Accepted Manuscript

Reaction of β -alkoxyvinyl α -ketoesters with acyclic *NCN* binucleophiles – Scalable approach to novel functionalized pyrimidines

Oleksandr O. Stepaniuk, Tymofii V. Rudenko, Bohdan V. Vashchenko, Vitalii O. Matvienko, Ivan S. Kondratov, Andrey A. Tolmachev, Oleksandr O. Grygorenko

PII: S0040-4020(19)30512-5

DOI: https://doi.org/10.1016/j.tet.2019.05.005

Reference: TET 30326

To appear in: Tetrahedron

Received Date: 21 February 2019

Revised Date: 29 April 2019

Accepted Date: 2 May 2019

Please cite this article as: Stepaniuk OO, Rudenko TV, Vashchenko BV, Matvienko VO, Kondratov IS, Tolmachev AA, Grygorenko OO, Reaction of β -alkoxyvinyl α -ketoesters with acyclic *NCN* binucleophiles – Scalable approach to novel functionalized pyrimidines, *Tetrahedron* (2019), doi: https://doi.org/10.1016/j.tet.2019.05.005.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.





ACCEPTED MANUSCRIPT



Tetrahedron journal homepage: www.elsevier.com



Reaction of β -alkoxyvinyl α -ketoesters with acyclic *NCN* binucleophiles – scalable approach to novel functionalized pyrimidines

Oleksandr O. Stepaniuk,^{a,b} Tymofii V. Rudenko,^{a,b} Bohdan V. Vashchenko,^{a,b} Vitalii O. Matvienko,^a Ivan S. Kondratov,^{a,c}* Andrey A. Tolmachev,^{a,b} Oleksandr O. Grygorenko^{a,b}*

^a Enamine Ltd., Chervonotkatska Street 78, Kyiv 02094, Ukraine, <u>www.enamine.net</u>

^b Taras Shevchenko National University of Kyiv, Volodymyrska Street, 60, Kyiv 01601, Ukraine

^c V.P. Kukhar Institute of Bioorganic Chemistry & Petrochemistry, NAS of Ukraine, Murmanska Street 1, Kyiv 02660, Ukraine

gregor@univ.kiev.ua (O. O. G.); kondratov@mail.enamine.net (I. S. K.)

ARTICLE INFO

Article history: Received Received in revised form Accepted Available online Keywords: nitrogen heterocycles pyrimidines amidines guanidines

ABSTRACT

Two protocols for synthesis of series of low-molecular-weight di- and tri-substituted pyrimidines bearing a functional group at the 4th position, which rely on a base-mediated condensation of amidines or guanidines with β -alkoxyvinyl α -keto esters, have been developed. This approach allowed for multigram preparation of novel pyrimidine-4-carboxylates in 21–90% yield. The synthetic utility of these compounds was demonstrated by some standard functional group transformations providing promising building blocks for organic synthesis and drug discovery.

2019 Elsevier Ltd. All rights reserved.

1. Introduction

building blocks

Pyrimidine is one of the most important heterocycles, which is an essential constituent of natural compounds such as nucleic acids and vitamins (thiamine, riboflavin and folic acid).^{1,2} It is not surprising therefore that pyrimidine derivatives are extremely popular in drug discovery;³ they are represented by numerous examples of marketed drugs with a broad range of pharmacological activities, *i.e.* anticancer,⁴⁻¹⁰ antiviral,¹¹⁻¹⁴ anxiolytic,¹⁵ antioxidant,¹⁶⁻¹⁸ antifungal,¹⁹ anticonvulsant,²⁰ and antibacterial agents.^{5,21} Within just 2017–2018, several pyrimidine-containing drugs were approved by FDA.¹⁹



Figure 1 Examples of pyrimidine-containing drugs



Scheme 1 Synthesis of pyrimidine-4-carboxylates by reactions of 1,3-dicarbonyl compounds and *NCN* binucleophiles

A general approach to pyrimidine derivatives relies on the principal two-component condensation of *NCN* binucleophiles with *CCC* bis-electrophiles. 1,3-Dicarbonyl compounds are commonly used for that; however, this variation of the method have found limited application for the preparation of pyrimidine-4-carboxylates. Most reported examples referred to reactions of oxaloacetic acid esters with amidines, thiuronium salts or guanidine (Scheme 1, **A**).^{22–27} In another work, benzoyl- or (2-thienoyl)pyruvate were first transformed into mixtures of the corresponding β -chlorovinylketones, which then reacted with 1-(β -d-ribofuranosyl)formamidine to give the corresponding pyrimidine derivatives in 32–36% overall yields (**B**).²⁸

β-Alkoxyvinyl carbonyl compounds are renown synthetic equivalents of 1,3-dicarbonyl compounds which were widely used for the synthesis of numerous heterocyclic systems, including pyrimidines, and often led to better yields and purity of the target products.^{29,30} Utility of these bis-electrophiles was demonstrated mainly by the preparation of substituted pyrimidines bearing alkyl, haloalkyl or aryl moiety at the C-4 position.³¹⁻⁶² It should be noted that the related methods were reported in the patent literature.⁶³⁻⁶⁷ Some published papers described synthesis of pyrimidine-5-carboxylates.^{29,55,56,68} In the case of pyrimidine-4-carboxylates, only a few isolated examples were published, in particular, reactions of β-alkoxyvinyl α-ketoesters with only two specific substrates: 2-methylisothiourea or 2-benzylisothiourea (Scheme 2, *C*).⁶⁹ In turn, the reaction of amidines or guanidine (*D*)⁷⁰⁻⁷³ led to the corresponding pyrimidines in generally poor yields (up to 36%). Therefore, no optimized protocols were developed.



 R^1 = Me, Bn; R^2 = Me, Ph, p-FC₆H₄, p-BrC₆H₄, p-MeC₆H₄, p-MeOC₆H₄, p-O₂NC₆H₄



Scheme 2 Synthesis of pyrimidine-4-carboxylates by reactions of β -alkoxyvinyl carbonyl compounds and *NCN* binucleophiles

Furthermore, aryl-substituted binucleophiles were used predominantly - they are generally more convenient to handle and often have distinct reactivity as compared to their aliphatic counterparts. This is an example of recent finding by Churcher and co-authors,⁷⁴ who outlined that in many publications, the scope of the synthetic methods have been shown only for the most "convenient" substrates. Further application of such results by medicinal and industrial chemists often requires additional optimization of the protocols.

In this work, we have aimed at thorough and comprehensive studying of the lowest-molecular-weight β-alkoxyvinyl carbonyl compounds 1–3 as bis-electrophiles in the reactions with classical NCN binucleophiles, *i.e.* amidines (4-10) S-methylthiuronium salt (11), and guanidines (12-15) (Figure 2). These biselectrophiles were scarcely studied in the pyrimidine synthesis to date; in fact, most of the resulting pyrimidine-4-carboxylates obtained in this work were not described in the literature so far. In addition to that, application of the resulting products for preparation of multipurpose building blocks relevant for drug discovery was envisaged. As a result of this, carboxylic acids (8 examples), alcohols (7 examples), aldehydes (3 examples), chlorides (4 examples) and amines (4 examples) were obtained on multigram scale; 43 out of 51 pyrimidines prepared were novel compounds. Nearly all compounds are functionalized lowmolecular-weight building blocks highly compatible with stringent compound quality criteria which are still important to drug discovery.⁷⁷



Figure 2 Substrates studied in this work

2. Results/Discussion

The compounds 1–3 were prepared according to the known literature method.⁷⁶ Typical procedure for the reaction of 1–3 with binucleophiles 4–15 included heating of the starting materials at 70 °C in the presence of K_2CO_3 in MeCN for 2 h (method A). The reaction proceeded smoothly only in the case of 3, which appeared to be the most reactive bis-electrophile in the series studied, providing the target pyrimidines $16\{3,4-10\}$ and $16\{3,13-15\}$ in 67–90% yield (Table 1, Entries 16–26). Reaction of 3 with trifluoroacetamidine (7) did not require a base, providing 2-(trifluoromethyl)pyrimidine $16\{3,7\}$, which was used in the next step without isolation and purification (Entry 19).

It should be noted that the reaction of 3 and guanidines 13-15 was accompanied with trans-esterification of the CO₂Et moiety with MeOH formed, so that the corresponding pyrimidines bearing the dialkylamino moiety $16\{3,13-15\}$ contained up to 33% of the corresponding methyl ester $17\{3,13-15\}$. Since formation of this side product was not a problem for further

transformations, the resulting mixtures of $16\{3,13-15\}$ and M $17\{3,13-15\}$ were used in the next steps without additional purification (Table 1, Entries 24-26).

In the case of guanidine 12, the reaction with 3 using method A appeared to be too slow; therefore, alternative reaction conditions were developed, *i.e.* heating of the starting compounds in the presence of Et_3N in 1,4-dioxane at 100 °C for 48 h (method B), which gave the target product $16\{3,12\}$ in 82% yield (Table 1, Entry 23).

It was found that the method **B** was superior to the method **A** in the case of the less reactive 1,3-bis-electrophile **1** and amidines **5**, **8** or **10** (**A**: 39–57% yield; **B**: 65–80% yield; Table 1, Entries 2, 4 and 5). Lower yields and purity of the target products in the case of method **A** could be related to partial hydrolysis of the starting amidine with H_2O formed in the reaction; moreover, **Table 1**. Synthesis of substituted ethyl pyrimidine-4-carboxyla

animonia which was also released reacted with 1 to give the corresponding enaminone, thus complicating isolation of the target product. Therefore, the method **B** was used for less reactive *NCN* binucleophiles in the reaction with 1 (75–85% yield, Entries 6–8), as well as for all experiments with the compound 2 (71–86% yield, Entries 9–15), which appeared to be the least reactive among the 1,3-bis-electrophiles studied, possibly due to the steric hindrance around the ketone moiety.

Unfortunately, the method **B** was unfruitful in all experiments with formamidine acetate (4) (Entries 1, 9 and 16); meanwhile, the method **A** was applied for the preparation of the corresponding pyrimidines $16\{1,4\}$ and $16\{3,4\}$ more or less successfully (21% and 72% yield, respectively).

Table 1. Synthesis of substituted ethyl pyrimidine-4-carboxylates 16{1–3,4–15}								
	R ² O	A ⁻	<i>method A:</i> K ₂ CO ₃ , MeCN, 70 °C, 2 h	R^1 CO_2Et	R ² CO ₂ Me			
	RO CO-Ft +	$H_2N \downarrow NH_2^+$	method B:		+ N_N			
	R ¹	R ³	Et₃N, 1,4-dioxane, 100 °C, 48 h 65–85%	${R^3}$	\mathbf{R}^{3}			
	$A^- = CI, OAc, or free base (7)$			16{1-3,4-15}	17{3,13–15}			
#	β -Alkoxyvinyl α -ketoester	NCN binucleophile	Product	R ³	Yield, %	Method		
1		4	16{1,4}	Н	21 (A), 0 (B) ^a	A/B		
2		5	16{1,5}	Ме	39 (A), 65 (B) ^a	A/B		
3	0	6	16{1,6}	CH ₂ Cl	69	Α		
4	\sim	8	16{1,8}	cyclopropyl	53 (A), 73 (B) ^a	A/B		
5	EtO CO ₂ Et	10	16{1,10}	p-ClC ₆ H ₄	57 (A), 80 (B) ^a	A/B		
6	1	11	16{1,11}	SMe	75	В		
7		12	16{1,12}	NH ₂	76	В		
8		14	16{1,14}	pyrrolidin-1-yl	85	В		
9		4	16{2,4}	Н	0	A/B		
10		5	16{2,5}	Me	71	В		
11	O II	8	16{2,8}	cyclopropyl	76	В		
12	EtO CO ₂ Et	10	16{2,10}	p-ClC ₆ H ₄	83	В		
13	2	11	16{2,11}	SMe	75	В		
14		12	16{2,12}	NH ₂	77	В		
15		14	$16{2,14}$	pyrrolidin-1-yl	85	В		
16		4	16{3,4}	Н	72	Α		
17		5	16{3,5}	Me	78	Α		
18		6	16{3,6}	CH ₂ Cl	67	Α		
19		7	16{3,7}	CF ₃	N/A^d	\mathbf{A}^{c}		
20	QMe Q	8	16{3,8}	cyclopropyl	74	Α		
21	CO ₂ Et	9	16{3,9}	Ph	81	Α		
22	3	10	16{3,10}	p-ClC ₆ H ₄	83	Α		
23		12	$16{3,12}$	NH ₂	82	В		
24		13	$16\{3,\!13\}\!+\!17\{3,\!13\}$	NMe ₂	90 (16 : 17 = 2.5 :1) ^b	Α		
25		14	$16\{3,\!14\}\!+\!17\{3,\!14\}$	pyrrolidin-1-yl	90 (16 : 17 = 2:1) ^b	Α		
26		15	$16\{3,\!15\}\!+\!17\{3,\!15\}$	piperidin-1-yl	89 (16 : 17 = 2:1) ^b	Α		

^aBoth methods \mathbf{A} and \mathbf{B} were evaluated

^b16:17 ratio by ¹H NMR – the product was obtained as a mixture of 16 and 17 at the given ratio and was used in the next step without additional purification ^cThe reaction was performed without K₂CO₃

^dThe compound was used in the next step without isolation

The least reactive β -alkoxyvinyl α -ketoester 2 gave no target product 16{2,4} under either reaction conditions. In the case of chloromethyl-substituted amidine 6 (Entries 3 and 18), the method B could not be used due to alkylation of Et₃N with 6; nevertheless, the corresponding products 16{1,6} and 16{3,6} could be obtained using the method A.

It is important to outline that 20 of 24 synthesized pyrimidine ester derivatives $16\{1-3,4-15\}$ are novel compounds which were not described in the literature to date.

To demonstrate utility of the target esters **16**, their transformations into building blocks **18–22** was performed *via* standard carboxylate group modifications. (Figure 3, see the Supporting Information for more details).



3. Conclusion

The reaction of β -alkoxyvinyl α -keto esters 1–3 with amidines 4-10, S-methylthiuronium salt 11, and guanidines 12-15 in the presence of K₂CO₃ in MeCN (method A, 21-90% yield) or upon treatment with Et₃N in 1,4-dioxane (method **B**, 65–85% yield) is approach to the preparation an efficient of 4carboxylalkylpyrimidines 16. The method A was fruitful for the most reactive substrates (i.e. 1,3-bis-electrophile 3 and amidines), whereas in most other cases, the method **B** was more expedient. Utility of these key intermediates was confirmed by some typical functional group transformations; 43 out of 51 products were novel compounds. Most of these functionalized pyrimidine derivatives are low-molecular-weight and hydrophilic; they were prepared on multigram scale and are therefore promising building blocks for organic synthesis, drug discovery and agrochemistry, which are readily available to scientific community and can significantly expand the currently accessible lead-like chemical space (Figure S1).

4. Experimental section

The solvents were purified according to the standard procedures.⁷⁷ The compounds 1-3,⁷⁶ 4,⁷⁸ 5,⁷⁹ 6,⁸⁰ 7,⁸¹ 8,⁸² 9 and 10,³³ 11,⁸³ 13,⁸³ 14 and 15^{84} were prepared according to the literature methods. All other reagents and starting materials were obtained from commercial sources. Melting points were measured on MPA100 OptiMelt automated melting point system. Analytical TLC was performed using Polychrom SI F254 plates. Column chromatography was performed using silica gel (230–400 mesh) as the stationary phase. ¹H and ¹³C NMR spectra were recorded on a Bruker 170 Avance 500 spectrometer (at 500 MHz for ¹H NMR, 126 MHz for ¹³C NMR and 470 MHz for ¹⁹F NMR) and Varian Unity Plus 400 spectrometer (at 400 MHz for ¹⁹F NMR). NMR

chemical shifts are reported in ppm (δ scale) downfield from TMS as an internal standard and are referenced using residual NMR solvent peaks at 7.26 and 77.16 ppm for ¹H and ¹³C in CDCl₃, 2.50 and 39.52 ppm for ¹H and ¹³C in DMSO- d_6 . Coupling constants (J) are shown in Hz. Spectra are reported as follows: chemical shift (δ , ppm), multiplicity, integration, coupling constants (Hz). Elemental analyses were performed at the Laboratory of Organic Analysis, Department of Chemistry, National Taras Shevchenko University of Kyiv. Mass spectra were recorded on an Agilent 1100 LCMSD SL instrument (chemical ionization (CI)) and Agilent 5890 Series II 5972 GCMS instrument (electron impact ionization (EI)). Elemental analysis was performed using Elementar Vario MICRO Cube CHNS/O/Cl analyzer. Instant JChem v. 17.2.27.0 was used for the calculation of physico-chemical parameters, Chemaxon, Hungary, www.chemaxon.com

4.1. General procedure for the preparation of esters **16** (*Method* **A**).

 K_2CO_3 (27.6 g. 0.180 mol) was added to a stirred solution of the corresponding β-alkoxyvinyl α-ketoester 1–3 (0.100 mol) in MeCN (300 mL) and the resulting mixture was heated to 70 °C. Then, the corresponding *NCN* binucleophile 4–15 (0.130 mol, or 0.180 mol in the case of 4) was added in portions. The reaction mixture was refluxed for 2 h, then cooled to rt and evaporated in *vacuo*. The residue was diluted with H₂O (200 mL) and extracted with *t*-BuOMe (2×100 mL) (in the case of 16{1,8}, 16{1,10} and 16{3,9–10}) or CH₂Cl₂ (2×100 mL) (in other cases). The organic phase was separated, washed with H₂O (50 mL), dried over Na₂SO₄, filtered through silica gel and evaporated in *vacuo*.

The products $16{3,13-15}$ contained the corresponding methyl esters $17{3,13-15}$ formed by trans-esterification upon the reaction with ethyl 4-methoxy-2-oxopent-3-enoate (3).

Ethyl pyrimidine-4-carboxylate 16{1,4}.⁸⁵

Yielg 3.19 g (21 %); yellow crystals; mp = 39–41 °C (hexanes) (lit.⁸⁵ 35–36 °C). ¹H NMR (500 MHz, CDCl₃) δ 9.36 (d, J = 1.4 Hz, 1H), 8.94 (d, J = 5.0 Hz, 1H), 7.97 (dd, J = 5.0, 1.4 Hz, 1H), 4.45 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 163.5, 158.8, 158.7, 154.5, 120.5, 62.3, 13.7. LC/MS (CI): m/z = 153 [M+H]⁺. Anal. calcd. for C₇H₈N₂O₂: C, 55.26; H, 5.30; N, 18.41. Found: C, 55.37; H, 5.25; N, 18.78.

Ethyl 2-(chloromethyl)pyrimidine-4-carboxylate 16{1,6}.

The compound was purified by distillation in *vacuo*. Yield 13.8 g (69%); brownish liquid; bp = 102-104 °C / 1 mmHg. ¹H NMR (500 MHz, CDCl₃) δ 8.97 (d, *J* = 4.9 Hz, 1H), 7.87 (d, *J* = 4.9 Hz, 1H), 4.80 (s, 2H), 4.45 (q, *J* = 7.1 Hz, 2H), 1.39 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 166.1, 163.2, 159.5, 155.3, 119.1, 62.4, 46.2, 13.7. GC/MS (EI): *m*/*z* = 156/158 [M-OEt+H]⁺, 173/175 [M-H₂C=CH(CH₃)+H]⁺, 201/203 [M+H]⁺. Anal. calcd. for C₈H₉CIN₂O₂: C, 47.90; H, 4.52; N, 13.96; Cl, 17.67. Found: C, 47.55; H, 4.75; N, 14.01; Cl, 17.71.

Ethyl 6-methylpyrimidine-4-carboxylate 16{3,4}.

Yield 12.0 g (72%); colorless crystals, mp = 32-34 °C (hexanes) (bp = 86-88 °C / 1 mmHg). ¹H NMR (500 MHz, CDCl₃) δ 9.22 (s, 1H), 7.85 (s, 1H), 4.46 (q, *J* = 7.2 Hz, 2H), 2.62 (s, 3H), 1.42 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 169.6, 164.3, 158.8, 154.6, 120.5, 62.6, 24.3, 14.1. LC/MS (CI): $m/z = 139 [\text{M-H}_2\text{C}=\text{CH}(\text{CH}_3)+\text{H}]^+$, 167 [M+H]⁺. Anal. calcd. for C₈H₁₀N₂O₂: C, 57.82; H, 6.07; N, 16.86. Found: C, 57.51; H, 6.43; N, 16.86.

*Ethyl 2,6-dimethylpyrimidine-4-carboxylate*EPTED M 465.4J 162.1, 154.9, 107.4, 61.2 36.4, 23.9. LC/MS (CI): $m/z = 16\{3,5\}$.

Yield 14.1 g (78%); colorless crystals, mp = 44–47 °C (hexanes) (bp = 94–96 °C / 1 mmHg). ¹H NMR (500 MHz, CDCl₃) δ 7.66 (s, 1H), 4.47 (q, J = 7.1 Hz, 2H), 2.79 (s, 3H), 2.59 (s, 3H), 1.42 (t, J = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 169.4, 168.7, 164.6, 155.0, 117.4, 62.5, 26.1, 24.3, 14.2. LC/MS (CI): m/z = 181[M+H]⁺. Anal. calcd. for C₉H₁₂N₂O₂: C, 59.99; H, 6.71; N, 15.55. Found: C, 59.91; H, 6.78; N, 15.34.

Ethyl 2-(chloromethyl)-6-methylpyrimidine-4-carboxylate 16{3,6}.

The compound was purified by distillation in *vacuo*. Yield 14.4 g (67%); yellow powder; mp = 36–38 °C; bp = 117–119 °C / 1 mmHg. ¹H NMR (500 MHz, CDCl₃) δ 7.77 (s, 1H), 4.78 (s, 2H), 4.48 (q, *J* = 7.2 Hz, 2H), 2.65 (s, 3H), 1.42 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 170.7, 166.2, 164.0, 155.5, 119.2, 62.7, 46.8, 24.4, 14.2. GC/MS (EI): *m/z* = 155/157 [M-OEt]⁺, 186/188 [M-H₂C=CH(CH₃)]⁺, 214/216 [M]⁺. Anal. calcd. for C₉H₁₁ClN₂O₂: C, 50.36; H, 5.17; N, 13.05; Cl, 16.52. Found: C, 50.25; H, 5.31; N, 12.73; Cl, 16.72.

Ethyl 2-cyclopropyl-6-methylpyrimidine-4-carboxylate 16{3,8}.

Yield 15.3 g (74%); yellowish liquid. ¹H NMR (400 MHz, CDCl₃) δ 7.53 (s, 1H), 4.42 (q, *J* = 7.1 Hz, 2H), 2.49 (s, 3H), 2.30 (tt, *J* = 8.2, 2.7 Hz, 1H), 1.39 (t, *J* = 7.1 Hz, 3H), 1.16 – 1.13 (m, 2H), 1.05 – 1.02 (m, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 172.6, 169.1, 164.8, 154.5, 116.9, 62.2, 24.3, 18.3, 14.2, 11.0. LC/MS (CI): *m*/*z* = 207 [M+H]⁺. Anal. calcd. for C₁₁H₁₄N₂O₂: C, 64.06; H, 6.84; N, 13.58. Found: C, 64.05; H, 6.92; N, 13.77.

Ethyl 6-methyl-2-phenylpyrimidine-4-carboxylate **16{3,9}**.

Yield 19.6 g (81%); yellowish solid; mp = 95–97 °C (hexanes). ¹H NMR (500 MHz, CDCl₃) δ 8.54 (s, 2H), 7.72 (s, 1H), 7.52 – 7.47 (m, 3H), 4.51 (q, *J* = 6.7 Hz, 2H), 2.69 (s, 3H), 1.48 (t, *J* = 6.7 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 169.6, 165.0, 164.8, 155.4, 137.1, 130.9, 128.5, 128.5, 118.0, 62.3, 24.6, 14.2. LC/MS (CI): *m*/*z* = 213 [M-Et]⁻, 243 [M+H]⁺. Anal. calcd. for C₁₄H₁₄N₂O₂: C, 69.41; H, 5.82; N, 11.56. Found: C, 69.47; H, 5.97; N, 11.35.

Ethyl 2-(4-chlorophenyl)-6-methylpyrimidine-4-carboxylate 16{3,10}.

Yield 23.0 g (83%); colorless powder; mp = 107–109 (hexanes). °C. ¹H NMR (500 MHz, CDCl₃) δ 8.49 (d, *J* = 8.5 Hz, 2H), 7.73 (s, 1H), 7.47 (d, *J* = 8.5 Hz, 2H), 4.51 (q, *J* = 7.0 Hz, 2H), 2.68 (s, 3H), 1.48 (t, *J* = 7.0 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 169.7, 164.7, 164.0, 155.4, 137.2, 135.6, 129.9, 128.7, 118.1, 62.4, 24.6, 14.2. LC/MS (CI): *m*/*z* = 277/279 [M+H]⁺. Anal. calcd. for C₁₄H₁₃ClN₂O₂: C,s 60.77; H, 4.74; N, 10.12; Cl, 12.81. Found: C, 60.55; H, 4.66; N, 10.01; Cl, 13.11.

Ethyl 2-(dimethylamino)-6-methylpyrimidine-4-carboxylate 16{3,13}.

The compound was obtained as a mixture with methyl 2-(dimethylamino)-6-methylpyrimidine-4-carboxylate **17{3,13}** (**16:17** = 2.5:1). Yield 18.8 g (90%); yellowish liquid. ¹H NMR (500 MHz, CDCl₃) δ 6.84 (s, 1H), 4.28 (q, *J* = 7.1 Hz, 2H), 3.11 (s, 6H), 2.29 (s, 3H), 1.29 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 168.9, 164.9, 162.1, 154.6, 107.3, 52.2, 36.3, 23.9, 13.7. LC/MS (CI): *m/z* = 210 [M+H]⁺.

Methyl 2-(dimethylamino)-6-methyl-pyrimidine-4carboxylate 17{3,13} (spectral data).

¹H NMR (500 MHz, CDCl₃) δ 6.86 (s, 1H), 3.82 (s, 3H), 3.11 (s, 6H), 2.29 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 169.0,

Ethyl 6-methyl-2-(pyrrolidin-1-yl)pyrimidine-4-carboxylate **16{3,14}**.

The compound was obtained as a mixture with methyl 6methyl-2-(pyrrolidin-1-yl)pyrimidine-4-carboxylate **17{3,14}** (**16:17** = 2:1). Yield 21.2 (90%); yellowish oil. ¹H NMR (400 MHz, CDCl₃) δ 6.92 (s, 1H), 4.35 (q, *J* = 6.9 Hz, 2H), 3.62 – 3.54 (m, 4H), 2.36 (s, 3H), 1.95 – 1.91 (m, 4H), 1.35 (t, *J* = 6.9 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 168.4, 165.5, 160.9, 155.2, 107.9, 52.8, 46.6, 25.4, 24.4, 14.2. LC/MS (CI): *m*/*z* = 236 [M+H]⁺.

Methyl 6-methyl-2-(pyrrolidin-1-yl)pyrimidine-4carboxylate 17{3,14} (spectral data).

¹H NMR (400 MHz, CDCl₃) δ 6.94 (s, 1H), 3.89 (s, 3H), 3.62 – 3.54 (m, 4H), 2.36 (s, 3H), 1.99 – 1.93 (m, 4H). ¹³C NMR (126 MHz, CDCl₃) δ 169.6, 166.1, 161.2, 155.6, 108.0, 61.8, 46.7, 25.5, 24.5. LC/MS (CI): m/z = 222 [M+H]⁺.

Ethyl 6-methyl-2-(piperidin-1-yl)pyrimidine-4-carboxylate 16{3,15}.

The compound was obtained as a mixture with methyl 6methyl-2-(piperidin-1-yl)pyrimidine-4-carboxylate **17{3,15}** (**16:17** = 2:1). Yield 22.2 g (89%); yellowish oil. ¹H NMR (500 MHz, CDCl₃) δ 6.94 (s, 1H), 4.40 (q, *J* = 7.0 Hz, 2H), 3.85 (s, 4H), 2.39 (s, 3H), 1.68 – 1.60 (m, 6H), 1.41 (t, *J* = 7.0 Hz, 3H).¹³C NMR (126 MHz, CDCl₃) δ 169.6, 165.5, 162.1, 155.6, 108.0, 61.8, 44.6, 25.8, 24.8, 24.5,14.2. LC/MS (CI): *m/z* = 250 [M+H]⁺.

Methyl 6-methyl-2-(piperidin-1-yl)pyrimidine-4carboxylate 17{3,15} (spectral data).

¹H NMR (500 MHz, CDCl₃) δ 6.96 (s, 1H), 3.94 (s, 3H), 3.87 - 3.86 (m, 4H), 2.39 (s, 3H), 1.68 - 1.60 (m, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 169.6, 165.5, 162.1, 155.6, 108.1, 61.8, 44.7, 25.8, 24.8, 24.5. LC/MS (CI): *m*/*z* = 236 [M+H]⁺.

4.2. General procedure for the preparation of esters **16** (*Method* **B**).

Et₃N (27.9 mL, 20.2 g, 0.200 mol) was added to a stirred mixture of β -alkoxyvinyl ester **1–3** (0.100 mol) and the corresponding amidine/guanidine **4–15** (0.120 mol) in 1,4-dioxane (200 mL). The resulting mixture was stirred at 100 °C for 48 h, then cooled to rt and evaporated in *vacuo*. The residue was diluted with H₂O (150 mL) and extracted with *t*-BuOMe (2×100 mL). The combined organic extracts were washed with water (50 mL), dried over Na₂SO₄, filtered through silica gel and evaporated in *vacuo*.

Ethyl 2-methylpyrimidine-4-carboxylate 16{1,5}.

Yield 10.8 g (65%; Method **A** – 39%); brownish liquid. ¹H NMR (400 MHz, CDCl₃) δ 8.81 (d, J = 4.9 Hz, 1H), 7.74 (d, J = 4.9 Hz, 1H), 4.44 (q, J = 7.1 Hz, 2H), 2.80 (s, 3H), 1.38 (t, J = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 169.1, 164.3, 159.0, 155.2, 117.7, 62.6, 26.2, 14.2. LC/MS (CI): m/z = 139 [M-H₂C=CH(CH₃)+H]⁺, 167 [M+H]⁺. Anal. calcd. for C₈H₁₀N₂O₂: C, 57.82; H, 6.07; N, 16.86. Found: C, 57.94; H, 5.68; N, 17.19.

Ethyl 2-cyclopropylpyrimidine-4-carboxylate **16**{**1**,**8**}.

Yield 14.0 g (73%; Method **A** – 53%); yellowish liquid. ¹H NMR (400 MHz, CDCl₃) δ 8.71 (d, *J* = 4.9 Hz, 1H), 7.63 (d, *J* = 4.9 Hz, 1H), 4.42 (q, *J* = 7.2 Hz, 2H), 2.38 – 2.30 (m, 1H), 1.38 (t, *J* = 7.2 Hz, 3H), 1.16 – 1.12 (m, 2H), 1.08 – 1.04 (m, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 173.1, 164.4, 158.9, 154.8, 117.2,

62.22; H, 6.54; N, 14.96.

62.4, 18.4, 14.1, 11.4. LC/MS (CI): $m/z = 193 [M+H]^{+}$. Anal. M *Ethyl* 2-(*methylthio*)-5-*methylpyrimidine-4*-calcd. for C₁₀H₁₂N₂O₂: C, 62.49; H, 6.29; N, 14.57. Found: C, *carboxylate* 16{2,11}.

Ethyl 2-(4-chlorophenyl)pyrimidine-4-carboxylate 16{1,10}.

Yield 21.0 g (80%; Method A – 57%); colorless crystals; mp = 90–93 °C (hexanes). ¹H NMR (500 MHz, CDCl₃) δ 8.99 (d, *J* = 4.9 Hz, 1H), 8.46 (d, *J* = 8.7 Hz, 2H), 7.83 (d, *J* = 4.9 Hz, 1H), 7.46 (d, *J* = 8.7 Hz, 2H), 4.50 (q, *J* = 7.1 Hz, 2H), 1.47 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 163.9, 158.9, 155.2, 137.1, 134.9, 129.8, 129.5, 128.5, 118.1, 62.1, 13.8. LC/MS (CI): *m*/*z* = 263/265 [M+H]⁺. Anal. calcd. for C₁₃H₁₁ClN₂O₂: C, 59.44; H, 4.22; N, 10.66; Cl, 13.49. Found: C, 59.56; H, 4.30; N, 10.86; Cl, 13.23.

*Ethyl 2-(methylthio)pyrimidine-4-carboxylate {1,11}.*⁸⁶

Yield 14.9 g (75%); yellowish liquid. ¹H NMR (500 MHz, CDCl₃) δ 8.59 (d, J = 4.9 Hz, 1H), 7.45 (d, J = 4.9 Hz, 1H), 4.30 (q, J = 7.1 Hz, 2H), 2.45 (s, 3H), 1.26 (t, J = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 173.3, 163.3, 158.6, 154.7, 115.0, 62.0, 13.7, 13.6. LC/MS (CI): m/z = 199 [M+H]⁺. Anal. calcd. for C₈H₁₀N₂O₂S: C, 48.47; H, 5.08; N, 14.13; S, 16.17. Found: C, 48.85; H, 4.75; N, 14.35; S, 16.54.

Ethyl 2-(pyrrolidin-1-yl)pyrimidine-4-carboxylate **16**{1,14}.

Yield 18.8 g (85%); yellowish crystals; mp = 54–57 °C (hexanes). ¹H NMR (500 MHz, CDCl₃) δ 8.45 (d, J = 4.7 Hz, 1H), 7.03 (d, J = 4.7 Hz, 1H), 4.38 (q, J = 7.2 Hz, 2H), 3.65 – 3.55 (m, 4H), 1.99 – 1.93 (m, 4H), 1.37 (t, J = 7.2 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 164.7, 160.2, 159.1, 155.5, 107.6, 61.5, 46.3, 25.0, 13.7. LC/MS (CI): m/z = 222 [M+H]⁺. Anal. calcd. for C₁₁H₁₅N₃O₂: C, 59.71; H, 6.83; N, 18.99. Found: C, 59.76; H, 6.97; N, 19.18.

Ethyl 2,5-dimethylpyrimidine-4-carboxylate **16{2,5}**.

Yield 13.0 g (71%); yellow liquid. ¹H NMR (500 MHz, CDCl₃) δ 8.45 (s, 1H), 4.30 (q, J = 7.1 Hz, 2H), 2.58 (s, 3H), 2.28 (s, 3H), 1.26 (t, J = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 165.4, 164.7, 160.1, 153.9, 126.3, 61.7, 25.0, 15.5, 13.6. LC/MS (CI): $m/z = 181[M+H]^+$. Anal. calcd. for C₉H₁₂N₂O₂: C, 59.99; H, 6.71; N, 15.55. Found: C, 60.17; H, 6.38; N, 15.45.

Ethyl 2-cyclopropyl-5-methylpyrimidine-4-carboxylate 16{2,8}.

Yield 15.7 g (76%); yellowish liquid. ¹H NMR (500 MHz, CDCl₃) δ 8.50 (s, 1H), 4.44 (q, J = 7.1 Hz, 2H), 2.39 (s, 3H), 2.28 (tt, J = 8.3, 3.9 Hz, 1H), 1.41 (t, J = 7.1 Hz, 3H), 1.12 – 1.09 (m, 2H), 1.07 – 1.03 (m, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 170.0, 165.4, 160.4, 154.3, 126.0, 62.0, 17.8, 15.8, 14.1, 10.8. LC/MS (CI): m/z = 135 [M-CO₂-H₂C=CH(CH₃)+H]⁺, 179 [M-H₂C=CH(CH₃)+H]⁺, 207 [M+H]⁺. Anal. calcd. for C₁₁H₁₄N₂O₂: C, 64.06; H, 6.84; N, 13.58. Found: C, 63.95; H, 6.68; N, 13.96.

Ethyl 2-(4-chlorophenyl)-5-methylpyrimidine-4-carboxylate 16{2,10}.

Yield 23.0 g (83%); brownish liquid. ¹H NMR (400 MHz, CDCl₃) δ 8.71 (s, 1H), 8.37 (d, J = 8.4 Hz, 2H), 7.46 – 7.38 (m, 2H), 4.47 (q, J = 7.1 Hz, 2H), 2.48 (s, 3H), 1.44 (t, J = 7.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 165.3, 161.7, 160.8, 155.1, 137.0, 135.4, 129.5, 128.8, 127.7, 62.2, 16.1, 14.2. LC/MS (CI): m/z = 277/279 [M+H]⁺. Anal. calcd. for C₁₄H₁₃ClN₂O₂: C, 60.77; H, 4.74; N, 10.12; Cl, 12.81. Found: C, 61.08; H, 4.50; N, 10.09; Cl, 13.04.

Yield 14.9 g (75%); brownish liquid. ¹H NMR (500 MHz, CDCl₃) δ 8.38 (s, 1H), 4.33 (q, J = 7.1 Hz, 2H), 2.46 (s, 3H), 2.28 (s, 3H), 1.31 (t, J = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 169.8, 164.4, 160.1, 154.5, 124.0, 61.6, 15.1, 13.7, 13.6. LC/MS (CI): m/z = 213 [M+H]⁺. Anal. calcd. for C₉H₁₂N₂O₂S: C, 50.93; H, 5.70; N, 13.20; S, 15.10. Found: C, 50.55; H, 6.07; N, 13.51; S, 15.17.

Ethyl 5-methyl-2-(pyrrolidin-1-yl)pyrimidine-4-carboxylate 16{2,14}.

Yield 20.0 g (85%); yellowish crystals; mp = 40–41 (hexanes) °C. ¹H NMR (500 MHz, CDCl₃) δ 8.21 (s, 1H), 4.37 (q, *J* = 7.1 Hz, 2H), 3.53 (s, 4H), 2.19 (s, 3H), 1.93 (s, 4H), 1.36 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 165.7, 160.5, 158.8, 155.3, 114.8, 61.2, 46.2, 25.1, 14.5, 13.7. LC/MS (CI): *m/z* = 236 [M+H]⁺. Anal. calcd. for C₁₂H₁₇N₃O₂: C, 61.26; H, 7.28; N, 17.86. Found: C, 61.56; H, 7.20; N, 17.89.

Ethyl 2-aminopyrimidine-4-carboxylate 16{1,12}.⁷³

After evaporation in *vacuo* of the reaction mixture, the residue was diluted with H₂O (100 mL) and *t*-BuOMe (30 mL). The precipitate formed was filtered, washed with H₂O (2×10 mL) and dried in *vacuo*. Yield 12.7 g (76%); beige powder; mp = 239–242 °C (EtOH). ¹H NMR (500 MHz, DMSO-*d*₆) δ 8.46 (dd, *J* = 4.9, 2.2 Hz, 1H), 7.05 (s, 2H), 7.04 (dd, *J* = 4.4, 1.5 Hz, 1H), 4.30 (q, *J* = 6.9 Hz, 2H), 1.29 (t, *J* = 6.9 Hz, 3H). ¹³C NMR (126 MHz, DMSO-*d*₆) δ 164.9, 164.4, 161.0, 156.1, 109.4, 61.9, 14.5. LC/MS (CI): *m*/*z*= 122 [M-OEt]⁺, 168 [M+H]⁺. Anal. calcd. for C₇H₉N₃O₂: C, 50.29; H, 5.43; N, 25.14. Found: C, 50.05; H, 5.15; N, 25.10.

Ethyl 2-amino-5-methylpyrimidine-4-carboxylate **16**{2,12}.

After evaporation in *vacuo* of the reaction mixture, the residue was diluted with H₂O (100 mL) and *t*-BuOMe (30 mL). The precipitate formed was filtered, washed with H₂O (2×10 mL) and dried in *vacuo*. Yield 13.8 g (75%); yellowish powder; mp = 195–199 °C (EtOH). ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.27 (s, 1H), 6.67 (s, 2H), 4.31 (q, *J* = 7.1 Hz, 2H), 2.14 (s, 3H), 1.30 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (126 MHz, DMSO-*d*₆) δ 165.8, 162.5, 161.9, 155.9, 116.38, 61.7, 14.7, 14.5. LC/MS (CI): *m*/z= 182 [M+H]⁺. Anal. calcd. for C₈H₁₁N₃O₂: C, 53.03; H, 6.12; N, 23.19. Found: C, 52.96; H, 6.44; N, 23.48.

Ethyl 2-amino-6-methylpyrimidine-4-carboxylate **16{3,12}**.⁸⁷

After evaporation in *vacuo* of the reaction mixture, the residue was diluted with H₂O (100 mL) and *t*-BuOMe (30 mL). The precipitate formed was filtered, washed with H₂O (2×10 mL) and dried in *vacuo*. Yield 14.9 g (82%); brownish powder; mp = 145–147 °C (EtOH) (lit.⁸⁷ 151 °C). ¹H NMR (500 MHz, DMSO-*d*₆) δ 6.96 (s, 1H), 6.90 (s, 2H), 4.29 (q, *J* = 7.0 Hz, 2H), 2.31 (s, 3H), 1.29 (t, *J* = 7.0 Hz, 3H). ¹³C NMR (126 MHz, DMSO-*d*₆) δ 170.3, 165.1, 164.3, 156.0, 109.0, 61.8, 24.1, 14.5. LC/MS (CI): *m*/*z*= 182 [M+H]⁺. Anal. calcd. for C₈H₁₁N₃O₂: C, 53.03; H, 6.12; N, 23.19. Found: C, 53.06; H, 6.43; N, 23.46.

Acknowledgments

The work was funded by Enamine Ltd.

References and notes

1. Gilchrist TL. Heterocyclic Chemistry, 3rd ed., Addison Wesley: Essex, England, 1997.

- 2. Joule JA, Mills K. Heterocyclic Chemistry, 5th ed., D MANUSHosagrahara V, Hameed S, Shinde V, Bathula C, Wiley-Blackwell, 2010. Humnabadkar V, Kumar N, Reddy J, Panduga V,
- Panneer Selvam T, Richa James C, Vijaysarathy Dniandev P, Karyn Valzita S. *Res Pharm*. 2012; 2: 1– 9.
- 4. Kaur R, Kaur P, Sharma S, Singh G, Mehndiratta S, Bedi PMS, Nepali K. Anti-cancer pyrimidines in diverse scaffolds: a review of patent literature., 2015, vol. 10.
- 5. Rostom SAF, Ashour HMA, Abd El Razik HA. *Arch Pharm.* 2009; 342: 299–310.
- 6. Reynolds RC, Tiwari a, Harwell JE, Gordon DG, Garrett BD, Gilbert KS, Schmid SM, Waud WR, Struck RF. *J Med Chem.* 2000; 43: 1484–8.
- Iyer V V., Griesgraber GW, Radmer MR, McIntee EJ, Wagner CR. J Med Chem. 2000; 43: 2266–2274.
- 8. El-Deeb IM, Lee SH. *Bioorg Med Chem.* 2010; 18: 3860–3874.
- 9. Yoon J seong, Jarhad DB, Kim G, Nayak A, Zhao LX, Yu J, Kim HR, Lee JY, Mulamoottil VA, Chandra G, Byun WS, Lee SK, Kim YC, Jeong LS. *Eur J Med Chem.* 2018; 155: 406–417.
- 10. Kumar S, Narasimhan B. Chem Cent J. 2018; 12: 38.
- Tarnchompoo B, Sirichaiwat C, Phupong W, Intaraudom C, Sirawaraporn W, Kamchonwongpaisan S, Vanichtanankul J, Thebtaranonth Y, Yuthavong Y. *J Med Chem.* 2002; 45: 1244–1252.
- 12. Kraljević TG, Klika M, Kralj M, Martin-Kleiner I, Jurmanović S, Milić A, Padovan J, Raić-Malić S. *Bioorg Med Chem Lett.* 2012; 22: 308–312.
- Choi Y, Li L, Grill S, Gullen E, Lee CS, Gumina G, Tsujii E, Cheng YC, Chu CK. J Med Chem. 2000; 43: 2538–2546.
- Hopkins AL, Ren J, Tanaka H, Baba M, Okamato M, Stuart DI, Stammers DK. J Med Chem. 1999; 42: 4500–4505.
- 15. Chinnasamy G, Subramani K, Srinivasan V. Biomed Res-India. 2017; 28: 525–531.
- 16. Mansouri M, Movahedian A, Rostami M, Fassihi A. *Res Pharm Sci.* 2012; 7: 257.
- 17. Bano T, Kumar N, Dudhe R. *Org Med Chem Lett*. 2012; 2: 34.
- 18. Kumar S, Narasimhan B. Chem Cent J. 2018; 12: 38.
- 19. De La Torre BG, Albericio F. *Molecules*. 2018; 23.
- 20. Alam O, Mullick P, Verma SP, Gilani SJ, Khan SA, Siddiqui N, Ahsan W. *Eur J Med Chem.* 2010; 45: 2467–2472.
- 21. Mohana Roopan S, Sompalle R. *Synth Commun.* 2016; 46: 645–672.
- 22. Shirude PS, Shandil R, Sadler C, Naik M,

Humnabadkar V, Kumar N, Reddy J, Panduga V, Sharma S, Ambady A, Hegde N, Whiteaker J, McLaughlin RE, Gardner H, Madhavapeddi P, Ramachandran V, Kaur P, Narayan A, Guptha S, Awasthy D, Narayan C, Mahadevaswamy J, Vishwas KG, Ahuja V, Srivastava A, Prabhakar K, Bharath S, Kale R, Ramaiah M, Choudhury NR, Sambandamurthy VK, Solapure S, Iyer PS, Narayanan S, Chatterji M. *J Med Chem*. 2013; 56: 9701–9708.

- 23. Webb TR, Moran T, Huang CQ, McCarthy JR, Grigoriadis DE, Chen C. *Bioorg Med Chem Lett*. 2004; 14: 3869–3873.
- Gillespie RJ, Bamford SJ, Clay A, Gaur S, Haymes T, Jackson PS, Jordan AM, Klenke B, Leonardi S, Liu J, Mansell HL, Ng S, Saadi M, Simmonite H, Stratton GC, Todd RS, Williamson DS, Yule IA. *Bioorg Med Chem.* 2009; 17: 6590–6605.
- Provenzani R, Tarvainen I, Brandoli G, Lempinen A, Artes S, Turku A, Jäntti MH, Talman V, Yli-Kauhaluoma J, Tuominen RK, Boije af Gennäs G. *PLoS One.* 2018; 13: 1–27.
- Masami Otsuka, Kobayashi S, Ohno M, Umezawa Y, Morishima H, Umezawa H. *Chem Pharm Bull*. 1985; 33: 515–519.
- 27. Ti JS, Steinfeld AS, Naider F, Gulumoglu A, Lewis S V., Becker JM. *J Med Chem.* 1980; 23: 913–918.
- Iaroshenko VO, Dudkin S, Sosnovskikh VY, Villinger A, Langer P. *Eur J Org Chem*. 2013: 3166– 3173.
- Kudyakova YS, Bazhin DN, Goryaeva M V, Burgart Y V, Saloutin VI. *Russ Chem Rev.* 2014; 83: 120– 142.
- Krapcho AP, Maresch MJ, Helgason AL, Rosner KE, Hacker MP, Oliva SS, Menta E, Oliva A. J Het Chem. 1993; 30: 1597–1606.
- 31. Franke W, Kraft R. Angew Chem. 1955; 67: 395–399.
- Zanatta N, Fagundes MB, Ellensohn R, Marques M, Bonacorso HG, Marcos A. P. Martins. *J Heterocycl Chem.* 1998; 35: 451–455.
- 33. Golubev PR, Pankova AS, Kuznetsov MA. *Eur J Org Chem.* 2014; 2014: 3614–3621.
- 34. Bellur E, Langer P. *Tetrahedron*. 2006; 62: 5426– 5434.
- Palanki MSS, Erdman PE, Gayo-fung LM, Shevlin GI, Sullivan RW, Suto MJ, Goldman ME, Ransone LJ, Bennett BL, Manning AM. J Med Chem. 2000; 43: 3995–4004.
- Fandrick DR, Reinhardt D, Desrosiers JN, Sanyal S, Fandrick KR, Ma S, Grinberg N, Lee H, Song JJ, Senanayake CH. *Org Lett.* 2014; 16: 2834–2837.
- 37. Gerus II, Vdovenko SI, Gorbunova MG, Kukhar' VP.

8 _	Tetrahedron						
	Chem Heterocycl Compd. 1991; 27: 398–406.PTED N	A .55 NU	SYurugi S, Hieda M, Fushimi T, Kawamatsu Y,				
38.	Bonacorso HG, Bortolotto GP, Navarini J, Porte LMF, Wiethan CW, Zanatta N, Martins MAP, Flores AFC. <i>J Fluor Chem.</i> 2010; 131: 1297–1301.	56.	 Sugihara H, Tomimoto M. <i>Chem Pharm Bull</i>. 1972; 20: 1528–1535. McCombie SW, Tagat JR, Vice SF, Lin SI, Steensma 				
39.	Liu B, Liu M, Xin Z, Zhao H, Serby MD, Kosogof C, Nelson LTJ, Szczepankiewicz BG, Kaszubska W, Schaefer VG, Falls HD, Lin CW, Collins CA, Sham		R, Palani A, Neustadt BR, Baroudy BM, Strizki JM, Endres M, Cox K, Dan N, Chou CC. <i>Bioorg Med</i> <i>Chem Lett.</i> 2003; 13: 567–571.				
	HL, Liu G. <i>Bioorg Med Chem Lett.</i> 2006; 16: 1864– 1868.	57.	Palani A, Shapiro S, Clader JW, Greenlee WJ, Vice S, McCombie S, Cox K, Strizki J, Baroudy BM. <i>Bioorg</i> <i>Med Chem Lett.</i> 2003: 13: 709–712.				
40.	Dorigo P, Fraccarollo D, Santostasi G, Maragno I, Floreani M, Borea PA, Mosti L, Sansebastiano L, Fossa P, Orsini F, Benetollo F, Bombieri G. <i>J Med</i>	58.	Mcfadden HG, Huppatz JL. Aust J Chem. 1992; 45: 1045–1050.				
41.	Cameron M, Foster BS, Lynch JE, Shi Y-J, Dolling	59.	Effenberger F, Barthelmess I. J Heterocycl Chem. 1995; 32: 599–602.				
42.	Zanatta N, Lopes ECS, Fantinel L, Bonacorso HG, Martins MAP. <i>J Heterocycl Chem</i> . 2002; 39: 943– 947.	60.	Lahm GP, Selby TP, Freudenberger JH, Stevenson TM, Myers BJ, Seburyamo G, Smith BK, Flexner L, Clark CE, Cordova D. <i>Bioorg Med Chem Lett.</i> 2005; 15: 4898–4906.				
43.	Yamada S. J-Stage. 1951; 71: 1349–1355.	61.	Zumbrunn A, Lamberth C, Schaub F. <i>Synth Commun</i> .				
44.	Flores DC, Fiss GF, Wbatuba LS, Martins MAP, Burrow RA, Flores AFC. <i>Synthesis</i> . 2006: 2349– 2356.	62.	1998; 28: 475–485. Lemcke T, Messinger P. <i>Arch Pharm</i> . 1995; 328: 269–270.				
45.	Zanatta N, Amaral SS, dos Santos JM, de Mello DL, Fernandes L da S, Bonacorso HG, Martins MAP, Andricopulo AD, Borchhardt DM. <i>Bioorg Med Chem</i> . 2008; 16: 10236–10243.	63.	Fauber B., Niel, M. B. Van, Cridland, A., Hurley, C., Killen, J., Ward, S. WO2016177686 A1, 2016, 203 pages.				
16	Rongoorso HG, Forla A, Cachinal C a, Zanatta N	64.	Schaus JM. WO2009131815 (A1), 2009, 42 pages.				
40.	Martins M a. P. <i>J Heterocycl Chem</i> . 2008; 45: 483– 487.	65.	Honda T, Kawashima K, Okamoto K, Yamamoto M EP1864977 (A1), 2007, 51 pages.				
47.	Bonacorso HG, Calheiro TP, Rodrigues MB, Stefanello ST, Soares FAA, Zanatta N, Martins MAP.	66.	Priestley ES, Posy SL, Tremblay F, Martel A, Marinier A. WO2013163241 A1, 2013, 308 pages.				
48	Monatsh Chem. 2016; 147: 0. Zanatta N. Cortelini M de FM. Caspes MIS	67.	Marra JM, Goehring RR, Perez J, Stasaitis LR, Liu Y. WO2004111011 A2, 2004, 42 pages.				
-0.	Bonacorso HG, Martins MAP. <i>J Heterocycl Chem</i> . 1997; 34: 509–513.	68.	Goryaeva M V., Burgart Y V., Ezhikova MA, Kodess MI, Saloutin VI. <i>Beilstein J Org Chem.</i> 2015; 11:				
49.	Madruga CDC, Clerici E, Marcos AP. J Heterocycl Chem. 1995; 32: 735–738.	69.	Zanatta N, Fortes AS, Bencke CE, Marangoni MA,				
50.	Aquino E da C, Lobo MM, Leonel G, Martins MAP, Bonacorso HG, Zanatta N, F. <i>Eur J Org Chem</i> . 2017:		Camargo AF, Fantinel CA, Bonacorso HG, Martins MAP. <i>Synthesis</i> . 2015; 47: 827–835.				
51.	306–312. Marcos R. Martins P. Sinhorin AP. Rosa A. Flores	70.	Nesi R, Chimichi S, Scotton M, Degl'Innocenti A, Adembri G. J Chem Soc, Perkin Trans 1. 1980; 0:				
	AFC, Wastowski AD, P CM, Flores DC, Beck P, Freitag RA, Brondani S, Cunico W, Bonacorso HG. <i>Synthesis</i> . 2002: 2353–2358.	71.	1667–1670. Reiner R, Eugster CH. <i>Helv Chim Acta</i> . 1967; 50: 128–136.				
52.	Jones RG, Whitehead CW. <i>J Org Chem.</i> 1955; 20: 1342–1347.	72.	Kuwano R, Hashiguchi Y, Ikeda R, Ishizuka K. Angew Chem. 2015; 54: 2423–2426.				
53.	Iaroshenko VO, Dudkin S, Sosnovskikh VY, Villinger A, Langer P. <i>Eur J Org Chem.</i> 2013; 201300107: 3166–3173.	73.	Moszczyński-Pętkowski R, Majer J, Borkowska M, Bojarski Ł, Janowska S, Matłoka M, Stefaniak F, Smuga D, Bazydło K, Dubiel K, Wieczorek M. <i>Eur J</i> <i>Med Chem.</i> 2018; 155: 96–116.				
J4.	A. W. Stolle W, M. Veurink J, T. M. Marcelis A, Plas HC van der. <i>Tetrahedron</i> . 1992; 48: 1643–1656.		Blakemore DC, Castro L, Churcher I, Rees DC, Thomas AW, Wilson DM, Wood A. <i>Nat Chem.</i> 2018;				

10. ACCEPTED	MANUSCRIPT
Goldberg FW, Kettle JG, Kogej T, Perry MWD, Tomkinson NP. <i>Drug Discov Today</i> . 2015; 20: 11–17.	
Stepaniuk OO, Matviienko VO, Kondratov IS, Vitruk I V., Tolmachev AO. <i>Synthesis</i> . 2013; 45: 925–930.	
Armarego WLF, Chai C. Purification of laboratory chemicals, Elsevier: Oxford, 5th edn., 2003.	
Krechl J, Böhm S, Smrčková S, Kuthan J. Collect Czechoslov Chem Commun. 1989; 54: 673–683.	
Liu MTH., Chishti NH, Burkholder CD, Jones WE, Wasson J.S. J Org Chem. 1980; 45: 4515–4515.	
Wharton CJ, Wrigglesworth R. J Chem Soc, Perkin Trans 1. 1981; 0: 433–436.	
Moss RA, Guo W, Denney DZ, Houk KN, Rondan NG. J Am Chem Soc. 1981; 103: 6164–6169.	R'
Shapiro R. WO 2006/121648 A2, 2016, 26 pages.	
Mousdis GA, Ganotopoulos NM, Barkaoui H, Abid Y, Psycharis V, Savvidou A, Raptopoulou CP. <i>Eur J Inorg Chem.</i> 2017; 2017: 3401–3408.	
Parveen H, Hayat F, Salahuddin A, Azam A. <i>Eur J Med Chem.</i> 2010; 45: 3497–3503.	
Sturala J, Bohacova S, Chudoba J, Metelkova R, Cibulka R. <i>J Org Chem.</i> 2015; 80: 2676–2699.	
Leprêtre A, Turck A, Plé N, Knochel P, Quéguiner G. <i>Tetrahedron</i> . 2000; 56: 265–273.	
Tsuda K, Ogawa Y. <i>J-Stage</i> . 1950; 70: 73–76.	
	 ACCEPTED Goldberg FW, Kettle JG, Kogej T, Perry MWD, Tomkinson NP. Drug Discov Today. 2015; 20: 11–17. Stepaniuk OO, Matviienko VO, Kondratov IS, Vitruk I V., Tolmachev AO. Synthesis. 2013; 45: 925–930. Armarego WLF, Chai C. Purification of laboratory chemicals, Elsevier: Oxford, 5th edn., 2003. Krechl J, Böhm S, Smrčková S, Kuthan J. Collect Czechoslov Chem Commun. 1989; 54: 673–683. Liu MTH., Chishti NH, Burkholder CD, Jones WE, Wasson J.S. J Org Chem. 1980; 45: 4515–4515. Wharton CJ, Wrigglesworth R. J Chem Soc, Perkin Trans I. 1981; 0: 433–436. Moss RA, Guo W, Denney DZ, Houk KN, Rondan NG. J Am Chem Soc. 1981; 103: 6164–6169. Shapiro R. WO 2006/121648 A2, 2016, 26 pages. Mousdis GA, Ganotopoulos NM, Barkaoui H, Abid Y, Psycharis V, Savvidou A, Raptopoulou CP. Eur J Inorg Chem. 2017; 2017: 3401–3408. Parveen H, Hayat F, Salahuddin A, Azam A. Eur J Med Chem. 2010; 45: 3497–3503. Sturala J, Bohacova S, Chudoba J, Metelkova R, Cibulka R. J Org Chem. 2015; 80: 2676–2699. Leprêtre A, Turck A, Plé N, Knochel P, Quéguiner G. Tetrahedron. 2000; 56: 265–273. Tsuda K, Ogawa Y. J-Stage. 1950; 70: 73–76.