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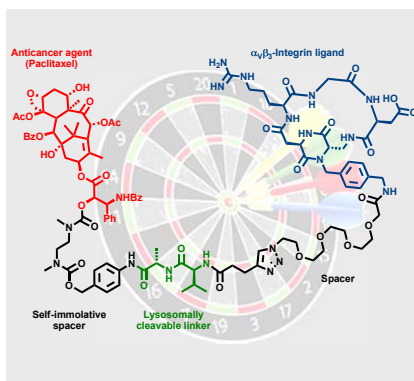
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FULL PAPER

Hitting the Bullseye with the best dart: RGD-peptidomimetics with high binding affinity for integrin $\alpha_v\beta_3$, which is overexpressed in the blood vessels of several human cancers, were conjugated to the cytotoxic drug Paclitaxel; variation of the $\alpha_v\beta_3$ -integrin ligand, lysosomally cleavable linker and spacer permits the identification of a conjugate with high tumor Targeting Index (TI = 533).



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Synthesis and Biological Evaluation of Paclitaxel Conjugates Involving Lysosomally Cleavable Linkers and $\alpha_v\beta_3$ -Integrin Ligands for Tumor Targeting

Key Topic: Tumor Targeting

Synthesis and Biological Evaluation of Paclitaxel Conjugates Involving Lysosomally Cleavable Linkers and $\alpha_v\beta_3$ -Integrin Ligands for Tumor Targeting

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Abstract: Two *cyclo*[DKP-RGD]-PTX (PTX = paclitaxel) and two *cyclo*[RGDFK]-PTX conjugates containing the lysosomally cleavable Gly-Phe-Leu-Gly (GFLG) linker were synthesized and compared to two *cyclo*[DKP-RGD]-Val-Ala-PTX conjugates. The conjugates were evaluated for their ability to inhibit biotinylated vitronectin binding to the isolated $\alpha_v\beta_3$ receptor, retaining good binding affinity, in the same nanomolar range of the free ligands. Cell viability assays were performed for the six conjugates in the $\alpha_v\beta_3$ + U87 and in the $\alpha_v\beta_3$ - HT29 cell lines. Loss of potency was observed for all the conjugates, attenuated by the presence of a tetraethylene glycol (PEG-4) spacer. A good Targeting Index (TI = Relative Potency in the $\alpha_v\beta_3$ + U87 / Relative Potency in the $\alpha_v\beta_3$ - HT29) was displayed by the conjugates, in particular by *cyclo*[DKP-RGD]-PEG-4-Val-Ala-PTX **9** (TI = 533). This conjugate was tested in the $\alpha_v\beta_3$ + U87 cell line in the presence of 50-fold excess free *cyclo*[DKP-RGD] ligand **2**. In this competition experiment, a fivefold decrease of the conjugate cytotoxicity was calculated, suggesting that the conjugate is possibly internalized by an $\alpha_v\beta_3$ integrin-mediated process.

Introduction

Cytotoxic drugs are characterized by a limited clinical efficacy in the cure of cancer due to a narrow therapeutic window, which means that the difference between the Maximum Tolerated Dose (MTD) and the Minimum Effective Dose (MED) is very small.^[1] One of the approaches to overcome this limitation is to increase the selectivity of the cytotoxic agents by conjugation to tumor-targeting devices, which leads to less severe side effects

and to the requirement of lower quantities.^[2] To date, Antibody-Drug Conjugates (ADCs) have been successfully used for this objective, combining humanized or chimeric antibodies with cytotoxic drugs, that allow the selective delivery of the antitumor agent into antigen-positive malignant cells. Indeed, four ADCs have been introduced into the market: the well-known KadcylaTM (azo-trastuzumab emtansine) and AdcetrisTM (brentuximab vedotin) and the recently approved MylotargTM (gemtuzumab-ozogamycin) and Besponsa^{TR} (inotuzumab ozogamicin).^[1, 3] However, several drawbacks are associated to this antibody targeting approach: high manufacturing costs, low tissue diffusion, possible immune system-induced alteration of the drug efficiency, etc..^[4] As an alternative, Small Molecule-Drug Conjugates (SMDCs) have emerged in the last decades, combining small receptor binders with cytotoxic agents. Just as in ADCs, also in SMDCs the choice of a proper linker system is crucial to avoid premature drug release in the blood stream and to achieve selective release of the payload at the tumor site.

One of the possible receptors that can be targeted by SMDCs is integrin $\alpha_v\beta_3$. This heterodimeric transmembrane receptor is overexpressed in the blood vessels of several human cancers (breast cancer, glioblastoma, pancreatic tumor, prostate carcinoma) but not in the healthy tissues.^[5] Integrin $\alpha_v\beta_3$ recognizes the tripeptide Arg-Gly-Asp (RGD)^[6] and for this reason a number of peptides and peptidomimetics containing this sequence and displaying high binding affinity to the receptor (in the low nanomolar range) have been prepared.^[7] Many of these RGD-peptidomimetics – such as, for example, *cyclo*[RGDFK] **1** (Figure 1) – have been used for the preparation of tumor-targeting drugs and imaging agents.^[8,9] In this context, our research group has developed a series of cyclic RGD ligands containing a diketopiperazine scaffold (DKP),^[7d-f] among which *cyclo*[DKP-RGD] **2** (Figure 1) showed a very good binding affinity and selectivity for integrin $\alpha_v\beta_3$ (Table 1). As a further step, compound **3** (Figure 1) – i.e. an analog of ligand *cyclo*[DKP-RGD] **2** bearing a primary amino group^[10] – was synthesized and conjugated to different drugs through various types of linker.^[10,11] In an initial attempt, we connected ligand *cyclo*[DKP-RGD] to paclitaxel (PTX) through an ester linkage, but the construct obtained suffered from limited stability in plasma.^[10] We then connected ligand *cyclo*[DKP-RGD] to paclitaxel (PTX) through a more stable carbamate linkage and the lysosomally cleavable Val-Ala (VA) dipeptide linker,^[12] obtaining a *cyclo*[DKP-RGD]-VA-PTX conjugate.^[11b,13] With the latter compound, cell viability assays were run in which a cell line expressing integrin $\alpha_v\beta_3$ ($\alpha_v\beta_3$ +) was compared with an $\alpha_v\beta_3$ non-expressing isogenic cell line ($\alpha_v\beta_3$ -). In these experiments, the *cyclo*[DKP-RGD]-VA-PTX conjugate^[11b,13] showed a fairly effective integrin targeting [Targeting Index (TI) up to 9.0],

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although the potency was slightly reduced compared to free PTX.^[11b] In another contribution from our group, the VA linker was also exploited for the development of *cyclo*[DKP-RGD] conjugates with α -amanitin.^[11e] Recently, we also explored the use of reductively cleavable disulfide linkers, synthesizing a *cyclo*[DKP-RGD]-SS-camptothecin conjugate^[11d] which, however, did not show any targeting due to the low stability of the α,α -unsubstituted disulfide bond in the cell medium.^[14]

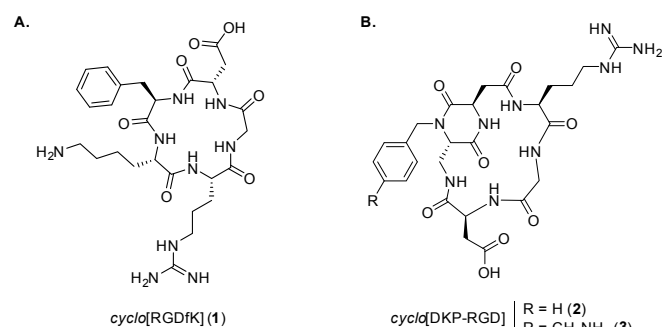


Figure 1. A: ligand *cyclo*[RGDFK] (1). B: ligand *cyclo*[DKP-RGD] (2) and its functionalized version (3).

Table 1. Inhibition of biotinylated vitronectin binding to the isolated $\alpha_v\beta_3$ and $\alpha_v\beta_5$ receptors.

Ligand	Structure	IC ₅₀ (nM) ^[a] $\alpha_v\beta_3$	IC ₅₀ (nM) ^[a] $\alpha_v\beta_5$
1	<i>cyclo</i> [RGDFK]	1.4 ± 0.2	117.5 ± 7.8
2	<i>cyclo</i> [DKP-RGD]	4.5 ± 1.1	149 ± 25

[a] IC₅₀ values were calculated as the concentration of compound required for 50% inhibition of biotinylated vitronectin binding as estimated by GraphPad Prism software. All values are the arithmetic mean ± the standard deviation (SD) of duplicate determinations.

Following the promising preliminary results outlined above,^[11b] in this paper we report a more systematic study on the RGD conjugates bearing intracellularly cleavable peptide linkers. Taking conjugate *cyclo*[DKP-RGD]-VA-PTX^[11b,13] as a reference, we set to assess the influence of the SMDC's different moieties on integrin binding and cell viability. As shown in Figure 2, keeping the PTX-self-immolative spacer system unchanged, we modified in turn: i) the peptide linker; ii) the spacer connecting the linker to the ligand; iii) the ligand itself. In conjugates 4–7 (Figure 3) we selected the well-known lysosomally cleavable tetrapeptide Gly-Phe-Leu-Gly (GFLG), that has been used in several different delivery systems,^[15] as alternative to the VA linker (present in conjugates 8 and 9). To replace the glutarate spacer (present in conjugates 4, 5 and 8), we used a PEG-4 chain connected to a triazole ring formed by the CuAAC “click” reaction^[16] (conjugates 6, 7 and 9). Finally, in conjugates 5 and 7 the well-known and easy-to-prepare ligand *cyclo*[RGDFK]^[17] was employed as replacement for ligand *cyclo*[DKP-RGD].

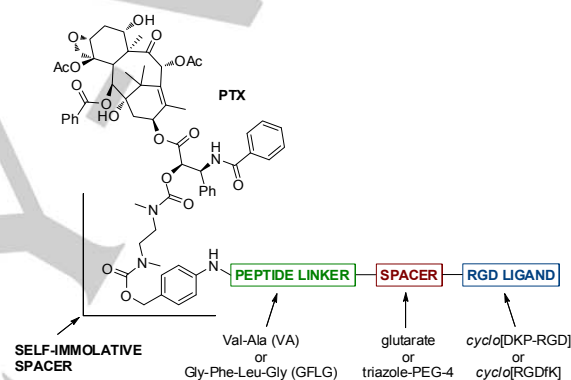


Figure 2. Synopsis of the RGD ligand-PTX conjugates used for the studies reported in this paper.

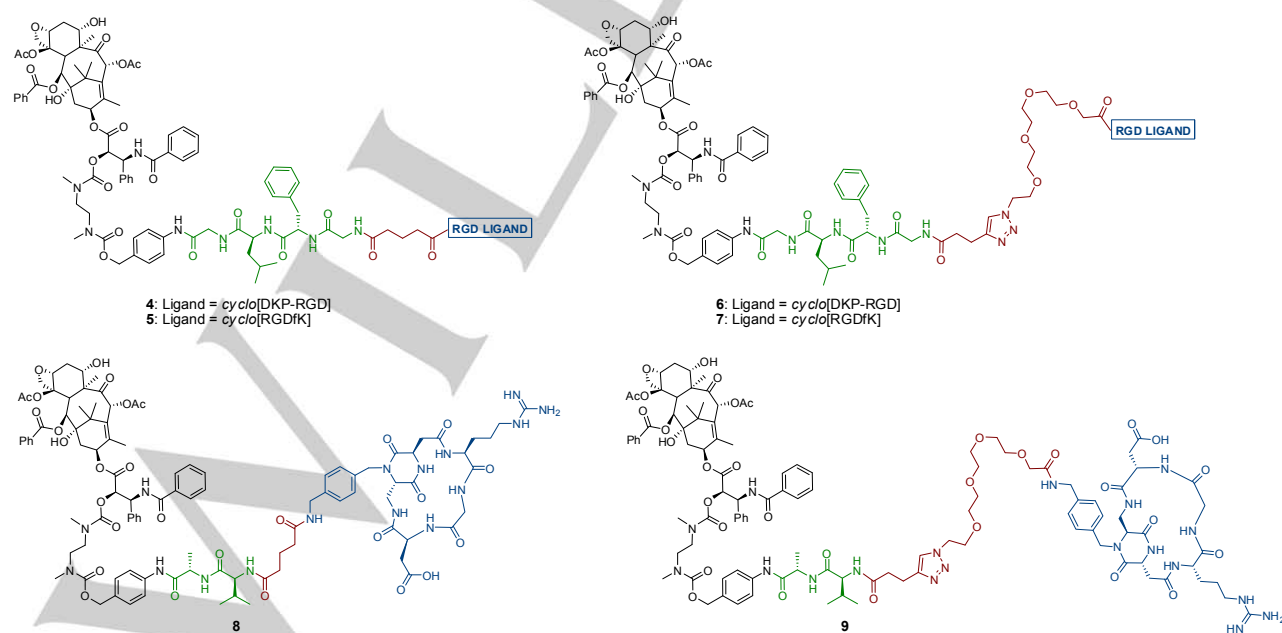


Figure 3. PTX conjugates: *cyclo*[DKP-RGD]-GFLG-PTX (4), *cyclo*[RGDFK]-GFLG-PTX (5), *cyclo*[DKP-RGD]-PEG-4-GFLG-PTX (6), *cyclo*[RGDFK]-PEG-4-GFLG-PTX (7), *cyclo*[DKP-RGD]-VA-PTX (8) and *cyclo*[DKP-RGD]-PEG-4-VA-PTX (9).

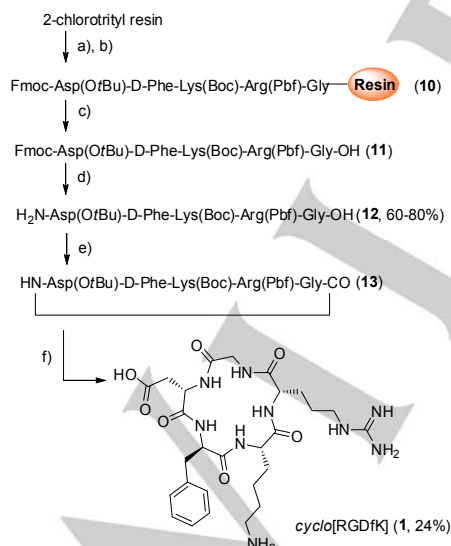
Results and Discussion

While conjugates **8**^[11b] and **9**^[18] (featuring the VA linker) had already been prepared in our laboratory, the synthesis of compounds **4–7** is reported here. In these new SMDCs, PTX is connected to the C-terminus of the GFLG linker through a *para*-aminobenzyl carbamate (PABC)-*N,N'*-dimethylethylenediamine self-immolative spacer, while the N-terminus of the tetrapeptide is connected to the integrin ligand (either *cyclo*[DKP-RGD] or *cyclo*[RGDfK] **1**, Scheme 1) with a spacer, see Scheme 2. The series is formed by two pairs of analogs (**4** / **6** and **5** / **7**) differing only for the spacer, which can be a glutarate (compounds **4** and **5**) or a triazole-tetraethylene glycol (PEG-4, compounds **6** and **7**). Integrin receptor competitive binding assays (Table 2) and cell viability assays (Table 3) were performed for the new GFLG conjugates (**4–7**) and for the VA conjugates **8** and **9** (Figure 3).^[11b,18] Additionally, a competition experiment was performed (Table 4), in which the $\alpha_v\beta_3$ U87 cell line was treated with conjugate **9** in the presence of 50-fold excess free ligand *cyclo*[DKP-RGD] (**2**).

Synthesis

Synthesis of compounds **1** and **14**

The linear protected RGDfK ligand [Fmoc-Asp(OrBu)-D-Phe-Lys(Boc)-Arg(Pbf)-Gly-OH **11**, Scheme 1] – precursor of *cyclo*[RGDfK] **1** – and the Fmoc-protected GFLG linker **14** (Scheme 2) were synthesized manually by solid phase peptide synthesis (SPPS) on 2-chlorotrityl chloride resin using the Fmoc protocol (see the Supporting Information). Cleavage from the resin (using a 8:1:1 CH₂Cl₂/MeOH/AcOH mixture for 2 h) followed by precipitation in water led to the protected peptides **11** (Scheme 1) and **14** (Scheme 2), which were used in the next step without purification.



Scheme 1. Synthesis of ligand **1**. Reagents and conditions: a) i. Fmoc-Gly-OH (1 equiv.), *i*Pr₂NEt (3 equiv.), 1:1 CH₂Cl₂/DMF, 2 h, r.t.; ii. Capping with 7:2:1 CH₂Cl₂/MeOH/*i*Pr₂NEt; b) i. Fmoc-deprotection: 2% DBU, 2% piperidine, DMF, 1 h; ii. Fmoc-AA-OH (3 equiv.), HOBT (4 equiv.), DIC (4 equiv.), 2 h; conditions (b) are repeated for the coupling of every amino acid of the sequence; c) 8:1:1 CH₂Cl₂/MeOH/AcOH, 2 h, precipitation in water; d) DMF, 20% piperidine, 2 h; e) 6:4:4 *i*Pr₂NEt/BOP/HOBt, 1 mM concentration in DMF, 24 h, precipitation in 5% NaHCO₃; f) TFA/thioanisole/EDT/phenol/TIS, 3 h. DBU = 1,8-Diazabicyclo[5.4.0]undec-7-ene; AA = amino acid; BOP = (benzotriazol-1-yloxy)tris(dimethylamino)phosphonium hexafluorophosphate; DIC = *N,N'*-diisopropylcarbodiimide; EDT = 1,2-ethanedithiol; TIS = triisopropylsilane.

Compound **11** was treated with piperidine to remove the Fmoc protecting group and the resulting free amino acid **12** was cyclized in the presence of a 6:4:4 *i*Pr₂NEt/BOP/HOBt mixture in DMF to give compound **13**. The cyclization reaction was run under diluted conditions (1 mM) to avoid possible intermolecular side reactions. The protecting groups of **13** were removed with a TFA/thioanisole/EDT/phenol/TIS cocktail to give pure *cyclo*[RGDfK] (**1**) after purification using preparative HPLC.

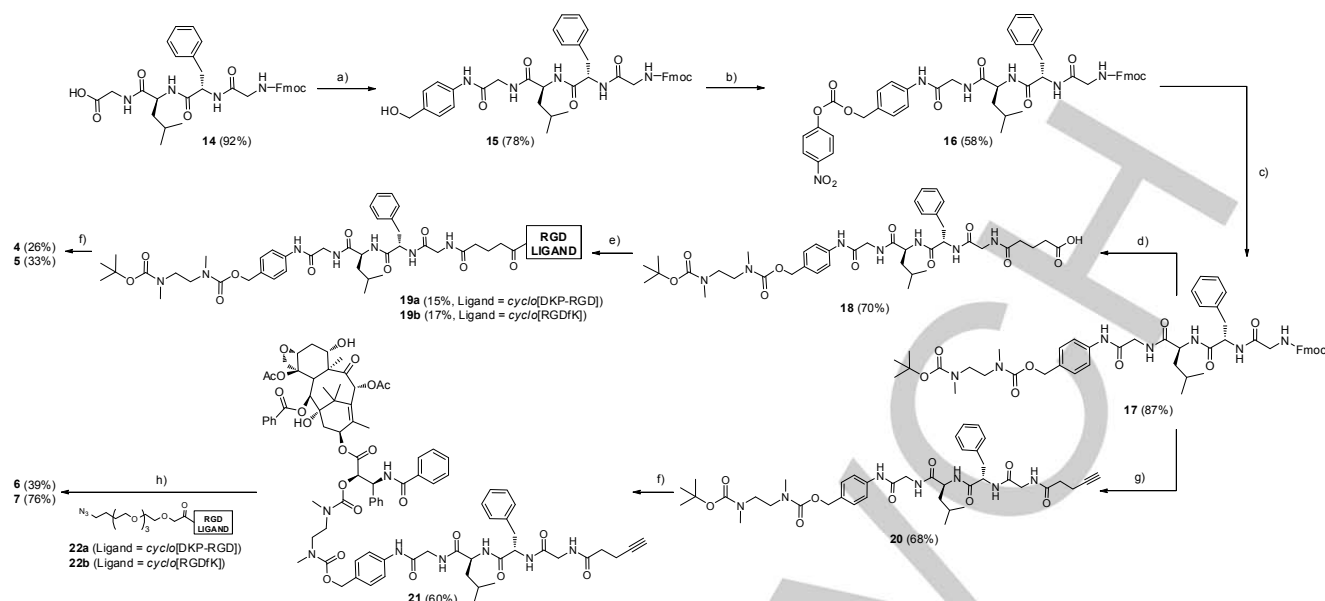
Synthesis of conjugates **4–7**

The *cyclo*[DKP-RGD]-GFLG-PTX and the *cyclo*[RGDfK]-GFLG-PTX conjugates (compounds **4–7**) were synthesized as shown in Scheme 2. The Fmoc-protected GFLG peptide **14** was coupled to 4-aminobenzyl alcohol with HOBT and *N,N'*-diisopropylcarbodiimide (DIC), leading to the benzylic alcohol **15**, which was activated with 4-nitrophenyl chloroformate and coupled to *N*-Boc-*N,N'*-dimethylethylenediamine to yield compound **17**. The latter served as a common intermediate for the synthesis of all the conjugates, either containing the glutarate or the PEG-4 spacer. In the first case, after Fmoc deprotection of **17** under basic conditions, the free amine was treated with glutaric anhydride to afford compound **18**, that was activated as a *N*-hydroxysuccinimide ester and coupled either to functionalized *cyclo*[DKP-RGD] (**3**) or to *cyclo*[RGDfK] (**1**) at controlled pH to afford intermediates **19a** and **19b**. The latter compounds were later Boc-deprotected with trifluoroacetic acid (TFA) and coupled to 2'-(4-nitrophenoxycarbonyl)paclitaxel,^[11b] giving the final conjugates **4** and **5**.

For the synthesis of the conjugates containing the PEG-4 spacer, compound **17** was Fmoc-deprotected and coupled to 4-pentynoic acid in the presence of HATU, HOAt and *i*Pr₂NEt to give alkyne **20**, which was later Boc-deprotected with TFA and coupled with 2'-(4-nitrophenoxycarbonyl)paclitaxel affording compound **21**. Finally, this compound was subjected to copper-catalyzed azide-alkyne cycloaddition ("click" reaction) with either *cyclo*[DKP-RGD]-PEG-4-azide (**22a**)^[18] or *cyclo*[RGDfK]-PEG-4-azide (**22b**), prepared in two steps from ligands **3** and **1**, respectively (see the Supporting Information). The click reaction gave the final conjugates **6** and **7**.

Integrin receptor competitive binding assay

Conjugates **4–9** were evaluated for their ability to inhibit biotinylated vitronectin binding to the isolated $\alpha_v\beta_3$ and $\alpha_v\beta_5$ receptors. The calculated half-maximal inhibitory concentrations (IC₅₀) are listed in Table 2. Screening assays were performed by incubating the immobilized integrin receptor with solutions of the *cyclo*[DKP-RGD]-paclitaxel and *cyclo*[RGDfK]-paclitaxel conjugates at different concentrations (10⁻¹²–10⁻⁵ M) in the presence of biotinylated vitronectin (1 µg mL⁻¹) and measuring bound vitronectin. Table 2 shows that conjugates **4–9** retain good binding affinity and high selectivity for $\alpha_v\beta_3$ integrin, with IC₅₀ values in the nanomolar range, only slightly worse compared to those obtained for the free ligands (see Table 1, compounds **1** and **2**). These results prompted us to proceed with cell viability assays.



Scheme 2. Synthesis of conjugates **4-7**. Reagents and conditions: a) HOBT, DIC, 4-aminobenzyl alcohol, DMF, overnight; b) 4-nitrophenylchloroformate, pyridine, 4:1 THF/DMF, 2 h; c) *N*-Boc-*N,N'*-dimethylethylenediamine, *i*Pr₂NEt, THF, overnight; d) i. piperidine, DMF, 4 h; ii. glutaric anhydride, DMAP, *i*Pr₂NEt, DMF, overnight; e) i. DIC, NHS, DMF; ii. **3** (for **19a**) or **1** (for **19b**), 1:1 DMF/PBS (phosphate-buffered saline, pH 7.3-7.6), overnight; f) i. TFA, CH₂Cl₂; ii. 2'-(4-nitrophenoxycarbonyl)PTX, *i*Pr₂NEt, DMF, overnight; g) i. piperidine, DMF, 4 h; ii. 4-pentynoic acid, HATU, HOAt, *i*Pr₂NEt, DMF, overnight; h) **22a** or **22b**, CuSO₄·5 H₂O, sodium ascorbate, DMF/H₂O. DIC = *N,N*-diisopropylcarbodiimide; DMAP = 4-dimethylaminopyridine; NHS = *N*-hydroxysuccinimide; PTX = paclitaxel.

Table 2. Inhibition of biotinylated vitronectin binding to the isolated $\alpha_v\beta_3$ and $\alpha_v\beta_5$ receptors.

Compound	Structure	IC ₅₀ (nM) ^[a] $\alpha_v\beta_3$	IC ₅₀ (nM) ^[a] $\alpha_v\beta_5$
4	cyclo[DKP-RGD]-GFLG-PTX	54.8 ± 14.0	> 1000 ^[b]
5	cyclo[RGDFK]-GFLG-PTX	62.6 ± 10.9	649 ± 136
6	cyclo[DKP-RGD]-PEG-4-GFLG-PTX	42.4 ± 7.4	> 1000 ^[b]
7	cyclo[RGDFK]-PEG-4-GFLG-PTX	12.1 ± 2.0	473 ± 25
8	cyclo[DKP-RGD]-VA-PTX	13.3 ± 3.6 ^[c]	924 ± 290 ^[c]
9	cyclo[DKP-RGD]-PEG-4-VA-PTX	14.8 ± 3.9 ^[d]	> 1000 ^[b]

[a] IC₅₀ values were calculated as the concentration of compound required for 50% inhibition of biotinylated vitronectin binding as estimated by GraphPad Prism software. All values are the arithmetic mean ± the standard deviation (SD) of duplicate determinations. [b] Biotinylated vitronectin binding was not completely inhibited in the concentration range tested. [c] See ref. [11b]. [d] See ref. [18].

Cell viability assays

The antiproliferative activity of conjugates **4-9** was evaluated in two cell lines expressing different levels of $\alpha_v\beta_3$ integrin. U87 cells (human glioblastoma) were selected as $\alpha_v\beta_3^+$, while HT29 cells (human colorectal adenocarcinoma) were used as $\alpha_v\beta_3^-$. The expression of $\alpha_v\beta_3$ integrin on the cell membrane was assessed by flow cytometry (see the Supporting Information), and the results were in good agreement with the literature.^[19] Both cell lines were treated for 96 hours^[20] with different concentrations of free PTX and conjugates **4-9** and the cell viability was evaluated with the MTT assay.

The calculated IC₅₀ values are shown in Table 3. All the conjugates proved less potent than free PTX in both cell lines.^[21] Interestingly, the conjugates containing the PEG-4 spacer (**6**, **7**

and **9**) proved 2.4-6.2 times more potent in the $\alpha_v\beta_3^+$ U87 cell line than the conjugates bearing the glutarate spacer (**4**, **5** and **8**). This might be due to improvement of the water solubility and flexibility provided by the tetraethylene glycol spacer, which is known to facilitate binding to the receptor.^[22] For all conjugates, the Relative Potencies (RP = IC₅₀ PTX / IC₅₀ Conjugate) in the $\alpha_v\beta_3^+$ U87 cell line are 1-2 orders of magnitude higher than in the $\alpha_v\beta_3^-$ HT29 cell line. This means that the loss of potency of the conjugates with respect to PTX is higher when there is no $\alpha_v\beta_3$ receptor in the cell line. Accordingly, we observed good Targeting Indexes (TI = RP in the $\alpha_v\beta_3^+$ U87 / RP in the $\alpha_v\beta_3^-$ HT29) throughout the series (TIs = 30-45, for compounds **4-8**), with compound **9** displaying the best value (TI = 533).

Table 3. Evaluation of anti-proliferative activity of free PTX and PTX conjugates **4-9** in U87 and HT29 cell lines.

Comp.	IC ₅₀ (nM) ^[a]				
	U87 ($\alpha_v\beta_3^+$)	HT29 ($\alpha_v\beta_3^-$)	RP _{U87} ^[b]	RP _{HT29} ^[c]	TI ^[d]
PTX	32.66 ± 21.81	1.82 ± 1.85	1	1	1
4	2031 ± 454	3413 ± 983	0.01608	0.00053	30
5	1250 ± 293.6	2692 ± 676	0.02613	0.000692	38
6	854.7 ± 165.1	1979 ± 252	0.03821	0.0009196	42
7	506.2 ± 113.6	1272 ± 156	0.06452	0.001431	45
8	2686 ± 589	6452 ± 1723	0.01216	0.0002821	43
9	432.6 ± 129.3	12840 ± 2730	0.07550	0.0001417	533

[a] IC₅₀ values were calculated as the concentration of compound required for 50% inhibition of cell viability. Both cell lines were treated with different concentrations of PTX and compounds **4-9** during 96 hours. The samples were measured in triplicate; [b] Relative Potency in U87 cell line (RP_{U87}): IC₅₀ PTX in U87 / IC₅₀ Conjugate in U87; [c] Relative Potency in HT29 cell line (RP_{HT29}): IC₅₀ PTX in HT29 / IC₅₀ Conjugate in HT29; [d] Targeting Index (TI): RP_{U87}/RP_{HT29}.

To determine whether the observed targeting is mediated by an integrin binding and internalization process, a competition experiment was carried out in the $\alpha_v\beta_3$ + U87 cell line. Conjugate **9** was tested in the presence of 50-fold excess free ligand *cyclo*[DKP-RGD] (**2**), with the aim of blocking integrins on the cell surface (see Table 4).

Table 4. Competition experiments of conjugate **9** in the presence of 50-fold excess *cyclo*[DKP-RGD] (**2**) in U87 cell line.

#	Compound(s)	IC ₅₀ (nM) ^[a] U87 ($\alpha_v\beta_3$ +)	Inhibition Ratio ^[b]	Corrected Inhibition Ratio ^[c]
1	<i>cyclo</i> [DKP-RGD] (2)	$342.8 \cdot 10^3 \pm 94.7 \cdot 10^3$	-	-
2	PTX	32.66 ± 21.81	-	-
3	PTX + 50-fold excess <i>cyclo</i> [DKP-RGD] (2)	10.66 ± 4.8	0.33	1
4	9	432.6 ± 129.3	-	-
5	9 + 50-fold excess <i>cyclo</i> [DKP-RGD] (2)	717.5 ± 216.3	1.66	5.03

[a] IC₅₀ values were calculated as the concentration of compound required for 50% inhibition of cell viability. Both cell lines were treated with different concentrations compounds **9** in the presence of 50-fold excess *cyclo*[DKP-RGD] (**2**) during 96 hours. The samples were measured in triplicates; [b] Inhibition Ratio = (IC₅₀ Compound + 50-fold excess **2**) / IC₅₀ Compound; [c] Corrected inhibition Ratio = [(IC₅₀ Compound + 50-fold excess **2**) / IC₅₀ Compound] / [(IC₅₀ PTX + 50-fold excess **2**) / IC₅₀ PTX].

As expected, the free ligand *cyclo*[DKP-RGD] (**2**) proved scarcely toxic (Table 4, entry 1: IC₅₀ = $343 \cdot 10^3$ nM = 343 μ M), but induced a distinct change in cell morphology and cell detachment. In the presence of excess *cyclo*[DKP-RGD] (**2**), PTX showed a threefold increased toxicity (Table 4, cf. entry 3 and entry 2). On the contrary, with conjugate **9** the presence of excess free ligand **2** induced a moderate increase of IC₅₀ (Table 4, cf. entry 5 and entry 4). Taking into account the effect produced by excess ligand **2** onto free PTX, a fivefold decrease of the conjugate cytotoxicity is calculated (Corrected Inhibition Ratio, see Table 4, entry 5), which might suggest that internalization is mediated by the $\alpha_v\beta_3$ integrin receptor.

Conclusions

The research reported in this article had the goal to investigate how the biological properties of integrin $\alpha_v\beta_3$ -targeting SMDs may vary depending on the type of ligand, spacer and linker employed, with a specific focus on the intracellularly cleavable peptide linkers. To this end, two *cyclo*[DKP-RGD]-GFLG-PTX (**4** and **6**) and two *cyclo*[RGDFK]-GFLG-PTX conjugates (**5** and **7**) were synthesized, the members of each pair differing for the spacer between ligand and peptide linker (glutarate in **4** and **5**, triazole-PEG-4 in **6** and **7**). Together with the two *cyclo*[DKP-RGD]-VA-PTX conjugates **8** and **9**, compounds **4-7** were used in (i) binding tests on the integrin $\alpha_v\beta_3$ isolated receptor, and (ii) viability tests using U87 ($\alpha_v\beta_3$ +) and HT29 cancer cells ($\alpha_v\beta_3$ -). As a general trend, all conjugates showed low-nanomolar affinity for the integrin $\alpha_v\beta_3$ receptor and reduced potency compared to

free PTX, with lower IC₅₀ values being obtained using the $\alpha_v\beta_3$ + cells compared to the $\alpha_v\beta_3$ - cells (TI in the 30-45 range for **4-8**, TI = 533 for **9**). While replacement of the *cyclo*[DKP-RGD] ligand moiety with *cyclo*[RGDFK] barely affected the SMDs' properties, the conjugates featuring a triazole-PEG-4 spacer proved 2.4-6.2 times more potent against the $\alpha_v\beta_3$ + cell line than those bearing the glutarate spacer. Replacing the peptide linker impacted on the conjugates' anti-proliferative effect in a non-easy-to-rationalize manner. The decrease of potency observed when the best compound (**9**) was incubated with U87 cells in the presence of a large excess of free ligand *cyclo*[DKP-RGD] (**2**) suggests that the conjugate is possibly internalized by an integrin $\alpha_v\beta_3$ -mediated process.

From these results it emerges that integrin $\alpha_v\beta_3$ -targeted SMDs can achieve promising TI values, although substantial improvements in terms of potency are needed in order for them to become therapeutically useful. To this end, modification of the spacer(s) and/or of the type of linker are being currently taken in consideration by our research group.

Experimental Section

All manipulations requiring anhydrous conditions were carried out in flame-dried glassware, with magnetic stirring and under a nitrogen atmosphere. All commercially available reagents were used as received. Anhydrous solvents were purchased from commercial sources and withdrawn from the container by syringe, under a slight positive pressure of nitrogen. The reactions were monitored by analytical thin-layer chromatography (TLC) using silica gel 60 F254 pre-coated glass plates (0.25 mm thickness). Visualization was accomplished by irradiation with a UV lamp and/or staining with Cerium/Molibdate reagent or ninhydrin. HPLC purifications and HPLC traces of final products were performed on Dionex Ultimate 3000 equipped with Dionex RS Variable Wavelength Detector (column: Atlantis Prep T3 OBDTM 5 μ m 19 \times 100 mm; flow 15 ml/min unless stated otherwise). The analysis of the integrals and the relative percentage of purity was performed with the software Chromeleon 6.80 SR11 Build 3161. Low resolution mass spectra (MS) were recorded on Thermo Finnigan LCQ Advantage (ESI source), Micro Waters Q-ToF (ESI source) and Bruker Daltonics Microflex LT (MALDI source) instruments. High-resolution mass spectra (HRMS) were performed with a Fourier Transform Ion Cyclotron Resonance (FT-ICR) Mass Spectrometer APEX II & Xmass software (Bruker Daltonics) – 4.7 T Magnet (Magnetex) equipped with ESI source, available at CIGA (Centro Interdipartimentale Grandi Apparecchiature) c/o Università degli Studi di Milano. Freeze-drying: the products were dissolved in water and frozen with dry ice. The freeze-drying was carried out at least for 48 h at -50 °C using the instrument 5Pascal Lio5P DGT.

Ligand *cyclo*[DKP-RGD] (**2**),^[7e] its functionalized analog (**3**)^[10] and *N*-Boc-*N*,*N*'-dimethylethylenediamine^[23] were prepared according to the literature. Also, conjugates **8**,^[11b] **9**^[18] and the activated drug 2'-(4-nitrophenoxy)carbonyl)paclitaxel^[11b] were prepared as previously described. The synthesis and characterization of ligand *cyclo*[RGDFK] (**1**), the Fmoc-protected linker **14**, intermediates **15-18**, intermediate **20** and azide-ligands **22a** and **22b** are described in the Supporting Information.

Cyclo[DKP-RGD]-Gly-Phe-Leu-Gly-N-[4-[[[(*N*-Boc)-*N*,*N*'-dimethylethylenediamine]carbonyl]oxy]methyl]phenyl] (19a). Compound **18** (11 mg, 0.0119 mmol, 2 equiv.) was dissolved in DMF (500 μ L) and cooled at 0 °C under nitrogen. *N*-Hydroxysuccinimide (NHS, 1.8 mg, 0.0155 mmol, 2.6 equiv.) and EDC-HCl (3.4 mg, 0.0179 mmol, 3 equiv.) were added and the reaction was stirred at 0 °C for a 5 min. The reaction was allowed to reach r.t. and stirred overnight under nitrogen atmosphere. Volatiles were then removed in vacuo and the crude was re-dissolved in DMF (375 μ L). A solution of compound **3** (5.2 mg, 0.006

mmol, 1 equiv.) in phosphate buffer (375 μ L, pH 7.5) was then added. During the first hours, the pH was kept in the 7.3-7.6 range by adding small aliquots of 0.2 M NaOH. The resulting solution was stirred overnight and then concentrated under vacuum. The crude residue was purified by semipreparative HPLC [Gradient: 100% (H_2O + 0.1% CF_3COOH) / 0% (CH_3CN + 0.1% CF_3COOH) to 25% (H_2O + 0.1% CF_3COOH) / 75% (CH_3CN + 0.1% CF_3COOH) in 15 min; t_R (product): 10.5 min]. Yield: 1.41 mg (15%) over 2 steps. HRMS (ESI+): m/z calcd. for $[\text{C}_{68}\text{H}_{96}\text{N}_{17}\text{O}_{18}]^+$: 1438.712 $[\text{M}+\text{H}]^+$; found: 1438.710; m/z calcd. for $[\text{C}_{68}\text{H}_{96}\text{N}_{17}\text{NaO}_{18}]^+$: 730.856 $[\text{M}+\text{Na}]^+$; found: 730.851; m/z calcd. for $[\text{C}_{68}\text{H}_{95}\text{N}_{17}\text{Na}_2\text{O}_{18}]^{2+}$: 741.840 $[\text{M}+2\text{Na}]^{2+}$; found: 741.842.

Cyclo[RGDFK]-Gly-Phe-Leu-Gly-N-[4-(((N-(Boc)-N,N'-dimethylethylenediamine)carbonyl)oxy)methyl]phenyl] (19b).

Compound **18** (13 mg, 0.016 mmol, 1.3 equiv.) was dissolved in DMF (350 μ L) and cooled at 0 $^\circ\text{C}$ under nitrogen. NHS (2.8 mg, 0.024 mmol, 2 equiv.) and DIC (3.7 μ L, 0.024 mmol, 2 equiv.) were added and the reaction was stirred at 0 $^\circ\text{C}$ for 5 min. The reaction was allowed to reach r.t. and stirred overnight under nitrogen atmosphere. Volatiles were then removed in vacuo and the crude was redissolved in DMF (750 μ L). A solution of cyclo[RGDFK] **1** (10 mg, 0.012 mmol, 1 equiv.) in phosphate buffer (750 μ L, pH 7.5) was then added. During the first hours, the pH was kept in the 7.3-7.6 range by adding small aliquots of 0.2 M NaOH. The resulting solution was stirred overnight and then concentrated under vacuum. The crude residue was purified by semipreparative HPLC [Gradient: 100% (H_2O + 0.1% CF_3COOH) / 0% (CH_3CN + 0.1% CF_3COOH) to 25% (H_2O + 0.1% CF_3COOH) / 75% (CH_3CN + 0.1% CF_3COOH) in 15 min; t_R (product): 11 min]. Yield: 3.15 mg (17%) over two steps. MS (MALDI): m/z calcd. for $[\text{C}_{68}\text{H}_{99}\text{N}_{16}\text{O}_{17}]^+$: 1412.61 $[\text{M}+\text{H}]^+$; found: 1412.2 (HCCA matrix); HRMS (ESI+): m/z calcd. for $[\text{C}_{68}\text{H}_{98}\text{N}_{16}\text{NaO}_{17}]^+$: 1433.719 $[\text{M}+\text{Na}]^+$; found: 1433.717; m/z calcd. for $[\text{C}_{68}\text{H}_{99}\text{N}_{16}\text{Na}_2\text{O}_{17}]^{2+}$: 728.360 $[\text{M}+2\text{Na}]^{2+}$; found: 728.355.

General Procedure for the Boc-deprotection of compounds 19a, 19b and 20.

A half volumen of TFA was added to a 0.03 M solution of intermediates **19a**, **19b** or **20** in CH_2Cl_2 and the reaction was stirred at room temperature for one hour. The solvent was evaporated and then the CH_2Cl_2 was added twice to the residue followed by evaporation under vacuum each each time to afford the amine TFA salt. The crudes were frozen-dried and used without further purification.

Alkyne-Gly-Phe-Leu-Gly-PTX (21). The TFA salt obtained from deprotection of compound **20** (7.5 mg, 0.0108 mmol, 1 equiv.) was dissolved in DMF (320 μ L) and cooled at 0 $^\circ\text{C}$ under nitrogen atmosphere. $i\text{Pr}_2\text{NEt}$ (7.5 μ L, 0.043 mmol, 4 equiv.) and 2'-(4-nitrophenoxy)carbonyl)paclitaxel (16.5 mg, 0.0162 mmol, 1.5 equiv.) were added in this order and the mixture was allowed to reach r.t. and stirred overnight. The reaction mixture was diluted with AcOEt and washed with a 1 M aqueous solution of KHSO_4 (2 x) and brine (1 x). The organic phase was dried over Na_2SO_4 , concentrated and purified by flash chromatography (eluent: 9:1 $\text{CH}_2\text{Cl}_2/\text{CH}_3\text{OH}$) to afford pure **21**. Yield: 10.18 mg (60%). HRMS (ESI+): m/z calcd. for $[\text{C}_{84}\text{H}_{98}\text{N}_8\text{O}_{22}\text{Na}]^+$: 1593.669 $[\text{M}+\text{Na}]^+$; found: 1593.667; m/z calcd. for $[\text{C}_{84}\text{H}_{98}\text{N}_8\text{O}_{22}\text{Na}_2]^{2+}$: 808.329 $[\text{M}+2\text{Na}]^{2+}$; found: 808.328.

Cyclo[DKP-RGD]-GFLG-PTX (4). The TFA salt obtained from deprotection of compound **19a** (6.4 mg, 0.0041 mmol, 1 equiv.) was dissolved in DMF (470 μ L) and cooled at 0 $^\circ\text{C}$ under nitrogen atmosphere. $i\text{Pr}_2\text{NEt}$ (3.6 μ L, 0.0205 mmol, 5 equiv.) and 2'-(4-nitrophenoxy)carbonyl)paclitaxel (12.5 mg, 0.0123 mmol, 3.5 equiv.) were added in this order and the mixture was allowed to reach r.t. and stirred overnight. The crude was concentrated, and the residue was purified by semipreparative HPLC [Gradient: 100% (H_2O + 0.1% CF_3COOH) / 0% (CH_3CN + 0.1% CF_3COOH) to 0% (H_2O + 0.1% CF_3COOH) / 100% (CH_3CN + 0.1% CF_3COOH) in 20 min; t_R (product): 12.5 min]. Yield: 2.09 mg (26%). MS (MALDI): m/z calcd. for $[\text{C}_{111}\text{H}_{137}\text{N}_{18}\text{O}_{31}]^+$: 2219.38 $[\text{M}+\text{H}]^+$; found: 2220.6 (HCCA matrix), 2219.1 (SA matrix); HRMS (ESI+):

m/z calcd. for $[\text{C}_{111}\text{H}_{137}\text{N}_{18}\text{O}_{31}\text{Na}]^{2+}$: 1120.480 $[\text{M}+1\text{H}+\text{Na}]^{2+}$; found: 1120.479; m/z calcd. for $[\text{C}_{111}\text{H}_{137}\text{N}_{18}\text{O}_{31}\text{Na}_2]^{3+}$: 754.650 $[\text{M}+1\text{H}+2\text{Na}]^{3+}$; found: 754.648.

Cyclo[RGDFK]-GFLG-PTX (5). The TFA salt obtained from deprotection of compound **19b** (2.5 mg, 0.00163 mmol, 1 equiv.) was dissolved in DMF (190 μ L) and cooled at 0 $^\circ\text{C}$ under nitrogen atmosphere. $i\text{Pr}_2\text{NEt}$ (2 μ L, 0.0115 mmol, 7 equiv.) and 2'-(4-nitrophenoxy)carbonyl)paclitaxel (5.8 mg, 0.0057 mmol, 3.5 equiv.) were added in this order and the mixture was allowed to reach r.t. and stirred overnight. The crude was concentrated, and the residue was purified by semipreparative HPLC [Gradient: 100% (H_2O + 0.1% CF_3COOH) / 0% (CH_3CN + 0.1% CF_3COOH) to 0% (H_2O + 0.1% CF_3COOH) / 100% (CH_3CN + 0.1% CF_3COOH) in 20 min; t_R (product): 13 min]. Yield: 1.22 mg (33%). MS (MALDI): m/z calcd. for $[\text{C}_{111}\text{H}_{140}\text{N}_{17}\text{O}_{30}]^+$: 2192.39 $[\text{M}+\text{H}]^+$; found: 2193.4 (HCCA matrix); 2194.1 (SA matrix); HRMS (ESI+): m/z calcd. for $[\text{C}_{111}\text{H}_{140}\text{N}_{17}\text{O}_{30}\text{Na}]^{2+}$: 1106.993 $[\text{M}+1\text{H}+\text{Na}]^{2+}$; found: 1106.991.

Cyclo[DKP-RGD]-PEG-4-GFLG-PTX (6). Compound **21** (3.9 mg, 0.0025 mmol, 1.5 equiv.) and **22b** (1.7 mg, 0.0017 mmol, 1 equiv.) were dissolved in a degassed 1:1 water/DMF mixture (170 μ L). Degassed aqueous solutions of $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ (0.2 mg, 0.000835 mmol, 0.5 equiv.) and sodium ascorbate (0.198 mg, 0.001 mmol, 0.6 equiv.) were added at room temperature and the mixture was stirred overnight at 30 $^\circ\text{C}$. The solution was concentrated, and the crude residue was purified by semipreparative HPLC [Gradient: 100% (H_2O + 0.1% CF_3COOH) / 0% (CH_3CN + 0.1% CF_3COOH) to 0% (H_2O + 0.1% CF_3COOH) / 100% (CH_3CN + 0.1% CF_3COOH) in 20 min; t_R (product): 12.5 min]. Yield: 1.67 mg (39%) over two steps. MS (ESI+): m/z calcd. for $[\text{C}_{121}\text{H}_{153}\text{N}_{21}\text{O}_{35}\text{Na}_2]^{2+}$: 1253.81 $[\text{M}+\text{H}]^+$, found 1254.06; MS (ESI-): m/z calcd. for $[\text{C}_{121}\text{H}_{151}\text{N}_{21}\text{O}_{35}]^{2-}$: 1229.8 $[\text{M}-2\text{H}]^{2-}$, found 1230.0.

Cyclo[RGDFK]-PEG-4-GFLG-PTX (7). Compound **21** (5 mg, 0.0032 mmol, 1.5 equiv.) and **22a** (1.84 mg, 0.0022 mmol, 1 equiv.) were dissolved in a degassed 1:1 water/DMF mixture (220 μ L). Degassed aqueous solutions of $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ (0.28 mg, 0.0011 mmol, 0.5 equiv.) and sodium ascorbate (0.26 mg, 0.00132 mmol, 0.6 equiv.) were added at room temperature and the mixture was stirred overnight at 30 $^\circ\text{C}$. The solution was concentrated, and the crude residue was purified by semipreparative HPLC [Gradient: 100% (H_2O + 0.1% CF_3COOH) / 0% (CH_3CN + 0.1% CF_3COOH) to 0% (H_2O + 0.1% CF_3COOH) / 100% (CH_3CN + 0.1% CF_3COOH) in 20 min; t_R (product): 13 min]. Yield: 4.13 mg (76 %) over two steps. MS (MALDI): m/z calcd. for $[\text{C}_{121}\text{H}_{157}\text{N}_{20}\text{O}_{34}]^+$: 2434.65 $[\text{M}+\text{H}]^+$, found: 2436.0 (HCCA matrix); 2435.7 (SA matrix).

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