Bioorganic & Medicinal Chemistry xxx (2015) xxx-xxx

Contents lists available at ScienceDirect

Bioorganic & Medicinal Chemistry

journal homepage: www.elsevier.com/locate/bmc

Discovery and structural analyses of *S*-adenosyl-L-homocysteine hydrolase inhibitors based on non-adenosine analogs

Akira Nakao*, Hiroko Suzuki, Hiroaki Ueno[†], Hiroshi Iwasaki[†], Tomofumi Setsuta[†], Akiko Kashima, Shinji Sunada

Research Division, Mitsubishi Tanabe Pharma Corporation, 1000, Kamoshida-cho, Aoba-ku, Yokohama 227-0033, Japan

ARTICLE INFO

Article history: Received 28 March 2015 Revised 9 May 2015 Accepted 11 May 2015 Available online xxxx

Keywords: Homocysteine S-Adenosyl-L-homocysteine hydrolase cis-Amide X-ray crystal structure Intramolecular eight-membered ring hydrogen bond interaction

1. Introduction

Homocysteine (Hcy) is a sulfur-containing amino acid, which is an intermediate metabolite of an essential amino acid, methionine (Met). Met is condensed with ATP to form *S*-adenosyl-L-methionine (AdoMet), and AdoMet utilized as a methyl donor for methylation reactions is converted to *S*-adenosyl-L-homocysteine (AdoHcy). AdoHcy is hydrolyzed to Hcy and adenosine. There are two metabolic pathways of Hcy: (i) remethylation through methionine synthase or betaine–homocysteine methyltransferase (BHMT) into Met, and (ii) degradation to cysteine thorough cystathionine beta-synthase (CBS) and cystathionine γ -lyase (CTH).¹ Intracellular Hcy is highly regulated at low levels and redundant Hcy is released into the blood (see Fig. 1).

In the late 1960s, McCully reported that Hcy caused vascular pathology such as arteriosclerosis and myocardial infarction.² A meta-analysis verified a 25% lower usual Hcy level (3 μ M in absolute level) was associated with about an 11% lower ischemic heart disease and about 19% lower stroke risk.³ Elevated plasma Hcy levels are now recognized as independent risk factors for

http://dx.doi.org/10.1016/j.bmc.2015.05.018 0968-0896/© 2015 Elsevier Ltd. All rights reserved.

ABSTRACT

Optimization of a new series of S-adenosyl-L-homocysteine hydrolase (AdoHcyase) inhibitors based on non-adenosine analogs led to very potent compounds **14n**, **18a**, and **18b** with IC_{50} values of 13 ± 3 , 5.0 ± 2.0 , and 8.5 ± 3.1 nM, respectively. An X-ray crystal structure of AdoHcyase with NAD⁺ and **18a** showed a novel open form co-crystal structure. **18a** in the co-crystals formed intramolecular eight membered ring hydrogen bond formations. A single crystal X-ray structure of **14n** also showed an intramolecular eight-membered ring hydrogen bond interaction.

© 2015 Elsevier Ltd. All rights reserved.

atherothrombotic diseases such as stroke. In addition, there is evidence that elevated plasma Hcy levels associate with other neuropsychiatric disorders.^{4–6} Lowering Hcy to normal levels therefore might be one of the possible approaches to prevent and treat diseases such as coronary artery disease, ischemic stroke, and neuropsychiatric diseases.

One of the strategies to lower the Hcy levels is the inhibition of the Hcy synthetic enzyme, *S*-adenosyl-L-homocysteine hydrolase (AdoHcyase; EC 3.3.1.1.).⁷ Almost all of the known AdoHcyase inhibitors are adenosine analogs.^{8–10} Some of them inhibit the enzyme irreversibly, and many of them lack selectivity against related enzymes for producing adenosine, suggesting that there could remain concerns about adverse side effects.⁸ A reversible inhibitor **4** has only a weak potency for the enzyme ($K_i \sim 10^{-6}$ M) (see Fig. 2).⁸

We hypothesize that reversible, competitive AdoHcyase inhibitors based on non-adenosine analogs can provide some distinct advantages in terms of selectivity and toxicity. In our previous paper, we reported that a high throughput screening using Automated Ligand Identification System (ALIS) resulted in discovery of a new series of *S*-adenosyl-L-homocysteine hydrolase inhibitors based on non-adenosine analogs.^{11,12} This paper describes detailed optimization strategies for lead discovery reported in the previous paper and an SAR study with X-ray crystal structural analyses, which elucidate a novel open form co-crystal structure and intramolecular eight-membered ring hydrogen bond formations.



^{*} Corresponding author at present address: Research Division, Mitsubishi Tanabe Pharma Corporation, 2-2-50, Kawagishi, Toda-shi, Saitama 335-8505, Japan. Tel.: +81 48 433 2685; fax: +81 48 433 2650.

E-mail address: Nakao.Akira@mc.mt-pharma.co.jp (A. Nakao).

[†] Present address: Chemistry, Manufacturing and Control Division, Mitsubishi Tanabe Pharma Corporation, 3-16-89, Kashima, Yodogawa-ku, Osaka 532-8505, Japan.

A. Nakao et al./Bioorg. Med. Chem. xxx (2015) xxx-xxx

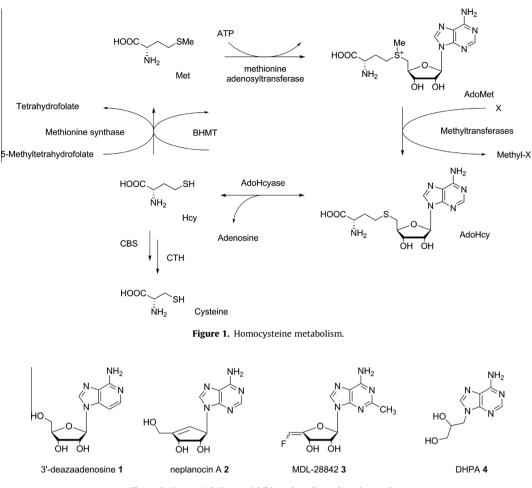


Figure 2. Known AdoHcyase inhibitors based on adenosine analogs.

The high throughput screening using ALIS led to a series of several lead candidate molecules (**5a–f**) illustrated in Figure 3. The structural features of the compounds are that they contain two amides: a hydrophobic amide and a hydrophilic amide with an amine group. **5a**, **5b**, and **5c** contained a methyl substituent at the hydrophobic amide nitrogen, and they were more potent than **5e** and **5f**, which did not have *N*-methyl groups in the hydrophobic amide parts.

In general, secondary amides present planar *trans*-peptide formations rather than *cis*-peptide formations, and the isomerization is restricted because of their partial double bond character.¹³ Once N-alkylations of the secondary amide nitrogen atoms occurred, an energy barrier between *cis* and *trans* configurations is lowered and the resulting tertiary amides can rotate to form less sterically-hindered or electrochemically stable structures.¹⁴ In the case of compound **5f**, its hydrophobic amide is presumed to form *trans*-amide conformation. We hypothesized that active conformations of **5** with AdoHcyase are *cis*-amide forms, and N-alkylation of less potent compound **5f** could easily access to *cis*-amide structure, resulting in stronger inhibitory activity against AdoHcyase (see Fig. 4). At the beginning of the optimization campaign, we addressed the replacement of the hydrophobic amide in **5** with various *N*,*N*-substituted amides, including the amide of compound **14e**.

2. Chemistry

The synthesis of intermediate **10** was shown in Scheme 1. Reaction of 1,4-dichloro-2-nitrobenzene (**6**) with 4-chlorophenol in the presence of NaH in DMF provided biphenylether **7**. Reduction of the nitro group of **7** was performed by using hydrazine hydrate as hydrogen sources with a catalytic amount of $FeCl_3$ and charcoal activated in MeOH under reflux conditions. Dialkylation of **8** with ethyl bromoacetate in *N*,*N*-diisopropylethylamine gave **9**, and hydrolysis of **9** with aqueous NaOH in MeOH and THF produced **10**.

Compounds **14a–i**, and **14n–p** were synthesized from **10** in one-pot reactions described in Scheme 2. Treatment of **10** with EDC in DMF followed by addition of various amines **12a–i** and **12n–p**, and subsequent addition of 2-(1-pyrrolidinyl)ethylamine, HOBt, and EDC gave **14a–i**, and **14n–p**, respectively. Deprotection of *N*-Boc in **14i** led to **14j**, and acylation with acetic anhydride, sulfonylation with methylsulfonyl chloride, and carbamation with methyl chloroformate resulted in **14k**, **14l**, and **14m**, respectively. Treatment of **14p** with *p*-toluenethiol in the present of Cs₂CO₃ in MeCN gave **14q**, which was converted into **14r** and **14m**, respectively.

Compounds **16a–d** and **16m–t** were synthesized from **10** through the intermediate **13e** described in Scheme 3. Treatment of **10** with EDC in DMF followed by addition of **12a** with DIPEA gave **13e** as a colorless solid. Reaction of **13e** with **15a–l** gave **16a–l**, respectively. Deprotection of *N*-Boc in **16e–l** led to **16m–t**, respectively.

Compounds **18a** and **18b** were synthesized as described in Scheme 4. Treatment of **10** with EDC in DMF followed by addition of **12n** with DIPEA, and subsequent addition of **15e** and **15f** gave **17a** and **17b**, respectively. Deprotection of *N*-Boc in **17a** and **17b**, followed by their salt formation gave **18a** and **18b**, respectively.

A. Nakao et al./Bioorg. Med. Chem. xxx (2015) xxx-xxx

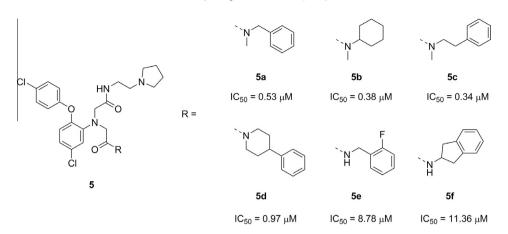


Figure 3. Representative compounds from ALIS screening and their IC₅₀ values for AdoHcyase inhibition.

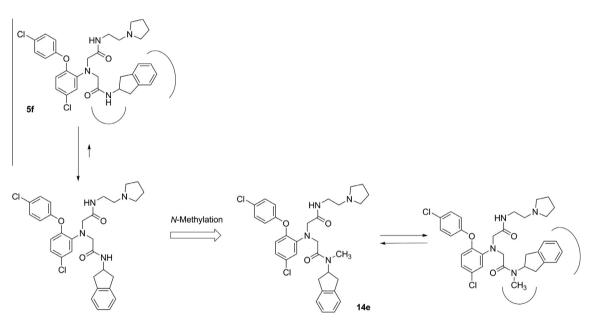
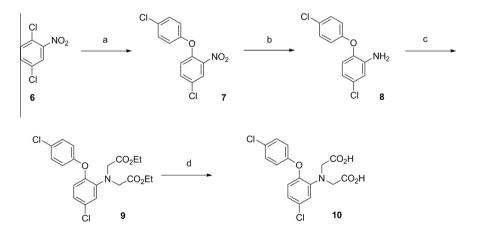
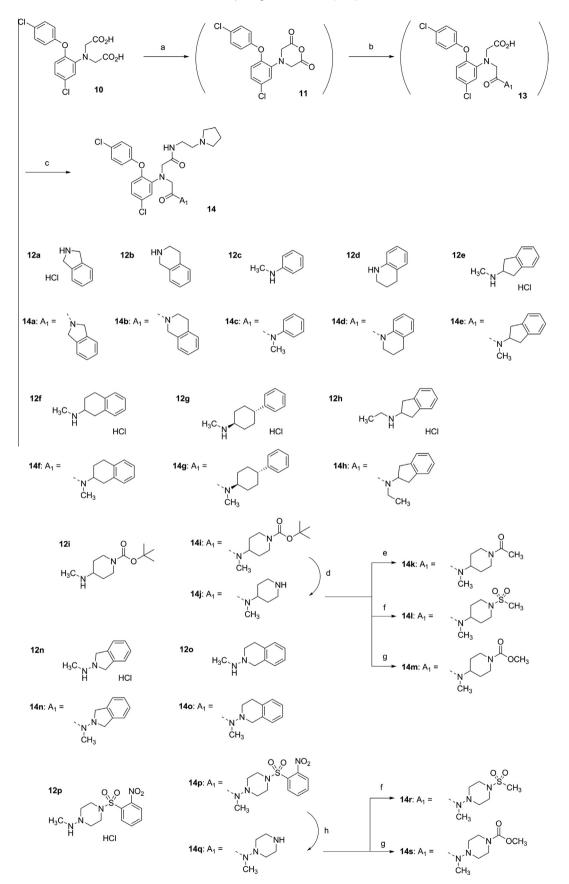


Figure 4. Proposed conformational changes by N-methylation of 5f to 14e.



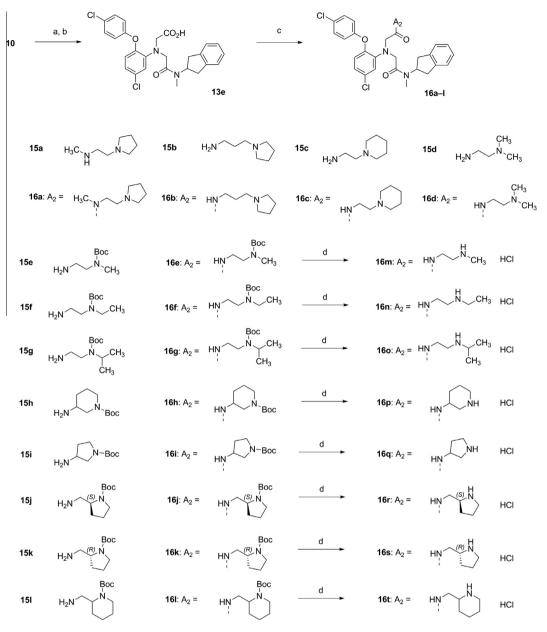
Scheme 1. Reagents and conditions: (a) 4-chlorophenol, NaH, DMF rt-80 °C; (b) FeCl₃, charcoal activated, NH₂NH₂·H₂O, MeOH, 50 °C to reflux; (c) ethyl bromoacetate, *N*,*N*-diisopropylethylamine, 140 °C; (d) 1 mol/L aq NaOH, MeOH, THF, rt.

A. Nakao et al./Bioorg. Med. Chem. xxx (2015) xxx-xxx

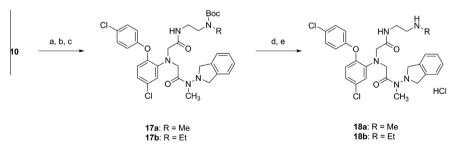


Scheme 2. Reagents and conditions: (a) EDC, DMF rt; (b) 12, or 12, DIPEA, 0 °C; (c) 2-(1-pyrrolidinyl)ethylamine, HOBt, EDC, 0 °C-rt; (d) 4 mol/L HCl in dioxane, CH₂Cl₂, rt; (e) Ac₂O, TEA, CH₂Cl₂, 0 °C-rt; (f) CH₃SO₂Cl, TEA, CH₂Cl₂, 0 °C-rt; (g) methyl chloroformate, TEA, CH₂Cl₂, 0 °C-rt; (h) *p*-toluenethiol, Cs₂CO₃, MeCN, rt.

A. Nakao et al./Bioorg. Med. Chem. xxx (2015) xxx-xxx



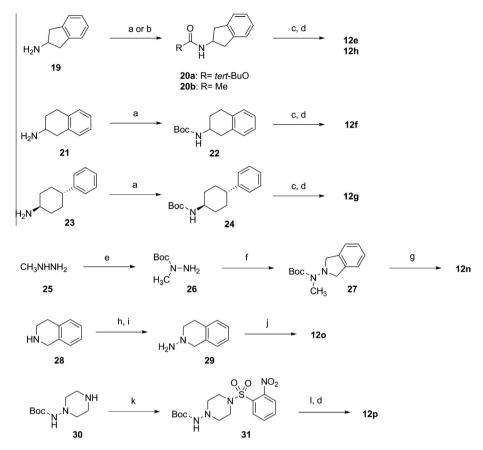
Scheme 3. Reagents and conditions: (a) EDC, DMF rt; (b) 12e, DIPEA, 0 °C; (c) 15a-I, HOBt, EDC, DMF, 0 °C-rt; (d) 4 mol/L HCl in dioxane, CH₂Cl₂, rt.



Scheme 4. Reagents and conditions: (a) EDC, DMF rt; (b) 12n, DIPEA, 0 °C; (c) 15e-f, HOBt, EDC, 0 °C-rt; (d) TFA, CH₂Cl₂, rt; (e) 4 mol/L HCl in dioxane, Et₂O, rt.

Compounds **12e–h** and **12n–p** were prepared shown in Scheme 5. Treatment of 2-aminoindane (**19**) with di*-tert*-butyl dicarbonate and acetic anhydride gave **20a** and **20b**, respectively. Reduction of **20a** and **20b** with LiAlH₄, followed by their salt formation afforded **12e** and **12h**, respectively. **12f** and **12g** were

synthesized by a similar method that used in the synthesis of **20a**. Boc protection of methylhydrazine (**25**) with di-*tert*-butyl dicarbonate, followed by cyclization with *o*-xylylene dibromide gave **27**. Deprotection of *N*-Boc in **27** afforded **12n**. Treatment of 1,2,3,4-tetrahydroisoquinoline (**28**) with sodium nitrite, followed



Scheme 5. Reagents and conditions: (a) Boc₂O, TEA, CH₂Cl₂, rt; (b) Ac₂O, TEA, CH₂Cl₂, 0 °C; (c) LiAlH₄, THF, 0 °C to reflux; (d) 4 mol/L HCl in dioxane, Et₂O, rt; (e) Boc₂O, EtOH, CH₂Cl₂, rt; (f) *o*-xylylene dibromide, TEA, NMP, 50 °C-rt; (g) 4 mol/L HCl in dioxane, CH₂Cl₂, rt; (h) NaNO₂, H₂O, AcOH, 5 °C-rt; (i) Zn, AcOH, MeOH, 0 °C-rt; (j) 37% formalin, AcOH, H₂O, 0 °C; LiAlH₄, THF, 0 °C to reflux; (k) *o*-nitrobenzenesulfonyl chloride, TEA, CH₂Cl₂, 0 °C-rt; (l) Mel, *t*-BuOK, THF, 0 °C.

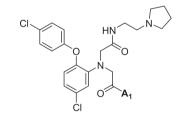
by reduction with Zn and acetic acid in MeOH gave **29**. Treatment of **29** with formalin followed by reduction with LiAlH₄ afforded **120**. Protection of **30** with *o*-nitrobenzenesulfonyl chloride gave **31**. Methylation of **31** with iodomethane, followed by removal of Boc provided **12p**.

3. Results and discussion

The compounds synthesized above were tested in an enzyme inhibition assay using human recombinant S-adenosyl-L-homocysteine hydrolase (Diazyme Laboratories) by measuring the hydrolysis activity of AdoHcy. In an effort to address the enhancement of the enzyme inhibition activity, we firstly investigated the replacement of the hydrophobic amide parts. Results are described in Table 1. Among compounds 5a-f, 5d only had a cyclic amine substituent in the hydrophobic amide parts. In order to make sure whether tertiary amides are simply required in the hydrophobic amide parts, we examined the effect of cyclic tertiary amides. Compounds 14a, and 14b decreased potency compared to 5a, and 5c. 1,2,3,4-Tetrahydroquinoline derivative 14d had similar inhibitory activity to **14c**. These results suggested that the tertiary amide bond in the hydrophobic parts play an important role in directions and locations of hydrophobic substituents. Compound 14e, designed for the conformational change of 5f, significantly increased the inhibitory activity against AdoHcyase (>200-fold). Replacement of the indane in 14e with 1,2,3,4-tetrahydronaphtalene maintained similar strong inhibitory activity (14f). On the

Table 1

AdoHcyase inhibition assay data of the conversion of tertiary amides



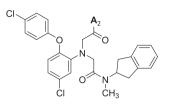
Compound	IC ₅₀ [*] (μM)
14a	>100
14b	17 ± 2
14c	2.7 ± 0.9
14d	2.3 ± 0.2
14e	0.052 ± 0.010
14f	0.081 ± 0.017
14g	0.53 ± 0.08
14h	0.64 ± 0.12
14j	>100
14k	4.6 ± 0.5
141	0.23 ± 0.05
14m	0.89 ± 0.18
14n	0.013 ± 0.003
140	0.20 ± 0.02
14r	0.049 ± 0.008
14s	0.044 ± 0.006

Each datum represents the mean ± standard error of triplicate determinations.

A. Nakao et al./Bioorg. Med. Chem. xxx (2015) xxx-xxx

Table 2

The conversion of hydrophilic amines



	IC ₅₀ * (μM)
14e	0.052 ± 0.010
16a	>100
16b	0.33 ± 0.05
16c	0.32 ± 0.05
16d	0.13 ± 0.03
16e	0.070 ± 0.012
16f	0.060 ± 0.006
16g	0.049 ± 0.005
16h	1.2 ± 0.1
16i	1.5 ± 0.2
16j	0.11 ± 0.01
16k	0.60 ± 0.07
161	0.15 ± 0.02

* Each datum represents the mean ± standard error of triplicate determinations.

Tal	ble	3	
-----	-----	---	--

Combination of the SAR on the two amide parts

	IC ₅₀ [*] (μM)
18a	0.0050 ± 0.0020
18b	0.0085 ± 0.0031

* Each datum represents the mean ± standard error of triplicate determinations.

other hand, replacement of the *N*-methyl group in **14e** with an *N*-ethyl group resulted in a 12-fold decrease in potency (**14h**). Introduction of a phenyl group into the 4-position of the cyclo-hexyl group in **5b** slightly decreased its potency (**14g**). Replacement of the cyclohexyl group in **5b** with piperidine (**14j**) led to a significant decrease in potency, but introduction of acetyl (**14k**), methanesulfonyl (**14l**), and methoxycarbonyl (**14m**) groups in **14j** regained the inhibitory activity. ¹H NMR spectra of the

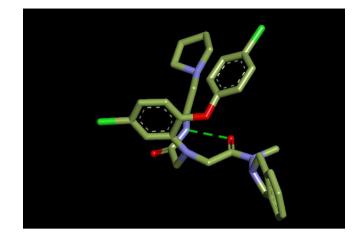


Figure 6. X-ray single crystal structure of 14n.

compounds having a methine proton next to a tertiary amide nitrogen, such as **14e**, **14f**, **14l**, and **14m** showed two distinct peaks of the methine proton, suggesting the existence of rotational isomers. In the case of **14e**, for example, the spectrum of the methine proton exhibited two multiplets of 2:3 intensities at 4.47–4.57 ppm and 5.37–5.47 ppm, respectively. The rotationally restricted isomers might have a disadvantageous effect on the interaction with the target enzyme. This result encouraged us to perform the modification of *N*-methyl amine derivatives to methylhydrazine analogs. Replacement of the indane in (**14e**) with an isoindoline (**14n**) led to a 4-fold increase in potency, and conversion of piperidine moieties (**14l**, **14m**) into piperazines (**14r**, **14s**) also improved the inhibitory activities. On the other hand, replacement of the tetrahydronaphthalene (**14f**) with a tetrahydroisoquinoline (**14o**) gave a 2.5-fold loss in potency.

Next, we conducted the conversion of hydrophilic amines with the *N*-methylindan-2-amine on the hydrophobic parts (Table 2). Nmethylation of the secondary amide in **14e** showed a marked decrease in potency (**16a**). Elongation of the methylene linker to give **16b** led to a 6-fold loss in potency. Replacement of the pyrrolidine with a piperidine (**16c**) and a dimethylamine (**16d**) showed 6 and 3 times losses in potency, respectively. The introductions of

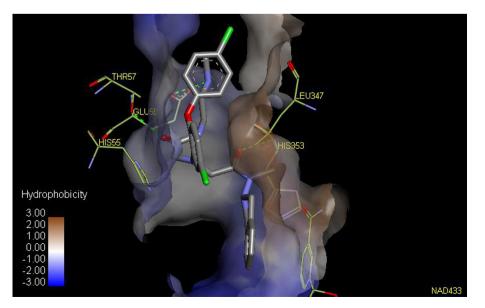


Figure 5. X-ray co-crystal structure of AdoHcyase, NAD⁺, and 18a.

N-monoalkylethylenediamines such as **16m**, **16n**, and **16o** showed comparable potency for AdoHcyase. The cyclic secondary amine series compounds **16p–t** showed weaker inhibitory activities than **14e**. Compounds **16j**, and **16l** were more potent than **16h**, **16i**, and **16k**.

Combined with the information of SAR on the two amides parts, we synthesized compounds **18a** and **18b**, which had the strongest inhibitory activities against AdoHcyase, with IC₅₀ values of 0.0050 ± 0.0020 , and $0.0085 \pm 0.0031 \mu$ M, respectively (Table 3).¹⁵

To elucidate the active conformation of our inhibitors with AdoHcyase, X-ray co-crystal structure analysis was employed. We were successful in obtaining plate-like co-crystals of the AdoHcyase complexed with the NAD⁺ coenzyme and compound 18a, which diffracted X-rays to 2.7 Å resolutions (see Fig. 5). The complex showed tetrameric structures, and each subunit contained one NAD⁺ molecule and one molecule of 18a. The AdoHcyase in the complexes has an open conformation similar to the apo enzyme structure and different from the reported X-ray co-structures.^{9b,16} **18a** was sandwiched between the catalytic domain and the NAD⁺ binding domain. The benzene ring of the isoindoline in 18a and the nicotinamide ring of the NAD⁺ were located in a parallel displaced fashion with a separation of 3.9 Å, suggesting that there should be a cation- π stacking interaction.¹⁷ 2-(4-Chlorophenoxyl)-5-chloroaniline part in 18a was positioned between the two domains, indicating hydrophobic interactions with Leu347 and π - π stacking with an imidazole substituent in His55 (3.7 Å).¹⁸ As expected, an active conformation of **18a** with AdoHcyase was a *cis*-amide form. There were four hydrogen bond interactions: (i) between the carbonyl oxygen of the hydrophobic amide part in **18a** and the amide of His353 in the NAD⁺ binding domain (2.8 Å), (ii) between the carbonyl oxygen of the hydrophilic amide part in 18a and the hydroxyl group of the side chain of Thr57 in the catalytic domain (2.5 Å), (iii) between the N-methylamine of the hydrophilic amide part in **18a** and the carboxyl group of the side chain of Glu59 in the catalytic domain (2.8 Å), and (iv) between the carbonyl oxygen of the hydrophobic amide part and the NH hydrogen of the hydrophilic amide part in **18a** (2.9 Å) as an unexpected intramolecular eight-membered ring hydrogen bond interaction.¹⁹ The loss of activity observed for compound 16a was consistent with the destruction of the intramolecular hydrogen bond interaction by N-methylation of 14e. The reduced activity observed for 14h was explained by the limited size of a hydrophobic pocket around the *N*-methyl amide region in **14e**. The increased activity observed for **14n** was thought to be due to the enhancement of a cation- π interaction by a small angle change from the indane in **14e** to the isoindoline in **14n**.

We speculated that the intramolecular eight-membered ring hydrogen bond formation in **18a** was induced only by the interaction of compound **18a** with the AdoHcyase and the NAD⁺. To solve the molecular conformation of **18a** itself, we made an effort to obtain a single crystal of **18a** or other derivatives. As a result, a single crystal of **18a** was not obtained, however a single crystal of compound **14n** having a very similar structure to **18a** was fortunately obtained as a colorless prismatic crystal from diethyl ether. The single X-ray crystal structure revealed that **14n** formed not only a *cis*amide formation, but also an intramolecular eight-membered ring hydrogen bond interaction (2.9 Å) similar to **18a** in the X-ray cocrystal structure (see Fig. 6), suggesting that the intramolecular eight-membered ring hydrogen bond interaction would be formed at least under hydrophobic environmental conditions such as binding pockets in enzymes or nonpolar solvents (see Fig. 6).²⁰

4. Conclusions

In conclusion, we identified a novel series of *S*-adenosyl-L-homocysteine hydrolase inhibitors based on non-adenosine analogs through ALIS screening. During the optimization campaign, we hypothesized that active conformations of the inhibitors with AdoHcyase were *cis*-amide forms. Our effort led to very potent compounds **18a** and **18b** with IC_{50} values of 5.0 ± 2.0 , and 8.5 ± 3.1 nM, respectively. A novel open form X-ray co-structure of AdoHcyase, **18a**, and NAD⁺ revealed that the active conformation of compound **18a** was a *cis*-amide form, and there were three intermolecular and one intramolecular hydrogen bond interactions. A single X-ray crystal structure of compound **14n** showed an intramolecular eight-membered ring hydrogen bond interaction seen in **18a**. Compound **18a** and **18b** could be a promising lead compound for research to reduce elevated homocysteine levels, and the information of the X-ray crystal structures will help our further lead optimization.

5. Experimentals

5.1. Chemistry

5.1.1. General

All melting points were obtained on a Büchi 535 melting point apparatus and are uncorrected. Silica gel column chromatography was performed on a SHOKO Scientific Purif- $\alpha 2$ Flash Chromatography System using Purif-Pack silica gel columns or Yamazen Hi-Flash columns, and the described solvents as eluent under gradient condition. ¹H and ¹³C NMR spectra were recorded on a Bruker ARX300 spectrometer (300 MHz) or a Bruker AVANCE400 (400 MHz) spectrometer. Chemical shifts are expressed in parts per million (ppm, δ units) relative to tetramethylsilane (TMS) as an internal standard, and the following abbreviations are used: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, dd = double doublet, dt = double triplet, br s = broad singlet. Mass spectra were measured in a combination with a Waters Acquity UPLC system (Acquity BEH C18 2.0×50 mm, mobile phase: A: H₂O + 0.05% HCOOH; B: CH₃CN + 0.05% HCOOH, a gradient of 5-98% B over 1 min, flow rate 0.6 mL/min, column temperature 40 °C) and a Micromass ZQ mass spectrometer in electrospray ionization (ESI) positive mode. Elemental analyses were performed on a Perkin-Elmer 2400 II CHN Elemental Analyzer. High resolution mass spectroscopy (HRMS) was measured in a combination with a Dionex UltiMate 3000 HPLC system (YMC Hydrosphere C18 (3 μ m) 2.0 \times 75 mm, mobile phase: A: H₂O + 0.1% HCOOH; B: CH₃CN, A/B = 50/50 over 3 min, flow rate 0.2 mL/min, column temperature 40 °C) and a Thermo Fisher Scientific LTQ Orbitrap Velos Pro mass spectrometer in ESI positive mode. Purity was determined by HPLC measured by an Agilent 1100 system (Sumipax ODS D-210SLP (3 $\mu m)$ 4.6 \times 50 mm, mobile phase: A: H₂O + 0.05% TFA; B: CH₃CN + 0.05% TFA, A/B = 40/60, 45/55, 50/50, 55/45, 60/40, or 70/30 over 12 min, flow rate 1.0 mL/min, column temperature 40 °C), and was >95% for all tested compounds. All chemicals and solvents were of reagent grade unless otherwise specified. The following abbreviations are used: DMF, N,N-dimethylformamide; DMSO, dimethyl sulfoxide; TEA, triethylamine; THF, tetrahydrofuran; EDC, 1-[3-(dimethylamino)propyl]-3-ethylcarbodiimide hydrochloride; HOBt, 3hydroxybenztriazole.

5.1.2. 4-Chloro-1-(4-chlorophenoxy)-2-nitrobenzene (7)²¹

To a solution of 4-chlorophenol (10.6 g, 82.5 mmol) in DMF (100 mL) was added NaH (3.32 g, 60% in mineral oil, 83.0 mmol), and the resulting mixture was stirred at room temperature for 40 min. To the mixture was added 1,4-dichloro-2-nitrobenzene (**6**) (14.5 g, 75.5 mmol), and the mixture was stirred at 80 °C for 75 min. The reaction mixture was cooled down, and poured into ice water (400 mL). The precipitated solid was filtered, washed with H₂O (100 mL), 1 mol/L aqueous NaOH (100 mL × 2), and H₂O (100 mL), and dried under reduced pressure to give **7**

(21.8 g, 93%) as a yellow solid. Mp 74–76 °C. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 6.98 (d, *J* = 8.7, 2H), 6.99 (d, *J* = 9.3, 1H), 7.35 (d, *J* = 8.7, 2H), 7.49 (dd, *J* = 2.6, 9.3, 1H), 7.96 (d, *J* = 2.6, 1H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 120.30, 121.81, 125.76, 128.85, 130.15, 130.23, 134.24, 141.50, 148.92, 154.22. MS (APCI) *m*/*z* 284, 286 [M+H]⁺.

5.1.3. 5-Chloro-2-(4-chlorophenoxy)aniline (8)²²

A solution of 7 (12.0 g, 42.2 mmol), FeCl₃ (702 mg, 4.33 mmol), and activated charcoal (3.70 g) in MeOH (360 mL) was stirred at 50 °C, and to the mixture was added hydrazine monohydrate (7.00 mL, 144 mmol) over 10 min, and then stirred under reflux for 1 h. The reaction mixture was cooled down, filtered through Celite, washed with MeOH, and the resulting solution was concentrated under reduced pressure. The residue oil was diluted with EtOAc, and the organic layer was washed with saturated aqueous NaHCO₃, brine, and dried over Na₂SO₄. The insoluble material was filtered off, and the solution was concentrated under reduced pressure to give **8** (10.6 g, 99%) as a colorless solid. Mp 59–61 °C. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 3.85 (br s, 2H), 6.67 (dd, J = 2.6, 8.2, 1H), 6.76 (d, J = 8.2, 1H), 6.80 (d, J = 2.6, 1H), 6.89 (d, J = 8.7, 2H), 7.26 (d, I = 8.7, 2H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 116.07, 118.33, 118.42, 121.04, 127.98, 129.76, 130.17, 139.75, 141.32, 155.81. LC-MS (ESI) m/z 254, 256 [M+H]⁺.

5.1.4. *N*-[5-Chloro-2-(4-chlorophenoxy)phenyl]iminodiacetic acid diethyl ester (9)

A solution of **8** (2.56 g, 10.1 mmol), ethyl bromoacetate (7.0 mL, 63 mmol), and *N*,*N*-diisopropylethylamine (7.0 mL, 41 mmol) was stirred at 140 °C for 5 h. The reaction mixture was cooled down, and diluted with EtOAc (100 mL). The organic layer was washed with 10% aqueous citric acid (100 mL × 2), saturated aqueous NaHCO₃, and brine, and dried over MgSO₄. The insoluble material was filtered off, and the solution was concentrated under reduced pressure. The residue oil was purified by silica gel column chromatography (hexane/EtOAc = 95:5 to 80:20) to afford **9** (3.68 g, 85%) as a pale yellow oil. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.20 (t, *J* = 7.2, 6H), 4.10 (s, 4H), 4.11 (q, *J* = 7.2, 4H), 6.76 (d, *J* = 8.7, 1H), 6.81–6.87 (m, 4H), 7.24 (d, *J* = 8.7, 2H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 14.15, 54.04, 60.95, 118.72, 119.42, 121.65, 122.31, 127.95, 129.56, 129.94, 142.40, 145.35, 155.99, 170.52. LC–MS (ESI) *m*/*z* 426, 428 [M+H]⁺.

5.1.5. *N*-[5-Chloro-2-(4-chlorophenoxy)phenyl]iminodiacetic acid (10)

To a solution of **9** (3.68 g, 8.63 mmol) in MeOH (20 mL) and THF (10 mL) was added 1 mol/L aqueous NaOH (30 mL, 30 mmol), and the resulting mixture was stirred at room temperature for 2 h. To the reaction mixture was added 1 mol/L aqueous HCl (40 mL, 40 mmol), and the precipitated solid was filtered, washed with H₂O, and dried under reduced pressure to give **10** (2.90 g, 91%) as a colorless solid. Mp 192–194 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ (ppm) 4.05 (s, 4H), 6.82–6.88 (m, 3H), 6.91 (d, *J* = 8.7, 2H), 7.36 (d, *J* = 8.7, 2H), 12.54 (br s, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ (ppm) 53.50, 118.10, 118.73, 120.01, 122.99, 126.42, 128.73, 129.41, 142.55, 143.94, 155.93, 171.47. LC–MS (ESI) *m/z* 370, 372 [M+H]⁺. Anal. Calculated for C₁₆H₁₃Cl₂NO₅: C, 51.91; H, 3.54; N, 3.78; Cl, 19.15. Found: C, 51.99; H, 3.49; N, 3.75; Cl, 19.07.

5.1.6. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[1,3-dihydro-2*H*-isoindol-2-yl(methyl)amino]-2-oxoethyl}- N^1 -(2-pyrrolidin-1-ylethyl)glycinamide (14n)

To a solution of **10** (371 mg, 1.00 mmol) in DMF (5 mL) was added EDC (195 mg, 1.02 mmol), and the resulting mixture was stirred at room temperature for 1 h. Then, to the reaction mixture stirred under ice-cooling was added **12n** (192 mg, 1.04 mmol) and

N,N-diisopropylethylamine (0.180 mL, 1.06 mmol), and the mixture was stirred at 0 °C for 1 h. To the reaction mixture was added 1-(2-aminoethyl)pyrrolidine (0.140 mL, 1.12 mmol). HOBt (174 mg, 1.14 mmol), and EDC (220 mg, 1.15 mmol), and the resulting mixture was stirred at room temperature for 6 h. The reaction mixture was diluted with EtOAc, and the organic layer was washed with H₂O, saturated aqueous NaHCO₃, and brine, and dried over Na₂SO₄. The insoluble material was filtered off, and the solution was concentrated under reduced pressure. The residue oil was purified by NH silica gel column chromatography (hexane/EtOAc = 50:50 to 0:100) to afford **14n** (497 mg, 83%) as a colorless solid. Mp 96–100 °C. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.63-1.74 (m, 4H), 2.34-2.48 (m, 6H), 2.88 (s, 3H), 3.25 (q, J = 6.4, 2H), 3.95 (s, 2H), 4.07 (d, J = 11.3, 2H), 4.20 (d, J = 11.3, 2H), 4.40 (s, 2H), 6.77 (d, J = 8.2, 1H), 6.79-6.87 (m, 3H), 6.98 (d, J = 2.1, 1H), 7.18–7.30 (m, 6H), 8.06 (t, J = 5.4, 1H). ¹³C NMR $(100 \text{ MHz}, \text{ CDCl}_3)$: δ (ppm) 23.50, 24.70, 38.36, 53.98, 54.76, 54.92. 55.45, 58.66, 118.49, 118.82, 121.23, 122.61, 122.89, 127.59, 127.85, 129.69, 130.39, 136.97, 142.28, 144.75, 156.65, 170.08, 172.75. LC-MS (ESI) m/z 596, 598 [M+H]⁺. HPLC purity: 98.87%. HRMS (ESI) m/z calculated for $C_{31}H_{36}Cl_2N_5O_3$ [M+H]⁺ 596.21897, found 596.21902.

9

5.1.7. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -[2-(1,3-dihydro-2*H*-isoindol-2-yl)-2-oxoethyl]- N^1 -(2-pyrrolidin-1-ylethyl)glycinamide (14a)

The title compound was synthesized according to the method described for the synthesis of **14n** to afford **14a** (350 mg, 72%) as a colorless oil. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.73–1.94 (m, 4H), 2.40–2.95 (m, 6H), 3.35 (q, *J* = 6.2, 2H), 4.07 (s, 2H), 4.37 (br s, 2H), 4.64 (s, 2H), 4.66 (s, 2H), 6.73–6.82 (m, 4H), 6.86–6.92 (m, 1H), 7.14 (d, *J* = 9.3, 2H), 7.18–7.25 (m, 1H), 7.26–7.35 (m, 3H), 8.75 (broad, 1H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 23.42, 37.27, 51.51, 52.38, 53.96, 54.66, 56.10, 59.26, 118.07, 120.97, 122.66, 122.94, 123.24, 127.75, 127.79, 128.02, 129.61, 130.66, 135.58, 141.59, 143.79, 156.56, 169.09, 170.59. LC–MS (ESI) *m/z* 567, 569 [M+H]⁺. HPLC purity 96.98%. HRMS (ESI) *m/z* calculated for C₃₀H₃₃Cl₂N₄O₃ [M+H]⁺ 567.19242, found 567.19306.

5.1.8. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[3,4-dihydroisoquinolin-2(1*H*)-yl]-2-oxoethyl}- N^1 -(2-pyrrolidin-1-ylethyl)glycinamide (14b)

The title compound was synthesized according to the method described for the synthesis of **14n** to afford **14b** (548 mg, 81%) as a colorless oil. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.65–1.77 (m, 4H), 2.36–2.59 (m, 6H), 2.76, 2.79 (2t, *J* = 6.2, 2H), 3.32 (q, *J* = 6.2, 2H), 3.46 (t, *J* = 6.2, 1.2H), 3.67 (t, *J* = 6.2, 0.8H), 3.98 (s, 2H), 4.28, 4.29 (2s, 2H), 4.40 (s, 0.8H), 4.55 (s, 1.2H), 6.68–6.75 (m, 3H), 6.76–6.82 (m, 1H), 6.88–6.95 (m, 1H), 6.97–7.25 (m, 6H), 8.39 (br s, 1H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 23.53, 28.23, 28.97, 38.27, 40.11, 42.24, 44.26, 46.16, 54.00, 54.83, 55.92, 58.97, 118.32, 118.65, 121.12, 122.76, 122.81, 126.01, 126.57, 126.60, 126.81, 126.93, 127.26, 127.91, 128.17, 128.99, 129.62, 129.67, 130.51, 131.58, 132.96, 133.75, 134.79, 141.58, 141.63, 144.26, 144.33, 156.28, 156.33, 168.59, 168.70, 169.86. LC–MS (ESI) *m/z* 581, 583 [M+H]⁺. HPLC purity: 96.39%. HRMS (ESI) *m/z* calculated for C₃₁H₃₅Cl₂N₄O₃ [M+H]⁺ 581.20807, found 581.20863.

5.1.9. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[methy-(phenyl)amino]-2-oxoethyl}- N^1 -(2-pyrrolidin-1-ylethyl)glycinamide (14c)

The title compound was synthesized according to the method described for the synthesis of **14n** to afford **14c** (548 mg, 86%) as a colorless oil. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.65–1.81 (m, 4H), 2.31–2.59 (m, 6H), 3.21 (s, 3H), 3.23 (q, *J* = 6.2, 2H), 3.82 (s, 2H), 3.88 (s, 2H), 6.73–6.84 (m, 5H), 7.13 (d, *J* = 7.7, 2H),

A. Nakao et al./Bioorg. Med. Chem. xxx (2015) xxx-xxx

7.22–7.28 (m, 2H), 7.35–7.49 (m, 3H), 8.21 (br s, 1H). 13 C NMR (100 MHz, CDCl₃): δ (ppm) 23.51, 37.57, 38.12, 53.98, 54.70, 55.49, 58.87, 118.59, 118.79, 121.52, 122.80, 124.34, 127.00, 127.95, 128.58, 129.70, 130.21, 130.27, 141.75, 142.28, 145.05, 156.50, 169.49, 170.11. LC–MS (ESI) *m*/*z* 555, 557 [M+H]⁺. HPLC purity: 99.50%. HRMS (ESI) *m*/*z* calculated for C₂₉H₃₃Cl₂N₄O₃ [M+H]⁺ 555.19242, found 555.19251.

5.1.10. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[3,4-dihydroquinolin-1(2H)-yl]-2-oxoethyl}- N^1 -(2-pyrrolidin-1-ylethyl)glycinamide (14d)

The title compound was synthesized according to the method described for the synthesis of **14n** to afford **14d** (970 mg, 77%) as a colorless oil. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.63–1.85 (m, 4H), 1.87–1.98 (m, 2H), 2.33–2.69 (m, 6H), 2.76 (t, *J* = 6.7, 2H), 3.23 (d, *J* = 6.2, 2H), 3.74 (br s, 2H), 3.96 (s, 2H), 4.30 (br s, 2H), 6.48–6.76 (m, 3H), 6.75 (d, *J* = 8.7, 1H), 6.81 (dd, *J* = 2.6, 8.7, 1H), 6.92–7.18 (broad, 1H), 7.15–7.31 (m, 5H), 8.30 (br s, 1H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 23.50, 23.79, 26.66, 37.93, 43.19, 54.01, 54.71, 55.11, 59.02, 118.24, 121.48, 123.16, 124.34, 126.41, 127.81, 129.02, 129.66, 130.38, 137.74, 141.80, 144.69, 156.52, 169.20, 170.24. LC–MS (ESI) *m*/*z* 581, 583 [M+H]⁺. HPLC purity: 99.42%. HRMS (ESI) *m*/*z* calculated for C₃₁H₃₅Cl₂N₄O₃ [M+H]⁺ 581.20807, found 581.20873.

5.1.11. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[2,3-dihydro-1*H*-inden-2-yl(methyl)amino]-2-oxoethyl}- N^1 -(2-pyrrolidin-1-ylethyl)glycinamide (14e)

The title compound was synthesized according to the method described for the synthesis of 14n to afford 14e (502 mg, 67%) as a colorless oil. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.65–1.81 (m, 4H), 2.37-2.59 (m, 6H), 2.66 (s, 1.8H), 2.70 (s, 1.2H), 2.78 (dd, *J* = 6.2, 16.4, 1.2H), 2.95 (dd, *J* = 6.2, 16.4, 0.8H), 3.04 (dd, *J* = 8.2, 16.4, 0.8H), 3.10 (dd, J = 8.2, 16.4, 1.2H), 3.32 (q, J = 6.2, 2H), 3.98 (s, 2H), 4.19 (s, 1.2H), 4.31 (s, 0.8H), 4.47-4.57 (m, 0.4H), 5.37-5.47 (m, 0.6H), 6.75 (d, J = 8.2, 2H), 6.78–6.98 (m, 3H), 7.14–7.23 (m, 4H), 7.28 (d, J = 8.2, 2H), 8.25, 8.36 (2 broad, 1H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 23.52, 27.92, 28.98, 36.11, 36.35. 38.23, 53.15, 54.00, 54.80, 55.64, 55.93, 56.12, 58.75, 59.00, 118.53, 118.60, 119.03, 121.07, 121.39, 122.68, 122.81, 124.44, 124.49, 126.85, 127.16, 128.06, 129.78, 129.81, 130.44, 130.50, 139.98, 140.91, 141.64, 141.79, 144.36, 144.75, 156.42, 156.48, 169.21, 169.57, 169.98. LC-MS (ESI) m/z 595, 597 [M+H]⁺. HPLC purity: 97.75%. HRMS (ESI) m/z calculated for C₃₂H₃₇Cl₂N₄O₃ [M+H]⁺ 595.22372, found 595.22444.

5.1.12. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[methyl(1,2,3,4-tetrahydronaphtalen-2-yl)amino]-2-oxoethyl}- N^1 -(2-pyrrolidin-1-ylethyl)glycinamide (14f)

The title compound was synthesized according to the method described for the synthesis of 14n to afford 14f (565 mg, 67%) as a colorless oil. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.64-2.00 (m, 6H), 2.36-2.76 (m, 8H), 2.78, 2.80 (2s, 3H), 2.84-3.00 (m, 2H), 3.24-3.37 (m, 2H), 3.74-3.85 (m, 0.4H), 3.98, 4.01 (2s, 2H), 4.30 (br s, 2H), 4.65-4.76 (m, 0.6H), 6.74 (d, J=8.7, 1H), 6.76-6.95 (m, 4H), 6.98-7.17 (m, 4H), 7.23-7.30 (m, 2H), 8.1-8.9 (broad, 1H). ¹³C NMR (100 MHz, $CDCl_3$): δ (ppm) 23.51, 26.75, 27.24, 27.85, 28.47, 29.18, 29.26, 31.76, 32.94, 37.94, 50.19, 52.74, 54.01, 54.77, 55.67, 56.24, 58.89, 59.25, 118.19, 118.74, 118.77, 120.71, 121.27, 122.58, 122.80, 125.90, 126.07, 126.20, 126.50, 128.07, 128.12, 128.67, 128.78, 129.27, 129.78, 130.38, 130.45, 133.97, 134.76, 134.82, 135.33, 141.49, 141.75, 144.31, 144.81, 156.41, 156.58, 169.41, 169.72, 170.16. LC-MS (ESI) m/z 609, 611 [M+H]⁺. HPLC purity: 98.34%. HRMS (ESI) m/z calculated for $C_{33}H_{39}Cl_2N_4O_3$ [M+H]⁺ 609.23937, found 609.24009.

5.1.13. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[methyl(*trans*-4-phenylcyclohexyl)amino]-2-oxoethyl}- N^1 -(2-pyrrolidin-1-ylethyl)glycinamide (14g)

The title compound was synthesized according to the method described for the synthesis of 14n to afford 14g (366 mg, 87%) as a colorless solid. Mp 151–153 °C. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.44-1.84 (m, 10H), 1.91-2.06 (m, 2H), 2.36-2.67 (m, 7H), 2.75, 2.77 (2s, 3H), 3.31 (q, J = 6.2, 2H), 3.37-3.48 (m, 0.35H), 3.99 (s, 2H), 4.21 (s, 1.3H), 4.29 (s, 0.7H), 4.35-4.44 (m, 0.65H), 6.75 (d, J = 8.7, 1H), 6.77-6.95 (m, 4H), 7.15-7.35 (m, 7H), 8.42 (broad, 1H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 23.50, 27.38, 28.51, 29.58, 30.63, 33.07, 33.20, 37.96, 43.19, 43.55, 52.72, 53.99, 54.74, 54.79, 55.46, 55.71, 55.90, 58.88, 59.11, 118.48, 118.58, 120.92, 121.22, 122.80, 122.91, 126.22, 126.39, 126.64, 126.73, 127.97, 128.45, 128.53, 129.68, 129.75, 130.42, 130.49, 141.66, 141.89, 144.22, 144.72, 145.77, 146.32, 156.52, 156.56, 169.14, 169.30, 170.17. LC-MS (ESI) *m*/*z* 637, 639 [M+H]⁺. HPLC purity: 99.71%. HRMS (ESI) m/z calculated for $C_{35}H_{43}Cl_2N_4O_3$ [M+H]⁺ 637.27067, found 637.27143.

5.1.14. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[2,3-dihydro-1*H*-inden-2-yl(ethyl)amino]-2-oxoethyl}- N^1 -(2-pyrrolidin-1-ylethyl)glycinamide (14h)

The title compound was synthesized according to the method described for the synthesis of **14n** to afford **14h** (351 mg, 46%) as a colorless amorphous solid. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.08 (t, J = 7.2, 1.5H), 1.14 (t, J = 7.2, 1.5H), 1.64–1.76 (m, 4H), 2.10-2.31 (m, 2H), 2.37-2.53 (m, 4H), 2.87-3.11 (m, 4H), 3.14-3.39 (m, 4H), 3.98 (s, 2H), 4.21 (s, 1H), 4.26 (s, 1H), 4.41-4.56 (m, 0.5H), 4.94–5.06 (m, 0.5H), 6.72–6.98 (m, 3H), 6.85 (d, J=9.3, 2H), 7.11-7.24 (m, 4H), 7.27 (d, J = 9.3, 2H), 8.16-8.26 (m, 0.5H), 8.27–8.36 (m, 0.5H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 14.33, 15.85, 23.51, 36.47, 36.91, 37.62, 38.21, 39.02, 53.99, 54.82, 55.71, 55.83, 56.07, 56.83, 58.76, 59.02, 118.72, 118.83, 119.07, 121.14, 121.34, 122.64, 122.74, 124.43, 124.53, 126.73, 127.17, 128.10, 129.76, 129.80, 130.39, 139.89, 140.89, 141.75, 141.85, 144.80, 156.43, 156.52, 168.96, 169.62, 170.04. LC-MS (ESI) m/z 609. 611 [M+H]⁺. HPLC purity: 98.56%. HRMS (ESI) *m*/*z* calculated for C₃₃H₃₉Cl₂N₄O₃ [M+H]⁺ 609.23937, found 609.23984.

5.1.15. N^2 -{2-{[1-(*tert*-Butoxycarbonyl)piperidin-4-yl](methyl)amino}-2-oxoethyl}- N^2 -[5-chloro-2-(4-chlorophenoxy)phenyl]- N^1 -(2-pyrrolidin-1-ylethyl)glycinamide (14i)

The title compound was synthesized according to the method described for the synthesis of **14n** to afford **14i** (351 mg, 94%) as a colorless amorphous solid. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.30–1.60 (m, 2H), 1.44 (s, 9H), 1.60–1.80 (m, 4H), 1.80–1.90 (m, 2H), 2.30–2.50 (m, 6H), 2.50–2.80 (m, 5H), 3.28 (q, *J* = 6.6, 2H), 3.35–3.50 (m, 0.2H), 3.96 (s, 2H), 4.05–4.30 (m, 4H), 4.30–4.50 (m, 0.8H), 6.65–6.90 (m, 5H), 7.25 (d, *J* = 8.7, 2H), 8.15 (broad t, 1H). LC–MS (ESI) *m*/*z* 662, 664 [M+H]⁺.

5.1.16. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[methyl(piperidin-4-yl)amino]-2-oxoethyl}- N^1 -(2-pyrrolidin-1ylethyl)glycinamide (14j)

To a solution of **14i** (650 mg, 0.980 mmol) in CHCl₃ (20 mL) was added 4 mol/L HCl in dioxane (2.0 mL, 8.0 mmol), and the resulting mixture was stirred at room temperature for 6 h. The reaction mixture was poured into diluted aqueous NaOH, and extracted with CHCl₃. The organic layer was washed with brine, dried over MgSO₄. The insoluble material was filtered off, and the solution was concentrated under reduced pressure to afford **14j** (387 mg, 70%) as a pale yellow solid. Mp 125–127 °C. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.40–1.60 (m, 4H), 1.63–1.73 (m, 4H), 1.73–2.12 (broad, 1H), 2.37–2.53 (m, 6H), 2.54–2.72 (m, 2H), 2.72, 2.74 (2s, 3H), 3.06–3.20 (m, 2H), 3.28 (q, *J* = 6.6, 2H), 3.32–3.43 (m, 0.3H),

3.96 (s, 2H), 4.15 (s, 1.4H), 4.21 (s, 0.6H), 4.33–4.45 (m, 0.7H), 6.74 (d, J = 8.7, 1H), 6.77–6.95 (m, 4H), 7.25 (d, J = 8.7, 2H), 8.24 (broad t, 1H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 23.53, 27.50, 28.52, 29.90, 31.05, 38.34, 45.93, 46.07, 51.67, 54.04, 54.60, 54.87, 55.62, 55.73, 58.78, 58.96, 118.48, 118.52, 118.82, 119.00, 121.16, 121.38, 122.81, 128.01, 129.69, 129.78, 130.48, 141.72, 141.92, 144.44, 144.80, 156.49, 168.93, 169.08, 169.91. LC–MS (ESI) m/z 562, 564 [M+H]⁺. HPLC purity: 98.94%. HRMS (ESI) m/z calculated for C₂₈H₃₈Cl₂N₅O₃ [M+H]⁺ 562.23462, found 562.23404.

5.1.17. N^2 -{2-[(1-Acetylpiperidin-4-yl)(methyl)amino]-2-oxoethyl}- N^2 -[5-chloro-2-(4-chlorophenoxy)phenyl]- N^1 -(2-pyrrolidin-1-ylethyl)glycinamide (14k)

To a solution of 14j (102 mg, 0.131 mmol) and TEA (0.060 mL, 0.43 mmol) in CH_2Cl_2 (3 mL) was added Ac_2O (0.030 mL, 0.32 mmol), and the resulting mixture was stirred at room temperature for 5 h. The reaction mixture was diluted with EtOAc, and the organic layer was washed with saturated aqueous NaHCO3 and brine, and dried over MgSO4. The insoluble material was filtered off, and the solution was concentrated under reduced pressure. The residue oil was purified by NH silica gel column chromatography (EtOAc/MeOH = 100:0 to 95:5) to afford 14k (118 mg, quant.) as a colorless oil. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.40–1.60 (m, 4H), 1.64–1.76 (m, 4H), 2.10 (s, 3H), 2.37-2.48 (m, 6H), 2.49-2.59 (m, 1H), 2.68 (s, 3H), 2.96-3.17 (m, 1H), 3.21-3.35 (m, 2H), 3.43-3.60 (m, 0.2H), 3.80-3.90 (m, 1H), 3.96 (s, 2H), 4.15 (s, 2H), 4.46-4.59 (m, 0.8H), 4.68–4.80 (m, 1H), 6.75 (d, J=8.7, 1H), 6.79–6.91 (m, 4H), 7.26 (d, J = 8.7, 2H), 8.04–8.15 (m, 1H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 21.42, 23.52, 28.36, 28.53, 29.24, 38.39, 40.87, 45.68, 51.22, 54.03, 54.88, 55.61, 58.61, 58.77, 118.48, 118.57, 119.00, 121.39, 121.60, 122.78, 128.12, 129.81, 130.50, 141.64, 141.79, 144.56, 144.93, 156.42, 168.84, 169.33, 169.78. LC-MS (ESI) *m*/*z* 604, 606 [M+H]⁺. HPLC purity: 98.24%. HRMS (ESI) m/z calculated for $C_{30}H_{40}Cl_2N_5O_4$ [M+H]⁺ 604.24519, found 604.24597.

5.1.18. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-{methyl-[1-(methylsulfonyl)piperidin-4-yl]amino}-2-oxoethyl}- N^1 -(2-pyrrolidin-1-ylethyl)glycinamide (141)

To a solution of 14j (350 mg, 0.620 mmol) and TEA (0.130 mL, 0.933 mmol) in CH₂Cl₂ (20 mL) was added methanesulfonyl chloride (0.060 mL, 0.775 mmol), and the resulting mixture was stirred overnight at room temperature. To the reaction mixture was added H₂O (100 mL), and extracted with CHCl₃ (100 mL). The organic layer was washed with brine, and dried over MgSO₄. The insoluble material was filtered off, and the solution was concentrated under reduced pressure. The residue oil was purified by silica gel column chromatography (CHCl₃/MeOH = 95:5 to 90:10) to afford **14l** (397 mg, 100%) as a colorless oil. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.52–1.61 (m, 2H), 1.65-1.77 (m, 2H), 1.79-2.00 (m, 2H), 2.50-3.04 (m, 8H), 2.74 (s, 3H), 2.79 (s, 3H), 3.29 (q, J = 6.6, 2H), 3.27-3.44 (m, 2H), 3.56-3.64 (m, 0.2H), 3.82-3.95 (m, 2H), 4.01 (s, 1.6H), 4.07 (s, 0.4H), 4.30 (s, 1.6H), 4.35-4.46 (m, 0.8H), 4.59 (br s, 0.4H), 6.73 (d, J = 8.2, 1H), 6.75-6.87 (m, 2H), 6.82 (d, J = 9.0, 2H, 7.26 (d, J = 9.0, 2H), 8.60 (broad, 1H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 23.38, 23.45, 27.57, 28.36, 28.67, 29.38, 29.70, 35.16, 35.31, 37.05, 45.54, 50.73, 53.46, 53.92, 53.98, 54.60, 55.98, 56.10, 59.02, 118.19, 118.67, 118.74, 120.90, 122.90, 127.99, 128.15, 129.69, 129.81, 130.46, 141.42, 141.75, 144.25, 156.51, 156.61, 170.00, 170.59. LC-MS (ESI) m/z 640, 642 [M+H]⁺. HPLC purity: 99.20%. HRMS (ESI) m/z calculated for $C_{29}H_{40}Cl_2N_5O_5S$ [M+H]⁺ 640.21217, found 640.21258.

5.1.19. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-{[1-(methoxycarbonyl)piperidin-4-yl](methyl)amino}-2-oxoethyl}- N^1 -(2-pyrrolidin-1-ylethyl)glycinamide (14m)

To a solution of **14***j* (101 mg, 0.130 mmol) and TEA (0.060 ml, 0.43 mmol) in CH₂Cl₂ (3 mL) was added methyl chloroformate (0.030 ml, 0.39 mmol), and the resulting mixture was stirred at room temperature for 5 h. The reaction mixture was diluted with EtOAc, and the organic layer was washed with saturated aqueous NaHCO₃ and brine, and dried over MgSO₄. The insoluble material was filtered off, and the solution was concentrated under reduced pressure. The residue oil was purified by NH silica gel column chromatography (EtOAc/MeOH = 100:0 to 95:5) to afford 14m (105 mg, 94%) as a colorless oil. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.41– 1.58 (m, 4H), 1.65-1.78 (m, 4H), 2.37-2.48 (m, 6H), 2.69, 2.70 (2s, 3H), 2.72–2.89 (m, 2H), 3.28 (q, J = 6.6, 2H), 3.40–3.54 (m, 0.2H), 3.70, 3.71 (2s, 3H), 3.95 (s, 2H), 4.14, 4.21 (2s, 2H), 4.10-4.37 (m, 2H), 4.41-4.53 (m, 0.8H), 6.74 (d, J = 8.2, 1H), 6.78-6.96 (m, 4H), 7.25 (d, I = 9.3, 2H), 8.10 (broad t, I = 5.1, 1H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 23.55, 27.44, 28.52, 28.62, 29.68, 38.39, 43.33, 51.25, 52.70, 52.84, 54.03, 54.15, 54.89, 55.67, 58.67, 58.84, 118.52, 118.61, 119.02, 119.20, 121.37, 121.58, 122.77, 128.14, 129.81, 130.51, 141.69, 144.60, 144.98, 155.77, 156.45, 168.97, 169.28, 169.79. LC-MS (ESI) m/z 620, 622 $[M+H]^+$. HPLC purity: 98.23%. HRMS (ESI) m/z calculated for $C_{30}H_{40}Cl_2N_5O_5$ [M+H]⁺ 620.24010, found 620.24091.

5.1.20. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[3,4-dihydroisoquinolin-2(1*H*)-yl(methyl)amino]-2-oxoethyl}- N^1 -(2-pyrrolidin-1-ylethyl)glycinamide (14o)

The title compound was synthesized according to the method described for the synthesis of **14n** to afford **14o** (1.20 g, 97%) as a colorless amorphous oil. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.64–1.80 (m, 4H), 2.32–2.64 (m, 6H), 2.77–3.07 (m, 3H), 2.95 (s, 3H), 3.10–3.22 (m, 1H), 3.26 (q, *J* = 6.6, 2H), 3.62 (d, *J* = 13.8, 1H), 3.94 (s, 2H), 4.07 (d, *J* = 13.8, 1H), 4.34 (d, *J* = 18.2, 1H), 4.45 (d, *J* = 18.2, 1H), 6.74 (d, *J* = 8.2, 1H), 6.79 (dd, *J* = 2.6, 8.2, 1H), 6.82 (d, *J* = 9.2, 2H), 6.92 (d, *J* = 2.6, 1H), 6.97–7.03 (m, 1H), 7.10–7.22 (m, 3H), 7.25 (d, *J* = 9.2, 2H), 8.29 (broad, 1H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 23.48, 23.81, 29.90, 38.00, 49.90, 52.57, 53.96, 54.67, 55.46, 58.77, 118.56, 121.04, 122.83, 126.14, 126.73, 126.81, 127.86, 128.65, 129.72, 130.32, 132.71, 132.87, 142.12, 144.69, 156.60, 170.31, 172.26. LC–MS (ESI) *m/z* 610, 612 [M+H]⁺. HPLC purity: 99.31%. HRMS (ESI) *m/z* calculated for C₃₂H₃₈Cl₂N₅O₃ [M+H]⁺ 610.23462, found 610.23456.

5.1.21. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[methyl-{4-[(2-nitrophenyl)sulfonyl]piperazin-1-yl}amino]-2-oxoethyl}- N^1 -(2-pyrrolidin-1-ylethyl)glycinamide (14p)

The title compound was synthesized according to the method described for the synthesis of **14n** to afford **14p** (725 mg, 97%) as a pale yellow oil. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.62–1.75 (m, 4H), 2.34–2.46 (m, 6H), 2.62 (d, *J* = 9.3, 2H), 2.84 (s, 3H), 2.87–3.03 (m, 4H), 3.24 (q, *J* = 6.5, 2H), 3.82 (d, *J* = 10.3, 2H), 3.90 (s, 2H), 4.25 (s, 2H), 6.74 (d, *J* = 8.7, 1H), 6.77–6.83 (m, 3H), 6.91 (d, *J* = 2.1, 1H), 7.24 (d, *J* = 8.7, 2H), 7.64–7.69 (m, 1H), 7.71–7.80 (m, 2H), 7.96–8.04 (m, 2H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 23.51, 24.14, 38.39, 45.66, 50.84, 53.98, 54.77, 55.52, 58.51, 118.45, 118.80, 121.44, 122.87, 124.35, 128.02, 129.75, 130.39, 131.09, 131.11, 131.81, 134.16, 142.07, 144.90, 148.31, 156.48, 169.86, 171.57. LC–MS (ESI) *m*/*z* 748, 750 [M+H]⁺.

5.1.22. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[methyl-(piperazin-1-yl)amino]-2-oxoethyl}- N^1 -(2-pyrrolidin-1-ylethyl)-glycinamide (14q)

To a solution of 14p (711 mg, 0.950 mmol) in CH₃CN (8 mL) were added Cs₂CO₃ (747 mg, 2.29 mmol) and 4-ethylthiophenol

(0.200 mL, 1.48 mmol), the resulting mixture was stirred at room temperature for 2 h. The reaction mixture was diluted with H₂O, and extracted with $CHCl_3$ (50 mL \times 3). The organic layers were combined, and dried over Na₂SO₄. The insoluble material was filtered off, and the solution was concentrated under reduced pressure. The residue oil was purified by NH silica gel column chromatography (EtOAc/MeOH = 100:0 to 90:10) to afford 14q (491 mg, 92%) as a colorless oil. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.61–1.79 (m, 5H), 2.37 (t, J = 6.9, 2H), 2.39–2.46 (m, 4H), 2.55 (d, J=9.8, 2H), 2.72-2.93 (m, 4H), 2.86 (s, 3H), 3.02 (d, J = 11.3, 2H), 3.23 (q, J = 6.5, 2H), 3.94 (s, 2H), 4.32 (s, 2H), 6.76 (d, J = 8.2, 1H), 6.79–6.89 (m, 3H), 6.94 (d, J = 2.6, 1H), 7.25 (d, J = 8.7, 2H), 8.11 (t, J = 5.4, 1H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 23.52, 23.74, 38.39, 45.91, 52.39, 54.01, 54.80, 55.36, 58.70, 118.40, 118.75, 121.26, 122.97, 127.83, 129.68, 130.38, 142.28, 144.82, 156.64, 170.09, 171.71. LC-MS (ESI) m/z 563, 565 $[M+H]^{+}$.

5.1.23. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-{methyl-[4-(methylsulfonyl)piperadin-1-yl]amino}-2-oxoethyl}- N^1 -(2-pyrrolidin-1-ylethyl)glycinamide (14r)

The title compound was synthesized from **14q** according to the method described for the synthesis of **14l** to afford **14r** (235 mg, 93%) as a colorless solid. Mp 144–147 °C. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.63–1.76 (m, 4H), 2.39 (t, *J* = 6.7, 2H), 2.39–2.47 (m, 4H), 2.65 (d, *J* = 10.3, 2H), 2.81–3.00 (m, 4H), 2.82 (s, 3H), 2.86 (s, 3H), 3.26 (q, *J* = 6.5, 2H), 3.78 (d, *J* = 10.8, 2H), 3.93 (s, 2H), 4.30 (s, 2H), 6.76 (d, *J* = 8.7, 1H), 6.79–6.86 (m, 3H), 6.93 (d, *J* = 2.1, 1H), 7.25 (d, *J* = 9.3, 2H), 8.04 (t, *J* = 5.1, 1H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 23.54, 24.17, 35.40, 38.42, 45.59, 50.78, 54.01, 54.81, 55.58, 58.63, 118.49, 118.85, 121.50, 122.88, 128.07, 129.75, 130.40, 142.14, 144.98, 156.52, 169.88, 171.57. LC–MS (ESI) *m*/*z* 641, 643 [M+H]⁺. HPLC purity: 98.96%. HRMS (ESI) *m*/*z* calculated for C₂₃H₃₉Cl₂N₆O₅S [M+H]⁺ 641.20742, found 641.20796.

5.1.24. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-{[4-(methoxycarbonyl)piperidin-1-yl](methyl)amino}-2-oxoethyl}- N^1 -(2-pyrrolidin-1-ylethyl)glycinamide (14s)

The title compound was synthesized from **14q** according to the method described for the synthesis of **14m** to afford **14s** (273 mg, 97%) as a colorless oil. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.64–1.77 (m, 4H), 2.38 (t, *J* = 6.9, 2H), 2.39–2.47 (m, 4H), 2.55 (d, *J* = 10.8, 2H), 2.75 (dt, *J* = 3.1, 11.3, 2H), 2.83 (s, 3H), 2.96 (t, *J* = 11.8, 2H), 3.25 (q, *J* = 6.3, 2H), 3.72 (s, 3H), 3.94 (s, 2H), 4.01–4.26 (broad, 2H), 4.32 (s, 2H), 6.76 (d, *J* = 8.2, 1H), 6.79–6.86 (m, 3H), 6.94 (d, *J* = 2.6, 1H), 7.25 (d, *J* = 9.3, 2H), 8.06 (t, *J* = 5.1, 1H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 23.53, 23.87, 38.40, 43.48, 50.98, 52.94, 54.01, 54.80, 55.45, 58.64, 118.45, 118.85, 121.40, 122.88, 127.99, 129.74, 130.40, 142.15, 144.90, 155.54, 156.54, 169.97, 171.67. LC–MS (ESI) *m*/*z* 621, 623 [M+H]⁺. HPLC purity: 99.35%. HRMS (ESI) *m*/*z* calculated for C₂₉H₃₉Cl₂N₆O₅ [M+H]⁺ 621.23535, found 621.23537.

5.1.25. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[1,3-dihydro-2*H*-isoindol-2-yl(methyl)amino]-2-oxoethyl}- N^1 -[2-(methylamino)ethyl]glycinamide hydrochloride (18a)

To a solution of **10** (276 mg, 0.746 mmol) in DMF (4 mL) was added EDC (146 mg, 0.762 mmol), and the resulting mixture was stirred at room temperature for 1 h. To the reaction mixture stirred under ice-cooling were added **12n** (142 mg, 0.769 mmol) and *N*,*N*-diisopropylethylamine (0.140 mL, 0.823 mmol), and the mixture was stirred at 0 °C for 40 min. To the reaction mixture was added *tert*-butyl (2-aminoethyl)methylcarbamate (160 mg, 0.918 mmol), HOBt·H₂O (145 mg, 0.947 mmol), and EDC (180 mg, 0.939 mmol), and the resulting mixture was stirred at room temperature for

2 h. The reaction mixture was diluted with EtOAc, and the organic layer was washed with H₂O, diluted aqueous NaOH, and brine, and dried over MgSO₄. The insoluble material was filtered off, and the solution was concentrated under reduced pressure. The residue oil was purified by silica gel column chromatography (hexane/EtOAc = 70:30 to 0:100) to afford 338 mg of **17a** as a pale yellow oil. To the obtained oil in CH₂Cl₂ (2 mL) was added TFA (2 mL), and the resulting mixture was stirred at room temperature for 2 h. A diluted aqueous NaOH solution was added to the reaction mixture to give an alkaline solution with a pH greater than 10. The solution was extracted with CH_2Cl_2 (50 mL \times 2), and the organic layers were combined, and dried over Na₂SO₄. The insoluble material was filtered off, and the solution was concentrated under reduced pressure. The residue oil was purified by NH silica gel column chromatography (EtOAc/MeOH = 100:0 to 90:10) to afford a colorless oil (227 mg). To the obtained oil dissolved with diethyl ether (2 mL) was added 4 mol/L HCl in dioxane (0.105 ml. 0.420 mmol), and the resulting mixture was stirred at room temperature for 20 min. The precipitated solid was filtered, washed with diethyl ether, and dried under reduced pressure to give 18a (175 mg, 40%) as a colorless solid. Mp 90–100 °C. ¹H NMR (400 MHz, DMSO- d_6): δ (ppm) 2.49 (s, 3H), 2.78–2.87 (m, 2H), 2.83 (s, 3H), 3.25 (d, *J* = 6.2, 2H), 3.94 (s, 2H), 4.02 (d, *J* = 11.8, 2H), 4.21 (d, J = 11.8, 2H), 4.51 (s, 2H), 6.75-6.85 (m, 3H), 6.88 (d, J = 8.7, 2H, 7.25 (br s, 4H), 7.40 (d, J = 8.7, 2H), 8.50 (t, J = 5.7, 2H), 8.50 (t, J =1H), 8.73 (br s, 2H). ¹³C NMR (100 MHz, DMSO- d_6): δ (ppm) 24.21, 32.23, 34.64, 47.45, 53.85, 54.77, 56.98, 117.05, 118.68, 119.14, 122.40, 123.31, 126.31, 127.02, 128.85, 129.46, 137.34, 142.90, 143.21, 156.52, 170.49, 172.13. LC-MS (ESI) m/z 556, 558 $[M+H]^+$. HPLC purity: 97.77%. HRMS (ESI) m/z calculated for C₂₈H₃₂Cl₂N₅O₃ [M+H]⁺ 556.18767, found 556.18816.

5.1.26. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[1,3-dihydro-2*H*-isoindol-2-yl(methyl)amino]-2-oxoethyl}- N^1 -[2-(ethylamino)ethyl]glycinamide hydrochloride (18b)

The title compound was synthesized according to the method described for the synthesis of **18a** to afford **18b** (350 mg, 73%) as a colorless solid. Mp 98–108 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ (ppm) 1.15 (t, *J* = 7.2, 3H), 2.79–2.95 (m, 4H), 2.83 (s, 3H), 3.25 (q, *J* = 5.8, 2H), 3.94 (s, 2H), 4.02 (d, *J* = 11.3, 2H), 4.21 (d, *J* = 11.3, 2H), 4.51 (s, 2H), 6.75–6.85 (m, 3H), 6.88 (d, *J* = 9.3, 2H), 7.25 (s, 4H), 7.40 (d, *J* = 9.3, 2H), 8.52 (t, *J* = 5.7, 1H), 8.74 (broad, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ (ppm) 10.79, 24.19, 34.75, 41.64, 45.32, 53.82, 54.77, 56.99, 117.01, 118.68, 119.11, 122.39, 123.31, 126.30, 127.01, 128.85, 129.46, 137.34, 142.90, 143.18, 156.54, 170.42, 172.15. LC–MS (ESI) *m*/*z* 570, 572 [M+H]⁺. HPLC purity: 97.22%. HRMS (ESI) *m*/*z* calculated for C₂₉H₃₄Cl₂N₅O₃ [M+H]⁺ 570.20332, found 570.20323.

5.1.27. *N*-[5-Chloro-2-(4-chlorophenoxy)phenyl]-*N*-{2-[2,3-dihydro-1*H*-inden-2-yl(methyl)amino]-2-oxoethyl}glycine (13e)

To a solution of **10** (556 mg, 1.50 mmol) in DMF (6 mL) was added EDC (294 mg, 1.53 mmol), and the resulting mixture was stirred at room temperature for 1 h. To the reaction mixture stirred under ice-cooling were added **12e** (284 mg, 1.55 mmol) and *N*,*N*-diisopropylethylamine (0.280 mL, 1.65 mmol), and the mixture was stirred at 0 °C for 1 h. To the reaction mixture were added H₂O (6 mL), 1 mol/L aqueous HCl (6 mL), and the precipitated solid was filtered, washed with H₂O, dried under reduced pressure, and washed with diethyl ether to give **13e** (684 mg, 91%) as a colorless solid. Mp 171–172 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ (ppm) 2.61 (s, 3H), 2.75–3.07 (m, 4H), 4.08 (s, 2H), 4.25 (s, 1.2H), 4.42 (s, 0.8H), 4.60–4.76 (m, 0.4H), 5.12–5.27 (m, 0.6H), 6.85 (br s, 3H), 6.91 (d, *J* = 8.7, 2H), 7.08–7.26 (m, 4H), 7.39 (d, *J* = 8.7, 2H), 12.91 (br s, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ (ppm) 27.35, 28.78, 35.19,

35.43, 52.96, 54.48, 55.58, 117.89, 118.19, 118.65, 118.79, 119.54, 119.83, 122.88, 123.14, 124.14, 126.37, 126.44, 128.70, 128.84, 129.50, 140.58, 140.98, 142.48, 143.42, 143.81, 156.07, 156.16, 169.19, 169.37, 171.53, 171.59. LC–MS (ESI) m/z 499, 501 [M+H]⁺. Anal. Calculated for $C_{26}H_{24}Cl_2N_2O_{4}$ ·1/3H₂O: C, 61.79; H, 4.92; N, 5.54; Cl, 14.03. Found: C, 61.56; H, 4.63; N, 5.51; Cl, 13.96.

5.1.28. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[2,3-dihydro-1*H*-inden-2-yl(methyl)amino]-2-oxoethyl}- N^1 -methyl (2-pyrrolidin-1-ylethyl)glycinamide (16a)

To a solution of 13e (64 mg, 0.13 mmol) and 1-(2-methylaminoethyl)pyrrolidine (28 mg, 0.22 mmol) in DMF (2 mL) under ice-cooling were added HOBt·H2O (39 mg, 0.25 mmol), and EDC (48 mg, 0.25 mmol), and the resulting mixture was stirred overnight at room temperature. The reaction mixture was diluted with EtOAc, and the organic laver was washed with H₂O, diluted aqueous NaOH, and brine, and dried over Na₂SO₄. The insoluble material was filtered off, and the solution was concentrated under reduced pressure. The residue oil was purified by NH silica gel column chromatography (hexane/EtOAc = 80:20 to 0:100) to afford **16a** (49 mg, 62%) as a colorless oil. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.72-1.85 (m, 2H), 2.02-2.18 (m, 2H), 2.47-2.65 (m, 5H), 2.58, 2.61 (2s, 3H), 2.70-3.17 (m, 8H), 3.23-3.32, 3.45-3.52 (2 m, 2H), 4.17-4.36 (m, 4H), 4.53-4.64 (m, 0.4H), 5.46-5.58 (m, 0.6H), 6.67–6.93 (m, 5H), 7.13–7.29 (m, 6H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ (ppm) 23.52, 23.58, 27.61, 28.95, 33.77, 34.80, 36.04, 36.08, 36.32, 36.38, 47.03, 48.35, 52.77, 53.02, 53.33, 53.72, 53.80, 53.95, 54.15, 54.24, 54.40, 54.54, 56.02, 118.64, 118.70, 118.75, 118.99, 119.16, 119.45, 120.32, 120.60, 122.02, 122.15, 122.24, 124.42, 126.74, 126.97, 127.87, 129.65, 129.68, 130.05, 140.27, 141.11, 143.68, 143.82, 144.48, 144.68, 156.40, 169.26, 169.38, 169.51, 169.64, 169.73. LC-MS (ESI) m/z 609, 611 $[M+H]^+$. HPLC purity: 97.84%. HRMS (ESI) m/z calculated for C₃₃H₃₉Cl₂N₄O₃ [M+H]⁺ 609.23937, found 609.23969.

5.1.29. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[2,3-dihydro-1*H*-inden-2-yl(methyl)amino]-2-oxoethyl}- N^1 -(3-pyr-rolidin-1-ylpropyl)glycinamide (16b)

The title compound was synthesized according to the method described for the synthesis of 16a to afford 16b (508 mg, 83%) as a colorless amorphous solid. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.64-1.78 (m, 2H), 1.78-1.89 (m, 4H), 2.42-2.70 (m, 6H), 2.64 (s, 1.8H), 2.69 (s, 1.2H), 2.77 (dd, J = 6.1, 16.4, 1.2H), 2.94 (dd, J = 6.1, 16.4, 0.8H), 3.03 (dd, *J* = 8.7, 16.4, 0.8H), 3.09 (dd, *J* = 8.7, 16.4, 1.2H), 3.23 (q, J = 6.1, 2H), 3.97 (s, 2H), 4.20 (s, 1.2H), 4.32 (s, 0.8H), 4.43-4.58 (m, 0.4H), 5.34-5.44 (m, 0.6H), 6.73-6.92 (m, 5H), 7.12–7.23 (m, 4H), 7.29 (d, J = 8.7, 2H), 8.51, 8.61 (2 broad, 1H). ^{13}C NMR (100 MHz, CDCl₃): δ (ppm) 23.41, 27.84, 28.04, 29.04, 36.08, 36.36, 37.29, 53.33, 53.68, 53.83, 56.10, 56.21, 56.47, 58.95, 59.17, 117.89, 118.22, 118.59, 118.66, 120.69, 121.00, 122.85, 122.94, 124.43, 124.49, 126.88, 127.19, 128.20, 129.83, 129.88, 130.39, 130.44, 139.91, 140.83, 141.34, 141.51, 144.04, 144.42, 156.39, 156.43, 169.41, 169.78, 169.97. LC-MS (ESI) *m*/*z* 609, 611 [M+H]⁺. HPLC purity: 98.19%. HRMS (ESI) *m*/*z* calculated for C₃₃H₃₉Cl₂N₄O₃ [M+H]⁺ 609.23937, found 609.24003.

5.1.30. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[2,3-dihydro-1*H*-inden-2-yl(methyl)amino]-2-oxoethyl}- N^1 -(2-pipe-ridin-1-ylethyl)glycinamide (16c)

The title compound was synthesized according to the method described for the synthesis of **16a** to afford **16c** (554 mg, 91%) as a colorless amorphous solid. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 1.31–1.44 (m, 2H), 1.45–1.59 (m, 4H), 2.20–2.45 (m, 6H), 2.65 (s, 1.8H), 2.70 (s, 1.2H), 2.78 (dd, *J* = 6.2, 16.4, 1.2H), 2.95 (dd, *J* = 6.2, 16.4, 0.8H), 3.03 (dd, *J* = 8.7, 16.4, 0.8H), 3.10 (dd, *J* = 8.7, 16.4, 1.2H), 3.28 (q, *J* = 6.2, 2H), 3.98 (s, 2H), 4.17 (s, 1.2H), 4.30 (s,

0.8H), 4.46–4.58 (m, 0.4H), 5.37–5.48 (m, 0.6H), 6.76 (d, J = 8.7, 1H), 6.78–6.97 (m, 2H), 6.86 (d, J = 9.3, 2H), 7.13–7.24 (m, 4H), 7.28 (d, J = 9.3, 2H), 8.14, 8.25 (2 broad, 1H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 24.22, 25.83, 27.90, 28.98, 36.11, 36.34, 36.46, 53.13, 54.34, 55.49, 55.78, 56.13, 57.52, 58.68, 58.92, 118.50, 118.58, 118.72, 119.10, 121.13, 121.43, 122.70, 122.82, 124.44, 124.48, 126.84, 127.16, 128.04, 129.78, 129.81, 130.44, 130.49, 139.97, 140.92, 141.72, 141.87, 144.37, 144.74, 156.42, 156.48, 169.05, 169.39, 169.83. LC–MS (ESI) *m*/*z* 609, 611 [M+H]⁺. HPLC purity: 99.92%. HRMS (ESI) *m*/*z* calculated for C₃₃H₃₉Cl₂N₄O₃ [M+H]⁺ 609.23937, found 609.24023.

5.1.31. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[2,3-dihydro-1*H*-inden-2-yl(methyl)amino]-2-oxoethyl}- N^1 -[2-(dimethylamino)ethyl]glycinamide (16d)

The title compound was synthesized according to the method described for the synthesis of **16a** to afford **16d** (494 mg, 87%) as a colorless amorphous solid. ¹H NMR (400 MHz, CDCl₃); δ (ppm) 2.15, 2.16 (2s, 6H), 2.23-2.29 (m, 2H), 2.66 (s, 1.8H), 2.70 (s, 1.2H), 2.78 (dd, / = 6.2, 16.4, 1.2H), 2.96 (dd, / = 6.2, 16.4, 0.8H), 3.05 (dd, J = 8.7, 16.4, 0.8H), 3.10 (dd, J = 8.7, 16.4, 1.2H), 3.25 (q, *J* = 6.6, 2H), 3.98 (s, 2H), 4.17 (s, 1.2H), 4.30 (s, 0.8H), 4.48–4.59 (m, 0.4H), 5.37–5.58 (m, 0.6H), 6.762 (d, J=8.7, 1H), 6.78–6.99 (m, 2H), 6.87 (d, *J* = 9.3, 2H), 7.13–7.22 (m, 4H), 7.28 (d, *J* = 9.3, 2H), 8.12 (t, J = 5.1, 0.4H), 8.25 (t, J = 5.1, 0.6H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 27.91, 28.98, 36.09, 36.35, 37.16, 45.38, 53.13, 55.50, 55.81, 56.11, 58.16, 58.69, 58.94, 118.59, 118.64, 118.89, 119.31, 121.21, 121.54, 122.62, 122.74, 124.44, 124.50, 126.85, 127.17, 128.07, 129.76, 129.81, 130.43, 130.48, 139.98, 140.90, 141.66, 141.82, 144.54, 144.93, 156.40, 156.47, 169.14, 169.50, 169.90. LC-MS (ESI) m/z 569, 571 [M+H]⁺. HPLC purity: 99.13%. HRMS (ESI) m/z calculated for C₃₀H₃₅Cl₂N₄O₃ [M+H]⁺ 569.20807, found 569.20824.

5.1.32. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[2,3-dihydro-1*H*-inden-2-yl(methyl)amino]-2-oxoethyl}- N^1 -[2-(methylamino)ethyl]glycinamide hydrochloride (16m)

To a solution of 13e (371 mg, 1.00 mmol) and tert-butyl (2aminoethyl)methylcarbamate (160 mg, 0.918 mmol) in DMF (3 mL) under ice-cooling were added HOBt H₂O (100 mg, 0.653 mmol), and EDC (119 mg, 0.621 mmol), and the resulting mixture was stirred overnight at room temperature. The reaction mixture was diluted with EtOAc, and the organic layer was washed with H₂O, diluted aqueous NaOH, and brine, and dried over MgSO₄. The insoluble material was filtered off, and the solution was concentrated under reduced pressure. The residue oil was purified by silica gel column chromatography (hexane/EtOAc = 70:30 to 0:100) to afford 233 mg of **16e** as a colorless oil. To the obtained oil in CH₂Cl₂ (1 mL) was added 4 mol/L HCl in dioxane (1.0 mL, 4.0 mmol), and the resulting mixture was stirred at room temperature for 1 h. To the reaction mixture was diluted with diethyl ether (20 mL) to allow precipitation of a solid. This solid was filtered, washed with diethyl ether, and dried under reduced pressure to give 16m (166 mg, 56%) as a colorless solid. Mp 98-109 °C. ¹H NMR (400 MHz, DMSOd₆): δ (ppm) 2.47-2.53 (m, 3H), 2.61 (s, 1.2H), 2.66 (s, 1.8H), 2.78-3.07 (m, 6H), 3.25 (d, J = 6.0, 2H), 3.94, 3.95 (2s, 2H), 4.35 (s, 1.2H), 4.51 (s, 0.8H), 4.61-4.72 (m, 0.4H), 5.16-5.27 (m, 0.6H), 6.77-6.93 (m, 5H), 7.12–7.24 (m, 4H), 7.41 (d, *J* = 9.2, 2H), 8.68 (t, *J* = 5.6, 1H), 8.73 (br s, 2H). ¹³C NMR (100 MHz, DMSO- d_6): δ (ppm) 27.39, 28.76, 32.23, 34.61, 35.17, 35.44, 47.44, 52.93, 54.75, 54.89, 55.54, 57.14, 57.32, 117.12, 117.36, 118.61, 118.75, 119.06, 119.33, 123.05, 123.18, 124.16, 126.39, 126.49, 128.76, 128.87, 129.51, 140.56, 140.98, 142.53, 142.61, 142.95, 143.39, 156.38, 169.22, 169.49, 170.37. LC-MS (ESI) m/z 555, 557 [M+H]⁺. HPLC purity: 98.38%. HRMS (ESI) m/z calculated for $C_{29}H_{33}Cl_2N_4O_3$ [M+H]⁺ 555.19242, found 555.19282.

5.1.33. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[2,3-dihydro-1*H*-inden-2-yl(methyl)amino]-2-oxoethyl}- N^1 -[2-(ethylamino)ethyl]glycinamide hydrochloride (16n)

The title compound was synthesized according to the method described for the synthesis of **16m** to afford **16n** (252 mg, 83%) as a colorless solid. Mp 97–108 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ (ppm) 1.11–1.22 (m, 3H), 2.62 (s, 1.2H), 2.66 (s, 1.8H), 2.76–3.09 (m, 8H), 3.26 (d, *J* = 6.0, 2H), 3.94, 3.95 (2s, 2H), 4.35 (s, 1.2H), 4.51 (s, 0.8H), 4.61–4.73 (m, 0.4H), 5.15–5.28 (m, 0.6H), 6.76–6.96 (m, 5H), 7.10–7.26 (m, 4H), 7.41 (d, *J* = 8.7, 2H), 8.64–8.75 (m, 1H), 8.77 (br s, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ (ppm) 10.82, 27.39, 28.76, 34.75, 35.18, 35.43, 41.68, 45.34, 52.96, 54.80, 54.91, 55.56, 57.14, 57.29, 117.16, 117.37, 118.63, 118.75, 119.10, 119.37, 123.05, 123.16, 124.15, 126.40, 126.49, 128.76, 128.86, 129.51, 140.54, 140.97, 142.53, 142.59, 143.01, 143.42, 156.38, 169.26, 169.48, 170.35. LC–MS (ESI) *m*/*z* 569, 571 [M+H]⁺. HPLC purity: 98.92%. HRMS (ESI) *m*/*z* calculated for C₃₀H₃₅Cl₂N₄O₃ [M+H]⁺ 569.20807, found 569.20839.

5.1.34. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[2,3-dihydro-1*H*-inden-2-yl(methyl)amino]-2-oxoethyl}- N^1 -[2-(isopropylamino)ethyl]glycinamide hydrochloride (16o)

The title compound was synthesized according to the method described for the synthesis of 16m to afford 16o (96 mg, 61%) as a colorless solid. Mp 93–101 °C. ¹H NMR (400 MHz, DMSO- d_6): δ (ppm) 1.19 (d, J = 6.2, 2.4H), 1.21 (d, J = 6.2, 3.6H), 2.62 (s, 1.2H), 2.66 (s, 1.8H), 2.72-3.07 (m, 6H), 3.19-3.34 (m, 3H), 3.94 (s, 1.2H), 3.95 (s, 0.8H), 4.34 (s, 1.2H), 4.51 (s, 0.8H), 4.61-4.72 (m, 0.4H), 5.16-5.27 (m, 0.6H), 6.78-6.86 (m, 3H), 6.87-6.95 (m, 2H), 7.11-7.25 (m, 4H), 7.41 (d, J = 9.3, 2H), 8.65-8.74 (m, 1H), 8.81 (br s, 2H). ¹³C NMR (100 MHz, DMSO- d_6): δ (ppm) 18.33, 27.38, 28.77, 34.92, 35.20, 35.42, 42.85, 49.19, 52.97, 54.73, 54.83, 55.59, 57.12, 57.25, 117.23, 117.42, 118.69, 118.81, 119.13, 119.39, 122.97, 123.07, 124.15, 126.41, 126.49, 128.72, 128.81, 129.48, 129.51, 140.54, 140.97, 142.55, 142.60, 143.12, 143.51, 156.32, 156.35, 169.19, 169.39, 170.32. LC-MS (ESI) m/z 583, 585 $[M+H]^+$. HPLC purity: 97.96%. HRMS (ESI) m/z calculated for C₃₁H₃₇Cl₂N₄O₃ [M+H]⁺ 583.22372, found 583.22415.

5.1.35. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[2,3-dihydro-1*H*-inden-2-yl(methyl)amino]-2-oxoethyl}- N^1 -(pipe-ridin-3-yl)glycinamide hydrochloride (16p)

The title compound was synthesized according to the method described for the synthesis of **16m** to afford **16p** (131 mg, 73%) as a colorless solid. Mp 124–136 °C. ¹H NMR (400 MHz, DMSO d_6): δ (ppm) 1.22–1.37 (m, 1H), 1.55–1.86 (m, 3H), 2.50–2.60 (m, 1H), 2.60 (s, 1.2H), 2.67 (s, 1.8H), 2.72-3.16 (m, 7H), 3.77-3.90 (m, 1H), 3.93 (s, 2H), 4.34 (d, J = 18.0, 0.6H), 4.39 (d, J = 18.0, 0.6H), 4.48 (d, J = 18.0, 0.4H), 4.54 (d, J = 18.0, 0.4H), 4.61-4.72 (m, 0.4H), 5.14-5.26 (m, 0.6H), 6.77-6.94 (m, 5H), 7.11-7.25 (m, 4H), 7.42 (d, J = 9.3, 2H), 8.78 (d, J = 7.2, 1H), 8.96 (br s, 1H), 9.12 (br s, 1H). ¹³C NMR (100 MHz, DMSO- d_6): δ (ppm) 20.27, 27.42, 27.67, 28.74, 35.16, 35.46, 42.49, 42.72, 45.80, 53.00, 54.98, 55.10, 55.49, 57.29, 57.52, 116.95, 117.16, 118.67, 118.79, 119.07, 119.31, 123.09, 123.17, 124.16, 126.39, 126.45, 126.50, 128.76, 128.86, 129.50, 129.56, 140.55, 140.94, 140.96, 142.18, 142.29, 143.03, 143.40, 156.37, 169.31, 169.44, 169.75. LC-MS (ESI) m/z 581, 583 [M+H]⁺. HPLC purity: 95.88%. HRMS (ESI) *m*/*z* calculated for C₃₁H₃₅Cl₂N₄O₃ [M+H]⁺ 581.20807, found 581.20836.

5.1.36. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[2,3-dihydro-1*H*-inden-2-yl(methyl)amino]-2-oxoethyl}- N^1 -(pyrro-lidin-3-yl)glycinamide hydrochloride (16q)

The title compound was synthesized according to the method described for the synthesis of **16m** to afford **16q** (106 mg, 68%) as a colorless solid. Mp 136–146 °C. ¹H NMR (400 MHz, DMSO-

*d*₆): δ (ppm) 1.56–1.70 (m, 1H), 1.92–2.07 (m, 1H), 2.62 (s, 1.2H), 2.69 (s, 1.8H), 2.77–3.30 (m, 8H), 3.84–4.00 (m, 2H), 4.06–4.18 (m, 1H), 4.31 (d, *J* = 17.9, 0.6H), 4.44 (d, *J* = 17.9, 0.6H), 4.47 (d, *J* = 17.4, 0.4H), 4.59 (d, *J* = 17.4, 0.4H), 4.65–4.75 (m, 0.4H), 5.18–5.29 (m, 0.6H), 6.77–6.93 (m, 5H), 7.11–7.25 (m, 4H), 7.41 (d, *J* = 9.2, 2H), 8.88 (d, *J* = 6.1, 0.6H), 8.91 (d, *J* = 6.1, 0.4H), 9.14 (br s, 1H), 9.36 (br s, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ (ppm) 27.44, 28.77, 29.58, 35.19, 35.50, 43.13, 43.20, 47.98, 48.04, 48.63, 52.91, 54.73, 54.90, 55.53, 57.35, 57.63, 116.96, 117.18, 118.59, 118.75, 119.12, 119.34, 123.13, 123.24, 124.16, 126.39, 126.49, 128.78, 128.89, 129.47, 129.51, 140.55, 140.96, 140.99, 142.34, 142.42, 142.99, 143.39, 156.40, 156.45, 169.41, 169.73, 169.89. LC–MS (ESI) *m*/*z* 567, 569 [M+H]⁺. HPLC purity: 99.30%. HRMS (ESI) *m*/*z* calculated for C₃₀H₃₃Cl₂N₄O₃ [M+H]⁺ 567.19242, found 567.19287.

5.1.37. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[2,3-dihydro-1*H*-inden-2-yl(methyl)amino]-2-oxoethyl}- N^1 -[(2*S*)-pyrrolidin-2-ylmethyl]glycinamide hydrochloride (16r)

The title compound was synthesized according to the method described for the synthesis of 16m to afford 16r (117 mg, 75%) as a colorless solid. Mp 113–125 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ (ppm) 1.43–1.59 (m, 1H), 1.71–1.93 (m, 3H), 2.61 (s, 1.2H), 2.65 (s, 1.8H), 2.82 (dd, J = 6.7, 16.4, 1.2H), 2.90 (dd, J = 6.7, 16.4, 0.8H), 2.95 (dd, J = 8.7, 16.4, 1.2H), 3.00 (dd, J = 8.7, 16.4, 0.8H), 3.05-3.19 (m, 2H), 3.20-3.51 (m, 3H), 3.96, 3.97 (2s, 2H), 4.36 (s, 1.2H), 4.52 (s, 0.8H), 4.59-4.71 (m, 0.4H), 5.15-5.26 (0.6H), 6.76-6.94 (m, 5H), 7.11-7.24 (m, 4H), 7.42 (d, J = 8.7, 2H), 8.69 (br s, 1H), 8.85 (t, J = 5.4, 1H), 9.44 (br s, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ (ppm) 22.70, 26.95, 27.37, 28.75, 35.16, 35.42, 44.37, 52.95, 54.90, 55.03, 55.52, 57.16, 57.35, 58.70, 66.25, 117.08, 117.33, 118.68, 118.81, 119.07, 119.33, 123.00, 123.11, 124.15, 126.40, 126.45, 126.49, 128.74, 128.84, 129.51, 129.55, 140.54, 140.96, 142.28, 142.39, 143.00, 143.42, 156.28, 156.31, 169.25, 169.53, 170.53. LC-MS (ESI) m/z 581, 583 [M+H]⁺. HPLC purity: 99.80%. HRMS (ESI) m/z calculated for $C_{31}H_{35}Cl_2N_4O_3$ [M+H]⁺ 581.20807, found 581.20828.

5.1.38. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[2,3-dihydro-1*H*-inden-2-yl(methyl)amino]-2-oxoethyl}- N^1 -[(2*R*)-pyrrolidin-2-ylmethyl]glycinamide hydrochloride (16s)

The title compound was synthesized according to the method described for the synthesis of 16m to afford 16s (387 mg, 76%) as a colorless solid. Mp 110–118 °C. ¹H NMR (400 MHz, DMSO- d_6): δ (ppm) 1.44–1.59 (m, 1H), 1.73–1.92 (m, 3H), 2.61 (s, 1.2H), 2.65 (s, 1.8H), 2.82 (dd, J = 6.7, 16.4, 1.2H), 2.90 (dd, J = 6.7, 16.4, 0.8H), 2.95 (dd, J = 8.7, 16.4, 1.2H), 3.00 (dd, J = 8.7, 16.4, 0.8H), 3.05-3.17 (m, 2H), 3.18-3.49 (m, 3H), 3.97 (s, 2H), 4.34 (s, 1.2H), 4.50 (s, 0.8H), 4.59-4.70 (m, 0.4H), 5.14-5.26 (0.6H), 6.76-6.94 (m, 5H), 7.11–7.24 (m, 4H), 7.42 (d, J = 8.7, 2H), 8.52 (br s, 1H), 8.87 (t, J = 5.1, 1H), 9.19 (br s, 1H). ¹³C NMR (100 MHz, DMSO*d*₆): δ (ppm) 22.71, 26.96, 27.36, 28.75, 35.16, 35.41, 44.34, 52.94, 54.89, 55.02, 55.52, 57.16, 57.35, 58.69, 117.08, 117.32, 118.67, 118.82, 119.07, 119.32, 122.99, 123.12, 124.15, 126.39, 126.44, 126.49, 128.73, 128.84, 129.50, 129.55, 140.54, 140.96, 142.29, 142.39, 142.99, 143.41, 156.31, 169.25, 169.52, 170.51. LC-MS (ESI) *m*/*z* 581, 583 [M+H]⁺. HPLC purity: 98.85%. HRMS (ESI) *m*/*z* calculated for C₃₁H₃₅Cl₂N₄O₃ [M+H]⁺ 581.20807, found 581.20824.

5.1.39. N^2 -[5-Chloro-2-(4-chlorophenoxy)phenyl]- N^2 -{2-[2,3-dihydro-1*H*-inden-2-yl(methyl)amino]-2-oxoethyl}- N^1 -(piperidin-2-ylmethyl)glycinamide hydrochloride (16t)

The title compound was synthesized according to the method described for the synthesis of **16m** to afford **16t** (369 mg, 84%) as a colorless solid. Mp 124–134 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ (ppm) 1.20–1.44 (m, 2H), 1.46–1.75 (m, 4H), 2.60 (s, 1.2H), 2.65

(s, 1.8H), 2.73–3.06 (m, 6H), 3.09–3.35 (m, 3H), 3.98 (s, 2H), 4.38 (s, 1.2H), 4.53 (s, 0.8H), 4.59–4.70 (m, 0.4H), 5.13–5.25 (m, 0.6H), 6.77–6.94 (m, 5H), 7.10–7.24 (m, 4H), 7.38–7.45 (m, 2H), 8.3–9.0 (broad, 2H), 8.77 (t, *J* = 6.2, 0.4H), 8.82 (t, *J* = 6.2, 0.6H). ¹³C NMR (100 MHz, DMSO- d_6): δ (ppm) 21.27, 21.53, 25.60, 27.38, 28.77, 35.16, 35.40, 40.33, 43.81, 52.99, 54.90, 55.11, 55.27, 55.54, 57.26, 57.47, 117.02, 117.36, 118.65, 118.76, 118.98, 119.30, 123.01, 123.13, 124.15, 126.41, 126.44, 126.50, 128.78, 128.88, 129.50, 129.55, 140.54, 140.96, 142.22, 142.41, 142.94, 143.40, 156.28, 156.32, 169.43, 169.71, 170.36. LC–MS (ESI) *m/z* 595, 597 [M+H]⁺. HPLC purity: 98.37%. HRMS (ESI) *m/z* calculated for C₃₂H₃₇Cl₂N₄O₃ [M+H]⁺ 595.22372, found 595.22394.

5.1.40. *N*-Methyl-1,3-dihydro-2*H*-isoindol-2-amine hydrochloride (12n)²³

Xylylene dibromide (160 g, 606 mmol) and tert-butyl 1-methylhydrazinecarboxylate (26) (88.5 g, 606 mmol) obtained according to the reported method²⁴ were dissolved in *N*-methylpyrrolidone (550 mL). To the stirred reaction mixture at 50-60 °C, TEA (190 mL, 1.36 mol) was gradually added dropwise. After the addition was completed, the reaction mixture was stood overnight at room temperature. To the reaction mixture was added 5% aqueous citric acid (700 mL), and the precipitate solid was collected by filtration, washed with H₂O, and dried under reduced pressure to give *tert*-butyl 1,3-dihydro-2*H*-isoindol-2-yl(methyl)carbamate (27) (126 g, 84%) as a pale pink solid. This compound was used in the next reaction without further purification. ¹H NMR (300 MHz, CDCl₃): δ (ppm) 1.41 (s, 9H), 3.09 (s, 3H), 4.44 (s, 4H), 7.1–7.2 (m, 4H). To a solution of 27 (126 g, 507 mmol) in CH₂Cl₂ (150 mL) and EtOH (150 mL) was added 4 mol/L HCl in dioxane (500 mL), and the reaction mixture was stirred at room temperature for 5 h. The reaction mixture was diluted with H₂O and CH₂Cl₂ and the aqueous layer was extracted. An ice-cooled aqueous NaOH solution was added to the ice-cooled aqueous layer to give a strongly-alkaline solution. The aqueous layer was extracted with CH₂Cl₂. The organic layer was dried over K₂CO₃, and the insoluble material was filtered off. The solution was concentrated under reduced pressure. The residue oil was dissolved in diethyl ether (500 mL), and 4 mol/L HCl in dioxane (140 mL) was added with stirring under ice-cooling to allow precipitation of a solid. This solid was filtered, washed with diethyl ether, and dried under reduced pressure to give 12n (75.9 g, 81%) as a gray solid. Mp 159–165 °C. ¹H NMR (400 MHz, DMSO- d_6): δ (ppm) 2.77 (s, 3H), 4.42 (br s, 4H), 7.23–7.43 (m, 4H), 10.97 (br s, 2H). ¹³C NMR (100 MHz, DMSO- d_6): δ (ppm) 32.81, 56.66, 122.67, 127.63, 136.10. LC-MS (ESI) m/z 149 [M+H]⁺. Anal. Calculated for C₉H₁₃ClN₂: C, 58.54; H, 7.10; N, 15.17; Cl, 19.20. Found: C, 58.31; H, 7.08; N, 14.87; Cl, 18.84.

5.1.41. 3,4-Dihydroisoquinolin-2(1*H*)-amine (29)²⁵

To a stirred solution of 1,2,3,4-tetrahydroisoquinoline (28) (2.00 g, 15.0 mmol) and sodium nitrite (2.07 g, 30.0 mmol) in H₂O (30 mL) was gradually added AcOH (1.30 mL, 22.7 mmol) at under 5 °C, and the mixture was stirred at room temperature for 90 min. The reaction mixture was diluted with EtOAc, and the organic layer was washed with H₂O, saturated aqueous NaHCO₃ and brine and dried over MgSO4. The insoluble material was filtered off, and the solution was concentrated under reduced pressure. The residue oil was purified by silica gel column chromatography (hexane/EtOAc = 90:10 to 50:50) to afford 2.03 g of a purple oil. This oil was dissolved with MeOH (15 mL), and to the mixture was added Zn (4.00 g, 61.2 mmol). To the mixture was gradually added AcOH (15.0 mL) under ice-cooling, and the mixture was stirred at room temperature for 80 min. The insoluble material in the reaction mixture was filtered off through Celite, and washed with MeOH and CHCl₃. The solution was neutralized with saturated aqueous NaHCO₃ and the solution was extracted with CHCl₃ (200 mL × 3). The organic layers were combined, and dried over Na₂SO₄. The insoluble material was filtered, and the solution was concentrated under reduced pressure. The residue oil was purified by silica gel column chromatography (EtOAc/MeOH = 100:0 to 80:20) to afford **29** (1.50 g, 67%) as a pale yellow oil. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 2.5–3.5 (broad, 2H), 2.94 (t, *J* = 5.7, 2H), 3.00 (t, *J* = 5.7, 2H), 3.82 (s, 2H), 7.00–7.05 (m, 1H), 7.09–7.19 (m, 3H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 29.27, 57.23, 62.01, 125.82, 126.46, 126.67, 128.37, 133.19, 134.00. LC–MS (ESI) *m/z* 149 [M+H]⁺.

5.1.42. N-Methyl-3,4-dihydroisoquinolin-2(1H)-amine (120)²⁶

To a stirred solution of 29 (1.50 g, 10.1 mmol) in H₂O (15 mL) and AcOH (0.700 mL, 12.2 mmol) was added 37% aqueous formalin (0.950 mL, 12.6 mmol), and the resulting mixture was stirred at 0 °C for 10 min. The reaction mixture was diluted with diluted aqueous NaOH, and the solution was extracted with diethyl ether (100 mL). The organic layer was washed with brine, and dried over MgSO₄. The insoluble material was filtered off, and the solution was concentrated under reduced pressure. The residue oil was purified by silica gel column chromatography (hexane/EtOAc = 100:0 to 80:20) to afford 1.34 g of a colorless oil. The obtained oil in THF (10 mL) was added to a stirred suspension of LiAlH₄ (460 mg, 12.1 mmol) in diethyl ether (50 mL), the resulting mixture was stirred under reflux for 40 min. The reaction mixture was ice-cooled, and H₂O (0.460 mL), 15% aqueous NaOH (0.460 mL), H₂O (1.38 mL) and Na₂SO₄ were successively added with stirring. The insoluble material was filtered off through Celite, and the solution was concentrated under reduced pressure. The residue oil was purified by NH silica gel column chromatography (hexane/EtOAc = 100:0 to 50:50) to afford **12o** (1.30 g, 79%) as a colorless oil. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 2.69 (s, 3H), 2.99 (s, 4H), 3.88 (s, 2H), 7.01–7.18 (m, 4H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 28.92, 35.48, 52.73, 57.96, 125.82, 126.44, 126.95, 128.46, 133.71, 133.99. LC-MS (ESI) m/z 163 [M+H]⁺.

5.1.43. *tert*-Butyl 4-[(2-nitrophenyl)sulfonylpiperazin-1-yl]carbamate (31)

To an ice-cooled solution of *tert*-butyl piperazin-1-ylcarbamate (30) (820 mg, 4.07 mmol, Bepharma. Ltd) and TEA (0.850 ml, 6.10 mmol) in CH₂Cl₂ (10 mL) was added 2-nitrobenzenesulfonyl chloride (947 mg, 4.27 mmol), and the resulting mixture was stirred overnight at room temperature. The reaction mixture was diluted with EtOAc, and the organic layer was washed with 10% aqueous citric acid and brine, and dried over MgSO₄. The insoluble material was filtered off, and the solution was concentrated under reduced pressure. The residue oil was purified by silica gel column chromatography (hexane/EtOAc = 90:10 to 0:100) to afford 31 (1.39 g, 86%) as a pale blue solid. Mp 183–184 °C. ¹H NMR (400 MHz, DMSO- d_6): δ (ppm) 1.44 (s, 9H), 2.89 (t, J = 4.9, 4H), 3.43 (t, J = 4.9, 4H), 5.54 (s, 1H), 7.62 (dd, J = 1.5, 7.7, 1H), 7.66-7.77 (m, 2H), 7.95 (dd, J = 1.5, 7.2, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ (ppm) 28.29, 45.74, 54.95, 80.75, 124.17, 130.81, 130.86, 131.52, 133.90, 148.48, 154.19. LC-MS (ESI) m/z 331 [M+H]⁺. Anal. Calculated for C₁₅H₂₂N₄O₆S: C, 46.62; H, 5.74; N, 14.50. Found: C, 46.64; H, 5.58; N, 14.44.

5.1.44. *N*-Methyl-4-(2-nitrophenyl)sulfonyl-piperazin-1-amine hydrochloride (12p)

To a stirred solution of **31** (775 mg. 2.01 mmol) in THF (8 mL) was added potassium *tert*-butoxide (331 mg, 2.95 mmol), the resulting mixture was stirred at 0 °C for 5 min. Then, to the reaction mixture was added iodomethane (0.300 ml, 4.82 mmol), and the mixture was stirred at 0 °C for 30 min. The reaction mixture was diluted with EtOAc, and the organic layer was washed with H_2O and brine, and dried over MgSO₄. The insoluble material was

A. Nakao et al./Bioorg. Med. Chem. xxx (2015) xxx-xxx

filtered off, and the solution was concentrated under reduced pressure. The residue oil was purified by silica gel column chromatography (hexane/EtOAc = 90:10 to 35:65) to afford 543 mg of a colorless oil. The obtained oil was dissolved with CH₂Cl₂ (2 mL), and to the solution was added 4 mol/L HCl in dioxane (2.0 mL, 8.0 mmol), and the resulting mixture was stirred at room temperature for 4 h. The reaction mixture was diluted with diethyl ether (8 mL). The precipitated solid was filtered, washed with diethyl ether and dried under reduced pressure to give 12p (3.53 g, 83%) as a colorless solid. Mp 224-226 °C. ¹H NMR (400 MHz, DMSO*d*₆): δ (ppm) 2.64 (br s, 3H), 3.09 (br s, 4H), 3.36 (br s, 4H), 7.86– 7.92 (m, 1H), 7.95 (t, J = 7.7, 1H), 8.01-8.06 (m, 2H), 10.85, 10.98 (2 br s, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ (ppm) 30.22, 44.65, 49.91, 124.25, 129.00, 130.32, 132.43, 135.02, 147.68. LC-MS (ESI) m/z 301 [M+H]⁺. Anal. Calculated for C₁₁H₁₇ClN₄O₄S·1/5H₂O: C, 38.81; H, 5.15; N, 16.46; Cl, 10.42; S, 9.42. Found: C, 38.99; H, 5.00: N. 16.09: Cl. 10.53: S. 9.23.

5.1.45. *N*-Methylindan-2-amine hydrochloride (12e)²⁷

2-Aminoindane (19) (3.10 g, 23.3 mmol) was dissolved in CH₂Cl₂ (50 mL), Boc₂O (5.20 g, 23.8 mmol) and TEA (5 mL) were added under ice-cooling, and the mixture was stirred at room temperature for 2 h. The reaction mixture was diluted with a 1:1 EtOAc/hexane (100 mL) solution, and the organic layer was washed with 10% aqueous citric acid solution, saturated aqueous NaHCO₃ and brine and dried over MgSO₄. The insoluble material was filtered off, and the solution was concentrated under reduced pressure to give 4.68 g of 20a as a colorless oil. This was added to a suspension of LiAlH₄ (2.56 g, 67.5 mmol) in THF (100 mL), and the mixture was heated under reflux conditions for 3 h. The reaction mixture was ice-cooled, and H₂O (2.56 mL), 15% aqueous NaOH (2.56 mL), H₂O (7.68 mL) and MgSO₄ were successively added with stirring. The insoluble material was filtered off through Celite, nd the solution was concentrated under reduced pressure to give an oil (3.15 g). This was dissolved in diethyl ether (50 mL), 4 mol/L HCl in dioxane (6 mL) was added, and the precipitated solid was filtered, washed with diethyl ether and dried under reduced pressure to give 12e (3.53 g, 83%) as a colorless solid. Mp 233–236 °C.²⁰ ¹H NMR (400 MHz, DMSO- d_6): δ (ppm) 2.57 (s, 3H), 3.11 (dd, J = 6.7, 16.4, 2H), 3.27 (dd, J = 8.0, 16.4, 2H), 3.87-4.00 (m, 1H), 7.16-7.23 (m, 2H), 7.23-7.30 (m, 2H), 9.39 (br s, 2H). ¹³C NMR (100 MHz, DMSO- d_6): δ (ppm) 30.76, 35.29, 58.17, 124.44, 126.89, 139.36. LC-MS (ESI) m/z 148 [M+H]⁺.

5.1.46. N-Ethylindan-2-amine hydrochloride (12h)²⁷

The title compound was synthesized according to the method described for the synthesis of **12e** from 2-aminoindane (**19**) (1.07 g, 8.03 mmol) and acetic anhydride (0.76 mL, 8.04 mmol) to afford **12h** (1.01 g, 94%) as a colorless solid. Mp 168–176 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ (ppm) 1.25 (t, *J* = 7.2, 3H), 2.92–3.06 (m, 2H), 3.08–3.19 (m, 2H), 3.22–3.33 (m, 2H), 3.90–4.07 (m, 1H), 7.15–7.31 (m, 4H), 9.32 (br s, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ (ppm) 11.06, 35.47, 40.60, 56.66, 124.40, 126.87, 139.38. LC–MS (ESI) *m*/*z* 162 [M+H]⁺.

5.1.47. *N*-Methyl-1,2,3,4-tetrahydronaphthalen-2-amine hydrochloride (12f)²⁷

The title compound was synthesized according to the method described for the synthesis of **12e** from 1,2,3,4-tetrahydronaph-thalen-2-amine (**21**) (2.50 g, 13.6 mmol) to afford **12f** (1.66 g, 62%) as a colorless solid. Mp 185–186 °C. ¹H NMR (400 MHz, DMSO- d_6): δ (ppm) 1.71–1.85 (m, 1H), 2.20–2.28 (m, 1H), 2.60 (s, 3H), 2.74–2.93 (m, 3H), 3.13–3.23 (m, 1H), 3.31–3.42 (m, 1H), 7.08–7.17 (m, 4H), 9.24 (br s, 2H). ¹³C NMR (100 MHz, DMSO- d_6): δ (ppm) 24.85, 26.78, 29.37, 30.93, 53.97, 125.90, 126.15, 128.42, 128.93, 132.50, 134.86. LC–MS (ESI) m/z 162 [M+H]⁺.

5.1.48. *trans-N*-Methyl-4-phenylcyclohexanamine hydrochloride (12g)

The title compound was synthesized according to the method described for the synthesis of **12e** from *trans*-4-phenylcyclohexanamine (**23**) (3.42 g, 16.2 mmol) to afford **12g** (3.11 g, 88%) as a colorless solid. Mp 269–272 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ (ppm) 1.38–1.58 (m, 4H), 1.81–1.95 (m, 2H), 2.08–2.22 (m, 2H), 2.44-2.60 (m, 4H), 2.91–3.09 (m, 1H), 7.15–7.33 (m, 5H), 8.88 (br s, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ (ppm) 28.28, 31.37, 42.27, 56.23, 126.01, 126.56, 128.23, 145.88. LC–MS (ESI) *m*/*z* 190 [M+H]⁺. Anal. Calculated for C₁₃H₂₀ClN·1/10H₂O: C, 68.61; H, 8.95; N, 6.16; Cl, 15.58. Found: C, 68.77; H, 8.78; N, 6.16; Cl, 15.38.

5.2. X-ray crystallography

5.2.1. Crystallization, data, and structural refinement of human recombinant AdoHcyase, NAD⁺, and 18a

The enzyme of human recombinant AdoHcyase purchased from Diazyme Laboratory was purified according to the method reported by Yuan et al.²⁸ The protein-inhibitor complex was produced by sitting-drop vapor diffusion method using a reservoir solution composed of 100 mM HEPES (pH 7.5), 13% (w/v) PEG 4000, 10% (v/v) 2-PrOH, and 0.5% (v/v) EtOAc. A plate-shape crystal suitable for X-rav diffraction having dimensions 0.1 mm \times 0.05 mm \times 0.005 mm in a drop was dipped into a cryoprotectant solution containing 100 mM HEPES (pH 7.5), 25% (w/v) PEG 4000, and 15% (v/v) glycerol for 2 h before it was frozen in liquid nitrogen. X-ray diffraction data were collected at SPring-8 BL24XU beam line. The diffraction data were measured up to 2.7 Å resolutions at -180 °C. The data were processed with the program HKL2000. The structure of AdoHcyase and inhibitor complex was solved by molecular replacement with the program AMoRe, utilizing the previously determined coordinates of AdoHcyase with Protein Data Bank accession code 1B3R. The structure was refined against all available data to 2.7 Å using Maximum likelihood (Refmac) to a crystallographic R-factor of 0.199 and free R-factor of 0.240. Data collection and model refinement statistics are summarized in Table 4. The crystallographic refinement parameters, final (2Fo Fc) maps, and conformational analysis by PROCHECK indicate that the crystal structure has been determined with acceptable statistics. Coordinates have been deposited with the Protein Data Bank, (Entry name: 4YVF).

5.2.2. X-ray crystal structure determination of 14n

A colorless prismatic crystal of compound **14n** suitable for X-ray single crystal analysis was obtained at room temperature by partial evaporation from diethyl ether. X-ray data (see Table 5) were collected at a temperature of 25 ± 1 °C on a Rigaku AFC7R diffractometer equipped with a graphite monochromated Cu-K α radiation ($\lambda = 1.5418$ Å) and a rotating anode generator. The crystal structure was solved by direct methods.²⁹ The non-hydrogen atoms were refined anisotropically by full-matrix least squares refinement on F^2 . The H atoms were refined using the riding model. All calculations were package.³⁰ CCDC 1049632 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

5.3. In vitro AdoHcyase inhibition assays

The enzyme inhibitory activity was measured using the hydrolysis activity of AdoHcy as an index. The measurement method was modification of the reported method.³¹ AdoHcy (10μ M) and adenosine deaminase (Roche) (4 units) were added to 50 mM phosphate buffer (pH 7.2, containing 1 mM EDTA) with the total

Table 4

Resolution (Å) No. of all reflections Completeness (working + test) (%) R^{b} (working) Free <i>R</i> Free <i>R</i> test set size (%) No. of reflections used for free <i>R</i>	47.81-2.7 31,525 99.5 0.199 0.240 5.0 1565	(Highest range ^a : 4826) (Highest range ^a : 98.5) (Highest range ^a : 0.229) (Highest range ^a : 0.282) (Highest range ^a : 5.5) (Highest range ^a : 280)
No. of non-hydrogen atoms used in refi AdoHcyase NAD ⁺ 18a Water	nement 6644 88 76 244	
B values From Wilson plot (Å ²) Mean B value all (Å ²) AdoHcyase NAD* 18a Water Estimated coordinate error ESD from Luzzati plot (Å) ESD from SIGMAA (Å) Low resolution cutoff (Å) RMS deviations from ideal values Bond length (Å) Bond angles (°)	22.1 23.9 24.1 18.6 20.4 21.5 0.25 0.19 5.00 0.006 1.2	
Dihedral angles (°) Improper angles (°)	22.0 0.73	

^a 2.87-2.70 Å resolution.

^b $R = \Sigma |Fo - Fc| / \Sigma |Fo|$.

Table 5

Crystal data and structure refinement of **14n**

Compound	14n
Empirical formula	$C_{31}H_{35}Cl_2N_5O_3$
Formula weight	596.56
Recryst. solvent	Diethyl ether
Crystal system	Monoclinic
Unit cell dimensions	$a = 18.9093(17)$ Å, $\alpha = 90^{\circ}$
	$b = 16.1873(9)$ Å, $\beta = 131.002(5)^{\circ}$
	$c = 13.2852(9)$ Å, $\gamma = 90^{\circ}$
Volume (Å ³)	3068.9(5)
Space group	Сс
Z value	4
Density (calculated) (g/cm ³)	1.291
Absorption coefficient (cm ⁻¹)	22.252
Crystal size (mm ³)	$0.300 \times 0.200 \times 0.100$
Temperature (K)	298
2θ max (°)	135.8
Reflections collected	2876
Independent reflections	2790 [<i>R</i> (int) = 0.0347]
Data/restraints/parameters	2344/0/406
Goodness-of-fit on F^2	1.003
Residuals: R, Rw	0.0438, 0.1257
CCDC reference number	1049632

amount being 200 µL, and to the solution were added a test substance and then human recombinant *S*-adenosyl-L-homocysteine hydrolase (50 ng, Diazyme Laboratories) to start the reaction, and the mixture was incubated at 37 °C for 8 min. The reaction was quenched by the addition of 1 mol/L aqueous perchloric solution (20 µl), and the mixture was centrifuged under the conditions of 10,000 rpm, 5 min, 4 °C. The supernatant was collected, and the amount of AdoHcy after the reaction was quantified by HPLC. The inhibitory rate was determined with the amount of decrease in AdoHcy before and after the reaction without using the test substance as 100%. Inhibitory rate (%) = [(amount of decrease of AdoHcy in the presence of test substance)] × 100.

Acknowledgements

The authors thank Dr. Kunitomo Adachi, Dr. Yoshinori Nakamura, Dr. Masako Okamoto, and Dr. Masami Yamashita for their helpful discussion for course of this work.

References and notes

- Tehlivets, O.; Malanovic, N.; Visram, M.; Pavkov-Keller, T.; Keller, W. Biochim. Biophys. Acta 2013, 1832, 204.
- 2. McCully, K. S. Am. J. Pathol. 1969, 56, 111.
- 3. Clarke, R. JAMA 2002, 288, 2015.
- (a) Nygård, O.; Vollset, S. E.; Refsum, H.; Stensvold, I.; Tverdal, A.; Nordrehaug, J. E.; Ueland, M.; Kvåle, G. JAMA 1995, 274, 1526; (b) Clarke, R. JAMA 2002, 288, 2015.
- (a) Seshadri, S.; Beiser, A.; Selhub, J.; Jacques, P. F.; Rosenberg, I. H.; D'Agostino, R. B.; Wilson, P. W.; Wolf, P. A. N. Eng. J. Med. 2002, 346, 476; (b) Van Dam, F.; Van Gool, W. A. Arch. Gerontol. Geriatr. 2009, 48, 425.
- Stanger, O.; Fowler, B.; Pietrzik, K.; Huemer, M.; Haschke-Becher, E.; Semmler, A.; Lorenzl, S.; Linnebank, M. Expert Rev. Neurother. 2009, 9, 1393.
- Langheinrich, A. C.; Braun-Dullaeus, R. C.; Walker, G.; Jeide, I.; Schiling, R.; Tammoscheit, K.; Dreyer, T.; Fink, L.; Bohle, R. M.; Haberbosch, W. Atherosclerosis 2003, 171, 181.
- 8. Yuan, C.-S.; Saso, Y.; Lazarides, E.; Borchardt, R. T.; Robins, M. J. Exp. Opin. Ther. Patents 1999, 9, 1197.
- (a) Wu, Q.-L.; Fu, Y.-F.; Zhou, W.-L.; Wang, J.-X.; Feng, Y.-H.; Liu, J.; Xu, J.-Y.; He, P.-L.; Zhou, R.; Tang, W.; Wang, G.-F.; Zhou, Y.; Yang, Y.-F.; Ding, J.; Li, X.-Y.; Chen, X.-R.; Yuan, C.; Lawson, B. R.; Zuo, J.-P. J. Pharmacol. Exp. Ther. 2005, 313, 705; (b) Yamada, T.; Komoto, J.; Lou, K.; Ueki, A.; Hua, D. H.; Sugiyama, K.; Takata, Y.; Ogawa, H.; Takusagawa, F. Biochem. Pharmacol. 2007, 73, 981; (c) Zhang, Y.-M.; Ding, Y.; Tang, W.; Luo, W.; Gu, M.; Lu, W.; Tang, J.; Zuo, J.-P.; Nan, F.-J. Bioorg. Med. Chem. 2008, 16, 9212; (d) Kim, B. G.; Chun, T. G. Bioorg. Med. Chem. 2009, 17, 6707; (e) Converso, A.; Hartingh, T.; Fraley, M. E.; Garbaccio, R. M.; Hartman, G. D.; Huang, S. Y.; Majercak, J. M.; McCampbell, A.; Na, S. J.; Ray, W. J.; Savage, M. J.; Wolffe, C.; Yeh, S.; Yu, Y.; White, R.; Zhang, R. Bioorg. Med. Chem. Lett. 2014, 24, 2737.
- (a) Nakao, A.; Suzuki, H.; Tatsumi, R.; Seki, M.; Tanaka, M.; Setsuta, T.; Iwasaki, H. WO 2,009,125,853, 2009; (b) Tan, X.; Wang, P.; Nian, S.; Wang, G. Chem. Pharm. Bull. 2014, 62, 112.
- 11. (a) Annis, D. A.; Athanasopoulos, J.; Curran, P. J.; Felsch, J. S.; Kalghatgi, K.; Lee, W. H.; Nash, H. M.; Orminati, J.-P. A.; Rosner, K. E.; Shipps, J. G. W.; Thaddupathy, G. R. A.; Tyler, A. N.; Vilenchik, L.; Wagner, C. R.; Wintner, E. Int. J. Mass Spectrom. 2004, 238, 77; (b) Annis, D. A.; Nickbarg, E.; Yang, X.; Ziebell, M. R.; Whitehurst, C. E. Curr. Opin. Chem. Biol. 2007, 11, 518.
- Nakao, A.; Suzuki, H.; Ueno, H.; Iwasaki, H.; Setsuta, T. Bioorg. Med. Chem. Lett. 2014, 24, 4336.
- Schulz, G. E.; Schirmer, R. H. Principles of Protein Structure; Springer: New York, 1979.
- (a) Itai, A.; Toriumi, Y.; Saito, S.; Kagechika, Hiroyuki; Shudo, K. J. Am. Chem. Soc.
 1992, 114, 10649; (b) Laursen, J. S.; Engel-Andreasen, J.; Fristrup, P.; Harris, P.; Olsen, C. A. J. Am. Chem. Soc. 2013, 135, 2835.
- 15. The previous paper presented that compound **18a** showed competitive inhibition against AdoHcyase with a K_i value of 1.5 nM.¹²
- 16 (a) Hu, Y.; Komoto, J.; Huang, Y.; Gomi, T.; Ogawa, H.; Takata, Y.; Fujioka, M.; Takusagawa, F. Biochemistry 1999, 38, 8323; (b) Turner, M. A.; Yuan, C. S.; Borchardt, R. T.; Hershfield, M. S.; Smith, G. D.; Howell, P. L. Nat. Struct. Biol. 1998, 5, 369; (c) Komoto, J.; Huang, Y.; Gomi, Y.; Ogawa, H.; Takata, Y.; Fujioka, M.; Takusagawa, F. J. Biol. Chem. 2000, 275, 32147; (d) Huang, Y.; Komoto, J.; Takata, Y.; Powell, D. R.; Gomi, T.; Ogawa, H.; Fujioka, M.; Takusagawa, F. J. Biol. Chem. 2002, 277, 7477; (e) Takata, Y.; Yamada, T.; Huang, Y.; Komoto, J.; Gomi, T.; Ogawa, H.; Fujioka, M.; Takusagawa, F. J. Biol. Chem. 2002, 277, 22670; (f) Yang, X.; Hu, Y.; Yin, D. H.; Turner, M. A.; Wang, M.; Borchardt, R. T.; Howell, P. L.; Kuczera, K.; Schowen, R. L. Biochemistry 2003, 42, 1900; (g) Yamada, T.; Takata, Y.; Komoto, J.; Gomi, T.; Ogawa, H.; Fujioka, M.; Takusagawa, F. Int. J. Biochem. Cell Biol. 2005, 37, 2417; (h) Yamada, T.; Komoto, J.; Lou, K.; Ueki, A.; Hua, D. H.; Sugiyama, K.; Takata, Y.; Ogawa, H.; Takusagawa, F. Biochem. Pharmacol. 2007, 37, 981; (i) Lee, K. M.; Choi, W. J.; Lee, Y.; Lee, H. J.; Zhao, L. X.; Lee, H. W.; Park, J. G.; Kim, H. O.; Hwang, K. Y.; Heo, Y. S.; Choi, S.; Jeong, L. S. J. Med. Chem. 2011, 54, 930.
- (a) Ma, J. C.; Dougherty, D. A. Chem. Rev. **1997**, 97, 1303; (b) Gallivan, J. P.; Dougherty, D. A. Proc. Natl. Acad. Sci. U.S.A. **1999**, 96, 9459; (c) Meyer, E. A.; Castellano, R. K.; Diederich, F. Angew. Chem., Int. Ed. **2003**, 42, 1210; (d) Salonen, L. M.; Ellermann, M.; Diederich, F. Angew. Chem., Int. Ed. **2011**, 50, 4808.
- (a) Hunter, C. A.; Sanders, J. K. M. J. Am. Chem. Soc. 1990, 112, 5525; (b) McGaughey, G. B.; Gagne, M.; Rappe, A. K. J. Biol. Chem. 1998, 273, 15458.
- 19. Kuhn, B.; Mohr, P.; Stahl, M. J. Med. Chem. 2010, 53, 2601.
- 20. (a) Bates, R. B.; Hruby, V. J.; Kriek, G. R. Acta Crystallogr. 1979, B35, 188; (b) Gellman, S. H.; Dado, G. P.; Liang, G.-B.; Adams, B. R. J. Am. Chem. Soc. 1991, 113, 1164; (c) Benjamin, W.-G.; Zhaohai, Z. Tetrahedron Lett. 1996, 37, 2189; (d) Yang, D.; Ng, F.-F.; Li, Z.-J.; Wu, Y.-D.; Chan, K. W. K.; Wang, D.-P. J. Am. Chem. Soc. 1996, 118, 9794; (e) Abele, S.; Seiler, P.; Seebach, D. Eur. J. Org. Chem. 2000, 20. 1.

21. Raiford, L. C.; Colbert, J. C. J. Am. Chem. Soc. 1926, 48, 2654.

18

ARTICLE IN PRESS

A. Nakao et al./Bioorg. Med. Chem. xxx (2015) xxx-xxx

- 22. McCombie, H.; Macmillan, W. G.; Scarborough, H. A. J. Chem. Soc. 1931, 529.
- 23. Gibbs, C. G. U.S. Patent 4,272,284, 1981.
- 24. Metz, H. J.; Neunhoeffer, H. Chem. Ber. 1982, 115, 2807.
- 25. Johannes, C. J.; Lotte, G. S.; Hans, T.; Poul, J. Bioorg. Med. Chem. Lett. 2004, 14, 1741.
- Biel, J. H.; Drukker, A. E.; Mitchell, T. F., Jr. J. Am. Chem. Soc. 1960, 82, 2204.
 Cannon, J. G.; Perez, J. A.; Pease, J. P.; Long, J. P.; Flynn, J. R.; Rusterholz, D. B.;
- Cannon, J. G.; Perez, J. A.; Pease, J. P.; Long, J. P.; Hynn, J. R.; Rusterholz, D. B. Dryer, S. E. J. Med. Chem. 1980, 23, 745.
- Yuan, C.-S.; Yeh, J.; Liu, S.; Borchardt, R. T. *J. Biol. Chem.* **1993**, *268*, 17030.
 SIR92: Altomare, A.; Cascarano, G.; Giacovazzo, C.; Guagliardi, A.; Burla, M.; Polidori, G.; Camalli, M. *J. Appl. Crystallogr.* **1994**, *27*, 435.
- (a) CrystalStructure 4.0: Crystal Structure Analysis Package; Rigaku Corporation: Tokyo 196-8666, Japan, 2000–2010; (b) Carruthers, J. R.; Rollett, J. S.; Betteridge, P. W.; Kinna, D.; Pearce, L.; Larsen, A.; Gabe, E. CRYSTALS Issue 11; Chemical Crystallography Laboratory: Oxford, UK, 1999.
- 31. Richards, H. H.; Chiang, P. K.; Cantoni, G. L. J. Biol. Chem. 1978, 253, 4476.