloride (21a).

Yield: 55%. **21a** was synthesized by the general procedure given above as yellow oil. ¹H-NMR (400 MHz, MeOD) δ 8.16 (s, 1H), 7.85 (dd, J = 7.1, 1.4 Hz, 1H), 7.66 – 7.59 (m, 1H), 7.47 – 7.39 (m, 6H), 6.97 – 6.91 (m, 1H), 6.43 – 6.31 (m, 1H), 4.70 (d, J = 13.9 Hz, 1H), 4.54 (d, J = 13.9 Hz, 1H), 4.19 – 4.10 (m, 1H), 3.99 – 3.89 (m, 1H), 2.92 (s, 3H), 1.34 (s, 9H); ¹³C NMR (126 MHz, MeOD) δ 157.29, 155.43, 147.80, 136.07, 130.02, 127.32, 126.67, 126.67, 125.35, 125.35, 123.69, 123.35, 120.29, 119.18, 116.55, 111.40, 110.76, 109.78, 58.12, 54.50, 38.29, 38.29, 38.29. ¹³C NMR (101 MHz, MeOD) δ 155.47, 152.12, 147.97, 140.54, 132.54, 129.90, 129.85, 126.57, 126.57, 126.51, 125.35, 125.35, 125.21, 123.44, 119.49, 115.54, 111.45, 110.00, 57.73, 47.93, 38.32, 34.16, 30.26, 30.26, 30.26. HRMS (ESI) m/z calcd for C₂₃H₂₈NO (M+H)⁺ 334.2171, found 334.2172.

(E)-N-((1H-indol-7-yl)methyl)-3-(4-(tert-butyl)phenyl)-N-methylprop-2-en-1 -amine hydrochloride (21b).

Yield: 80%. **21b** was synthesized by the general procedure given above as white solid. m.p. 119-122 °C. ¹H NMR (400 MHz, MeOD) δ 7.59 (d, J = 7.8, 1H), 7.37 (d, J = 3.2 Hz, 4H), 7.32 (d, J = 3.2 Hz, 1H), 7.15 (d, J = 7.2 Hz, 1H), 7.09 – 7.03 (m, 1H), 6.69 (d, J = 12.8 Hz, 1H), 6.51 (d, J = 3.2 Hz, 1H), 6.31 (d, J = 7.2 Hz, 1H), 4.25 (d, J

= 6.4 Hz, 2H), 3.82 – 3.62 (m, 2H), 2.58 – 2.54 (m, 2H), 1.32 – 1.09 (m, 9H); ¹³C NMR (101 MHz, MeOD) δ 151.13, 136.49, 135.48, 133.42, 128.78, 126.12, 126.12, 125.39, 125.17, 125.17, 124.94, 124.68, 122.93, 120.68, 118.86, 116.83, 58.76, 57.24, 39.59, 34.05, 30.29, 30.29, 30.29. HRMS (EI) m/z calcd for $C_{23}H_{28}N_2$ (M)⁺ 332.2252, found 332.2253.

Methyl(E)-4-(3-((benzofuran-3-ylmethyl)(methyl)amino)prop-1-en-1-yl)benz oate hydrochloride (22a).

Yield: 55%. **22a** was synthesized by the general procedure given above as yellow oil. ¹H NMR (400 MHz, MeOD) δ 8.13 (s, 1H), 8.02 (d, J = 8.4 Hz, 2H), 7.82 (d, J =7.5 Hz, 1H), 7.62 (t, J = 8.8 Hz, 3H), 7.48 – 7.30 (m, 2H), 7.00 (d, J = 15.8 Hz, 1H), 6.60 – 6.43 (m, 1H), 4.69 (d, J = 13.9 Hz, 1H), 4.54 (d, J = 13.7 Hz, 1H), 4.15 (d, J =7.1 Hz, 1H), 4.04 – 3.94 (m, 1H), 3.91 (s, 3H), 2.92 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 166.60, 155.52, 148.02, 139.81, 139.38, 130.15, 129.60, 129.60, 126.82, 126.82, 126.42, 125.28, 123.47, 119.33, 111.50, 109.82, 57.32, 51.31, 48.16, 38.52. HRMS (ESI) m/z calcd for C₂₁H₂₂NO₃ (M+H)⁺ 336.1600, found 336.1599.

methyl(E)-4-(3-(((1H-indol-7-yl)methyl)(methyl)amino)prop-1-en-1-yl)benzo ate hydrochloride (22b).

Yield: 73%. **22b** was synthesized by the general procedure given above as white oil. ¹H NMR (400 MHz, MeOD) δ 7.97 (s, 2H), 7.54 – 7.45 (m, 3H), 7.29 (s, 1H), 7.09 (s, 1H), 7.02 (s, 1H), 6.72 (d, J = 6.4 Hz, 1H), 6.49 (s, 2H), 4.10 (s, 2H), 3.89 (s, 3H), 3.50 (s, 2H), 2.45 (s, 3H); ¹³C NMR (126 MHz, MeOD) δ 135.59, 133.26, 132.91,

129.46, 129.46, 127.14, 126.58, 126.58, 125.24, 124.18, 123.18, 122.35, 121.98, 119.02, 105.24, 102.01, 57.73, 56.16, 51.18, 38.62. HRMS (EI) m/z calcd for $C_{21}H_{22}N_2O_2$ (M)⁺ 334.1681, found 334.1682.

(E)-3-([1,1'-biphenyl]-4-yl)-N-(benzofuran-3-ylmethyl)-N-methylprop-2-en-1 -amine hydrochloride (23a).

Yield: 61%. **23a** was synthesized by the general procedure given above as white solid. m.p. 157-158 °C. ¹H-NMR (400 MHz, MeOD) δ 8.13 (s, 1H), 7.86 – 7.77 (m, 1H), 7.67 – 7.58 (m, 7H), 7.41 (dddd, J = 23.2, 15.9, 9.3, 4.8 Hz, 5H), 6.99 (d, J =15.7 Hz, 1H), 6.50 – 6.34 (m, 1H), 4.68 – 4.44 (m, 2H), 4.06 (d, J = 55.9 Hz, 2H), 2.92 (s, 3H); ¹³C NMR (101 MHz, DMSO) δ 155.03, 148.79, 140.67, 139.90, 138.50, 135.09, 129.47, 129.47, 128.16, 127.91, 127.91, 127.43, 127.43, 127.40, 127.04, 127.04, 125.58, 123.79, 121.03, 118.87, 112.12, 110.51, 56.77, 47.37, 38.73. HRMS (ESI) m/z calcd for C₂₅H₂₄NO (M+H)⁺ 354.1858, found 354.1857.

(E)-N-((1H-indol-7-yl)methyl)-3-([1,1'-biphenyl]-4-yl)-N-methylprop-2-en-1amine hydrochloride (23b).

Yield: 69%. **23b** was synthesized by the general procedure given above as white solid. m.p. 199-200 °C. ¹H NMR (400 MHz, MeOD) δ 7.72 (d, *J* = 7.7 Hz, 1H), 7.67 – 7.57 (m, 6H), 7.44 (t, *J* = 7.6 Hz, 2H), 7.40 (d, *J* = 3.2 Hz, 1H), 7.34 (t, *J* = 7.4 Hz, 1H), 7.29 (d, *J* = 7.2 Hz, 1H), 7.16 (t, *J* = 7.6 Hz, 1H), 6.94 (d, *J* = 15.7 Hz, 1H), 6.59 (d, *J* = 3.2 Hz, 1H), 6.46 – 6.34 (m, 1H), 4.67 (s, 2H), 4.05 (d, *J* = 6.2 Hz, 2H), 2.83 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 140.23, 140.15, 135.61, 134.28, 129.45, 128.57,

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128.57, 127.38, 127.28, 127.28, 126.92, 126.92, 126.44, 126.44, 125.52, 124.35, 122.44, 119.20, 116.16, 115.62, 112.18, 102.23, 57.92, 55.74, 38.16. HRMS (EI) m/z calcd for $C_{25}H_{24}N_2$ (M)⁺ 352.1939, found 352.1937.

(E)-N-(benzofuran-3-ylmethyl)-3-(2-chlorophenyl)-N-methylprop-2-en-1-am ine hydrochloride (24a).

Yield: 43%. **24a** was synthesized by the general procedure given above as white solid. m.p. 145-146 °C. ¹H-NMR (400 MHz, MeOD) δ 8.11 (s, 1H), 7.82 (d, *J* = 7.2 Hz, 1H), 7.74 – 7.69 (m, 1H), 7.60 (d, *J* = 8.0 Hz, 1H), 7.42 (dd, *J* = 17.3, 7.5 Hz, 3H), 7.37 – 7.29 (m, 3H), 6.40 (dt, *J* = 15.4, 7.6 Hz, 1H), 4.58 (s, 2H), 4.04 (s, 2H), 2.91 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.53, 148.02, 136.42, 133.33, 133.09, 130.03, 129.52, 127.27, 127.11, 126.41, 125.29, 123.49, 119.77, 119.34, 111.51, 109.81, 57.35, 48.16, 38.48. HRMS (ESI) m/z calcd for C₁₉H₁₉CINO (M+H)⁺ 312.1155, found 312.1156.

(E)-N-((1H-indol-7-yl)methyl)-3-(2-chlorophenyl)-N-methylprop-2-en-1-ami ne hydrochloride (24b).

Yield: 80%. **24b** was synthesized by the general procedure given above as white solid. m.p. 185-188 °C. ¹H NMR (400 MHz, MeOD) δ 7.66 (d, J = 7.9 Hz, 2H), 7.43 (d, J = 6.4 Hz, 1H), 7.37 (d, J = 2.4 Hz, 1H), 7.32 (d, J = 7.8 Hz, 2H), 7.21 (t, J = 6.4 Hz, 2H), 7.12 (t, J = 4.8 Hz, 1H), 6.56 (d, J = 2.4 Hz, 1H), 6.45 – 6.34 (m, 1H), 4.40 (s, 2H), 3.82 (d, J = 6.9 Hz, 2H), 2.68 (s, 3H); ¹³C NMR (126 MHz, MeOD) δ 135.65, 135.47, 133.36, 132.91, 129.76, 129.36, 129.28, 127.14, 126.94, 125.33, 124.18,

122.18, 120.29, 119.04, 112.49, 102.04, 57.64, 55.95, 38.34. HRMS (EI) m/z calcd for $C_{19}H_{19}ClN_2$ (M)⁺ 310.1237, found 310.1233.

(E)-N-(benzofuran-3-ylmethyl)-3-(2-fluorophenyl)-N-methylprop-2-en-1-am ine hydrochloride (25a).

Yield: 47%. **25a** was synthesized by the general procedure given above as white solid. m.p. 140-141 °C. ¹H-NMR (400 MHz, MeOD) δ 8.12 (s, 1H), 7.85 – 7.76 (m, 1H), 7.68 – 7.54 (m, 2H), 7.49 – 7.29 (m, 3H), 7.27 – 6.99 (m, 3H), 6.56 – 6.40 (m, 1H), 4.61 (d, *J* = 45.6 Hz, 2H), 4.08 (d, *J* = 69.5 Hz, 2H), 2.91 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.56, 148.07, 136.47, 134.12, 133.08, 130.23, 129.49, 127.25, 127.15, 126.44, 125.21, 123.39, 119.69, 119.29, 111.48, 109.77, 57.32, 48.14, 38.51. HRMS (ESI) m/z calcd for C₁₉H₁₉FNO (M+H)⁺ 296.1451, found 296.1452.

(E)-N-((1H-indol-7-yl)methyl)-3-(2-fluorophenyl)-N-methylprop-2-en-1-ami ne hydrochloride (25b).

Yield: 54%. **25b** was synthesized by the general procedure given above as white solid. m.p. 167-168 °C ¹H NMR (500 MHz, MeOD) δ 7.72 (d, J = 7.8 Hz, 1H), 7.64 – 7.58 (m, 1H), 7.39 (d, J = 3.2 Hz, 1H), 7.36 (d, J = 12.4 Hz, 1H), 7.29 (d, J = 7.2 Hz, 1H), 7.20 (t, J = 7.5 Hz, 1H), 7.17 – 7.10 (m, 2H), 7.05 (s, 1H), 6.59 (d, J = 3.1 Hz, 1H), 6.50 – 6.42 (m, 1H), 4.66 (s, 2H), 4.15 – 3.96 (m, 2H), 2.83 (s, 3H); ¹³C NMR (126 MHz, MeOD) δ 135.49, 132.57, 130.34, 130.27, 129.31, 127.84, 125.39, 124.29, 124.17, 122.31, 119.39, 119.06, 115.41, 115.23, 112.08, 102.06, 57.89, 55.78, 38.12. HRMS (EI) m/z calcd for C₁₉H₁₉FN₂ (M)⁺ 294.1532, found 294.1532.

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(E)-N-(benzofuran-3-ylmethyl)-N-methyl-3-(o-tolyl)prop-2-en-1-amine hydrochloride (26a).

Yield: 32%. **26a** was synthesized by the general procedure given above as white solid. m.p. 164-165 °C. ¹H-NMR (400 MHz, CDCl₃) δ 7.70 (dd, J = 15.7, 7.5 Hz, 1H), 7.56 (d, J = 8.7 Hz, 1H), 7.47 (t, J = 7.1 Hz, 2H), 7.32 – 7.27 (m, 1H), 7.26 – 7.22 (m, 1H), 7.18 – 7.11 (m, 3H), 6.77 (d, J = 15.7 Hz, 1H), 6.20 (dt, J = 15.7, 6.6 Hz, 1H), 3.71 (s, 2H), 3.27 (d, J = 6.6 Hz, 2H), 2.34 (s, 3H), 2.32 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.50, 147.99, 138.61, 135.96, 134.24, 130.13, 128.59, 126.49, 125.98, 125.66, 125.24, 123.45, 119.37, 117.66, 111.48, 109.88, 57.71, 47.88, 38.34, 18.38. HRMS (ESI) m/z calcd for C₂₀H₂₂NO (M+H)⁺ 292.1701, found 292.1702.

(E)-N-((1H-indol-7-yl)methyl)-N-methyl-3-(o-tolyl)prop-2-en-1-amine hydrochloride (26b).

Yield: 73%. **26b** was synthesized by the general procedure given above as white solid. m.p. 167-169 °C. ¹H NMR (400 MHz, MeOD) δ 7.67 (d, J = 7.5 Hz, 1H), 7.54 – 7.48 (m, 1H), 7.36 (d, J = 3.2 Hz, 1H), 7.23 (d, J = 7.2 Hz, 1H), 7.20 – 7.15 (m, 3H), 7.10 (d, J = 8.6 Hz, 2H), 6.56 (d, J = 3.2 Hz, 1H), 6.27 – 6.16 (m, 1H), 4.48 (s, 2H), 3.87 (d, J = 6.6 Hz, 2H), 2.72 (d, J = 6.8 Hz, 3H), 2.35 (d, J = 3.4 Hz, 3H); ¹³C NMR (101 MHz, MeOD) δ 136.99, 135.71, 135.56, 134.63, 130.03, 129.13, 128.25, 125.91, 125.60, 125.48, 125.13, 123.85, 121.74, 119.65, 119.04, 113.93, 58.26, 56.33, 38.69, 18.40. HRMS (EI) m/z calcd for C₂₀H₂₂N₂ (M)⁺ 290.1783, found 290.1784.

(E)-N-(benzofuran-3-ylmethyl)-N-methyl-3-(2-nitrophenyl)prop-2-en-1-ami ne hydrochloride (27a).

Yield: 32%. **27a** was synthesized by the general procedure given above as yellow oil. ¹H-NMR (400 MHz, MeOD) δ 8.14 (s, 1H), 8.07 – 8.02 (m, 1H), 7.85 – 7.80 (m, 1H), 7.76 – 7.71 (m, 2H), 7.63 – 7.56 (m, 2H), 7.46 – 7.34 (m, 3H), 6.38 – 6.27 (m, 1H), 4.70 (d, *J* = 13.9 Hz, 1H), 4.56 (d, *J* = 13.2 Hz, 1H), 4.17 (d, *J* = 7.3 Hz, 1H), 4.04 (d, *J* = 8.0 Hz, 1H), 2.95 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.50, 148.11, 147.96, 136.34, 133.44, 130.93, 129.47, 129.06, 126.44, 125.26, 124.26, 123.48, 121.62, 119.43, 111.48, 109.77, 57.01, 48.14, 38.57. HRMS (ESI) m/z calcd for C₁₉H₁₉N₂O₃ (M+H)⁺ 323.1396, found 323.1397.

(E)-N-((1H-indol-7-yl)methyl)-N-methyl-3-(2-nitrophenyl)prop-2-en-1-amin e hydrochloride (27b).

Yield: 69%. **27b** was synthesized by the general procedure given above as white solid. m.p. 179-181 °C. ¹H NMR (500 MHz, MeOD) δ 7.99 (d, J = 8.0 Hz, 1H), 7.68 (dd, J = 14.9, 7.2 Hz, 2H), 7.58 – 7.51 (m, 2H), 7.36 (d, J = 3.1 Hz, 1H), 7.30 – 7.20 (m, 2H), 7.10 (d, J = 6.8 Hz, 1H), 6.55 (d, J = 2.4 Hz, 1H), 6.38 – 6.27 (m, 1H), 4.53 (d, J = 7.2 Hz, 2H), 3.90 (d, J = 6.7 Hz, 2H), 2.74 (s, 3H); ¹³C NMR (126 MHz, MeOD) δ 147.74, 135.42, 134.13, 133.16, 133.16, 131.06, 128.82, 128.82, 125.07, 124.02, 124.02, 121.69, 118.99, 118.99, 113.82, 101.85, 57.55, 56.43, 38.85. HRMS (EI) m/z calcd for C₁₉H₁₉N₃O₂ (M)⁺ 321.1477, found 321.1476.

(E)-N-(benzofuran-3-ylmethyl)-N-methyl-3-(2-(trifluoromethyl)phenyl)prop -2-en-1-amine hydrochloride (28a).

Yield: 43%. **28a** was synthesized by the general procedure given above as white solid. m.p. 144-146 °C. ¹H-NMR (400 MHz, MeOD) δ 8.15 (d, J = 5.1 Hz, 1H), 7.89 – 7.75 (m, 3H), 7.61 (dt, J = 15.2, 7.7 Hz, 3H), 7.40 (dt, J = 18.9, 7.3 Hz, 2H), 7.01 (d, J = 15.8 Hz, 1H), 6.62 – 6.46 (m, 1H), 4.70 (d, J = 14.0 Hz, 1H), 4.56 (d, J = 13.7 Hz, 1H), 4.17 (dd, J = 12.9, 7.1 Hz, 1H), 3.99 (dd, J = 12.8, 7.8 Hz, 1H), 2.92 (s, 3H); ¹³C NMR (126 MHz, MeOD) δ 155.44, 147.80, 136.00, 134.12, 132.24, 128.59, 127.83, 126.30, 125.45, 125.41, 125.30, 125.17, 123.36, 121.75, 119.20, 111.40, 109.84, 57.18, 48.21, 38.42. HRMS (ESI) m/z calcd for C₂₀H₁₉F₃NO (M+H)⁺ 346.1419, found 346.1418.

(E)-N-((1H-indol-7-yl)methyl)-N-methyl-3-(2-(trifluoromethyl)phenyl)prop-2-en-1-amine hydrochloride (28b).

Yield: 83%. **28b** was synthesized by the general procedure given above as white solid. m.p. 179-180 °C. ¹H NMR (400 MHz, MeOD) δ 7.66 (d, J = 7.8 Hz, 2H), 7.56 (t, J = 7.6 Hz, 1H), 7.50 (d, J = 7.7 Hz, 1H), 7.40 (t, J = 7.6 Hz, 1H), 7.25 (d, J = 3.2 Hz, 1H), 7.04 (d, J = 7.0 Hz, 1H), 7.01 – 6.96 (m, 1H), 6.93 (d, J = 6.8 Hz, 1H), 6.46 (d, J = 3.2 Hz, 1H), 6.41 – 6.31 (m, 1H), 3.91 (s, 2H), 3.32 (d, J = 4.6 Hz, 2H), 2.30 (d, J = 6.4 Hz, 3H); ¹³C NMR (101 MHz, MeOD) δ 136.02, 135.33, 131.96, 131.48, 128.66, 128.42, 127.46, 127.15, 125.22, 125.16, 124.12, 122.54, 121.61, 120.73,

119.31, 118.60, 101.13, 59.34, 58.88, 41.03. HRMS (EI) m/z calcd for $C_{20}H_{19}F_3N_2$ (M)⁺ 344.1500, found 344.1498.

(E)-N-(benzofuran-3-ylmethyl)-3-(2-methoxyphenyl)-N-methylprop-2-en-1-a mine hydrochloride (29a).

Yield: 53%. **29a** was synthesized by the general procedure given above as white solid. m.p. 151-152 °C. ¹H-NMR (400 MHz, MeOD) δ 8.12 (s, 1H), 7.81 (d, J = 7.6 Hz, 1H), 7.60 (d, J = 7.9 Hz, 1H), 7.52 (dd, J = 7.7, 1.5 Hz, 1H), 7.46 – 7.29 (m, 3H), 7.21 (d, J = 15.9 Hz, 1H), 7.02 (d, J = 8.3 Hz, 1H), 6.96 (t, J = 7.5 Hz, 1H), 6.40 (dt, J = 15.1, 7.5 Hz, 1H), 4.58 (d, J = 50.9 Hz, 2H), 4.18 – 3.90 (m, 2H), 3.87 (s, 3H), 2.90 (s, 3H); ¹³C NMR (126 MHz, MeOD) δ 157.29, 155.43, 147.80, 136.07, 130.02, 127.32, 126.29, 125.17, 123.69, 123.35, 120.29, 119.18, 116.55, 111.40, 110.76, 109.78, 58.12, 54.50, 48.12, 38.29. HRMS (ESI) m/z calcd for C₂₀H₂₂NO₂ (M+H)⁺ 308.1651, found 308.1649.

(E)-N-((1H-indol-7-yl)methyl)-3-(2-methoxyphenyl)-N-methylprop-2-en-1-a mine hydrochloride (29b).

Yield: 74%. **29b** was synthesized by the general procedure given above as white solid. m.p. 154-155 °C. ¹H NMR (400 MHz, MeOD) δ 7.67 (d, J = 5.2 Hz, 1H), 7.50 (s, 1H), 7.36 (s, 1H), 7.26 (d, J = 2.4 Hz, 2H), 7.12 (d, J = 4.8 Hz, 2H), 7.04 – 6.87 (m, 2H), 6.56 (s, 1H), 6.37 (s, 1H), 4.48 (s, 1H), 3.86 – 3.25 (m, 6H), 2.71 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 157.19, 135.57, 134.44, 129.74, 129.14, 127.16, 125.17,

124.18, 123.96, 121.84, 120.34, 119.07, 118.40, 113.66, 110.77, 101.97, 58.64, 56.12, 54.59, 38.51. HRMS (EI) m/z calcd for C₂₀H₂₂N₂O (M)⁺ 306.1732, found 306.1733.

(E)-N-(benzofuran-3-ylmethyl)-3-(3-chlorophenyl)-N-methylprop-2-en-1-am ine hydrochloride (30a).

Yield: 53%. **30a** was synthesized by the general procedure given above as white solid. m.p. 136-137 °C . ¹H NMR (400 MHz, MeOD) δ 8.14 (s, 1H), 7.83 (d, *J* = 7.0 Hz, 1H), 7.63 – 7.54 (m, 2H), 7.43 (t, *J* = 8.1 Hz, 2H), 7.40 – 7.32 (m, 3H), 6.91 (d, *J* = 15.8 Hz, 1H), 6.52 – 6.36 (m, 1H), 4.61 (d, *J* = 36.8 Hz, 2H), 4.04 (d, *J* = 55.0 Hz, 2H), 2.90 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.51, 147.98, 139.02, 137.38, 134.43, 129.99, 128.53, 128.04, 126.46, 125.25, 125.23, 123.45, 119.39, 118.29, 111.48, 109.90, 57.32, 48.11, 38.50. HRMS (ESI) m/z calcd for C₁₉H₁₉CINO (M+H)⁺ 312.1155, found 312.1154.

(E)-N-((1H-indol-7-yl)methyl)-3-(3-chlorophenyl)-N-methylprop-2-en-1-ami ne hydrochloride (30b).

Yield: 64%. **30b** was synthesized by the general procedure given above as white solid. m.p. 186-190 °C. ¹H NMR (400 MHz, MeOD) δ 7.58 (d, J = 7.8 Hz, 1H), 7.46 (s, 1H), 7.36 – 7.24 (m, 5H), 7.13 (d, J = 6.9 Hz, 1H), 7.08 – 7.02 (m, 1H), 6.67 (d, J = 12.8 Hz, 1H), 6.51 (d, J = 3.2 Hz, 1H), 6.40 (d, J = 7.2 Hz, 1H), 4.22 (d, J = 12.0 Hz, 2H), 3.58 (d, J = 7.5 Hz, 2H), 2.51 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 138.89, 137.41, 135.58, 134.42, 129.96, 129.43, 128.50, 126.42, 125.50, 125.23,

124.36, 122.43, 119.18, 118.24, 112.20, 102.20, 57.59, 55.83, 38.31. HRMS (EI) m/z calcd for $C_{19}H_{19}CIN_2$ (M)⁺ 310.1237, found 310.1234.

(E)-N-(benzofuran-3-ylmethyl)-3-(3-fluorophenyl)-N-methylprop-2-en-1-am ine hydrochloride (31a).

Yield: 43%. **31a** was synthesized by the general procedure given above as white solid. m.p. 161-162 °C . ¹H NMR (400 MHz, MeOD) δ 8.11 (s, 1H), 7.81 (d, J = 7.5 Hz, 1H), 7.64 – 7.58 (m, 2H), 7.45 – 7.34 (m, 3H), 7.24 – 7.11 (m, 2H), 7.08 (d, J = 15.9 Hz, 1H), 6.56 – 6.41 (m, 1H), 4.58 (s, 2H), 4.02 (s, 2H), 2.90 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.45, 147.81, 138.59, 136.98, 134.41, 129.45, 128.49, 128.64, 126.45, 125.15, 125.35, 123.58, 119.28, 118.36, 111.31, 109.79, 57.82, 47.89, 38.51 . HRMS (ESI) m/z calcd for C₁₉H₁₉FNO (M+H)⁺ 296.1451, found 296.1452.

(E)-N-((1H-indol-7-yl)methyl)-3-(3-fluorophenyl)-N-methylprop-2-en-1-ami ne hydrochloride (31b).

Yield: 52%. **31b** was synthesized by the general procedure given above as white solid. m.p. 186-187 °C. ¹H NMR (500 MHz, MeOD) δ 7.72 (d, J = 7.9 Hz, 1H), 7.43 – 7.35 (m, 2H), 7.34 – 7.26 (m, 3H), 7.16 (t, J = 7.6 Hz, 1H), 7.11 – 7.05 (m, 1H), 6.90 (d, J = 6.8 Hz, 1H), 6.59 (d, J = 3.1 Hz, 1H), 6.46 – 6.37 (m, 1H), 4.67 (s, 2H), 4.15 – 3.94 (m, 2H), 2.82 (d, J = 6.4 Hz, 3H); ¹³C NMR (126 MHz, MeOD) δ 139.05, 137.69, 135.49, 130.14, 129.32, 125.39, 124.32, 122.83, 122.80, 122.32, 119.07, 118.00, 115.25, 112.88, 112.05, 102.06, 57.45, 55.71, 38.14. HRMS (EI) m/z calcd for C₁₉H₁₉FN₂ (M)⁺ 294.1532, found 294.1533. (E)-N-(benzofuran-3-ylmethyl)-N-methyl-3-(m-tolyl)prop-2-en-1-amine hydrochloride (32a).

Yield: 47%. **13c** was synthesized by the general procedure given above as yellow oil .¹H-NMR (400 MHz, MeOD) δ 8.12 (s, 1H), 7.81 (d, *J* = 7.2 Hz, 1H), 7.60 (d, *J* = 8.1 Hz, 1H), 7.47 – 7.33 (m, 4H), 7.19 (d, *J* = 7.9 Hz, 2H), 6.90 (d, *J* = 15.8 Hz, 1H), 6.32 (dt, *J* = 15.5, 7.6 Hz, 1H), 4.67 (d, *J* = 13.9 Hz, 1H), 4.51 (d, *J* = 13.9 Hz, 1H), 4.11 (dd, *J* = 13.0, 7.0 Hz, 1H), 3.93 (dd, *J* = 12.9, 7.9 Hz, 1H), 2.89 (s, 3H), 2.34 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.53, 147.95, 139.12, 137.35, 134.23, 129.87, 128.54, 128.29, 126.39, 125.27, 125.14, 123.25, 119.45, 118.15, 111.36, 109.84, 57.45, 48.19, 38.75, 18.29. HRMS (ESI) m/z calcd for C₂₀H₂₂NO (M+H)⁺ 292.1701, found 292.1702.

(E)-N-((1H-indol-7-yl)methyl)-N-methyl-3-(m-tolyl)prop-2-en-1-amine hydrochloride (32b).

Yield: 78%. **32b** was synthesized by the general procedure given above as white solid. m.p. 196-197 °C. ¹H NMR (400 MHz, MeOD) δ 7.61 (d, J = 7.1 Hz, 1H), 7.33 (d, J = 3.2 Hz, 1H), 7.29 – 7.14 (m, 4H), 7.13 – 7.04 (m, 2H), 6.71 (d, J = 6.8 Hz, 1H), 6.53 (d, J = 3.2 Hz, 1H), 6.39 – 6.28 (m, 1H), 4.29 (s, 2H), 3.67 (d, J = 6.4 Hz, 2H), 2.57 (s, 3H), 2.33 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 138.02, 137.31, 136.03, 135.49, 130.95, 128.86, 128.70, 128.20, 126.98, 124.77, 123.56, 123.11, 121.11, 120.78, 118.90, 116.21, 58.61, 57.07, 39.43, 19.99. HRMS (EI) m/z calcd for C₂₀H₂₂N₂ (M)⁺ 290.1783, found 290.1784.

(E)-N-(benzofuran-3-ylmethyl)-N-methyl-3-(3-nitrophenyl)prop-2-en-1-ami ne hydrochloride (33a).

Yield: 35%. **33a** was synthesized by the general procedure given above as yellow oil. ¹H-NMR (400 MHz, MeOD) δ 8.37 (t, J = 1.8 Hz, 1H), 8.19 (dd, J = 8.2, 2.1 Hz, 1H), 8.11 (s, 1H), 7.91 (d, J = 7.8 Hz, 1H), 7.85 – 7.80 (m, 1H), 7.63 (t, J = 8.0 Hz, 1H), 7.58 (d, J = 7.5 Hz, 1H), 7.44 – 7.33 (m, 2H), 7.01 (d, J = 15.7 Hz, 1H), 6.64 – 6.53 (m, 1H), 4.55 (s, 2H), 4.04 (t, J = 14.6 Hz, 2H), 2.88 (s, 3H); ¹³C NMR (126 MHz, MeOD) δ 155.42, 148.61, 147.52, 137.35, 137.20, 132.46, 129.64, 126.45, 125.07, 123.26, 122.81, 121.01, 120.67, 119.30, 111.33, 110.39, 57.22, 48.27, 38.77. HRMS (ESI) m/z calcd for C₁₉H₁₉N₂O₃ (M+H)⁺ 323.1396, found 323.1399.

(E)-N-((1H-indol-7-yl)methyl)-N-methyl-3-(3-nitrophenyl)prop-2-en-1-amin e hydrochloride hydrochloride (33b).

Yield: 86%. **33b** was synthesized by the general procedure given above as white solid. m.p. 186-188 °C. ¹H NMR (400 MHz, MeOD) δ 8.23 (t, J = 1.9 Hz, 1H), 8.08 (d, J = 6.9 Hz, 1H), 7.78 (d, J = 7.8 Hz, 1H), 7.58 – 7.46 (m, 2H), 7.26 (d, J = 3.2 Hz, 1H), 7.04 (d, J = 6.6 Hz, 1H), 6.98 (d, J = 3.6 Hz, 1H), 6.67 (d, J = 6.4 Hz, 1H), 6.54 (d, J = 6.6 Hz, 1H), 6.47 – 6.43 (m, 1H), 3.91 (s, 2H), 3.32 (d, J = 6.4 Hz, 1H), 2.31 (d, J = 8.0 Hz, 4H); ¹³C NMR (126 MHz, MeOD) δ 148.50, 138.66, 135.23, 134.46, 131.84, 131.15, 129.28, 129.10, 128.34, 124.10, 121.75, 121.53, 120.31, 119.89, 119.49, 118.56, 58.85, 58.56, 40.86. HRMS (EI) m/z calcd for C₁₉H₁₉N₃O₂ (M)⁺ 321.1477, found 321.1476.

(E)-N-(benzofuran-3-ylmethyl)-N-methyl-3-(3-(trifluoromethyl)phenyl)prop -2-en-1-amine hydrochloride (34a).

Yield: 58%. **34a** was synthesized by the general procedure given above as white solid. m.p. 121-123 °C. ¹H-NMR (400 MHz, MeOD) δ 8.15 (d, J = 5.1 Hz, 1H), 7.88 – 7.73 (m, 3H), 7.61 (dt, J = 15.2, 7.7 Hz, 3H), 7.40 (dt, J = 18.9, 7.3 Hz, 2H), 7.01 (d, J = 15.8 Hz, 1H), 6.62 – 6.45 (m, 1H), 4.70 (d, J = 14.0 Hz, 1H), 4.56 (d, J = 13.7 Hz, 1H), 4.23 – 4.13 (m, 1H), 3.99 (dd, J = 12.8, 7.8 Hz, 1H), 2.92 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.48, 148.06, 138.84, 136.41, 130.64, 130.24, 129.35, 126.48, 125.47, 125.23, 125.03, 123.44, 123.34, 119.46, 118.89, 111.46, 109.88, 57.28, 48.14, 38.52. HRMS (ESI) m/z calcd for C₂₀H₁₉F₃NO (M+H)⁺ 346.1419, found 346.1418.

(E)-N-((1H-indol-7-yl)methyl)-N-methyl-3-(3-(trifluoromethyl)phenyl)prop-2-en-1-amine hydrochloride (34b).

Yield: 82%. **34b** was synthesized by the general procedure given above as white solid. m.p. 129-131 °C. ¹H NMR (400 MHz, MeOD) δ 7.65 (s, 2H), 7.49 (dd, J = 7.5, 3.8 Hz, 3H), 7.25 (d, J = 3.1 Hz, 1H), 7.03 (d, J = 7.2 Hz, 1H), 6.97 (t, J = 7.4 Hz, 1H), 6.63 (d, J = 15.8 Hz, 1H), 6.53 – 6.42 (m, 2H), 3.88 (s, 2H), 3.28 (s, 2H), 2.31–2.10 (m, 3H); ¹³C NMR (101 MHz, MeOD) δ 138.12, 135.33, 131.38, 129.40, 128.98, 128.43, 125.15, 124.13, 123.51, 123.47, 122.53, 122.49, 121.61, 120.78, 119.28, 118.58, 101.12, 59.26, 58.88, 41.11. HRMS (EI) m/z calcd for C₂₀H₁₉F₃N₂ (M)⁺ 344.1500, found 344.1503.

(E)-N-(benzofuran-3-ylmethyl)-3-(3-methoxyphenyl)-N-methylprop-2-en-1-a mine hydrochloride (35a).

Yield: 23%. **35a** was synthesized by the general procedure given above as white solid. m.p. 153-154 °C. ¹H-NMR (400 MHz, MeOD) δ 8.13 (s, 1H), 7.82 (d, *J* = 7.3 Hz, 1H), 7.60 (d, *J* = 8.2 Hz, 1H), 7.49 – 7.34 (m, 2H), 7.29 (t, *J* = 7.9 Hz, 1H), 7.08 (dd, *J* = 10.7, 4.8 Hz, 2H), 6.95 – 6.86 (m, 2H), 6.47 – 6.29 (m, 1H), 4.68 (d, *J* = 14.0 Hz, 1H), 4.52 (d, *J* = 13.7 Hz, 1H), 4.13 (dd, *J* = 12.9, 7.2 Hz, 1H), 4.01 – 3.89 (m, 1H), 3.81 (s, 3H), 2.90 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 160.13, 155.51, 147.99, 140.75, 136.62, 129.48, 128.45, 126.43, 125.26, 123.45, 119.31, 116.50, 114.31, 112.05, 111.49, 109.86, 57.55, 54.36, 48.05, 38.38. HRMS (ESI) m/z calcd for C₂₀H₂₂NO₂ (M+H)⁺ 308.1651, found 308.1652.

(E)-N-((1H-indol-7-yl)methyl)-3-(3-methoxyphenyl)-N-methylprop-2-en-1-a mine hydrochloride (35b).

Yield: 75%. **35b** was synthesized by the general procedure given above as white solid. m.p. 154-155 °C. ¹H NMR (400 MHz, MeOD) δ 7.71 (d, *J* = 7.9 Hz, 1H), 7.39 (d, *J* = 3.2 Hz, 1H), 7.27 (t, *J* = 7.7 Hz, 2H), 7.14 (t, *J* = 7.6 Hz, 1H), 7.07 (d, *J* = 7.7 Hz, 1H), 7.04 (s, 1H), 6.88 (s, 2H), 6.58 (d, *J* = 3.2 Hz, 1H), 6.40 – 6.29 (m, 1H), 4.64 (s, 2H), 4.01 (d, *J* = 7.2 Hz, 2H), 2.81 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 160.09, 140.35, 136.72, 135.60, 129.43, 129.37, 125.47, 124.38, 122.32, 119.26, 119.16, 116.83, 114.22, 112.39, 112.00, 102.12, 57.80, 55.77, 54.36, 38.17. HRMS (EI) m/z calcd for C₂₀H₂₂N₂O (M)⁺ 306.1732, found 306.1733.

(E)-N-(benzofuran-3-ylmethyl)-3-(2,4-dichlorophenyl)-N-methylprop-2-en-1 -amine hydrochloride (36a).

Yield: 46%. **36a** was synthesized by the general procedure given above as white solid. m.p. 146-148 °C. ¹H NMR (400 MHz, MeOD) δ 8.13 (s, 1H), 7.82 (d, *J* = 7.5 Hz, 1H), 7.71 (d, *J* = 8.5 Hz, 1H), 7.60 (d, *J* = 7.7 Hz, 1H), 7.53 (d, *J* = 2.0 Hz, 1H), 7.40 (ddt, *J* = 9.9, 8.5, 4.1 Hz, 3H), 7.27 (d, *J* = 15.7 Hz, 1H), 6.43 (dt, *J* = 15.6, 7.7 Hz, 1H), 4.69 (d, *J* = 13.9 Hz, 1H), 4.55 (d, *J* = 13.8 Hz, 1H), 4.19 (dd, *J* = 13.3, 6.8 Hz, 1H), 4.02 (dd, *J* = 13.5, 7.8 Hz, 1H), 2.92 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.51, 148.05, 135.04, 134.90, 133.74, 132.20, 129.12, 128.39, 127.44, 126.42, 125.27, 123.48, 120.65, 119.37, 111.49, 109.79, 57.20, 48.15, 38.53. HRMS (ESI) m/z calcd for C₁₉H₁₈Cl₂NO (M+H)⁺ 346.0765, found 346.0763.

(E)-N-((1H-indol-7-yl)methyl)-3-(2,4-dichlorophenyl)-N-methylprop-2-en-1amine hydrochloride (36b).

Yield: 68%. **36b** was synthesized by the general procedure given above as white solid. m.p. 176-177 °C. ¹H NMR (400 MHz, MeOD) δ 7.61 (d, J = 8.5 Hz, 1H), 7.56 (d, J = 7.8 Hz, 1H), 7.47 (s, 1H), 7.35 – 7.29 (m, 2H), 7.12 (d, J = 7.0 Hz, 1H), 7.04 (t, J = 3.6 Hz, 1H), 6.98 (s, 1H), 6.50 (d, J = 3.2 Hz, 1H), 6.46 – 6.35 (m, 1H), 4.09 (s, 2H), 3.50 (s, 2H), 2.46 (s, 3H); ¹³C NMR (126 MHz, MeOD) δ 135.38, 134.20, 133.35, 133.12, 132.51, 128.88, 128.88, 128.09, 128.09, 127.18, 127.18, 125.22, 124.35, 123.41, 121.32, 118.88, 57.96, 56.76, 39.18. HRMS (EI) m/z calcd for C₁₉H₁₈Cl₂N₂ (M)⁺ 344.0847, found 344.0838.

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(E)-N-(benzofuran-3-ylmethyl)-3-(2-fluoro-4-(trifluoromethyl)phenyl)-N-me thylprop-2-en-1-amine hydrochloride (37a).

Yield: 35%. **37a** was synthesized by the general procedure given above as yellow oil. ¹H-NMR (400 MHz, MeOD) δ 8.15 (d, J = 5.1 Hz, 1H), 7.89 – 7.75 (m, 3H), 7.61 (dt, J = 15.2, 7.7 Hz, 3H), 7.40 (dt, J = 18.9, 7.3 Hz, 2H), 7.01 (d, J = 15.8 Hz, 1H), 6.61 – 6.46 (m, 1H), 4.70 (d, J = 14.0 Hz, 1H), 4.56 (d, J = 13.7 Hz, 1H), 4.23 – 4.12 (m, 1H), 3.99 (dd, J = 12.8, 7.8 Hz, 1H), 2.92 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.54, 148.05, 135.04, 133.74, 132.20, 129.89, 129.12, 128.35, 127.44, 126.42, 125.27, 124.58, 123.48, 120.65, 119.39, 111.49, 109.79, 57.20, 48.19, 38.53. HRMS (ESI) m/z calcd for C₂₀H₁₈F₄NO (M+H)⁺ 364.1325, found 364.1326.

(E)-N-((1H-indol-7-yl)methyl)-3-(2-fluoro-4-(trifluoromethyl)phenyl)-N-met hylprop-2-en-1-amine hydrochloride (37b).

Yield: 69%. **37b** was synthesized by the general procedure given above as white solid. m.p. 154-155 °C. ¹H NMR (400 MHz, MeOD) δ 7.76 (t, J = 8.0 Hz, 1H), 7.61 (d, J = 7.7 Hz, 1H), 7.47 (s, 2H), 7.33 (d, J = 3.2 Hz, 1H), 7.16 (d, J = 7.2 Hz, 1H), 7.07 (t, J = 7.6 Hz, 1H), 6.92 (d, J = 6.4 Hz, 1H), 6.64 – 6.54 (m, 1H), 6.52 (d, J = 3.2Hz, 1H), 4.29 (s, 2H), 3.71 (s, 2H), 2.56 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 160.91, 158.42, 135.44, 128.82, 128.58, 128.54, 127.00, 124.71, 124.36, 122.96, 120.94, 120.81, 118.87, 116.57, 112.87, 112.61, 101.63, 58.56, 57.52, 39.86. HRMS (EI) m/z calcd for C₂₀H₁₈F₄N₂ (M)⁺ 362.1406, found 362.1408.

(E)-N-(benzofuran-3-ylmethyl)-3-(3-fluoro-4-(trifluoromethyl)phenyl)-N-me thylprop-2-en-1-amine hydrochloride (38a).

Yield: 42%. **38a** was synthesized by the general procedure given above as white solid. m.p. 166-168 °C. ¹H-NMR (400 MHz, MeOD) δ 8.07 (s, 1H), 7.80 (d, J = 7.1 Hz, 1H), 7.69 (t, J = 7.6 Hz, 1H), 7.58 (d, J = 8.0 Hz, 1H), 7.54 – 7.34 (m, 4H), 6.92 (d, J = 16.4 Hz, 1H), 6.63 – 6.50 (m, 1H), 4.63 (s, 2H), 4.49 (s, 1H), 3.96 (s, 1H), 3.48 (s, 1H), 2.83 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.52, 148.08, 137.63, 128.58, 127.30, 126.42, 125.28, 124.71, 123.48, 123.06, 123.02, 121.21, 119.32, 114.62, 114.41, 111.50, 109.78, 56.97, 47.52, 38.62. HRMS (ESI) m/z calcd for C₂₀H₁₈F₄NO (M+H)⁺ 364.1325, found 364.1326.

(E)-N-((1H-indol-7-yl)methyl)-3-(3-fluoro-4-(trifluoromethyl)phenyl)-N-met hylprop-2-en-1-amine hydrochloride (38b).

Yield: 35%. **38b** was synthesized by the general procedure given above as white solid. m.p. 119-120 °C. ¹H NMR (400 MHz, MeOD) δ 7.69 – 7.56 (m, 2H), 7.41 (d, *J* = 12.4 Hz, 2H), 7.32 (d, *J* = 3.2 Hz, 1H), 7.15 (d, *J* = 7.2 Hz, 1H), 7.06 (t, *J* = 7.5 Hz, 1H), 6.77 (d, *J* = 6.8 Hz, 1H), 6.55 (s, 1H), 6.51 (t, *J* = 7.4 Hz, 1H), 4.26 (s, 2H), 3.67 (d, *J* = 6.7 Hz, 2H), 2.55 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 160.71, 158.40, 135.37, 128.77, 128.51, 128.42, 127.01, 124.69, 124.28, 122.92, 120.91, 120.79, 118.83, 116.53, 112.82, 112.59, 101.61, 58.53, 57.49, 39.81. HRMS (EI) m/z calcd for C₂₀H₁₈F₄N₂ (M)⁺ 362.1406, found 362.1405.

(E)-N-(benzofuran-3-ylmethyl)-3-(2-fluoro-4-methoxyphenyl)-N-methylprop -2-en-1-amine hydrochloride (39a).

Yield: 47%. **39a** was synthesized by the general procedure given above as white solid. m.p. 150-151 °C. ¹H-NMR (400 MHz, MeOD) δ 8.14 (s, 1H), 7.83 (d, *J* = 7.3 Hz, 1H), 7.60 (d, *J* = 8.1 Hz, 1H), 7.39 (tt, *J* = 17.3, 8.7 Hz, 2H), 7.32 – 7.21 (m, 1H), 7.08 (dd, *J* = 9.8, 5.0 Hz, 2H), 6.93 (s, 1H), 6.47 – 6.31 (m, 1H), 4.60 (d, *J* = 43.9 Hz, 2H), 3.99 (dd, *J* = 41.1, 32.3 Hz, 2H), 3.82 (d, *J* = 6.1 Hz, 3H), 2.91 (d, *J* = 9.1 Hz, 3H); ¹³C NMR (126 MHz, MeOD) δ 160.03, 155.41, 147.87, 140.65, 136.52, 129.38, 126.33, 125.15, 123.35, 119.26, 119.15, 116.40, 114.21, 111.95, 111.38, 109.76, 57.45, 54.26, 47.91, 38.29. HRMS (ESI) m/z calcd for C₂₀H₂₁FNO₂ (M+H)⁺ 326.1556, found 326.1554.

(E)-N-((1H-indol-7-yl)methyl)-3-(2-fluoro-4-methoxyphenyl)-N-methylprop-2-en-1-amine hydrochloride (39b).

Yield: 45%. **39b** was synthesized by the general procedure given above as white solid. m.p. 108-111 °C. ¹H NMR (400 MHz, MeOD) δ 7.71 (d, *J* = 7.8 Hz, 1H), 7.52 (t, *J* = 8.7 Hz, 1H), 7.39–7.31 (m, 1H), 7.27 (d, *J* = 7.2 Hz, 1H), 7.14 (t, *J* = 7.6 Hz, 1H), 6.95 (d, *J* = 15.9 Hz, 1H), 6.75–6.60 (m, 2H), 6.58 (d, *J* = 3.2 Hz, 1H), 6.35–6.25 (m, 1H), 4.64 (s, 2H), 4.05 (d, *J* = 6.7 Hz, 2H), 3.81 (s, 3H), 2.80 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 135.60, 132.71, 129.40, 128.63, 125.48, 124.37, 122.38, 119.16, 116.48, 115.40, 115.28, 112.21, 110.48, 102.15, 101.20, 100.94, 58.23, 55.70, 54.85, 38.05. HRMS (EI) m/z calcd for C₂₀H₂₁FN₂O (M)⁺ 324.1638, found 324.1635.

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(E)-N-(benzofuran-3-ylmethyl)-3-(3-fluoro-4-methoxyphenyl)-N-methylprop -2-en-1-amine hydrochloride (40a).

Yield: 51%. **40a** was synthesized by the general procedure given above as yellow oil. ¹H-NMR (400 MHz, MeOD) δ 8.07 (s, 1H), 7.80 (d, J = 7.1 Hz, 1H), 7.69 (t, J = 7.6 Hz, 1H), 7.58 (d, J = 8.0 Hz, 1H), 7.54 – 7.34 (m, 4H), 6.92 (d, J = 16.4 Hz, 1H), 6.63 – 6.50 (m, 1H), 4.63 (s, 2H), 4.49 (s, 1H), 3.96 (s, 1H), 3.48 (s, 1H), 2.83 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 155.51, 148.15, 148.09, 146.41, 136.73, 130.94, 129.29, 128.47, 126.96, 124.83, 123.32, 123.01, 119.50, 113.21, 113.03, 111.25, 58.13, 55.29, 48.53, 39.36. HRMS (ESI) m/z calcd for C₂₀H₁₈F₄NO (M+H)⁺ 364.1325, found 364.1326.

(E)-N-((1H-indol-7-yl)methyl)-3-(3-fluoro-4-methoxyphenyl)-N-methylprop-2-en-1-amine hydrochloride (40b).

Yield: 64%. **40b** was synthesized by the general procedure given above as white solid. m.p. 117-118 °C. ¹H NMR (400 MHz, MeOD) δ 7.49 (d, J = 7.7 Hz, 1H), 7.26 (d, J = 3.2 Hz, 1H), 7.19 (d, J = 8.8 Hz, 1H), 7.11 (d, J = 8.6 Hz, 1H), 7.00 (t, J = 6.6Hz, 2H), 6.47 (s, 3H), 6.20– 6.04 (m, 2H), 3.90 (s, 2H), 3.84 (s, 3H), 3.28 (d, J = 6.9Hz, 2H), 2.30 (s, 3H); ¹³C NMR (101 MHz, MeOD) δ 147.42, 135.39, 133.16, 130.24, 128.52, 124.31, 123.55, 122.76, 122.72, 122.17, 119.84, 119.16, 118.71, 113.14, 112.95, 112.77, 59.03, 58.09, 55.27, 40.42. HRMS (EI) m/z calcd for C₂₀H₂₁FN₂O (M)⁺ 324.1638, found 324.1637.

3-([1,1'-biphenyl]-4-yl)-N-(benzofuran-3-ylmethyl)-N-methylprop-2-yn-1-a mine hydrochloride (41a).

Yield: 51%. **41a** was synthesized by the general procedure given above as yellow oil. ¹H NMR (400 MHz, MeOD) δ 7.70 – 7.56 (m, 7H), 7.46 (dd, J = 10.3, 4.8 Hz, 2H), 7.37 (dd, J = 8.3, 6.4 Hz, 1H), 7.32 (dd, J = 7.6, 1.6 Hz, 1H), 7.27 – 7.18 (m, 1H), 6.97 – 6.82 (m, 2H), 5.64 (t, J = 9.1 Hz, 2H), 4.41 (s, 2H), 2.92 (s, 3H). ¹³C NMR (101 MHz, MeOD) δ 155.37, 145.47, 140.35, 138.70, 135.72, 134.56, 130.25, 130.25, 129.31, 129.31, 127.51, 127.51, 126.24, 127.17, 126.34, 126.34, 126.08, 126.08, 124.30, 124.27, 85.24, 82.59, 65.58, 59.64, 39.89. HRMS (EI) m/z calcd for C₂₅H₂₁NO (M)⁺ 351.1623, found 351.1624.

N-((1H-indol-7-yl)methyl)-3-([1,1'-biphenyl]-4-yl)-N-methylprop-2-yn-1-ami ne hydrochloride (41b).

Yield: 29%. **41b** was synthesized by the general procedure given above as white solid. m.p. 163-167 °C. ¹H NMR (400 MHz, MeOD) δ 7.71 – 7.59 (m, 7H), 7.46 (t, *J* = 7.6 Hz, 2H), 7.40 – 7.34 (m, 2H), 7.27 (d, *J* = 7.1 Hz, 1H), 7.13 (t, *J* = 7.6 Hz, 1H), 6.57 (d, *J* = 3.2 Hz, 1H), 4.59 (s, 2H), 4.20 (s, 2H), 2.92 (s, 3H). ¹³C NMR (101 MHz, MeOD) δ 140.69, 140.29, 137.28, 136.44, 130.87, 130.32, 130.32, 130.27, 130.27, 129.76, 128.89, 127.75, 127.75, 127.80, 127.80, 127.32, 126.58, 123.21, 120.27, 114.15, 85.49, 82.56, 56.72, 39.68, 18.80. HRMS (EI) m/z calcd for C₂₅H₂₂N₂ (M)⁺ 350.1783, found 350.1782.
3-([1,1'-biphenyl]-4-yl)-N-(benzofuran-3-ylmethyl)-N-methylpropan-1-amin e hydrochloride (42a).

Yield: 35%. **42a** was synthesized by the general procedure given above as yellow oil. ¹H NMR (400 MHz, MeOD) δ 7.62 – 7.53 (m, 4H), 7.43 (t, *J* = 7.6 Hz, 2H), 7.23 (dddd, *J* = 13.9, 11.2, 9.5, 4.5 Hz, 5H), 6.88 – 6.81 (m, 2H), 4.62 (s, 1H), 4.27 (s, 1H), 3.17 – 3.02 (m, 2H), 2.78 (s, 3H), 2.69 (dd, *J* = 9.4, 5.6 Hz, 2H), 2.12 – 1.94 (m, 2H). ¹³C NMR (101 MHz, MeOD) δ 155.28, 145.41, 140.31, 137.70, 135.59, 134.21, 128.56, 128.56, 127.51, 127.51, 126.24, 127.17, 126.34, 126.34, 126.08, 126.08, 124.30, 124.27, 112.03, 110.25, 65.58, 59.64, 39.89, 31.58, 28.56. HRMS (EI) m/z calcd for C₂₅H₂₅NO (M)⁺ 355.1936, found 355.1936.

N-((1H-indol-7-yl)methyl)-3-([1,1'-biphenyl]-4-yl)-N-methylpropan-1-amine hydrochloride (42b).

Yield: 42%. 42b was synthesized by the general procedure given above as white solid. m.p. 154-155 °C. ¹H NMR (400 MHz, MeOD) δ 7.64 – 7.61 (m, 1H), 7.59 – 7.56 (m, 2H), 7.52 – 7.48 (m, 2H), 7.42 (d, *J* = 6.8 Hz, 2H), 7.32 (d, *J* = 6.9 Hz, 2H), 7.21 (d, *J* = 8.3 Hz, 2H), 7.13 (d, *J* = 7.3 Hz, 1H), 7.05 (dd, *J* = 8.7, 6.4 Hz, 1H), 6.53 (d, *J* = 3.2 Hz, 1H), 4.38 (s, 2H), 3.05 – 2.99 (m, 2H), 2.70 (d, *J* = 7.5 Hz, 2H), 2.68 (s, 3H), 2.09 – 2.03 (m, 2H). ¹³C NMR (101 MHz, MeOD) δ 140.81, 140.39, 137.18, 136.24, 130.87, 130.19, 130.19, 129.76, 129.28, 129.28, 128.89, 128.05, 128.05, 127.32, 126.58, 124.39, 123.21, 120.27, 114.15, 102.41, 56.72, 42.15, 39.68, 31.89, 27.58. HRMS (EI) m/z calcd for C₂₅H₂₂N₂ (M)⁺ 354.2096, found 354.2097.

(E)-3-([1,1'-biphenyl]-4-yl)-N-(benzofuran-3-ylmethyl)-N,2-dimethylprop-2en-1-amine hydrochloride (43a).

Yield: 41%. **43a** was synthesized by the general procedure given above as yellow oil. ¹H NMR (400 MHz, MeOD) δ 7.72 – 7.57 (m, 5H), 7.44 (dd, J = 7.9, 6.3 Hz, 5H), 7.37 – 7.31 (m, 2H), 7.29 – 7.22 (m, 1H), 6.92 (ddd, J = 12.5, 9.5, 4.6 Hz, 1H), 6.82 (d, J = 24.1 Hz, 1H), 2.89 (s, 3H), 2.04 (dd, J = 39.4, 1.2 Hz, 3H). ¹³C NMR (101 MHz, MeOD) δ 154.35, 140.47, 140.30, 138.70, 135.78, 134.90, 130.26, 130.20, 129.31, 129.31, 128.56, 128.56, 127.24, 127.17, 126.54, 126.54, 126.49, 126.49, 124.30, 120.22, 115.59, 65.25, 59.54, 39.76, 15.74. HRMS (EI) m/z calcd for C₂₆H₂₅NO (M)⁺ 367.1936, found 367.1938.

(E)-N-((1H-indol-7-yl)methyl)-3-([1,1'-biphenyl]-4-yl)-N,2-dimethylprop-2-e n-1-amine hydrochloride (43b).

Yield: 38%. **43b** was synthesized by the general procedure given above as white solid. m.p. 164-165 °C. ¹H NMR (400 MHz, MeOD) δ 7.73 (d, J = 7.9 Hz, 1H), 7.63 (t, J = 7.2 Hz, 4H), 7.47 – 7.38 (m, 5H), 7.37 – 7.30 (m, 2H), 7.17 (t, J = 7.6 Hz, 1H), 6.82 (s, 1H), 6.57 (t, J = 8.0 Hz, 1H), 4.72 (s, 2H), 3.99– 2.90 (m, 2H), 2.87 (s, 3H), 2.02 (d, J = 6.2 Hz, 3H). ¹³C NMR (101 MHz, MeOD) δ 140.79, 140.31, 137.24, 136.42, 135.55, 130.85, 130.32, 130.32, 130.27, 130.27, 129.76, 128.89, 127.86, 127.86, 127.80, 127.80, 126.58, 123.21, 120.25, 114.20, 103.07, 65.89, 56.72, 39.68, 18.80. HRMS (EI) m/z calcd for C₂₅H₂₂N₂ (M)⁺ 366.2096, found 366.2097.

(E)-4-([1,1'-biphenyl]-4-yl)-N-(benzofuran-3-ylmethyl)-N-methylbut-3-en-1amine hydrochloride (44a).

Yield: 32%. **44a** was synthesized by the general procedure given above as yellow oil. ¹H NMR (400 MHz, MeOD) δ 7.72 (d, *J* = 7.9 Hz, 1H), 7.64 – 7.55 (m, 4H), 7.49 – 7.37 (m, 5H), 7.35 – 7.24 (m, 2H), 7.15 (t, *J* = 7.6 Hz, 1H), 6.58 – 6.35 (m, 2H), 6.28 – 6.15 (m, 1H), 4.65 (s, 2H), 3.37 (t, *J* = 6.4 Hz, 2H), 2.92 – 2.82 (m, 3H), 2.72 – 2.62 (m, 2H). ¹³C NMR (101 MHz, MeOD) δ 140.16, 135.59, 135.70, 133.16, 129.29, 129.29, 128.50, 127.98, 127.98, 127.83, 127.83, 126.94, 126.94, 126.44, 126.36, 125.50, 124.43, 123.38, 122.41, 119.18, 112.14, 102.19, 57.89, 53.59, 41.28, 31.28. HRMS (ESI) m/z calcd for C₂₆H₂₅NO (M+H)⁺ 367.1936, found 367.1937.

(E)-N-((1H-indol-7-yl)methyl)-4-([1,1'-biphenyl]-4-yl)-N-methylbut-3-en-1-a mine hydrochloride (44b).

Yield: 64%. **44b** was synthesized by the general procedure given above as white solid. m.p. 169-172 °C. ¹H NMR (400 MHz, MeOD) δ 7.72 (d, J = 7.9 Hz, 1H), 7.64 – 7.55 (m, 4H), 7.49 – 7.37 (m, 5H), 7.35 – 7.24 (m, 2H), 7.15 (t, J = 7.6 Hz, 1H), 6.58 – 6.35 (m, 2H), 6.28 – 6.15 (m, 1H), 4.65 (s, 2H), 3.37 (t, J = 6.4 Hz, 2H), 2.92 – 2.82 (m, 3H), 2.72 – 2.62 (m, 2H). ¹³C NMR (101 MHz, MeOD) δ 140.44, 140.36, 135.77, 135.70, 133.16, 129.40, 128.50, 128.50, 127.03, 126.74, 126.74, 126.44, 126.44, 126.36, 126.36, 125.50, 124.43, 123.38, 122.41, 119.18, 112.14, 102.19, 56.46, 55.22, 38.87, 27.76. HRMS (EI) m/z calcd for C₂₆H₂₆N₂ (M)⁺ 366.2096, found 366.2090.

(2E,4E)-5-([1,1'-biphenyl]-4-yl)-N-(benzofuran-3-ylmethyl)-N-methylpenta-2,4-dien-1-amine hydrochloride (45a).

Yield: 52%. **45a** was synthesized by the general procedure given above as yellow solid. m.p. 155-158 °C. ¹H NMR (400 MHz, MeOD) δ 7.60 (m, 5H), 7.51 (d, *J* = 8.1 Hz, 2H), 7.42 (t, *J* = 7.5 Hz, 2H), 7.33 (d, *J* = 7.6 Hz, 1H), 7.14 (dd, *J* = 20.8, 7.8 Hz, 2H), 6.97 – 6.88 (m, 1H), 6.75 (d, *J* = 7.6 Hz, 2H), 6.64 (d, *J* = 16.0 Hz, 1H), 6.55 – 6.40 (m, 1H), 5.94 (s, 1H), 4.61 (s, 2H), 3.38 (d, *J* = 6.5 Hz, 2H), 2.42 (s, 3H). ¹³C NMR (101 MHz, MeOD) δ 155.47, 147.93, 138.69, 135.74, 135.59, 135.46, 129.44, 128.55, 128.51, 127.11, 126.95, 126.95, 126.87, 126.87, 126.58, 126.18, 126.18, 125.53, 124.40, 122.46, 119.27, 119.18, 112.01, 102.21, 57.37, 48.17, 38.10. HRMS (EI) m/z calcd for C₂₇H₂₅NO (M)⁺ 379.1936, found 379.1937.

(2E,4E)-N-((1H-indol-7-yl)methyl)-5-([1,1'-biphenyl]-4-yl)-N-methylpenta-2, 4-dien-1-amine hydrochloride (45b).

Yield: 33%. **45b** was synthesized by the general procedure given above as white solid. m.p. 193-199 °C ¹H NMR (400 MHz, MeOD) δ 7.72 (d, J = 7.7 Hz, 1H), 7.67 – 7.59 (m, 4H), 7.56 (d, J = 8.3 Hz, 2H), 7.46 – 7.38 (m, 3H), 7.33 (t, J = 7.3 Hz, 1H), 7.28 (d, J = 7.1 Hz, 1H), 7.16 (t, J = 7.6 Hz, 1H), 7.00 (dd, J = 12.6, 10.5 Hz, 1H), 6.80 (d, J = 12.8 Hz, 1H), 6.73 (dd, J = 15.0, 10.5 Hz, 1H), 6.59 (d, J = 3.2 Hz, 1H), 5.94 (dt, J = 15.2, 7.7 Hz, 1H), 4.63 – 4.38 (m, 4H), 2.80 (s, 3H). ¹³C NMR (101 MHz, MeOD) δ 141.27, 141.00, 140.30, 135.64, 135.59, 135.86, 129.44, 128.53, 128.53, 127.18, 126.97, 126.97, 126.88, 126.88, 126.58, 126.38, 126.38, 125.53, 124.40, 128.53, 124.40, 128.53, 128.53, 124.40, 128.53, 128.53, 124.40, 128.54, 128.54, 128.54, 126.58, 126.58, 126.38, 125.53, 124.40, 128.54, 128.54, 128.54, 125.54, 125.54, 125.55, 124.40, 128.55, 126.55

122.46, 119.26, 119.18, 112.03, 102.21, 57.59, 55.51, 37.98. HRMS (EI) m/z calcd for $C_{27}H_{26}N_2$ (M)⁺ 378.2096, found 378.2098.

1-(3a,7a-dihydrobenzofuran-2-yl)-N-methylmethanamine (48a)

To a solution of 3a,7a-dihydrobenzofuran-3-carbaldehyde (18.2 g, 123 mmol) in methanol (20 mL) was slowly treated with methanamine (31g, 33%, in methanol), and then the reaction was stirred at room temperature for 4 h. Thereafter, NaBH₄ (9.4g, 246 mmol) was added in batches at 0 °C and the reaction was heated to r.t. The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, filtered and condensed. The residue was then purified via flash chromatography on silica gel, eluting with EtOAc/petroleum ether (1/25, v/v) to give **48a** as yellow oil. Yield: 42%.¹H-NMR (400 MHz, CDCl₃) δ 7.63 (dt, *J* = 15.3, 4.5 Hz, 1H), 7.58 (s, 1H), 7.34-7.27 (m, 1H), 7.26-7.24 (m, 1H), 3.90(d, *J* = 0.9 Hz, 2H), 2.51(d, *J* = 4.2 Hz, 3H). ¹³C NMR (101 MHz, DMSO) δ 157.06, 145.21, 143.44, 130.15, 129.52, 124.73, 118.02, 117.62, 55.59, 33.90.

(E)-3-(4-(trifluoromethyl)phenyl)prop-2-en-1-ol (49a).

To a solution of cinnamaldehyde (400.0 mg, 2 mmol) in methanol (10 mL) was treated with sodium borohydride (76.0 mg, 2 mmol) in batches at 0 °C. The reaction mixture was stirred at room temperature for 15 min and concentrated. The residue was poured into water and extracted with EtOAc. The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, filtered and condensed. The crude was used for next step without further separation giving the title **49a** as yellow oil. Yield: 59%. ¹H NMR (400 MHz, DMSO) δ 7.65 (m, 4H), 6.67 (d, J = 16.1 Hz, 1H), 6.58 (dt, J = 16.0, 4.4 Hz, 1H), 5.14 – 4.92 (m, 1H), 4.18 (m, 2H). ¹³C NMR (101 MHz, DMSO) δ 141.50, 134.67, 127.14, 127.14, 127.11, 127.11, 125.88, 125.85, 124.35, 61.69.

(E)-1-(3-bromoprop-1-en-1-yl)-4-(trifluoromethyl)benzene (51a).

To a solution of **49a** (303.0 mg, 1.5 mmol) in anhydrous ether (20 mL) was treated with phosphorus tribromide (84.0 μ L, 0.9 mmol) at 0 °C under N₂ atmosphere. The reaction mixture was stirred at room temperature overnight and poured into ice water containing sodium bicarbonate. The mixture was partitioned between EtOAc and water. The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, filtered and condensed at 30 °C. The crude was used for next step without further purification affording the **51a** as white solid. Yield: 85%. ¹H NMR (400 MHz, DMSO) δ 7.82 (d, *J* = 8.0 Hz, 2H), 7.77 – 7.73 (d, 2H), 6.99 (d, *J* = 21.3, 12.3 Hz, 1H), 6.61 – 6.55 (m, 1H), 4.38 (d, *J* = 7.6 Hz, 2H). ¹³C NMR (101 MHz, DMSO) δ 138.72, 134.70, 127.56, 127.47, 127.47, 127.12, 127.12, 125.90, 119.51, 61.65.

Associated Content

Supporting Information.

Experimental procedures and characterizations of derivatives, bacterial growth assays of *S. aureus* Newman and MRSA strains, the hERG inhibition assay, MIC values of derivatives against MRSA strains, pigment inhibition assay, CrtN enzyme 77

inhibition assay, hydrogen peroxide killing and human whole blood killing, *S. aureus* systemic infection models, anti-fungal assay, cytochrome P450 inhibition assay, pharmacokinetic methods: This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The authors declare no competing financial interests.

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Abbreviations

S. aureus, *Staphylococcus aureus*; MRSA, methicillin-resistant *Staphylococcus aureus*; STX, staphyloxanthin; CrtN, diapophytoene desaturases; ND4BB, New Drugs for Bad Bugs; CrtM, dehydrosqualene synthase; PK, pharmacokinetics; SAR, structure-activity relationship; IC₅₀, half maximal inhibitory concentration; MIC, minimum inhibitory concentration; HPLC, high-performance liquid chromatography; MS, mass chromatography; CFU, colony-forming unit; PBS, phosphate-buffered saline; DMF, *N,N*-dimethylformamide; DMSO, dimethyl sulfoxide; EtOH, ethanol; EtOAc, ethyl acetate; MeOH, methanol; THF, tetrahydrofuran; CH₂Cl₂, dichloromethane.

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FIGURES

Figure 1. Chemical modification strategies and purpose for lead compound 1 and

pigment inhibition activities of analogues 2a-j.



Figure 2. Effects of 23a-b on the bacterial growth of S. aureus Newman (A and E),

USA400 MW2 (B and F), USA300 LAC (C and G), and Mu50 (D and H). Data are

presented as means \pm SEM, n = 2 independent experiments.



Figure 3. Effects of **23a-b** on susceptibility to hydrogen peroxide killing. *S. aureus* Newman (A), USA400 MW2 (B), USA300 LAC (C), and Mu50 (D); *** p < 0.001 via two-tailed t-test (n = three biological replicates, each with two technical replicates).



Figure 4. Effects of **23a-b** on susceptibility to human whole blood killing. *S. aureus* Newman (A), USA400 MW2 (B), USA300 LAC (C), and Mu50 (D); *** p < 0.001 via two-tailed t-test (n = three biological replicates, each with two technical replicates).



Figure 5. Effects of analogues **23a-b** on *S. aureus* bacteria (A and B), Mu50 (C and D) and NRS271 (E and F) survival in hearts and livers of mice (n=10 for each group) challenged with 2.3×10^7 CFU Newman bacteria, 1.1×10^9 CFU Mu50 bacteria and 2.3×10^8 CFU NRS271 bacteria. **P** = pretreatment, drugs or compounds were intraperitoneally administered 24 h before infection compared with the normal treatment (drugs or compounds were intraperitoneally administered 6 h after infection). Statistical significance was determined by a Mann–Whitney test (two-tailed): ** p < 0.01, *** p < 0.001. Each symbol represents the value for an individual mouse. Horizontal bars indicate the observation means, and dashed lines mark the limits of detection.



Figure 6. Effects of **23a-b** on Mu50 survival in the kidneys (A) and hearts (B) of mice challenged with 1.3×10^9 CFU Mu50 bacteria. **P** = pretreatment, drugs or compounds were intraperitoneally administered 24 h before infection compared with the normal treatment (drugs or compounds were intraperitoneally administered 6 h after infection). Statistical significance was determined by a Mann-Whitney test (two-tailed): * p < 0.05, ** p < 0.01, *** p < 0.001, n.s. indicates no significant difference. Each symbol represents the value for an individual mouse. Horizontal bars indicate the observation means, and dashed lines mark the limits of detection.



Figure 7. Effects of **1** and **23a** on *S. aureus* survival in protecting mice (n=15) from lethal *S. aureus* infection challenged with 2×10^8 CFU Newman bacteria.



Figure 8. Effects of **23a** on LRSA45 (5.0×10^7 CFU), LRSA202 (7.0×10^7 CFU), NF65Y (3.5×10^8 CFU) and XN108 (2.1×10^8 CFU) survival in the kidneys (A) and hearts (B) of mice. **P** = pretreatment, drugs or compounds were intraperitoneally administered 24 h before infection compared with the normal treatment (drugs or compounds were intraperitoneally administered 6 h after infection). Statistical significance was determined by a Mann–Whitney test (two-tailed): * p < 0.05, ** p < 0.01, *** p < 0.001, n.s. indicates no significant difference. Each symbol represents the value for an individual mouse. Horizontal bars indicate the observation means, and dashed lines mark the limits of detection.



Figure 9. Effects of **23a** on Newman $(3.1 \times 10^7 \text{ CFU})$, Mu50 $(4.6 \times 10^9 \text{ CFU})$, and NRS271 $(3.5 \times 10^8 \text{ CFU})$ survival in the kidneys (A, C and E) and hearts (B, D and F) of mice. **P** = pretreatment, drugs or compounds were intraperitoneally administered 24 h before infection compared with the normal treatment (drugs or compounds were intraperitoneally administered 6 h after infection). Statistical significance was determined by a Mann–Whitney test (two-tailed): * p < 0.05, ** p < 0.01, *** p < 0.001, n.s. indicates no significant difference. Each symbol represents the value for an individual mouse. Horizontal bars indicate the observation means, and dashed lines mark the limits of detection.



Figure 10. Development of bacterial resistance in Newman towards erythromycin and H_2O_2 .



Figure 11. Images of the treated and nontreated (control) TSA plates of 12 passages Newman (1000 CFU), which were under the sequential H_2O_2 + CrtN inhibitors selection pressure. The concentrations of 23a and 23b were 7.5 nM/mL and 10.0 nM/mL (IC₉₀ of pigment inhibition).



12 passages Newman



12 passages Newman + 23a



12 passages Newman + 23b

SCHEMES

Scheme 1. Syntheses of Derivatives 2a-d and 5a-40a^a

	R ¹	R^2		R ¹	ł
49a, 51a, 2a	4-trifluoromethylphenyl	H 49r	, 51r, 21a	4-(tert-butyl)phenyl	ł
49a, 51a, 2b	4-trifluoromethylphenyl	H 49s	s, 51s, 22a	4-(methoxycarbonyl)pheny	/ ł
49a, 51a, 2c	4-trifluoromethylphenyl	H 49t	, 51t, 23a	4-phenylphenyl	ł
49a, 51a, 2d	4-trifluoromethylphenyl	H 49u	ı, 51u, 24a	2-chiorophenyl	ł
49b, 51b, 5a	cyclopentyl	H 49v	v, 51v, 25a	2-fluorophenyl	ł
49c, 51c, 6a	cyclohexyl	H 49v	v, 51w, 26a	2-methylphenyl	ł
49d, 51d, 7a	furan-2-yl	H 49X	(, 51X, 2/a	2-nitropnenyi	1
49e, 51e, 8a	thiophene-2-yl	H 49y	7, 51y, 28a		
49f, 51f, 9a	naphthalen-1-yl	H 492	2, 51Z, 29a	2-methoxyphenyl	
49g, 51g, 10a	naphthalen-1-yi	H 508	1, 52a, 50a	3-fluorophenyl	
490, 510, 11a	A chiorophonyl	H 500	520, 37a	3-methylphenyl	
491, 511, 12a 49i 51i 13a	4-chlorophenyl	н 50c	1. 52d. 33a	3-nitrophenyl	i
49k 51k 14a	4-homophenyl	н 50е	e. 52e. 34a	3-trifluoromethylphenyl	i
491. 511. 15a	4-methylphenyl	н 50f	, 52f, 35a	3-methoxyphenyl	i
49m.51m. 16a	4-methoxyphenyl	H 50g	, 52g, 36a	2,4-dichlorophenyl	1
49n,51n, 17a	4-ethyoxyphenyl	H 50	n, 52h, 37a	2-fluoro-4-trifluoromethylp	henyll
49o, 51o, 18a	4-nitrophenyl	H 50i	, 52i, 38a	3-fluoro-4-trifluoromethylp	henyll
49p, 51p, 19a	4-difluoromethylphenyl	Н 50ј	, 52j, 39a	2-fluoro-4-methoxyphenyl	1
49q, 51q, 20a	4-cyanolphenyl	H 201	k, 52k, 40a	3-fluoro-4-methoxyphenyl	1
	\mathbf{R}^2	E	3r	ŎН	
	N, \overline{R}^{1}	□ 1	∫ f ⊳1≎	е _1 _CHO)
	• HCI		∢ ∩	$ \leftarrow R' = P^2 = $	
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\	j g	52a-k.	50)a-k.	
(\sim 2a, X=0; \sim	1 ~	\sim		
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ľ.	∕—x́		7		
2b	, 5-40a, X=O:		- X		
2d	. X=S.	48	b Acab A	46b	
	, 		40a-D,4	оа-ы, х= 0,5 а	
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^aReagents and conditions: (a) ammonia in THF, DCM, NaBH(OAc)₃, r. t, 2 h, 30%; (b) (1) K₂CO₃, DMF, r.t., overnight, 43-85%, (2) bubbled into hydrogen chloride gas. (c) iodoethane or 2-iodopropane, NaH, DMF 0 °C to r.t, overnight, under

N₂, 50%; (d) methylamine, MeOH, NaBH₄, r. t, 0 °C, 10 mins, 55%; (e) NaBH₄, methanol, 0 °C, 30 min; (f) phosphorus tribromide, diethyl ether, 0 °C to r.t., overnight, under N₂, 57-84% (2 steps); (g) (1) K₂CO₃, DMF, r.t., overnight, 43-85%, (2) bubbled into hydrogen chloride gas.

Scheme 2. Syntheses of Derivatives 2e-j, 3d-f and 5b-40b^a

2h		R ¹	R^2		R ¹ R ²
49a. 51a.	2e	4-trifluoromethylphenyl	Н	49q, 51q, 20b	4-cyanolphenyl H
49a. 51a.	2f	4-trifluoromethylphenyl	H	49r, 51r, 21b	4-(tert-butyl)phenyl H
49a. 51a.	2a	4-trifluoromethylphenyl	Н	49s, 51s, 22b	4-(methoxycarbonyl)pheny H
49a, 51a,	2ň	4-trifluoromethylphenyl	Н	49t, 51t, 23b	4-phenylphenyl H
49a, 51a,	2i	4-trifluoromethylphenyl	Н	49u, 51u, 24b	2-chiorophenyl H
49a, 51a,	2i	4-trifluoromethylphenyl	Н	49v, 51v, 25b	2-fluorophenyl H
49b, 51b,	5b	cyclopentyl	н	49w, 51w, 26b	2-methylphenyl H
49c, 51c,	6b	cýclohexyl	н	49x, 51x, 27b	2-nitrophenyl H
49d, 51d,	7b	furan-2-yl	н	49y, 51y, 28b	2-trifluoromethylphenyl H
49e, 51e,	8b	thiophene-2-yl	н	49z, 51z, 29b	2-methoxyphenyl H
49f, 51f,	9b	naphthalen-1-yl	н	50a, 52a, 30b	3-chiorophenyl H
49g, 51g,	10b	naphthalen-1-yl	Н	50b, 52b, 31b	3-fluorophenyl H
49h, 51h,	11b	phenyl	Н	50c, 52c, 32b	3-methylphenyl H
49i, 51i,	12b	4-chiorophenyl	Н	50d, 52d, 33b	3-nitrophenyl H
49j, 51j,	13b	4-fluorophenyl	н	50e, 52e, 34b	3-trifluoromethylphenyl H
49k, 51k,	14b	4-bromophenyl	Н	50f, 52f, 35b	3-methoxyphenyl H
49I, 51I,	15b	4-methylphenyl	Н	50g, 52g, 36b	2,4-dichlorophenyl H
49m,51m,	16b	4-methoxyphenyl	Н	50h, 52h, 37b	2-fluoro-4-trifluoromethylphenyl H
49n,51n,	17b	4-ethyoxyphenyl	Н	50i, 52i, 38b	3-fluoro-4-trifluoromethylphenyl H
490, 510,	18b	4-nitrophenyl	Н	50j, 52j, 39b	2-fluoro-4-methoxyphenyl H
49p, 51p,	19b	4-difluoromethylphenyl	н	50k, 52k, 40b	3-fluoro-4-methoxyphenyl H
	subs	ititued R ²		Br	ŎН



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^{*a*}Reagents and conditions: (a) ammonia in THF, DCM, NaBH(OAc)₃, r. t, 2h, 30%; (b) (1) K₂CO₃, DMF, r.t., overnight, 43-85%, (2) bubbled into hydrogen chloride gas. (c) iodoethane or 2-iodopropane, NaH, DMF 0 °C to r.t, overnight, under N₂, 50%; (d) methylamine, MeOH, NaBH₄, r. t, 0 °C, 10 mins, 55%; (e) NaBH₄, methanol, 0 °C, 30 min; (f) phosphorus tribromide, diethyl ether, 0 °C to r.t., overnight, under N₂, 57-84% (2 steps); (g) (1) K₂CO₃, DMF, r.t., overnight, 43-85%, (2) bubbled into hydrogen chloride gas.

Scheme 3. Syntheses of Derivatives 41a-b^a



^aReagents and conditions: (a) CuI, Pd(PPh₃)₄, DMF, TEA, 90 °C; (b) TBAF/THF;
(c). PBr₃, Et₂O; (d) (1) K₂CO₃, DMF, r.t., overnight, 41-45%, (2) bubbled into hydrogen chloride gas.

Scheme 4. Syntheses of Derivatives 42a-b^a



^aReagents and conditions: (a) H₂, Pd/C, MeOH; (b) PBr₃, CCl₄, reflux, 4 h; (c).

(1) K₂CO₃, DMF, r.t., overnight, 25-35%, (2) bubbled into hydrogen chloride gas..





^{*a*}Reagents and conditions: (a) ethyl 2-(triphenyl-15-phosphanylidene)propanoate, toluene, 100 °C; (b) DIABL-H, DCM, 0 °C; (c). PBr₃, Et₂O; (d)(1) K₂CO₃, DMF, r.t., overnight, 41-55%, (2) bubbled into hydrogen chloride gas.





^{*a*}Reagents and conditions: (a)(1) (2-carboxyethyl)triphenylphosphonium, NaH in THF/DMSO, (2) EtOH/H₂SO₄; (b) DIABL-H, DCM, 0 °C; (c). CBr₄, PPh₃, DCM; (d) (1) K₂CO₃, DMF, r.t., overnight, (2) bubbled into hydrogen chloride gas.

Scheme 7. Syntheses of Derivatives 45a-b^a



1	
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3	^a Reagents and conditions: (a) ethyl 2-(triphenyl-l5-phosphanylidene)acetate,
4 5	
6	toluene 100 °C· (b) DIABL-H_DCM_0 °C· (c)_PBr2_EtaO· (d)(1) K2CO2_DME_r t
7	(0) (0)
8	avarnight 29.270/(2) hubbled into hudrogon ableride gas
9	overnight, 28-3776, (2) bubbled into hydrogen chloride gas.
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TABLES

 Table 1. Chemical structures of drivatives 3a-f and their pigment inhibitory activities

 against S. aureus Newman.



Compd	R^1	R^2	$IC_{50}^{a}(nM)$
3 a	Н		> 1000
3b	ethyl		> 1000
3c	isopropyl		> 1000
3d	Н	HZ C	> 1000
3e	ethyl	K T	117.9 ± 8.9
3f	isopropyl	K K	> 1000

^{*a*}The values given are the IC₅₀ values for pigment inhibition in *S. aureus* Newman. The values are reported as the average \pm S.D.

Table 2. Structures and pigment inhibition activities of the analogs of compound4a-40a



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	Compd	R ³	IC_{50}^{a} (nM)	Compd	R ³	IC_{50}^{a} (nM)
_	4a	ž	91.3 ± 3.8	23a	, () ()	2.0 ± 0.1
	5a	z	397.3 ± 20.3	24a	τ τ CI	359.8 ± 57.8
	6a	×	> 1000	25a	·ze F	> 1000
	7a	× 50	> 1000	26a	*z	86.4 ± 0.6
	8a	34 S	> 1000	27a	₹ NO2	74.1 ± 2.7
	9a	24	> 1000	28a	₹ CF3	> 1000
	10a	¥	6.4 ± 0.6	29a	Ne Ne	> 1000
	11a	4	> 1000	30a	3 CI	> 1000
	12a	₹, CI	4.9 ± 0.1	3 1a	₹	> 1000
	1 3 a	F	74.1 ± 2.7	32a	4	> 1000
	14a	, ↓ ↓ Br	2.5 ± 0.3	33 a	3-2- NO2	> 1000
	15a	ł	9.9 ± 1.8	34a	¹ /2 CF3	> 1000
	16a	2 OMe	15.8 ± 2.8	35a	Ne OMe	> 1000
	17a	•=OEt	> 1000	36a	₹ Ţ	6.2 ± 0.5
	18 a	NO2	9.5 ± 3.2	37a	CF3	9.2 ± 0.4
	19a	CHF ₂	6.4 ± 0.1	38 a	F CF3	> 1000
	20a		80.1 ± 4.7	39a	·₂₂ ₂₂	> 1000
	21a	J) k	> 1000	40a	F OMe	> 1000
		5 ×				

22a
$$9.5 \pm 0.8$$

^{*a*}The values given are the IC₅₀ values for pigment inhibition in *S. aureus* Newman. The values are reported as the average \pm S.D.

 Table 3. Structures and pigment inhibition activities of the analogs of compound

 4b-40b



Compd	R^3	$\mathrm{IC}_{50}^{a}(\mathrm{nM})$	Compd	R^3	IC_{50}^{a} (nM)
4 b	Safety and S	> 1000	23b	4	3.3 ± 0.4
5b	2	> 1000	24b	ζ. CI	> 1000
6b	·2	> 1000	25b	₹ F	> 1000
7b	3/0	> 1000	26b	¥	> 1000
8b	2 S	> 1000	27b	24 NO2	> 1000
9b	*	> 1000	28b	₹ CF3	> 1000
10b	2	131.8 ± 11.6	29b	No.	> 1000
11b	ł	> 1000	30b	ξ. CI	> 1000
12b	₹, CI	142.6 ± 5.6	31b	× F	> 1000
13b	,₂F	> 1000	32b	ł.	> 1000
14b	, Br	83.8 ± 2.5	33b	3 NO2	> 1000

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15b	24	199.0 ± 4.2	34b	₹. ℃F3	> 1000
16b	., ₹	167.0 ± 24.2	35b	₹.COMe	> 1000
17b	₹2 OEt	64.8 ± 3.2	36b	₹ CI	279.6 ± 0.3
18b	NO2	> 1000	37b	₹ F	48.4 ± 0.2
19b	CHF2	43.5 ± 5.4	38b	کر CF3 ۴	> 1000
20b	'łż	784.7 ± 20.0	39b	∿OMe F	73.1 ± 0.3
21b	4 C	700.0 ± 57.4	40b	°4₂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂⊂	> 1000
22b	₹, CO₂Me	87.8 ± 0.2			

^{*a*}The values given are the IC₅₀ values for pigment inhibition in *S. aureus* Newman. The values are reported as the average \pm S.D.

Table 4. Structures and pigment inhibition activities of the analogs of compound41-45a and 41-45b.



Compd.	R ²	Linker (X)	S. aureus Newman $IC_{50} (nM)^{a}$
41 a	o - C	and the second s	> 1000
42a		e ⁵ 2	> 1000
		103	

43 a	O	phi to the second se	> 1000
44a		w to	> 1000
45a		r the second sec	> 1000
41b	H N	22 Jacobson Start	101.8 ± 5.2
42b	H K	, e ^s	> 1000
43b	H. T	, of the second se	> 1000
44b	H. T	r h	> 1000
45b	HZ C	phi and a second s	19.4 ± 0.2

^{*a*}The values given are the IC₅₀ values for pigment inhibition in *S. aureus* Newman. The values are reported as the average \pm S.D.

Table 5. Cytotoxicity of **12a**, **14a** and **23a-b** on human embryonic kidney cell(HEK-293) and human hepatocellular carcinoma cell (HepG2).

	HEK-293	HepG2
Compd.		
	$\text{CC}_{50}^{a}(\mu\text{M})$	$IC_{50}^{b}(\mu M)$
Amphotericin B	4.2	3.1
12a	42.1	60.2
	101	

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14a	41.9	33.0
23a	48.1	105.8
23b	26.5	21.9

^{*a*}The values given are the IC₅₀ values for Cytotoxicity of HEK-293. ^{*b*}The values given are the IC₅₀ values for Cytotoxicity of HepG2.

Table 6. Effects of **23a-b** on pigment production of *S. aureus* USA400 MW2,USA300 LAC, Mu50 and NRS271.

	USA400 MW2	USA300 LAC	Mu50	NRS271
Compd.	IC_{50}^{a} (nM)	$\mathrm{IC}_{50}^{b}(\mathrm{nM})$	$\mathrm{IC}_{50}^{c}(\mathrm{nM})$	$\mathrm{IC}_{50}^{d}(\mathrm{nM})$
1	3.4 ± 0.02	5.5 ± 0.4	0.4 ± 0.02	6.7 ± 0.3
23a	7.9 ± 1.2	1.9 ± 0.2	0.8 ± 0.01	5.6 ± 0.9
23b	8.0 ± 0.1	4.0 ± 0.3	0.4 ± 0.04	10.1 ± 0.6

^{*a*}The values given are the IC₅₀ values against USA400 MW2. The values are reported as the average \pm S.D ^{*b*}The values given are the IC₅₀ values against USA300 LAC. The values are reported as the average \pm S.D. ^{*c*}The values given are the IC₅₀ values against Mu50. The values are reported as the average \pm S.D. ^{*d*}The values given are the IC₅₀ values against NRS271. The values are reported as the average \pm S.D.

Table 7. The results of water solubility, hERG and CrtN inhibition activities (IC₅₀) of **1** and **23a-b.**

G 1	Water solubility ^a	hERG inhibition	CrtN IC ₅₀ ^c
Compd.	(mg/mL)	activities IC_{50}^{b} (μ M)	(µM)

1	10.0	3.7	0.3
23a	0.2	34.8	0.4 ± 0.01
23b	0.2	> 40	0.2 ± 0.01

^{*a*}The values given are the values of water solubility. ^{*b*}The values given are the IC₅₀ values of hERG inhibition activity. ^{*c*}The values given are the IC₅₀ values against CrtN. The values are reported as the average \pm S.D.

Table 8. Human miarosome stability of 23a-b.

Compd.	k	CL _{int, in vitro} (ml/min/mg)	CL _{int, in vivo} (ml/min/mg)	t _{1/2} (min)
Midazolam	0.165	0.33	381.0	4.2
23a	0.007	1.23	31.6	50.7
23b	0.014	0.02	17.0	93.7

Table 9. Antifungal activities of 1 and 23a-b.

Antifungal Activities, MIC ₈₀ (µg /mL)					
Compd.	Trichophyton rubrum	Microsporum gypseum	Tinea barbae		
Ketoconazole	0.5	2	0.0625		
Voriconazole	0.03125	0.25	0.03125		
Fluconazole	1	8	2		
1	16	32	> 64		

23a	64	32	> 64
23b	> 64	> 64	> 64

Table 10. Effects of **23a** on pigment production of NRS271, NRS70, NRS100,NRS108, LRSA56, LRSA202, LRSA205, HS663, NF65Y and XN108.

Pigment production, IC ₅₀ (nM)				
MRSA strains (country)	IC ₅₀ (nM)	MRSA strains (country)	IC ₅₀ (nM)	
NRS70 (JPN)	1.7 ± 0.01	LRSA205 (CHN)	10.5 ± 0.4	
NRS100 (USA)	1.2 ± 0.1	HS663 (CHN)	0.02 ± 0.01	
NRS108 (FRA)	5.6 ± 0.02	NF65Y (CHN)	1.0 ± 0.02	
LRSA56 (CHN)	1.8 ± 0.1	XN108 (CHN)	0.2 ± 0.04	
LRSA202 (CHN)	1.6 ± 0.2			

Table 11. Effects of **23a** on inhibition of CYP1A2, CYP2B6, CYP2C8, CYP2C9,CYP2C19, CYP2D6, CYP3A4M and CYP3A4T.

СҮР	Selective Inhibitors	Inhibition Activity	Compd.	Inhibition Activity
		IC ₅₀ (µM)		IC ₅₀ (µM)
1A2	α -Naphthoflavon	30	23a	5.7
2B6	Ticlopidine	70	23a	10.0
2C8	Montelukast	1	23a	10.0
2C9	Sulfaphenazole	10	23a	10.0
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2C19

2D6

3A4M

3A4T

Omeprazole

Quinidine

Ketoconazole

Ketoconazole

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dose of 23a mg/kg	AUC _(0-24h) (h*ng/mL)	AUC _(0-∞) (h*ng/mL)	C _{max} (ng/mL)	T _{max} (h)	CL (mL/h/kg)	T _{1/2} (h)	F (%)
5 (I.V.)	2805.48	2862.41	4608.78	0.08	1750.00	2.46	-
10 (P.O.)	898.70	930.33	212.56	2.00	-	1.29	16.30
10 (I.P.)	974951.40	1086152.29	130484.03	3.00	-	7.61	

 Table 13. Pharmacokinetic parametears for 23a after single dosing in mice.

dose of 23a	AUC _(0-24h)	AUC _(0-∞)	C _{max}	T _{max}	CL	T _{1/2}	E(0/)
mg/kg	(h*ng/mL)	(h*ng/mL)	(ng/mL)	(h)	(mL/h/kg)	(h)	F (%)
2 (I.V.)	1067.27	1086.95	2230.35	0.08	1840.00	2.05	-

10 (P.O.)	4528.41	4556.40	801.19	2.00 -	3.35	83.84
			110			

Table 14. Effects of **23a-b** on pigment production of the original and 12 passages of *S*. *aureus* Newman under IC_{90} concentrations of **23a** or **23b**.

	Pigment production, IC	C ₅₀ (nM)
Compd.	Original <i>S. aureus</i> Newman,	12 passages of <i>S. aureus</i> Newman,
	IC_{50}^{a} (nM)	IC_{50}^{b} (nM)
23a	2.0 ± 0.1	1.8 ± 0.1
23b	3.3 ± 0.4	2.6 ± 0.2

^{*a*}The values given are the IC₅₀ against original *S. aureus* Newman. The values are reported as the average \pm S.D. ^{*a*}The values given are the IC₅₀ against 12 passages of *S. aureus* Newman. The values are reported as the average \pm S.D..

 Table 15. MIC values of 23a for enterococcus faecium, S. aureus Newman, and

 pseudomonas aeruginosa.

	MIC()	ıg/mL)	
Connel	enterococcus	S. aureus	pseudomonas
Compd.	faecium	Newman	aeruginosa
23a	>500µg/mL	>500µg/mL	>500µg/mL

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