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## Synthesis and antibacterial activity of novel and potent DNA gyrase inhibitors with azole ring

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Abstract—The 4-piperidyl moiety and the pyrazole ring in 1-(3-chlorophenyl)-5-(4-phenoxyphenyl)-3-(4-piperidyl)pyrazole 2, which has previously shown improved DNA gyrase inhibition and target-related antibacterial activity, were transformed to other groups and the in vitro antibacterial activity of the synthesized compounds was evaluated. The selected pyrazole, oxazole and imidazole derivatives showed moderate inhibition against DNA gyrase and topoisomerase IV with similar  $IC_{50}$  values ( $IC_{50} = 9.4-25 \mu g/$ mL). In addition, many of the pyrazole, oxazole and imidazole derivatives synthesized in this study exhibited potent antibacterial activity against quinolone-resistant clinical isolates and coumarin-resistant laboratory isolates of Gram-positive bacteria with minimal inhibitory concentration values equivalent to those against susceptible strains. © 2004 Elsevier Ltd. All rights reserved.

## 1. Introduction

The emergence and spread of multidrug resistant Grampositive bacteria, such as methicillin-resistant Staphylococcus aureus (MRSA), penicillin-resistant Streptococcus pneumoniae (PRSP) and vancomycin-resistant enterococci (VRE), have made treatment of infectious diseases difficult and have over the last decades become a serious medical problem. As pathogenic bacteria continuously evolve mechanisms of resistance to currently used antibacterial agents, the discovery of novel and potent bactericides is the best way to overcome bacterial resistance and develop effective therapies. Unfortunately, finding novel antibacterial agents has been difficult with the oxazolidinones, identified in 1980, being the last example to successfully reach the clinic.<sup>1</sup>

Bacterial DNA gyrase is a proven target for antibacterial chemotherapy.<sup>2</sup> Quinolones, such as sparfloxacin<sup>3</sup> (SPFX, Fig. 1), inhibit bacterial DNA gyrase and topoisomerase IV and cause bacterial cell death. Besides the quinolones, other naturally occurring bacterial DNA gyrase inhibitors, such as the coumarins, which include novobiocin (NB, Fig. 1), clorobiocin and coumermycin



Figure 1.

Keywords: DNA gyrase inhibitor; Pyrazole, oxazole and imidazole derivatives; Multidrug resistant strains.

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 $A_1$  have also been known as antibacterial agents.<sup>4</sup> The coumarins inhibit ATPase activity of DNA gyrase by competing with ATP for binding to the B subunit of the enzyme. However, no pharmaceutically useful drug has so far been derived from the coumarins. Although many efforts have been dedicated to finding potent antibacterial agents that can overcome bacterial resistance, promising lead structures of DNA gyrase inhibitors have not been found.<sup>5–10</sup>

Using a new screening system for specific inhibitors of chromosome partitioning in *Escherichia* (*E.*)  $coli^{11,12}$  we have previously reported that one of our hit compounds (compound **1**, Fig. 1) having a piperidine ring represents a new class of bacterial DNA gyrase inhibitors that have potent antibacterial activity against *Staphylococcus* (*S.*) *aureus* and *Enterococcus* (*E.*) *faecalis.*<sup>13</sup> We have also demonstrated that compound **2** (Fig. 1), a derivative of **1**, shows improved DNA gyrase inhibitory concentration (MIC) values against clinically isolated multidrug resistant Gram-positive bacteria as against drug susceptible strains.

Our aim in this study was to optimize the piperidine moiety and pyrazole ring of compound 2 and find new potent DNA gyrase inhibitors with antibacterial activity against MRSA, PRSP and VRE. Here we report the synthesis and structure-activity relationships (SARs) of a series of pyrazole and other five-membered heterocyclic derivatives.

## 2. Chemistry

The novel pyrazole derivatives 8–16 were synthesized as shown in Scheme 1. The requisite intermediate 1,3-diketones 4a–f were prepared by coupling reaction of the *N*-(*tert*-butoxycarbonyl) amino acids 3 with various benzophenone derivatives in THF using 1,1'-carbonyldi-1*H*-imidazole (CDI) and lithium hexamethyldisilazide (LHMDS).<sup>14,15</sup> Condensation of 4a–f with 3-chlorophenylhydrazine hydrochloride with aqueous NaOH gave a mixture of the 1-(3-chlorophenyl)pyrazoles 5 and 6. Cleavage of the protecting group under acidic conditions, followed by separation using CHP-20P (reverse phase) column chromatography and/or recrystallization gave the 3-substituted 1-(3-chlorophenyl)pyrazoles 8, 10–12, 14, 15 and the regioisomers 9, 13, 16 as acid salts. Compound 7 having a cyclohexyl group at the 3-position instead of the 4-piperidyl group of 2 was synthesized from cyclohexanecarboxylic acid and 4-phenoxyacetophenone in a similar manner to that described above. The corresponding 5-substituted isomers of 1-(3-chlorophenyl)pyrazole derivatives 7, 10, 11 and 14 were not isolated.

The position of 3-chlorophenyl group in 7–16 was determined on the basis of nuclear Overhauser effects (NOE) experiments. For example, in compound 8, a correlation of NOE was observed between the protons of the phenyl moiety on R<sup>1</sup>-Ph and the *ortho* protons of the 3-chlorophenyl moiety. On the other hand, in compound 9, a correlation of NOE was observed between the 2-H protons of the 3-piperidyl moiety on R<sup>2</sup> and the *ortho* protons of the 3-chlorophenyl moiety.

The novel oxazole derivatives 21 and 22 and the imidazole derivatives 24 and 25 were synthesized as shown in Scheme 2. The 2-trimethylsilyloxyacetonitrile 18a was prepared by condensation of 4-phenoxybenzalde-hyde ( $R^3 = 4$ -phenoxy) with trimethylsilylcyanide and zinc iodide in CH<sub>2</sub>Cl<sub>2</sub>.<sup>16</sup> Coupling reaction of **18a** with 3-chlorobenzaldehyde ( $\mathbb{R}^4 = 3$ -chlorophenyl) using LHMDS in THF, followed by treatment with aqueous H<sub>2</sub>SO<sub>4</sub> afforded the 2-hydroxyethane-1-one 19a. Reaction of 19a with 1-(tert-butoxycarbonyl)piperidine-4carbxylic acid in the presence of 1-ethyl-3-[3-(dimethylamino)propyl]carbodiimide hydrochloride (EDC) and 4-dimethylaminopyridine (DMAP) gave the ester 20a, which was treated with NH4OAc in AcOH to afford the desired **21**.<sup>17</sup> In a similar manner to that described above, 22 was obtained from 3-chlorobenzaldehyde  $(R^3 = 3$ -chloro) as starting material via 18b–20b  $(\mathbf{R}^4 = 4$ -benzyloxy).

The imidazole analogues 24 and 25 were obtained by condensation of 23a,b, which were prepared by oxidation of 19a,b using CuSO<sub>4</sub> in aqueous pyridine, with 1-(*tert*-butoxycarbonyl)-4-formylpiperidine in the presence of  $NH_4OAc$  in AcOH.<sup>18</sup>



Scheme 1. General synthesis of the 1-(3-chlorophenyl)pyrazole derivatives. Reagents: (a) i. CDI, THF, ii. R<sup>1</sup>-PhCOCH<sub>3</sub>, LHMDS, THF; (b) 3-chlorophenylhydrazine HCl, aq NaOH, EtOH; (c) HCl, EtOH or TFA, CH<sub>2</sub>Cl<sub>2</sub>.



Scheme 2. Synthesis of the oxazole and imidazole derivatives. Reagents: (a) TMSCN, ZnI, CH<sub>2</sub>Cl<sub>2</sub>; (b) i. R<sup>4</sup>-CHO, LHMDS, THF, ii. aq H<sub>2</sub>SO<sub>4</sub>, MeOH; (c) 1-(*tert*-butoxycarbonyl)piperidine-4-carboxylic acid, EDC, DMAP, CH<sub>2</sub>Cl<sub>2</sub>; (d) NH<sub>4</sub>OAc, AcOH; (e) CuSO<sub>4</sub>, pyridine, H<sub>2</sub>O; (f) 1-(*tert*-butoxycarbonyl)-4-formylpiperidine, NH<sub>4</sub>OAc, AcOH.

## 3. Results and discussion

### **3.1.** Antibacterial activity

To investigate the effect of basic amine appended on the pyrazole ring on the antibacterial activity, the 4-piperidyl moiety in 2 was transformed to other groups (7-14) and the in vitro antibacterial activity of the compounds obtained was evaluated against two Gram-positive bacteria, that is a drug susceptible strain, *S. aureus* FDA 209P and a multidrug resistant strain, *S. aureus* KMP9 (MRSA), and against two Gram-negative bacteria that is a susceptible strain, *E. coli* NIHJ JC-2 and a multidrug efflux pump mutant, *E. coli* W3110  $\Delta acrA$ . For comparison, the in vitro antibacterial activity of sparfloxacin and novobiocin has also been evaluated. As shown in Table 1, the cyclohexyl derivative 7 having no basic amine did not exhibit any antibacterial activity. This result suggests that basic amines in the  $R^2$  moiety appended on the pyrazoles are essential for antibacterial activity. The 3-piperidyl analogue 8 showed 2-fold more potent antibacterial activity against the four strains of bacteria than the corresponding 4-piperidyl derivative 2. However, the 2-piperidyl analogue 10 showed fourto eightfold less potent antibacterial activity against *S. aureus* than the 4-piperidyl derivative 2 and the 3-piperidyl derivative 8. These results suggest that the position of NH group in the  $R^2$  moiety is important for antibacterial activity.

Compound 11 with an acyclic substituent, N-methylaminopropyl, exhibited twofold and fourfold more potent

Table 1. Antibacterial activity of the pyrazole derivatives



Compd	$\mathbb{R}^1$	H-R <sup>2</sup>	MIC (µg/mL)				
			S. aureus		E. coli		
			FDA 209P	KMP9 <sup>a</sup>	NIHJ JC-2	W3110 ∆acrA <sup>b</sup>	
2	4-PhO	4-Piperidyl	4	4	32	4	
7	4-PhO	c-Hexyl	>128	>128	>128	>128	
8	4-PhO	3-Piperidyl	2	2	16	2	
9	4-PhO	3-Piperidyl	4	4	32	16	
10	4-PhO	2-Piperidyl	16	16	>128	8	
11	4-PhO	CH <sub>3</sub> NHCH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub>	2	2	8	2	
12	4-PhO	CH <sub>3</sub> NHCH <sub>2</sub>	4	4	16	2	
13	4-PhO	CH <sub>3</sub> NHCH <sub>2</sub>	8	8	>128	4	
14	4-PhO	PhCH <sub>2</sub> NHCH <sub>2</sub>	64	128	>128	>128	
15	3-PhCH <sub>2</sub> O	4-Piperidyl	2	2	32	1	
16	3-PhCH <sub>2</sub> O	4-Piperidyl	4	4	32	2	
1	_		64	64	128	64	
SPFX			0.125	128	0.032	0.004	
NB	_	_	0.25	0.25	64	0.5	

<sup>a</sup> Multidrug resistant S. aureus.

<sup>b</sup>A multidrug efflux pump mutant.

antibacterial activity against *S. aureus* and *E. coli* NIHJ JC-2, respectively, than the lead compound **2**. Compound **12** having another acyclic substituent, *N*-methyl-aminomethyl, showed the same or twofold more potent antibacterial activity against the four strains of bacteria studied compared to compound **2**. However the *N*-benz-ylaminomethyl derivative **14** showed 16–32-fold less potent antibacterial activity against *S. aureus* than compound **2**. The SARs of these acyclic substituent suggest that contrary to cyclic amines, the position of NH group in the  $R^2$  moiety is not important for antibacterial activity. It is also noted that a small group, such as a methyl on NH retains strong potency whereas a bulky group, such as benzyl weaken the potency.

The 3-benzyloxyphenyl pyrazole **15** revealed twofold more potent antibacterial activity against *S. aureus* and the same potency against *E. coli* NIHJ JC-2 as compound **2**. When regioisomers of the synthesized compounds were compared, the 3-aminoalkyl-1-(3chlorophenyl)pyrazoles **8**, **12** and **15** showed slightly more potent antibacterial activity than the corresponding 5-aminoalkyl-1-(3-chlorophenyl)pyrazoles **9**, **13** and **16**. Among all pyrazole analogues synthesized in this study, compounds **8**, **11** and **15** had comparatively strong antibacterial activity against the four strains of bacteria studied.

Next, the effects of a transformation of the pyrazole ring of compound **2** on the antibacterial activity were examined (Table 2). The oxazole and imidazole derivatives configured with three substituents in the structure of the pyrazole derivatives **2** and **15** (i.e., 4-piperidyl, 3chlorophenyl and a substituted phenyl) were synthesized. The oxazole derivative **21** and **22** showed the same antibacterial activity against *S. aureus* and less potent antibacterial activity against *E. coli* NIHJ JC-2 compared to the corresponding pyrazole derivatives **2** and **15**.

On the other hand, the imidazole derivatives 24 and 25 showed similar antibacterial activity against *S. aureus* and four- to eightfold more potent antibacterial activity against *E. coli* NIHJ JC-2 compared to the corresponding pyrazole derivatives 2 and 15. From these results, it

is assumed that the imidazole ring induces potent antibacterial activity against not only Gram-positive bacteria but against Gram-negative bacteria. Since the imidazole derivatives showed nearly the same antibacterial activity against *E. coli* NIHJ JC-2 and W3110  $\Delta acrA$ , it is presumed that these derivatives are not affected by bacterial outer membrane pump.

The SARs of the pyrazole derivatives described in this study can be summarized as follows: a basic amine structure on the  $R^2$  moiety of the pyrazoles is essential for antibacterial activity. In addition, the position of the nitrogen atom in the cyclic amine and the size of the substituents on the nitrogen atom of the acyclic amine are important for potent antibacterial activity. Although no significant difference in the antibacterial activity against *S. aureus* among the three types of azole, the imidazole derivatives had more potent antibacterial activity against *E. coli* NIHJ JC-2 than the corresponding pyrazoles. On the whole, the order of antibacterial activity for these five-membered ring derivatives can be summarized as follows: imidazole > pyrazole > oxazole.

## 3.2. Inhibitory effects against DNA gyrase and topoisomerase IV

To elucidate the mechanism by which the pyrazole, oxazole and imidazole derivatives induce their antibacterial activity, the inhibitory activity of selected compounds (15, 22 and 25) against DNA gyrase and topoisomerase IV isolated from E. coli was examined.<sup>19,20</sup> As shown in Table 3, the IC<sub>50</sub> values of the initial lead compound 1 against DNA gyrase and topoisomerase IV were >128 and 128 µg/mL, respectively. Compounds 15, 22 and 25 having the same three substituents (i.e., 4-piperidyl, 3-chlorophenyl and 3-benzyloxyphenyl) showed moderate inhibition against the two enzymes (IC<sub>50</sub> =  $9.4-25 \,\mu\text{g/mL}$ ). The oxazole derivative 22 and the imidazole derivative 25 showed twofold more potent inhibitory activity against DNA gyrase than the pyrazole derivative 15. On the other hand, compounds 22 and 25 showed an inhibitory activity against topoisomerase IV almost similar to that of the pyrazole derivative 15.

Table 2. A	Antibacterial	activity	of the	e oxazole and	imidazole	derivative
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Compd	Х	R <sup>3</sup>	$\mathbb{R}^4$	MIC (µg/mL)			
				S. aureus		E. coli	
				FDA 209P	KMP9 <sup>a</sup>	NIHJ JC-2	W3110 ∆acrA <sup>b</sup>
21	0	4-PhO	3-Cl	4	4	128	4
22	Ο	3-C1	3-PhCH <sub>2</sub> O	2	2	64	4
24	NH	4-PhO	3-C1	2	4	8	2
25	NH	3-C1	3-PhCH <sub>2</sub> O	2	2	4	2

<sup>a</sup> Multidrug resistant S. aureus.

<sup>b</sup>A multidrug efflux pump mutant.

 Table 3. Inhibitory effects of selected compounds against DNA gyrase

 and topoisomerase IV

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Compd	$IC_{50} (\mu g/mL)^a$		
	Gyrase <sup>b</sup>	Topo IV <sup>c</sup>	
1	>128	128	
15	18.8	25	
22	9.4	25	
25	9.4	18.8	
SPFX	0.25	6.9	
NB	0.25	3.5	

<sup>a</sup> Isolated from *E. coli*.

<sup>b</sup> DNA gyrase supercoiling activity.

<sup>c</sup> Topoisomerase IV decatenation activity.

There was a good correlation between the MICs and the  $IC_{50}$ s (Tables 1 and 2), suggesting that inhibition of the DNA gyrase and topoisomerase IV by the pyrazole, oxazole and imidazole derivatives suppresses bacterial cell growth. While sparfloxacin ratio of inhibition of DNA gyrase and topoisomerase IV was more than 20, that of the pyrazole, oxazole and imidazole derivatives ranged between 1.3 and 2.7. These results suggest that the pyrazole, oxazole and imidazole derivatives would not be easily tolerated by bacteria.

## 3.3. Effect against multidrug resistant Gram-positive bacteria

To examine whether the pyrazole, oxazole and imidazole derivatives are effective against multidrug resistant Gram-positive bacteria, MIC values of selected compounds against quinolone-resistant clinical isolates and coumarin-resistant laboratory isolates of Gram-positive bacteria were determined and compared with those of sparfloxacin and novobiocin. The strains of bacteria used for MIC values determination were: S. aureus KMP9 (MRSA; a sparfloxacin-, clarithromycin- and ampicillin-resistant strain), S. aureus N175 (a novobiocin-resistant strain) derived from RN4220,<sup>21</sup> Streptococcus (S.) pneumoniae KT2524 (PRSP; a sparfloxacin-, clarithromycin- and ampicillin-resistant strain) and Enterococcus (E.) faecium KU1778 (VRE; a sparfloxacin-, clarithromycin-, ampicillin- and vancomycin-resistant strain).

As shown in Table 4, MIC values of sparfloxacin against the susceptible strains (MSSA, PSSP and VSE) were 0.125, 0.125 and 0.25µg/mL, respectively, and those against MRSA, PRSP and VRE were 128, 4 and 64µg/mL, respectively. MIC values of novobiocin against *S. aureus* RN4220 and *S. aureus* N175 derived from RN4220 were 0.125 and 4µg/mL, respectively. On the other hand, the pyrazole derivatives **8**, **11** and **15**, the oxazole derivative **22** and the imidazole derivative **25** revealed almost the same antibacterial activity against both susceptible and resistant Gram-positive bacteria. Although sparfloxacin and novobiocin ratio of MIC values against susceptible and resistant strains was more than 32, that of the pyrazole, oxazole and imidazole derivatives ranged between 1 and 2.

The pyrazole, oxazole and imidazole derivatives were also effective against quinolone- and coumarin-resistant Gram-positive bacteria with compounds 8, 11, 15, 22 and 25 showing a more potent antibacterial activity against clinically isolated quinolone-resistant Gram-positive bacteria than sparfloxacin, and compounds 22 and 25 revealing the same antibacterial activity against laboratory isolated coumarin-resistant Gram-positive as novobiocin. Compounds 8 and 15 demonstrated the most potent antibacterial activity against susceptible and resistant Gram-positive bacteria with across-theboard MIC values of  $1-4\mu g/mL$ .

It is therefore clear that the pyrazole, oxazole and imidazole derivatives have potent antibacterial activity against both susceptible and quinolone- and coumarinresistant Gram-positive bacteria. Additionally, the results above suggest that the mode of action of these analogues against DNA gyrase and topoisomerase IV is different from that of the quinolones or the coumarins.

## 4. Conclusions

In this study, we have described the synthesis and SARs of new pyrazole, oxazole and imidazole analogues. Among the pyrazole analogues, compounds **8** and **11** had comparatively strong antibacterial activity, whereas

Table 4. Antibacterial activity of selected pyrazole derivatives against susceptible and resistant Gram-positive bacteria

Organism <sup>a</sup>	MIC (µg/mL)							
	8	11	15	22	25	SPFX	NB	
S. aureus FDA 209P (MSSA) <sup>b</sup>	2	2	2	2	2	0.125	0.25	
S. aureus KMP9 (MRSA) <sup>c</sup>	2	2	2	2	2	128	0.25	
S. aureus RN4220 <sup>b</sup>				4	4	0.125	0.125	
S. aureus N175 <sup>d</sup>				4	4	0.125	4	
S. pneumoniae ATCC49619 (PSSP) <sup>b</sup>	1	4	2	4	4	0.125	0.5	
S. pneumoniae KT2524 (PRSP) <sup>e</sup>	2	4	2	4	4	4	0.5	
E. faecium ATCC19434 (VSE) <sup>b</sup>	4	8	4	16	4	0.25	2	
E. faecium KU1778 (VRE) <sup>f</sup>	2	4	4	8	2	64	2	

<sup>a</sup> SPFX: sparfloxacin, CAM: clarithromycin, ABPC: ampicillin, VCM: vancomycin.

<sup>b</sup> Susceptible strain.

<sup>c</sup> SPFX-, CAM-, ABPC-resistant strain.

<sup>d</sup> S. aureus N175 (GyrB; R144I) was derived from RN4220 with selection of novobiocin.

<sup>e</sup>SPFX-, CAM-, ABPC-resistant strain.

<sup>f</sup>SPFX-, CAM-, ABPC-, VCM-resistant strain.

among the oxazole and imidazole analogues, compounds 22 and 25 showed comparatively strong antibacterial activity. These selected compounds 8, 11, 22 and 25 possessed potent antibacterial activity against not only susceptible strains, but also against multidrug resistant strains. In addition, 8, 11, 22 and 25 showed a more potent antibacterial activity against clinically isolated quinolone-resistant Gram-positive bacteria than sparfloxacin. Compounds 22 and 25 revealed the same antibacterial activity against laboratory isolated coumarin-resistant Gram-positive bacteria compared to novobiocin.

Though the inhibitory activity of sparfloxacin against DNA gyrase was more than 20-fold that against topoisomerase IV, the pyrazole, oxazole and imidazole derivatives synthesized in this study showed almost the same inhibitory activity against both enzymes. These results suggest that the pyrazole, oxazole and imidazole derivatives would not be easily resisted by bacteria. We are pursuing further modifications of the pyrazole, oxazole and imidazole scaffold to obtain more potent inhibitors of both DNA gyrase and topoisomerase IV.

## 5. Experimental

## 5.1. Spectroscopic measurements

All melting points were determined on a Yanaco micromelting point apparatus and are uncorrected. <sup>1</sup>H NMR spectra were taken at 200 MHz on a Varian Gemini spectrometer, at 300 MHz on a JEOL spectrometer, or at 400 MHz on a Varian Mercury-400 spectrometer in DMSO- $d_6$  or CDCl<sub>3</sub>. All chemical shift values are reported as ppm ( $\delta$ ) values with tetramethylsilane as internal standard. The abbreviations used are as follows: s; singlet, d; doublet, t; triplet, q; quartet, m; multiplet, and br; broad. Mass spectra were obtained on a JEOL JMS-SX 102A QQ for fast atom bombardment mass spectra (FABMS) or a Hitachi M-1000 LC APCI mass spectrometer for atmospheric pressure chemical ionization mass spectra (APCIMS). For elements analysis, C, H and N were analyzed using a CE instruments EA1110 CHN elements analyzer, and F, Cl and Br were analyzed using a Yokogawa IC-7000 ion chromatograph. All analytical results were within  $\pm 0.4\%$  of the theoretical values obtained by a given formula.

## 5.2. Starting material

The following compounds **3** were commercially available from Aldrich or Watanabe Chemical Industries, Ltd, Japan: 1-(*tert*-butoxycarbonyl)piperidine-4-carboxylic acid, 1-(*tert*-butoxycarbonyl)piperidine-3-carboxylic acid, 1-(*tert*-butoxycarbonyl)-*N*-methylglycine and *N*-(*tert*-butoxycarbonyl)-*N*-methylglycine. The following compounds were synthesized according to published procedures: *N*-(*tert*-butoxycarbonyl)-*N*-methyl- $\gamma$ -aminobutylic acid<sup>22</sup> and 1-(*tert*-butoxycarbonyl)-4-formylpiperidine.<sup>23</sup>

#### 5.3. General procedure for 1,3-diketones 4a-g

To a solution of lithium hexamethyldisilazide (11 mmol) in THF (30mL) was added benzophenone derivatives (10mmol) in THF (10mL) at -78°C under argon atmosphere, and the resulting yellow suspension was stirred for 30min at the same temperature (solution A). A solution of 1,1'-carbonyldi-1H-imidazole (CDI, 16.2g, 10mmol) and an N-(tert-butoxycarbonyl) amino acid derivatives 3 [e.g., 1-(tert-butoxycarbonyl)piperidine-4-carboxylic acid] (10mmol) in THF (30mL) was stirred for 45 min at room temperature (solution B). The resulting solution B was added dropwise to solution A over  $30 \min at - 78 \degree C$ . After removal of the cold bath, the reaction mixture was warmed to room temperature and stirred for 12h. To this reaction mixture was added 10% aqueous citric acid (30 mL), and the resulting solution was taken up in AcOEt (100mL), washed with brine, dried over MgSO<sub>4</sub>, and concentrated to dryness. The residue was purified by silica gel column chromatography (MeOH/CHCl<sub>3</sub>) to give 4 as an oil.

**5.3.1.** *tert*-Butyl 3-[1,3-dioxo-3-(4-phenoxyphenyl)-1-propyl]piperidine-1-carboxylate (4a). 56% Yield. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  1.48 (s, 9H), 1.61–1.67 (m, 3H), 2.31–2.36 (m, 2H), 2.94–2.98 (m, 2H), 4.09 (br, 2H), 6.46 (s, 1H), 6.99 (d, 2H, J = 8.7 Hz), 7.04–7.06 (m, 2H), 7.15–7.20 (m, 1H), 7.35–7.40 (m, 2H), 7.83 (d, 2H, J = 8.7 Hz), 16.02 (br, 1H); HRMS calcd for 424.2124, found 424.2127.

**5.3.2.** *tert*-Butyl **2-[1,3-dioxo-3-(4-phenoxyphenyl)pro**pyl]piperidine-1-carboxylate (**4b**). 29% Yield. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  1.44 (s, 9H), 1.65–1.69 (m, 3H), 1.93–1.97 (m, 1H), 2.43–2.51 (m, 1H), 2.82–3.07 (m, 2H), 3.90–4.13 (m, 2H), 6.17 (s, 1H), 6.96 (d, 2H, J = 8.8 Hz), 7.01–7.04 (m, 2H), 7.12–7.17 (m, 1H), 7.27–7.37 (m, 2H), 7.85 (d, 2H, J = 8.8 Hz), 15.95 (br, 1H); HRMS calcd for 424.2124, found 424.2119.

**5.3.3. 1-[(***tert***-Butoxycarbonyl)-***N***-methyl]amino-6-(4phenoxyphenyl)hexane-4,6-dione (4c). 29% Yield. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): \delta 1.45 (s, 9H), 1.81–1.95 (m, 2H), 2.42 (t, 2H, J = 7.3 Hz), 2.86 (s, 3H), 3.29 (t, 2H, J = 7.0 Hz), 6.13 (s, 1H), 6.99 (d, 2H, J = 8.8 Hz), 7.02–7.08 (m, 2H), 7.16–7.21 (m, 1H), 7.36–7.41 (m, 2H), 7.86 (d, 2H, J = 8.8 Hz), 16.08 (br, 1H); HRMS calcd for 412.2124, found 412.2119.** 

**5.3.4. 1-[(***tert***-Butoxycarbonyl)-***N***-methyl]amino-4-(4phenoxyphenyl)butane-2,4-dione (4d). 49% Yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): \delta 1.45 (s, 9H), 2.99 (s, 3H), 4.03 (s, 2H), 6.12 (s, 1H), 7.01 (d, 2H, J = 8.7 Hz), 7.04–7.23 (m, 3H), 7.39–7.42 (m, 2H), 7.83 (d, 2H, J = 8.7 Hz), 15.98 (br, 1H); HRMS calcd for 383.1732, found 383.1735.** 

**5.3.5. 1-[N-Benzyl-(***tert***-butoxycarbonyl)]amino-4-(4-phenoxyphenyl)butane-2,4-dione (4e).** 33% Yield. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  1.46 (s, 9H), 3.93 (s, 2H), 4.58 (s, 2H), 6.03 (s, 1H), 6.99 (d, 2H, J = 8.6Hz), 7.05–7.08 (m, 2H), 7.17–7.42 (m, 8H), 7.79 (d, 2H,

*J* = 8.6 Hz), 16.16 (br, 1H); HRMS calcd for 460.2124, found 460.2129.

**5.3.6.** *tert*-Butyl **4-[3-(3-benzyloxyphenyl)-1,3-dioxo-1**propyl]piperidine-1-carboxylate (4f). 68% Yield. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  1.39 (s, 9H), 1.45–1.60 (m, 2H), 1.73–1.76 (m, 2H), 2.28–2.38 (m, 1H), 2.62–2.70 (m, 2H), 3.98–4.08 (m, 2H), 4.95 (s, 2H), 6.05 (s, 1H), 6.98 (m, 1H), 7.19–7.42 (m, 8H), 16.13 (br, 1H); HRMS calcd for 437.2202, found 437.2204.

**5.3.7. 1-Cyclohexyl-3-(4-phenoxyphenyl)propane-1,3dione (4g).** 69% Yield. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$ 1.20–1.51 (m, 5H), 1.69–1.95 (m, 5H), 2.30 (tt, 1H, J = 11.4 Hz, J = 3.1 Hz), 6.12 (s, 1H), 7.00 (d, 2H, J = 8.9 Hz), 7.05–7.08 (m, 2H), 7.15–7.20 (m, 1H), 7.35–7.42 (m, 2H), 7.86 (d, 2H, J = 8.9 Hz), 15.95 (br, 1H); HRMS calcd for 323.1647, found 323.1645.

# 5.4. General procedure for 1-(3-chlorophenyl)pyrazole derivatives 8–16

(a) A solution of 1,3-diketone derivatives 4 (10 mmol), 3chlorophenylhydrazine hydrochloride (3.58g, 20 mmol) and 2mol/L NaOH (19 mmol) in EtOH (30 mL) was heated to reflux for 12 h and cooled to 0 °C. To this reaction mixture was added 10% aqueous citric acid (30 mL) and the resulting solution was taken up in AcOEt (100 mL), washed successively with saturated aqueous NaHCO<sub>3</sub> and brine, and dried over MgSO<sub>4</sub>. The residue was purified by silica gel column chromatography (MeOH/CHCl<sub>3</sub>) to give a mixture of the 1-(3-chlorophenyl)-3-[*N*-(*tert*-butoxycarbonyl)]aminoalkylpyrazoles **5** and the 1-(3-chlorophenyl)-5-[*N*-(*tert*-butoxycarbonyl)]aminoalkylpyrazoles **6**.

(b) A solution of the mixture of **5** and **6** (10 mmol) and  $CF_3COOH$  (0.5 mL) in  $CH_2Cl_2$  (30 mL) was stirred for 4h at room temperature. The reaction mixture was concentrated to dryness, and the resulting residue was separated using CHP-20P (reverse phase, Mitsubishi Chemical Co., Ltd, Japan;  $CH_3CN/H_2O$ ) column chromatography and/or recrystallization from suitable solvents to give the 3-aminoalkyl-1-(3-chlorophenyl)pyrazole derivatives.

**5.4.1. 1-(3-Chlorophenyl)-5-(4-phenoxyphenyl)-3-(3-piperidyl)pyrazole hydrochloride (8).** 38% Yield from **4a**, prepared by lyophilization. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  1.68–1.86 (m, 3H), 2.11–2.14 (m, 1H), 2.85–2.92 (m, 1H), 3.04–3.10 (m, 1H), 3.17–3.29 (m, 2H), 3.49–3.51 (m, 1H), 6.63 (s, 1H), 6.99 (d, 2H, *J* = 8.8 Hz), 7.04 (d, 2H, *J* = 7.6 Hz), 7.15–7.20 (m, 2H), 7.24 (d, 2H, *J* = 8.8 Hz), 7.38–7.43 (m, 5H), 8.81–9.19 (m, 2H); MS (APCI<sup>+</sup>), *m*/*z* 430 (MH<sup>+</sup>). Anal. Calcd for C<sub>26</sub>H<sub>24</sub>ClN<sub>3</sub>O·HCl·2H<sub>2</sub>O: C, 62.15; H, 5.82; N, 8.36; Cl, 14.11. Found: C, 62.38; H, 5.70; N, 8.32; Cl, 14.12.

5.4.2. 1-(3-Chlorophenyl)-3-(4-phenoxyphenyl)-5-(3-piperidyl)pyrazole hydrochloride (9). 3% Yield from 4a, prepared by lyophilization. <sup>1</sup>H NMR (200 MHz, DMSO- $d_6$ ):  $\delta$  1.64–2.04 (m, 4H), 2.83–3.09 (m, 2H), 3.17–3.51 (m, 3H), 6.96 (s, 1H), 7.04–7.22 (m, 5H), 7.39–7.78 (m, 6H), 7.85 (d, 2H, J = 8.3 Hz), 8.76–9.16 (m, 2H); MS (APCI<sup>+</sup>), m/z 430 (MH<sup>+</sup>). Anal. Calcd for C<sub>26</sub>H<sub>24</sub>ClN<sub>3</sub>O·1.2HCl·2H<sub>2</sub>O: C, 61.55; H, 5.77; N, 8.24; Cl, 15.30. Found: C, 61.55; H, 5.60; N, 8.17; Cl, 15.12.

**5.4.3. 1-(3-Chlorophenyl)-5-(4-phenoxyphenyl)-3-(2-piperidyl)pyrazole hydrochloride (10).** 8% Yield from **4b**, mp 245–248 °C (acetone–Et<sub>2</sub>O). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  1.56 (br, 1H), 1.76–1.94 (m, 3H), 2.11–2.15 (m, 2H), 3.08 (br, 2H), 4.27 (br, 1H), 7.07–7.09 (m, 4H), 7.16–7.26 (m, 2H), 7.38–7.48 (m, 2H), 7.55–7.69 (m, 4H), 7.79–7.82 (m, 2H), 9.31 (br, 1H), 9.37 (br, 1H); MS (APCI<sup>+</sup>), *m/z* 430 (MH<sup>+</sup>). Anal. Calcd for C<sub>26</sub>H<sub>24</sub>ClN<sub>3</sub>O·1.25HCl·0.75H<sub>2</sub>O: C, 63.86; H, 5.51; N, 8.59; Cl, 16.31. Found: C, 63.96; H, 5.29; N, 8.58; Cl, 16.02.

**5.4.4. 1-(3-Chlorophenyl)-3-[3-(***N***-methyl)aminopropyl]-5-(4-phenoxyphenyl)pyrazole hydrochloride (11). 53% Yield from 4c, prepared by lyophilization. <sup>1</sup>H NMR (300 MHz, DMSO-***d***<sub>6</sub>): \delta 1.99–2.09 (m, 2H), 2.52–2.56 (m, 3H), 2.73 (t, 2H,** *J* **= 7.3 Hz), 2.98 (br, 2H), 6.54 (s, 1H), 6.98–7.01 (m, 2H), 7.04 (d, 2H,** *J* **= 8.4 Hz), 7.16– 7.21 (m, 2H), 7.26 (d, 2H,** *J* **= 8.4 Hz), 7.35–7.45 (m, 5H), 9.03 (br, 2H); MS (APCI<sup>+</sup>),** *m***/***z* **418 (MH<sup>+</sup>). Anal. Calcd for C<sub>25</sub>H<sub>24</sub>ClN<sub>3</sub>O·1.25HCl·0.75H<sub>2</sub>O: C, 63.19; H, 5.66; N, 8.84; Cl, 16.41. Found: C, 63.13; H, 5.80; N, 8.76; Cl, 16.23.** 

**5.4.5. 1-(3-Chlorophenyl)-3-(***N***-methylaminomethyl)-5-(4phenoxyphenyl)pyrazole fumarate (12).** 14% Yield from **4d**, mp 145–148 °C (*i*-PrOH–Et<sub>2</sub>O). <sup>1</sup>H NMR (200 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  2.48–2.54 (m, 3H), 4.05 (br, 2H), 6.49 (s, 2H), 6.72 (s, 1H), 6.98–7.08 (m, 4H), 7.15–7.28 (m, 4H), 7.38–7.46 (m, 5H), 9.87 (br, 3H); MS (APCI<sup>+</sup>), *m*/*z* 390 (MH<sup>+</sup>). Anal. Calcd for C<sub>23</sub>H<sub>20</sub>ClN<sub>3</sub>O·C<sub>4</sub>H<sub>4</sub>O<sub>4</sub>: C, 64.10; H, 4.78; N, 8.31; Cl, 7.01. Found: C, 64.11; H, 4.76; N, 8.31; Cl, 6.91.

**5.4.6. 1-(3-Chlorophenyl)-5-(***N***-methylaminomethyl)-3-(4phenoxyphenyl)pyrazole hydrochloride (13).** 57% Yield from **4d**, mp 184–187 °C (CH<sub>3</sub>CN). <sup>1</sup>H NMR (200 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  2.48–2.53 (m, 3H), 4.25 (m, 2H), 7.05– 7.31 (m, 6H), 7.42 (d, 2H, *J* = 8.4Hz), 7.47–7.76 (m, 4H), 7.83 (d, 2H, *J* = 8.4Hz), 9.93 (br, 2H); MS (APCI<sup>+</sup>), *m*/*z* 390 (MH<sup>+</sup>). Anal. Calcd for C<sub>23</sub>H<sub>20</sub>ClN<sub>3</sub>O·0.95-HCl·0.10H<sub>2</sub>O: C, 64.80; H, 5.00; N, 9.86; Cl, 16.22. Found: C, 64.53; H, 4.91; N, 10.00; Cl, 16.07.

**5.4.7. 3-**(*N*-Benzylaminomethyl)-1-(**3**-chlorophenyl)-**5**-(**4**-phenoxyphenyl)pyrazole hydrochloride (14). 25% Yield from **4e** as an oil. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  4.21 (s, 2H), 4.24 (s, 2H), 6.91 (s, 1H), 7.00–7.10 (m, 4H), 7.16–7.26 (m, 4H), 7.37–7.49 (m, 8H), 7.54–7.60 (m, 2H), 9.99 (br, 2H); HRMS calcd for 466.1686, found 466.1693.

5.4.8. 5-(3-Benzyloxyphenyl)-1-(3-chlorophenyl)-3-(4-piperidyl)pyrazole hydrochloride (15). 19% Yield from 4f, mp 90–93 °C (acetone–Et<sub>2</sub>O). <sup>1</sup>H NMR (200 MHz, DMSO- $d_6$ ):  $\delta$  1.84–2.03 (m, 2H), 2.13–2.21 (m, 2H),

2.96–3.10 (m, 3H), 3.31–3.34 (m, 2H), 5.05 (s, 2H), 6.59 (s, 1H), 6.76–6.82 (m, 1H), 6.94–7.18 (m, 3H), 7.25–7.44 (m, 9H), 9.02 (br, 1H), 9.18 (br, 1H); MS (APCI<sup>+</sup>), m/z 444 (MH<sup>+</sup>). Anal. Calcd for C<sub>27</sub>H<sub>26</sub>ClN<sub>3</sub>O·HCl·0.50-H<sub>2</sub>O: C, 66.26; H, 5.77; N, 8.59; Cl, 14.49. Found: C, 66.37; H, 5.84; N, 8.55; Cl, 14.79.

**5.4.9. 3-(3-Benzyloxyphenyl)-1-(3-chlorophenyl)-5-(4-piperidyl)pyrazole hydrochloride (16).** 41% Yield from **4f**, mp 135–138°C (CH<sub>3</sub>CN). <sup>1</sup>H NMR (200 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  1.75–1.98 (m, 4H), 2.85–2.99 (m, 2H), 3.07–3.32 (m, 3H), 5.17 (s, 2H), 6.88 (s, 1H), 6.97–7.04 (m, 1H), 7.31–7.70 (m, 12H), 9.11 (br, 2H); MS (APCI<sup>+</sup>), *m*/*z* 444 (MH<sup>+</sup>). Anal. Calcd for C<sub>27</sub>H<sub>26</sub>ClN<sub>3</sub>O·1.40-HCl·0.25H<sub>2</sub>O: C, 64.92; H, 5.63; N, 8.41; Cl, 17.03. Found: C, 65.16; H, 5.45; N, 8.57; Cl, 16.90.

5.4.10. 1-(3-Chlorophenyl)-3-(cyclohexyl)-5-(4-phenoxyphenyl)pyrazole (7). A solution of 1-cyclohexyl-3-(4phenoxyphenyl)propane-1,3-dione (2.11g, 6.54 mmol) and 3-chlorophenylhydrazine hydrochloride (2.34g, 13.1 mmol) in EtOH (20mL) was heated to reflux for 12h and cooled to 0°C. To this reaction mixture was added 10% aqueous citric acid (20 mL) and the resulting solution was taken up in AcOEt (70 mL), washed successively with saturated aqueous NaHCO<sub>3</sub> and brine, dried over MgSO<sub>4</sub> and evaporated to dryness. The residue was then purified by silica gel column chromatography (AcOEt/n-hexane) to give 1.15g (41%) of 7 as an oil. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  1.21–1.48 (m, 5H), 1.70-1.91 (m, 6H), 6.48 (s, 1H), 7.00-7.18 (m, 5H), 7.28–7.53 (m, 6H), 7.81 (d, 2H, J = 8.8 Hz); HRMS calcd for 429.1734, found 429.1730.

## 5.5. Synthesis of 2-hydroxyethane-1-one 19a,b

5.5.1. 2-(3-Chlorophenyl)-1-(4-phenoxyphenyl)-2-hydroxyethane-1-one (19a). (a) To a solution of 17a (6.33g, 31.3 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (30 mL) was added dropwise trimethylsilylcyanide (3.11 g, 31.3 mmol) at 0 °C and added zinc iodide (15 mg) at the same temperature. The mixture was stirred for 48 h at room temperature. The solvent was evaporated and the residue was purified by silica gel column chromatography (CHCl<sub>3</sub>) to give 18a as an oil, which was used in the next step without further purification.

(b) To a solution of 1 mol/L lithium hexamethyldisilazide (25mmol) in THF (50mL) was added dropwise 18a (7.44g, 25mmol) in THF (15mL) at -78°C under argon atmosphere. The yellow suspension was stirred for 30min at the same temperature. To the resulting solution was added dropwise 3-chlorobenzaldehyde (3.51 g, 25 mmol) in THF (20 mL) at the same temperature. After removal of the cold bath, the reaction mixture was raised to room temperature and stirred for 12h. To this reaction mixture was added saturated aqueous NH<sub>4</sub>Cl (60mL), and the solution was taken up in AcOEt (100 mL), washed with brine, dried over MgSO<sub>4</sub>, and concentrated to dryness. The residue was dissolved in MeOH (30mL) and was added 5% aqueous H<sub>2</sub>SO<sub>4</sub> (15mL). The reaction mixture was stirred for 30min at room temperature. The resulting solution was taken

up in Et<sub>2</sub>O (50mL), washed with 0.5 mol/L NaOH (15mL), dried over MgSO<sub>4</sub>, and concentrated to dryness. The residue was purified by silica gel column chromatography (CHCl<sub>3</sub>) to give 2.01 g (19% from **17a**) of **19a** as an oil. <sup>1</sup>H NMR (200 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  4.61 (d, 1H, J = 6.0 Hz) 5.84 (d, 1H, J = 6.0 Hz), 6.94 (d, 2H, J = 8.6 Hz), 7.01–7,42 (m, 9H), 7.88 (d, 2H, J = 8.6 Hz); MS (APCI<sup>+</sup>), *m/z* 339 (MH<sup>+</sup>).

**5.5.2. 2-(3-Benzyloxyphenyl)-1-(3-chlorophenyl)-2**hydroxyethane-1-one (19b). In a similar manner to that described above, 19b was prepared from 17b via 18b in 78% overall yield. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  4.40 (d, 1H, J = 6.1 Hz), 5.05 (s, 2H), 5.85 (d, 1H, J = 6.1 Hz), 6.89–6.94 (m, 2H), 7.23–7.51 (m, 9H), 7.70–7.90 (m, 2H); MS (APCI<sup>+</sup>), m/z 353 (MH<sup>+</sup>).

## 5.6. Synthesis of oxazole derivatives 21, 22

5.6.1. 5-(3-Chlorophenyl)-4-(4-phenoxyphenyl)-2-(4-piperidyl)oxazole fumarate (21). A solution of 19a (1.60g, 4.72 mmol), 1-(*tert*-butoxycarbonyl)piperidine-4-carboxylic acid (1.19g, 5.18 mmol), 1-ethyl-3[3-(dimethyl-(EDC, amino)propyl]carbodiimide hydrochloride 5.18 mmol) and 4-dimethylaminopyridine 0.92g, (DMAP, 0.58g, 4.72 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (40 mL) was stirred for 6h at room temperature. To this reaction mixture was added 10% aqueous citric acid (30mL) and the resulting solution was taken up in CHCl<sub>3</sub> (50 mL), dried over MgSO<sub>4</sub> and concentrated to dryness. The residue was dissolved in AcOH (40mL) and was added NH<sub>4</sub>OAc (0.91g, 11.8 mmol). The reaction mixture was heated to reflux for 6h and cooled to room temperature. The solvent was evaporated, and the residue was purified by silica gel column chromatography (2:1 CHCl<sub>3</sub>/MeOH) and recrystallization from EtOH-Et<sub>2</sub>O to give 1.16g (51% from 19a) of 21, mp 145-148°C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  1.62–1.74 (m, 1H), 1.91-2.02 (m, 2H), 2.22-2.26 (m, 1H), 2.83-2.92 (m, 1H), 2.99-3.07 (m, 1H), 3.18-3.32 (m, 3H), 6.48 (s, 2H), 6.63 (br, 2H), 7.04-7.10 (m, 4H), 7.16-7.21 (m, 1H), 7.42–7.51 (m, 5H), 7.54–7.61 (m, 3H), 8.12 (br, 1H); HRMS calcd for 430.1448, found 430.1454.

**5.6.2.** 5-(3-Benzyloxyphenyl)-4-(3-chlorophenyl)-2-(4-piperidyl)oxazole fumarate (22). In a similar manner to that described above, **22** was prepared from **19b** via **20b** in 44% overall yield, mp 123–126 °C (CHCl<sub>3</sub>). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  1.67 (br, 3H), 1.81–1.93 (m, 2H), 2.10–2.14 (m, 2H), 2.74–2.81 (m, 2H), 2.97–3.05 (m, 1H), 3.19–3.23 (m, 2H), 5.00 (s, 2H), 6.95–6.97 (m, 1H), 7.15–7.18 (m, 2H), 7.23–7.39 (m, 10H), 7.50–7.70 (m, 2H); MS (APCI<sup>+</sup>), *m*/*z* 445 (MH<sup>+</sup>). Anal. Calcd for C<sub>27</sub>H<sub>25</sub>CIN<sub>2</sub>O<sub>2</sub>Cl·C<sub>4</sub>H<sub>4</sub>O<sub>4</sub>: C, 66.37; H, 5.21; N, 4.99; Cl, 6.32. Found: C, 66.06; H, 4.86; N, 4.93; Cl, 6.43.

#### 5.7. Synthesis of imidazole derivatives 24, 25

5.7.1. 5-(3-Chlorophenyl)-4-(4-phenoxyphenyl)-2-(4-piperidyl)imidazole difumarate (24). (a) A solution of CuSO<sub>4</sub> (1.35 g, 8.50 mmol) in pyridine (5 mL) and H<sub>2</sub>O (1 mL) was heated to reflux for 10 min and cooled to

room temperature. To the resulting solution was added **19a** (1.44g, 4.25 mmol) and reheated to reflux for 1 h and cooled to room temperature. The solvent was evaporated and the residue was taken up  $Et_2O$  (20 mL) and  $H_2O$  (10 mL). The organic layer was separated and dried over MgSO<sub>4</sub>, and concentrated to dryness. The residue was purified by silica gel column chromatography (3:1 CHCl<sub>3</sub>/*n*-hexane) to give **23a** as an oil, which was used in the next step without further purification.

(b) A solution of **23a** (0.65 g, 1.93 mmol), 1-(*tert*-butoxycarbonyl)-4-formylpiperidine (0.41 g, 1.93 mmol) and NH<sub>4</sub>OAc (1.19 g, 1.54 mmol) in AcOH (15 mL) was stirred for 4h at 90 °C. The solvent was evaporated, and the residue was taken up in CHCl<sub>3</sub> (20 mL), washed with saturated aqueous NaHCO<sub>3</sub>, and dried over MgSO<sub>4</sub>. The solvent was evaporated. The residue was purified by CHP-20P (3:1 CH<sub>3</sub>CN/H<sub>2</sub>O) column chromatography to give 0.22 g (12% from **19a**) of **24**, mp 93–95 °C (EtOH–Et<sub>2</sub>O). <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>):  $\delta$ 1.89–2.02 (m, 2H), 2.10–2.16 (m, 2H), 2.95–3.06 (m, 3H), 3.30 (br, 5H), 3.32–3.36 (m, 2H), 6.51 (s, 4H), 7.00–7.06 (m, 4H), 7.13–7.18 (m, 1H), 7.24–7.49 (m, 9H); HRMS calcd for 429.1608, found 429.1599.

**5.7.2. 5-(3-Benzyloxyphenyl)-4-(3-chlorophenyl)-2-(4-piperidyl)imidazole dihydrochloride (25).** In a similar manner to that described above, **25** was prepared from **19b** via **23b** in 24% overall yield, mp 140–141 °C (CH<sub>3</sub>CN). <sup>1</sup>H NMR (200 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  2.06–2.34 (m, 4H), 2.97–3.16 (m, 2H), 3.39–3.53 (m, 3H), 5.11 (s, 2H), 7.03 (d, *J* = 8.0 Hz, 1H), 7.09–7.21 (m, 2H), 7.35–7.64 (m, 11H), 9.24 (br, 3H); MS (APCI<sup>+</sup>), *m*/*z* 444 (MH<sup>+</sup>). Anal. Calcd for C<sub>27</sub>H<sub>26</sub>ClN<sub>3</sub>OCl·2HCl·1.30H<sub>2</sub>O: C, 60.02; H, 5.71; N, 7.78; Cl, 19.68. Found: C, 59.65; H, 5.33; N, 7.75; Cl, 19.71.

## 5.8. Microbiology

**5.8.1. In vitro antibacterial activity.** The minimal inhibitory concentration (MIC) of each test-compound was determined by a microdilution method according to guidelines of the National Committee for Clinical Laboratory Standard.

**5.8.2. Enzyme inhibition.** DNA gyrase supercoiling and topoisomerase IV decatenation assays using *E. coli* enzymes were carried out by the methods of Sato et al.<sup>19</sup> and Peng and Marians,<sup>20</sup> respectively.

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### **References and notes**

- Brickner, S. J.; Hutchinson, D. K.; Barbachyn, M. R.; Manninen, P. R.; Ulanowicz, D. A.; Garmon, S. A.; Grega, K. C.; Hendges, S. K.; Toops, D. S.; Ford, C. W.; Zurenko, G. E. J. Med. Chem. 1996, 39, 673–679.
- (a) Ferrero, L.; Cameron, B.; Manse, B.; Lagneaux, D.; Crouzet, J.; Famechon, A.; Blanche, F. *Mol. Microbiol.* **1994**, *13*, 641–653; (b) Kim, O. K.; Ohemeng, K. A. *Expert Opin. Ther. Pat.* **1998**, *8*, 959–969.
- Miyamoto, T.; Matsumoto, J.; Chiba, K.; Egawa, H.; Shibamori, K.; Minamida, A.; Nishimura, Y.; Okada, H.; Kataoka, M.; Fujita, M.; Hirose, T.; Nakano, J. J. Med. Chem. 1990, 33, 1645–1656.
- 4. Maxwell, A. Mol. Microbiol. 1993, 9, 681-686.
- (a) Nakada, N.; Gmuender, H.; Hirata, T.; Arisawa, M. Antimicrob. Agents Chemother. 1994, 38, 1966–1973; (b) Nakada, N.; Gmunder, H.; Hirata, T.; Arisawa, M. J. Biol. Chem. 1995, 270, 14286–14291.
- (a) Rudolph, J.; Theis, H.; Hanke, R.; Endermann, R.; Johannsen, L.; Geschke, F. J. Med. Chem. 2001, 44, 619– 626; (b) Angehrn, P.; Buchmann, S.; Funk, C.; Goetschi, E.; Gmuender, H.; Hebeisen, P.; Kostrewa, D.; Link, H.; Luebbers, T.; Masciadri, R.; Nielsen, J.; Reindl, P.; Ricklin, F.; Schmitt-Hoffmann, A.; Theil, F. J. Med. Chem. 2004, 47, 1487–1513.
- Boehm, H.; Boehringer, M.; Bur, D.; Gmuender, H.; Hunber, W.; Klaus, W.; Kostrewa, D.; Kuehne, H.; Luebbers, T.; Meunier-Keller, N.; Mueller, F. J. Med. Chem. 2000, 43, 2664–2674.
- Lubbers, T.; Angehrn, P.; Gmunder, H.; Herzig, S.; Kulhanek, J. *Bioorg. Med. Chem. Lett.* 2000, 10, 821– 826.
- Galm, U.; Heller, S.; Shapiro, S.; Page, M.; Li, S.; Heide, L. Antimicrob. Agents Chemother. 2004, 48, 1307–1312.
- (a) Tanitame, A.; Oyamada, Y.; Ofuji, K.; Kyoya, Y.; Suzuki, K.; Ito, H.; Kawasaki, M.; Nagai, K.; Wachi, M.; Yamagishi, J. *Bioorg. Med. Chem. Lett.* 2004, 14, 2857– 2862; (b) Tanitame, A.; Oyamada, Y.; Ofuji, K.; Suzuki, K.; Ito, H.; Kawasaki, M.; Wachi, M.; Yamagishi, J. *Bioorg. Med. Chem. Lett.* 2004, 14, 2863–2866.
- 11. Wachi, M.; Iwai, N.; Kunihisa, A.; Nagai, K. *Biochimie* **1999**, *81*, 909–913.
- 12. Hiraga, S.; Niki, H.; Ogura, T.; Ichinose, C.; Mori, H.; Ezaki, B.; Jaffe, A. J. Bacteriol. **1989**, *171*, 1496–1505.
- Tanitame, A.; Oyamada, Y.; Ofuji, K.; Fujimoto, M.; Iwai, N.; Hiyama, Y.; Suzuki, K.; Ito, H.; Terauchi, H.; Kawasaki, M.; Nagai, K.; Wachi, M.; Yamagishi, J. J. Med. Chem. 2004, 14, 3693–3696.
- Rowley, M.; Broughton, H. B.; Collins, I.; Baker, R.; Emms, F.; Marwood, R.; Patel, S.; Patel, S.; Ragan, C. I.; Freedman, S. B.; Leeson, P. D. J. Med. Chem. 1996, 39, 1943–1945.
- Rowley, M.; Collins, I.; Broughton, H. B.; Davey, W. B.; Baker, R.; Emms, F.; Marwood, R.; Patel, S.; Patel, S.; Ragan, C. I.; Freedman, S. B.; Ball, R.; Leeson, P. D. J. Med. Chem. 1997, 40, 2374–2385.
- Cameron, J. F.; Willson, C. G.; Frechet, J. M. J. J. Chem. Soc., Perkin Trans. 1 1997, 16, 2429–2442.
- Meanwell, N. A.; Rosenfeld, M. J.; Trehan, A. K.; Romine, J. L.; Wright, J. J. K.; Brassard, C. L.; Buchanan, J. O.; Federici, M. E.; Fleming, J. S.; Gamberdella, M.; Zavoico, G. B.; Seiler, S. M. J. Med. Chem. 1992, 35, 3498–3512.
- Brackeen, M. F.; Stafford, J. A.; Feldman, P. L.; Karanewsky, D. S. *Tetrahedron Lett.* 1994, 35, 1635–1638.
- Sato, K.; Inoue, Y.; Fujii, T.; Aoyama, H.; Inoue, M.; Mitsuhashi, S. Antimicrob. Agents Chemother. 1986, 30, 777–780.

- 20. Peng, H.; Marians, K. J. J. Biol. Chem. 1993, 268, 24481-24490.
- Stieger, M.; Angehrn, P.; Wohlgensinger, B.; Gmunder, H. Antimicrob. Agents Chemother. 1996, 40, 1060–1062.
- 22. Galakatos, N. G.; Kemp, D. S. J. Org. Chem. 1985, 50, 1302–1304.
- Hoekstra, W. J.; Maryanoff, B. E.; Damiano, B. P.; Andrade-Gordon, P.; Cohen, J. H.; Costanzo, M. J.; Haertlein, B. J.; Hecker, L. R.; Hulshizer, B. L.; Kauffman, J. A.; Keane, P.; McComsey, D. F.; Mitchell, J. A.; Scott, L.; Shah, R. D.; Yabut, S. C. J. Med. Chem. 1999, 42, 5254–5265.