

Heterocyclic sulfoxide and sulfone inhibitors of fatty acid amide hydrolase

Wu Du, Christophe Hardouin, Heng Cheng, Inkyu Hwang and Dale L. Boger*

*Department of Chemistry and The Skaggs Institute for Chemical Biology, The Scripps Research Institute,
10550 North Torrey Pines Road, La Jolla, CA 92037, USA*

Received 23 July 2004; revised 5 October 2004; accepted 9 October 2004
Available online 28 October 2004

Abstract—A novel series of heterocyclic sulfoxides and sulfones was prepared and examined as potential inhibitors of fatty acid amide hydrolase (FAAH), the enzyme responsible for inactivation of neuromodulating fatty acid amides including anandamide and oleamide.

© 2004 Elsevier Ltd. All rights reserved.

1. Introduction

Oleamide (1)^{1,2} and anandamide (2)^{3,4} are prototypical members of a class of endogenous fatty acid amides that serve as chemical messengers.⁵ Anandamide is an endogenous fatty acid ethanolamide that binds and activates the central (CB1) and peripheral (CB2) cannabinoid receptors, and the vanilloid (VR1) receptor through which it is thought to exhibit its analgesic and cannabinoid effects.⁴ Oleamide was found to accumulate in the cerebrospinal fluid under conditions of sleep deprivation and to induce physiological sleep in animals where it reduces motility, shortens the sleep induction period, and lengthens the time spent in slow wave sleep 2 at the expense of waking.^{1,6}

Fatty acid amide hydrolase (FAAH)⁷ is an integral membrane protein that degrades fatty acid primary amides and ethanolamides including oleamide and anandamide (Fig. 1).^{8,9} The distribution of FAAH in the central nervous system suggests that it degrades neuromodulating fatty acid amides at their sites of action and is responsible for or intimately involved in their regulation.¹⁰ FAAH hydrolyzes a wide range of fatty acid substrates and it appears to work most effectively on arachidonyl and oleyl substrates.¹¹ This enzyme belongs to the amidase family of hydrolytic enzymes for which FAAH is the only member found in mammals.¹² This unique mammalian distribution,^{12–14} its attractive and

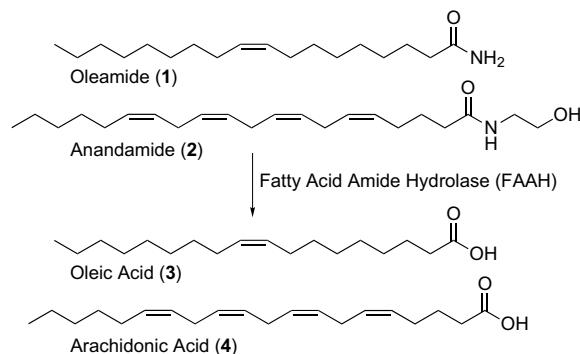


Figure 1.

targetable catalytic mechanism (Ser-Ser-Lys triad), and the consequences of inhibition (increase endogenous level of anandamide and/or oleamide) have made FAAH an attractive therapeutic target^{8,9} for the development of a new class of analgesics, nonhypnotic sleep-aids, and a range of additional clinical disorders.

In spite of this interest, only a selected series of FAAH inhibitors have been disclosed. These include the discovery that the endogenous sleep-inducing compound 2-octyl- γ -bromoacetoacetate is an effective inhibitor of FAAH,¹⁵ an early series of irreversible FAAH inhibitors (fatty acid sulfonyl fluorides,¹⁶ fluorophosphonates,¹⁷ and α -diazoketones¹⁸), and an early series of fatty acid-based reversible FAAH inhibitors bearing non-selective electrophilic carbonyls.^{19–21} More recently, two additional promising classes of FAAH inhibitors

* Corresponding author. Tel.: +1 8587847522; fax: +1 8587847550;
e-mail: boger@scripps.edu

3. Inhibition studies

Enzyme assays were performed at 20–23 °C with purified recombinant rat FAAH expressed in *E. coli*²⁸ in a 125 mM Tris/1 mM EDTA/0.2% glycerol/0.2% Triton X-100/0.4 mM Hepes, pH 9.0 buffer. The initial rates of hydrolysis (≤ 10 –20% reaction) were monitored using enzyme concentrations at least 3 times below the measured K_i by following the hydrolysis of ¹⁴C-oleamide

Compd	Aryl	X	K_i
—		CO ^{22d}	0.018 ± 0.005 μM
8a		S	160 ± 7 μM
10a		SO	8 ± 0.5 μM
—		CO ^{22d}	0.012 ± 0.001 μM
8b		S	130 ± 10 μM
10b		SO	2.2 ± 0.2 μM
—		CO ^{22d}	0.054 ± 0.004 μM
8c		S	180 ± 10 μM
10c		SO	8 ± 1 μM

Compd	Aryl	X	K_i
—		CO ^{22d}	0.0047 ± 0.0013 μM
9a		S	110 ± 10 μM
11a		SO	11 ± 0.4 μM
12a		SO ₂	16 ± 4.5 μM
—		CO ^{22d}	0.0046 ± 0.0004 μM
9b		S	80 ± 5 μM
11b		SO	20 ± 4 μM
12b		SO ₂	10 ± 2 μM
—		CO ^{22d}	0.012 ± 0.001 μM
9c		S	120 ± 10 μM
11c		SO	30 ± 10 μM
12c		SO ₂	40 ± 10 μM

Compd	X	K_i
—	CO ^{22a}	0.22 ± 0.01 μM
14	S	150 ± 10 μM
15	SO	2.6 ± 0.1 μM
16	SO ₂	3.3 ± 0.7 μM

Compd	X	K_i
—	CO ^{22f}	0.003 ± 0.0002 μM
18a	S	140 ± 10 μM
19a	SO	13 ± 1 μM ²⁹

Compd	n	X	K_i
18b	1	S	720 ± 50 μM
19b	1	SO	8 ± 1 μM
20b	1	SO ₂	7.5 ± 1 μM
18c	2	S	110 ± 10 μM
19c	2	SO	4.9 ± 1.0 μM
20c	2	SO ₂	6.8 ± 0.7 μM ³⁰
18d	3	S	860 ± 50 μM
19d	3	SO	7 ± 0.3 μM
20d	3	SO ₂	3.3 ± 0.5 μM

Figure 3. FAAH inhibition.

and K_i 's established as described (Dixon plot).^{22a} Lineweaver–Burk analysis established reversible, competitive inhibition (Fig. 2).

4. Results and discussion

The results of the examination of the sulfoxide and sulfone FAAH inhibitors are summarized in Figure 3.^{29,30} In each case, the sulfoxides and sulfones proved to be 10–100-fold more potent than the corresponding sulfides, consistent with an expected enhanced binding affinity derived from a polarized oxygen binding an enzyme oxyanion pocket mimicking the tetrahedral intermediate of the amide hydrolysis reaction. Moreover, little distinction and no discernable trends were observed in comparing the sulfoxides with the corresponding sulfones and both inhibit FAAH to the same extent.

Unlike the behavior of the corresponding α -ketoheterocycles, the potency of the corresponding sulfoxides and sulfones was relatively insensitive to the nature of the attached heterocycle as well as the side chain, although each system examined represent those known to convey effective FAAH active site binding.²² Finally, the sulfides were roughly 10^4 – 10^5 -fold less potent than the corresponding α -ketoheterocycles, whereas the sulfoxides and sulfones were roughly 10^2 – 10^3 -fold less potent than the α -ketoheterocycles. The notable exceptions are **15** and **16**, which proved to be only 10-fold less potent than the corresponding α -ketobenzoxazole, which constitutes one of the least potent α -ketoheterocycles examined in this series. Thus, the heterocyclic sulfoxides and sulfones exhibit a potency intermediate of the corresponding sulfides and α -ketoheterocycles illustrating that the polarized oxygen binding in an enzyme oxyanion binding pocket increases affinity up to 100-fold and that the reversible covalent binding of the α -ketoheterocycles to an active site serine increases binding up to 1000-fold beyond that of the corresponding sulfoxide and sulfone.

Acknowledgements

We gratefully acknowledge the financial support of the National Institute of Health (DA156480) and the Skaggs Institute for Chemical Biology. We wish to especially thank Professor B. F. Cravatt and his group for a generous supply of recombinant rat FAAH.

Supplementary data

Supplementary data associated with this article can be found, in the online version, at [doi:10.1016/j.bmcl.2004.10.025](https://doi.org/10.1016/j.bmcl.2004.10.025). Details of the synthesis and characterization of all new compounds is provided.

References and notes

- (a) Cravatt, B. F.; Prospero-Garcia, O.; Siuzdak, G.; Gilula, N. B.; Henriksen, S. J.; Boger, D. L.; Lerner, R. A.

- Science* **1995**, 268, 1506; (b) Cravatt, B. F.; Lerner, R. A.; Boger, D. L. *J. Am. Chem. Soc.* **1996**, 118, 580.
2. Review: Boger, D. L.; Henriksen, S. J.; Cravatt, B. F. *Curr. Pharm. Des.* **1998**, 4, 303.
 3. Devane, W. A.; Hanus, L.; Breuer, A.; Pertwee, R. G.; Stevenson, L. A.; Griffin, G.; Gibson, D.; Mandelbaum, A.; Etinger, A.; Mechoulam, R. *Science* **1992**, 258, 1946.
 4. Review: Di Marzo, V.; Bisogno, T.; De Petrocellis, L.; Melck, D.; Martin, B. R. *Curr. Med. Chem.* **1999**, 6, 721.
 5. (a) Schmid, H. H.; Schmid, P. C.; Natarajan, V. *Chem. Phys. Lipids* **1996**, 80, 133; (b) Schmid, H. H.; Schmid, P. C.; Natarajan, V. *Prog. Lipid Res.* **1990**, 29, 1.
 6. Huitron-Resendiz, S.; Gombart, L.; Cravatt, B. F.; Henriksen, S. J. *Exp. Neurol.* **2001**, 172, 235.
 7. (a) Giang, D. K.; Cravatt, B. F. *Proc. Natl. Acad. Sci., U.S.A.* **1997**, 94, 2238; (b) Cravatt, B. F.; Giang, D. K.; Mayfield, S. P.; Boger, D. L.; Lerner, R. A.; Gilula, N. B. *Nature* **1996**, 384, 83.
 8. (a) Cravatt, B. F.; Lichtman, A. H. *Curr. Opin. Chem. Biol.* **2003**, 7, 469; (b) Cravatt, B. F.; Lichtman, A. H. *Chem. Phys. Lipids* **2002**, 127, 135; (c) Patricelli, M. P.; Cravatt, B. F. *Vitam. Horm.* **2001**, 62, 95.
 9. (a) Reviews: Wendeler, M.; Kolter, T. *Angew. Chem., Int. Ed.* **2003**, 42, 2938; (b) Gaetani, S.; Cuomo, V.; Piomelli, D. *Trends Mol. Med.* **2003**, 9, 474; (c) Bisogno, T.; De Petrocellis, L.; Di Marzo, V. *Curr. Pharm. Des.* **2002**, 8, 533; (d) Deutsch, D. G.; Ueda, N.; Yamamoto, S. *Prostaglandins, Leukot. Essent. Fatty Acids* **2002**, 66, 201; (e) Maccarrone, M.; Van der Stelt, M.; Veldink, G. A.; Finazzi-Agro, A. *Curr. Med. Chem.: Anti-Inflammatory Anti-Allergy Agents* **2002**, 1, 103; (f) Fowler, C. J.; Jonsson, K.-O.; Tiger, G. *Biochem. Pharmacol.* **2001**, 62, 517; (g) Ueda, N.; Puffenbarger, R. A.; Yamamoto, S.; Deutsch, D. G. *Chem. Phys. Lipids* **2000**, 108, 107.
 10. (a) Lichtman, A. H.; Hawkins, E. G.; Griffin, G.; Cravatt, B. F. *J. Pharmacol. Exp. Ther.* **2002**, 302, 73; (b) Egertova, M.; Cravatt, B. F.; Elphick, M. R. *Neuroscience* **2003**, 119, 481; (c) Egertova, M.; Giang, D. K.; Cravatt, B. F.; Elphick, M. R. *Proc. Royal Soc. London, B: Biol. Sci.* **1998**, 265, 2081; (d) Thomas, E. A.; Cravatt, B. F.; Danielson, P. E.; Gilula, N. B.; Sutcliffe, J. G. *J. Neurosci. Res.* **1997**, 50, 1047.
 11. (a) Boger, D. L.; Fecik, R. A.; Patterson, J. E.; Miyauchi, H.; Patricelli, M. P.; Cravatt, B. F. *Bioorg. Med. Chem. Lett.* **2000**, 10, 2613; (b) Lang, W.; Qin, C.; Lin, S.; Khanolkar, A. D.; Goutopoulos, A.; Fan, P.; Abouzid, K.; Meng, Z.; Biegel, D.; Makriyannis, A. *J. Med. Chem.* **1999**, 42, 896.
 12. (a) Patricelli, M. P.; Lovato, M. A.; Cravatt, B. F. *Biochemistry* **1999**, 38, 9804; (b) Patricelli, M. P.; Cravatt, B. F. *Biochemistry* **1999**, 38, 14125; (c) Patricelli, M. P.; Cravatt, B. F. *J. Biol. Chem.* **2000**, 275, 19177.
 13. McKinney, M. K.; Cravatt, B. F. *J. Biol. Chem.* **2003**, 278, 37393.
 14. X-ray: Bracey, M. H.; Hanson, M. A.; Masuda, K. R.; Stevens, R. C.; Cravatt, B. F. *Science* **2002**, 298, 1793.
 15. Patricelli, M. P.; Patterson, J. E.; Boger, D. L.; Cravatt, B. F. *Bioorg. Med. Chem. Lett.* **1998**, 8, 613.
 16. Deutsch, D. G.; Lin, S.; Hill, W. A. G.; Morse, K. L.; Salehani, D.; Arreaza, G.; Omeir, R. L.; Makriyannis, A. *Biochem. Biophys. Res. Commun.* **1997**, 231, 217.
 17. (a) Deutsch, D. G.; Omeir, R.; Arreaza, G.; Salehani, D.; Prestwich, G. D.; Huang, Z.; Howlett, A. *Biochem. Pharmacol.* **1997**, 53, 255; (b) De Petrocellis, L.; Melck, D.; Ueda, N.; Maurelli, S.; Kurahashi, Y.; Yamamoto, S.; Marino, G.; Di Marzo, V. *Biochem. Biophys. Res. Commun.* **1997**, 231, 82; (c) Fernando, S. R.; Pertwee, R. G. *Br. J. Pharmacol.* **1997**, 121, 1716.
 18. Edgemond, W. S.; Greenberg, M. J.; McGinley, P. J.; Muthian, S.; Campbell, W. B.; Hillard, C. J. *J. Pharmacol. Exp. Ther.* **1998**, 286, 184.
 19. Koutek, B.; Prestwich, G. D.; Howlett, A. C.; Chin, S. A.; Salehani, D.; Akhavan, N.; Deutsch, D. G. *J. Biol. Chem.* **1994**, 269, 22937.
 20. Patterson, J. E.; Ollmann, I. R.; Cravatt, B. F.; Boger, D. L.; Wong, C.-H.; Lerner, R. A. *J. Am. Chem. Soc.* **1996**, 118, 5938.
 21. Boger, D. L.; Sato, H.; Lerner, A. E.; Austin, B. J.; Patterson, J. E.; Patricelli, M. P.; Cravatt, B. F. *Bioorg. Med. Chem. Lett.* **1999**, 9, 265.
 22. (a) Boger, D. L.; Sato, H.; Lerner, A. E.; Hedrick, M. P.; Fecik, R. A.; Miyauchi, H.; Wilkie, G. D.; Austin, B. J.; Patricelli, M. P.; Cravatt, B. F. *Proc. Natl. Acad. Sci., U.S.A.* **2000**, 97, 5044; (b) Boger, D. L.; Miyauchi, H.; Hedrick, M. P. *Bioorg. Med. Chem. Lett.* **2001**, 11, 1517; (c) Leung, D.; Hardouin, C.; Boger, D. L.; Cravatt, B. F. *Nat. Biotechnol.* **2003**, 21, 687; (d) Boger, D. L.; Miyauchi, H.; Du, W.; Hardouin, C.; Fecik, R. A.; Cheng, H.; Hwang, I.; Hedrick, M. P.; Leung, D.; Acevedo, O.; Guimaraes, C. R. W.; Jorgensen, W. L.; Cravatt, B. F. *J. Med. Chem.*, in press; (e) Lichtman, A. H.; Leung, D.; Schelton, C. C.; Saghatelian, A.; Hardouin, C.; Boger, D. L.; Cravatt, B. F. *J. Pharmacol. Exp. Ther.* **2004**, 311, 441; (f) Leung, D.; Du, W.; Hardouin, C.; Cheng, H.; Hwang, I.; Cravatt, B. F.; Boger, D. L., submitted for publication.
 23. (a) Kathuria, S.; Gaetani, S.; Fegley, D.; Valino, F.; Duranti, A.; Tontini, A.; Mor, M.; Tarzia, G.; La Rana, G.; Calignano, A.; Giustino, A.; Tattoli, M.; Palmery, M.; Cuomo, V.; Piomelli, D. *Nat. Med.* **2003**, 9, 76; (b) Tarzia, G.; Duranti, A.; Tontini, A.; Piersanti, G.; Mor, M.; Rivara, S.; Plazzi, P. V.; Park, C.; Kathuria, S.; Piomelli, D. *J. Med. Chem.* **2003**, 46, 2352.
 24. Full details are provided in [Supporting Information](#), which may be obtained upon request from the authors or online.
 25. Bobosik, V.; Piklerova, A.; Martvon, A. *Collect. Czech. Chem. Commun.* **1983**, 48, 3421.
 26. Poigny, S.; Guyot, M.; Samadi, M. *J. Chem. Soc., Perkin Trans. 1* **1997**, 2175.
 27. Ahmad, R.; Iqbal, R.; Akhtar, H.; Zia-ul-Haq; Duddeck, H.; Stefaniak, L.; Sitkowski, J. *Nucleosides, Nucleotides Nucleic Acids* **2001**, 20, 1671.
 28. Patricelli, M. P.; Lashuel, H. A.; Giang, D. K.; Kelly, J. W.; Cravatt, B. F. *Biochemistry* **1998**, 37, 15177.
 29. The corresponding epoxide exhibited a $K_i = 1.0 \pm 0.1 \mu\text{M}$.
 30. The corresponding pyridine N-oxide exhibited a $K_i = 590 \pm 50 \mu\text{M}$.