ChemComm



COMMUNICATION

View Article Online
View Journal | View Issue



Cite this: *Chem. Commun.,* 2015, **51**, 7012

Received 21st February 2015, Accepted 16th March 2015

DOI: 10.1039/c5cc01572c

www.rsc.org/chemcomm

The flavan-isoflavan rearrangement: bioinspired synthetic access to isoflavonoids *via* 1,2-shift-alkylation sequence†

Kayo Nakamura, Ken Ohmori* and Keisuke Suzuki*

An approach to 2-substituted isoflavonoids is reported based on the 1,2-shift of the aryl group in the catechin skeleton followed by the *in situ* alkylation. Synthesis of (–)-equol, a natural isoflavan with estrogenic activities, was achieved.

Isoflavonoids constitute a class of natural products widely found in leguminous plants. In addition to the phytoalexin activity in the original plants, some compounds have been attracting special attention in the field of human health care, e.g., the estrogen activities identified in the soybean-derived compounds 1–3. Furthermore, several elaborated compounds with stereogenicity are found in nature, including isoflavans, pterocarpans, and rotenonoids, such as 4–6 (Fig. 1). Due to the diverse range of bioactivities, isoflavonoids have become one of the current targets for chemical synthesis.

The isoflavonoid biosynthesis shares its early stage with that of the flavonoid, branching at the flavanone stage by P-450 mediated oxygenation to induce the aryl 1,2-shift within the chroman skeleton to form the 3-aryl derivatives (Scheme 1). Although the following dehydration gives isoflavones, 4 stereogenic compounds, e.g., 4-6 are

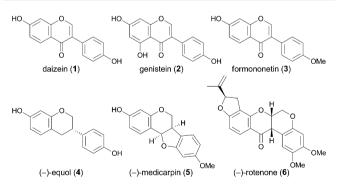
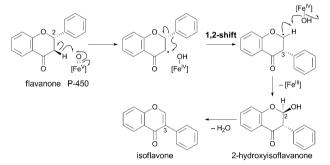


Fig. 1 Natural isoflavonoids.

Department of Chemistry, Tokyo Institute of Technology, O-okayama, Meguro, Tokyo 152-8551, Japan. E-mail: kohmori@chem.titech.ac.jp, ksuzuki@chem.titech.ac.ip



Scheme 1 Biosynthesis of isoflavonoids.

generated by further biosynthetic elaborations. This biogenetic 1,2-shift and the presence of the intriguing natural products 4–6 prompted us to devise a synthesis route for obtaining isoflavonoids.

In this communication, we describe a synthetic access to isoflavonoids *via* a 1,2-shift and alkylation sequence within the flavonoid scaffolds.

We took inspiration from our previous study (Scheme 2).⁵ Eqn (1) is a pinacol-type shift of a phenyl group by the activation

Scheme 2 Organoaluminum-mediated stereospecific 1,2-shift.

 $[\]dagger$ Electronic supplementary information (ESI) available. See DOI: 10.1039/c5cc01572c

Communication ChemComm

of a mesylate with Et₃Al to effect a stereospecific 1,2-shift.^{5a} A related process involves trapping of the intermediary oxonium species by an organoaluminum ligand (eqn (2)).^{5b} By analogy, we asked ourselves whether such a 1,2-shift was viable within a catechin framework, and the model study started with a catechin-derived substrate, *i.e.* mesylate 7.⁶

Mesylate 7 was prepared from tetra-O-benzyl catechin⁷ (CH₃SO₂Cl, Et₃N, CH₂Cl₂, 0 °C, 96% yield). Upon treatment of 7 with Me₃Al (2 equiv.) in CH₂Cl₂ at -78 °C followed by gradually warming to 0 °C, a single new product 8a was produced in 90% yield (eqn (3)). The *trans* stereochemistry of the aryl and the methyl groups was assigned using the coupling constant ($J_{2,3} = 9.2$ Hz) and the ROESY experiment.⁸ The stereochemical outcome could be attributed to the inversive 1,2-shift followed by trapping by a methyl nucleophile from the opposite side. Furthermore, the enantiomeric purity of 8a (>99% e.e.) was verified by HPLC analysis using a chiral stationary phase.⁹

$$\begin{array}{c} \text{BnO} \\ \text{OBn} \\ \text{OMs} \\ \text{OMs} \\ \text{OMs} \\ \text{OHs} \\ \text{OBn} \\ \text{CH}_2\text{Cl}_2 \\ -78 \rightarrow 0\,^{\circ}\text{C} \\ \text{BnO} \\ \text{8a} \\ \text{OBn} \\ \text{OBn} \\ \text{90% (>99\% e.e.)} \\ \end{array}$$

A control experiment showed the well-known importance of the anti-relationship of the leaving group and the migrating group (eqn (4)). Epicatechin derivative $\bf 9$, upon reaction with Me₃Al, led only to a slow 1,2-shift of the hydride to give $\bf 10$ in 21% yield, and mesylate $\bf 9$ was largely recovered. The ee of $\bf 10$ was 0%, not surprisingly, suggesting that the reactive species that underwent trapping by a methyl nucleophile was the oxonium species after the hydride shift was fully completed.

Table 1 shows the generality of the process, giving various 2-substituted isoflavans. Reaction of 7 with Et₃Al proceeded smoothly to give the ethylated product **8b** in excellent yield (run 1). AlH₃, *in situ* prepared from LiAlH₄ and AlCl₃, ¹⁰ induced the 1,2-shift of 7 followed by hydride trapping, giving isoflavan **8c** in 73% yield (run 2). Reaction of 7 with i-Bu₃Al gave the expected product **8d** with an i-butyl group. A small amount of **8c** was obtained, arising from the β-hydride delivery from i-Bu₃Al (run 3). Furthermore, reactions of other organoaluminum reagents gave various isoflavans **8e–8i** in moderate to high yields and rigorous *trans* selectivities. In runs 4–6, triorganoaluminum reagents were generated *in situ* from the respective organolithium and AlCl₃. ¹¹ In runs 7 and 8, EtAlCl₂ was used for this purpose, where the alkynyl ligands were exclusively transferred. ¹²

Table 1 Conversion of catechin mesylate 7 to various isoflavans

Run	Reagent	R	Product	Yield [%]
1	Et ₃ Al	Et	8b	86
2	AlH_3	Н	8c	73
3	i-Bu ₃ Al	i-Bu	8d	74
4	$Al(CH_2SiMe_3)_3^a$	CH ₂ SiMe ₃	8e	83
5	PhAl ^a	Ph	8f	95
	$\langle \sim _{c} t$ -Bu \rangle^{a}	∿√√ t-Bu		
6	Al 🗸 💛	· vy 🗸	8g	54
7	$EtAl(C \equiv CPh)_2^b$	$C \equiv CPh$	8h	76
8	$Al \leftarrow t$ -Bu $\binom{a}{3}$ EtAl $(C \equiv CPh)_2^b$ EtAl $(C \equiv CSiMe_3)_2^b$	$C \equiv CSiMe_3$	8i	96

 $[^]a$ Prepared from the corresponding organolithium and AlCl $_3.$ b Prepared from the corresponding alkynyllithium and EtAlCl $_2.$

As a variation of the migrating group, an *ortho*-substituted phenyl group was tested (Scheme 3). Mesylate 17 was prepared *via* our method for the flavan synthesis. The Mitsunobu reaction of epoxy alcohol 11^8 and iodophenol 12 gave ether 13 as a single product, and the subsequent cyclization gave flavan 16. After the removal of the TES group in 16, mesylation of the resulting alcohol gave mesylate 17. Treatment of 17 with Me₃Al ($-78 \rightarrow -10$ °C) induced a smooth 1,2-shift, giving isoflavan 18 in 84% yield as a single product.

Previously, we noted that o,o'-disubstituted phenyl groups are often sluggish to undergo the 1,2-shift, particularly when the substrate has steric hindrance. To address this point, we prepared mesylate 19 with an o,o'-dibenzyloxyphenyl group in a similar manner. To our delight, the reaction of 19^8 with Me₃Al $(-78 \rightarrow -30 \, ^{\circ}\text{C})$ gave 79% yield of the rearranged product 20

Scheme 3 Keys: (a) TMAD, $n\text{-Bu}_3\text{P}$, toluene, 0 °C, 2 h (83%, single diastereomer), (b) Li $_2\text{NiBr}_4$, THF, 0 °C \rightarrow RT, 80 h (91%), (c) TESOTf, 2,6-lutidine, CH $_2\text{Cl}_2$, 0 °C, 20 min (95%), (d) PhMgBr, PhLi, HMPA, THF, $-78 \rightarrow 0$ °C, 30 min (62%), (e) $n\text{-Bu}_4\text{NF}$, THF, 0 °C, 20 min (quant.), (f) MsCl, Et $_3\text{N}$, CH $_2\text{Cl}_2$, 0 °C, 40 min (94%), (g) Me $_3\text{Al}$, CH $_2\text{Cl}_2$, $-78 \rightarrow -10$ °C, 1.5 h (84%). TMAD = N,N,N',N'-tetramethylazodicarboxamide. HMPA = hexamethylphosphoric triamide.

(eqn (5)). This is a promising result in view of the synthesis of many natural isoflavonoids with an aryl group possessing ortho-hydroxy group(s).

ChemComm

Finally the enantiospecific synthesis of (-)-equol (4), a soy-derived isoflavonoid known since 1932 is described. 14 Recently, sizable phytoestrogenic activity has been found in 4,15 making it the current target of chemical synthesis.¹⁶

Scheme 4 outlines the synthesis of 4. The key intermediate 27 was prepared from the resorcinol derivative 21. The union of 21 with epoxide 22 $(>99\% \text{ e.e.})^7$ via the Mitsunobu reaction gave ether 23 in 78% yield as an inseparable mixture of diastereomers (93:7 ratio), 13b which was used for the next step. Regioselective cleavage of oxirane 23 gave bromohydrin 24 (87% yield) and its epimer 24' (4% yield), which were separated using flash column chromatography (hexane/toluene/EtOAc = 5/5/1). After protection of 24 as a TES ether, the cyclization precursor 25, thus obtained, was treated with Ph3MgLi to give flavan 26 in 88% yield. 13 After two-step conversion of 26 into mesylate 27, treatment with AlH₃ (CH₂Cl₂, 0 °C \rightarrow room temp., 2.5 h) cleanly afforded the desired isoflavan 28 in 85% yield. Finally, two benzyl groups were removed by hydrogenolysis [H₂, $Pd(OH)_2/C$, THF, MeOH, $H_2O(2/2/1)$, room temp., 45 min], giving (-)-equol (4) as a white solid (99% e.e.). ¹⁷ All the physical

Scheme 4 Keys: (a) TMAD, n-Bu₃P, toluene, 0 °C, 1 h (78%, dr = 93/7), (b) Li_2NiBr_4 , THF, 0 °C, 24 h (87%), (c) TESOTf, 2,6-lutidine, CH_2Cl_2 , 0 °C, 20 min (97%), (d) PhMgBr, PhLi, HMPA, THF, $-78 \rightarrow 0$ °C, 45 min (88%), (e) n-Bu₄NF, THF, 0 °C \rightarrow RT, 15 min (95%), (f) MsCl, Et₃N, CH₂Cl₂, 0 °C, 10 min (99%), (g) LiAlH₄, AlCl₃, CH₂Cl₂, 0 °C \rightarrow RT, 2.5 h (85%), (h) H₂, ASCA-2 [5% Pd(OH)₂/C], THF, MeOH, H₂O, RT, 45 min (quant.).

data of the synthetic sample of 4 (1 H and 13 C NMR, IR, $[\alpha]_{D}$) coincided with the reported data.16

In conclusion, an approach to the stereoselective synthesis of isoflavans has been established based on the 1,2-shift of aryl groups in flavan-3-ol derivatives and in situ alkylation by organoaluminum reagents. The method was applied in the synthesis of (-)-equol (4).

This work was supported by a Grant-in-Aid for Specially Promoted Research (No. 23000006) from ISPS.

Notes and references

- 1 D. A. Smith and S. W. Banks, Phytochemistry, 1986, 25, 979.
- 2 G. G. J. M. Kuiper, J. G. Lemmen, B. Carlsson, J. C. Corton, S. H. Safe, P. T. Saag, B. Burg and J.-A. Gustafsson, Endocrinology, 1998, 139, 4252
- 3 (a) A. Lévai, J. Heterocycl. Chem., 2004, 41, 449; (b) A. Goel, A. Kumar and A. Raghuvanshi, Chem. Rev., 2013, 113, 1614; (c) M. A. Selepe and F. R. V. Heerden, Molecules, 2013, 18, 4739; (d) R. L. Farmer and K. A. Scheidt, Chem. Sci., 2013, 4, 3304; (e) Z.-G. Feng, W.-J. Bai and T. R. R. Pettus, Angew. Chem., Int. Ed., 2015, 54, 1864, and references therein.
- 4 M. F. Hashim, T. Hakamatsuka, Y. Ebizuka and U. Sankawa, FEBS Lett., 1990, 271, 219,
- 5 (a) K. Suzuki, E. Katayama and G. Tsuchihashi, Tetrahedron Lett., 1983, 24, 4997; (b) Y. Honda, E. Morita and G. Tsuchihashi, Chem. Lett., 1985, 1153; (c) T. Saito, T. Suzuki, M. Morimoto, C. Akiyama, T. Ochiai, K. Takeuchi, T. Matsumoto and K. Suzuki, J. Am. Chem. Soc., 1998, 120, 11633.
- 6 For early examples on the use of the 1,2-shift to construct 3-aryl isoflavones, see (a) A. C. Jain, P. D. Sarpal and T. R. Seshadri, Indian J. Chem., 1965, 3, 369; (b) W. D. Ollis, K. L. Ormand and I. O. Sutherland, Chem. Commun., 1968, 1237; (c) O. Prakash, S. Pahuja, S. Goyal, S. N. Sawhney and M. Moriarty, Synlett, 1990, 337; (d) O. V. Singh, C. P. Garg and R. P. Kapoor, Tetrahedron Lett., 1990, 31, 2747; (e) T. Kinoshita, K. Ichinose and U. Sankawa, Tetrahedron Lett., 1990, **31**, 7355; (f) C. D. Gabbutt, J. D. Hepworth, B. M. Heron and J.-L. Thomas, Tetrahedron Lett., 1998, 39, 881.
- 7 H. Kawamoto, F. Nakatsubo and K. Murakami, Synth. Commun., 1996, 26, 531.
- 8 See the ESI†.
- 9 An authentic sample of ent-7 was prepared by the C2-epimerization of (-)-epicatechin (EC) (pH 8 phosphate buffer, 80 °C, 2 h), giving a mixture of (-)-catechin and (-)-EC. Peracetylation followed by the Kawamoto protocol, separation and mesylation gave ent-7. See the ESI†.
- 10 (a) A. E. Finholt, A. C. Bond and H. I. Schlesinger, J. Am. Chem. Soc., 1947, 69, 1199; (b) S. Raghavan, S. R. Reddy, K. A. Tony, C. N. Kumar, A. K. Varma and A. Nangia, J. Org. Chem., 2002, 67, 5838.
- 11 In run 6, the isoflavene (3-aryl-4H-chromene) derivative was obtained as a byproduct (16% yield). See the ESI†.
- 12 (a) J. Fried, C.-H. Lin and S. H. Ford, Tetrahedron Lett., 1969, 1379; (b) R. T. Hansen, D. B. Carr and J. Schwartz, J. Am. Chem. Soc., 1978, 100, 2244; (c) K. Taya, T. Nagasawa and K. Suzuki, Synlett, 1997, 304.
- 13 (a) T. Higuchi, K. Ohmori and K. Suzuki, Chem. Lett., 2006, 35, 1006; (b) K. Ohmori, M. Takeda, T. Higuchi, T. Shono and K. Suzuki, Chem. Lett., 2009, 38, 934.
- 14 G. F. Marrian and G. A. D. Haslewood, Biochem. J., 1932, 1227.
- 15 (a) J.-P. Yuan, J.-H. Wang and X. Liu, Mol. Nutr. Food Res., 2007, 51, 765; (b) P. J. Magee, Proc. Nutr. Soc., 2011, 70, 10; (c) R. L. Jackson, J. S. Greiwe and R. J. Schwen, Nutr. Rev., 2011, 69, 432; (d) D. Shor, T. Sathyapalan, S. L. Atkin and N. J. Thaycher, Eur. J. Nutr., 2012, 51, 389.
- 16 (a) J. M. Heemstra, S. A. Kerrigan, D. R. Doerge, W. G. Helferich and W. A. Boulanger, Org. Lett., 2006, 8, 5441; (b) Y. Takashima and Y. Kobayashi, Tetrahedron Lett., 2008, 49, 5156; (c) Y. Takashima, Y. Kaneko and Y. Kobayashi, *Tetrahedron*, 2010, **66**, 197; (d) J.-W. Lee and B. List, J. Am. Chem. Soc., 2012, 134, 1824; (e) S. Yang, S.-F. Zhu, C.-M. Zhang, S. Song, Y.-B. Yu, S. Li and Q.-L. Zhou, Tetrahedron, 2012, 68, 5172.
- 17 An authentic sample of (\pm) -4 was purchased from Tokyo Chemical Industry Co., Ltd. (TCI) and the enantiomeric purity of synthetic 4 was assessed by HPLC analysis. See the ESI†.