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Novel triazole derivatives containing different ester skeleton: Design,

synthesis, biological evaluation and molecular docking

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Abstract

Invasive fungal disease constitutes a growing health problem and development of novel antifungal drugs with high potency and selectivity are in an urgent need. In this study, a novel series of triazole derivatives containing different ester skeleton were designed and synthesized. Microdilution broth method was used to investigate antifungal activity. Significant inhibitory activity of compounds **5c**, **5d**, **5e**, **5f**, **5m** and **5n** was evaluated against the *Candida albicans* (I), *Candida albicans clinical isolate* (II), *Candida glabrata clinical isolate* (I), and *Candida glabrata* (II) with MIC₈₀ values ranging from 2 μ g/mL to 16 μ g/mL. Notably, compounds **5e** and **5n** showed the best inhibition against *Candida albicans* (II), *Candida glabrata* (II), and *Candida glabrata* (II) at the concentrations of 2 μ g/mL and 8 μ g/mL, respectively. Molecular docking study revealed that the target compounds interacted with CYP51 mainly through hydrophobic and Van der Waals interactions. The results indicated that these novel triazole derivatives could serve as promising leads for development of antifungal agents.

Key words: Triazole; Synthesis; Antifungal activity; Structure-activity relationship; Molecular docking

1. Introduction

Global morbidity due to life-threatening fungal infections has dramatically increased in recent decades, particularly among individuals suffering from cancer, autoimmune diseases, organ transplants or AIDS.¹⁻³⁾ Clinically, *Aspergillus fumigatus* (mortality rate: 50-90%), *Cryptococcus neoformans* (mortality rate: 20-70%), and *Candida albicans* (mortality rate: 20-40%) are the major pathogenic agents for systemic fungal infections.⁴⁾ Numerous antifungal drugs including polyenes (e.g., amphotericin B), fluorinated pyrimidines (e.g., 5-fluorocytosine), azoles (e.g., fluconazole), and echinocandins (e.g., caspofungin) (**Fig. 1**) are available for the treatment of invasive fungal infections.⁵⁾ For their high therapeutic index, broad spectrum activities and superior safety profile, the azole antifungals are considered as first-line therapy in current clinic. However, some of them still suffer from serious drug resistance issues, there is a real need for developing novel antifungal triazoles with improved profiles.^{6,7}



Fig. 1. Chemical structures of representative antifungal agents

Azole antifungal agents competitively inhibit the lanosterol 14 α -demethylase (CYP51), which catalyzes the oxidative removal of the 14 α -methyl group of lanosterol to give \triangle 14,15-desaturated intermediates in ergosterol biosynthesis. This selective inhibition of CYP51 would result in depletion of ergosterol and accumulation of lanosterol, thereby inhibiting the growth and proliferation of fungi.^{8,9)} Moreover, CYP51 has been regarded by many researchers as a target for molecular docking and widely applied in the rational design of antifungal compounds.¹⁰⁻¹⁵⁾

Studies showed that¹⁶⁾ the antifungal triazoles featured a core structure composed of a triazole, a halophenyl ring, while differing in the side chains. The development of triazole drugs is primarily driven through structure optimization especially of the side chains as well as SAR study. Itraconazole (**Fig. 2**) is the triazole antifungal drug available in the market and widely used for candidiasis. The long side chain of itraconnazole containing four linearly linked cyclic rings might attribute to its low solubility and erratic bioavailability, which drove the research to design novel agents with medium side chains.¹⁷⁾ Therefore, replacing the long side chain in the itraconazole structure with different ester moieties containing aryl rings and halogenated alkyl chain might result in novel triazole antifungal drugs. Furthermore, molecular docking study indicated that the triazole bound to the heme iron of CYP51, and the

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halophenyl ring could locate into the hydrophobic pocket.¹⁸⁾ We hypothesized that the ester moieties might accommodate in the deep cavity of CYP51, as well as providing in hydrophobic pockets of the biological targets which contribute to improve the potency, stability and specificity of the binding site. In addition, the ester groups were to be introduced in the target compounds because they could be further removed to reduce molecular weight. The halogen as a strong electron-withdrawing group could affect the charge distribution and hydrophobility of drug molecules and the time of drug action. Thus, the halogenated alkyl chain was to be introduced to leave large space for further optimization.

The above mentioned promoted us to design and synthesize a series of 1-(2,4-dichlorophenyl)-2-(1H-1,2,4-triazol-1-yl) ethyl derivatives containing a triazole ring, a dichlorophenyl group, and ester groups in the side chain. In our design, the structure of itraconazole was altered as a platform and different ester groups were insert into the side chain to find out whether these groups were necessary for the antifungal activity (**Fig. 2**).



Fig. 2. Design of compounds 5a-5q

2. Results and Discussion

2.1. Chemistry

1-(2,4-dichlorophenyl)-2-(1*H*-1,2,4-triazol-1-yl)ethyl derivatives (**5a-5q**) were synthesized in four steps as illustrated in **Chart 1**. At first, 1,3-dichlorobenzene (1.0 equiv), chloroacetyl chloride (ClCH₂COCl) (1.2 equiv) and anhydrous aluminum chloride (AlCl₃) (1.0 equiv) in an anhydrous dichloromethane (DCM) solution were heated at reflux for 4 h to obtain compound **2** in 80% yield.¹⁹⁾ Secondly, intermediate **3** was prepared by condensation of compound **2** (1.0 equiv) with triazole (1.5 equiv) in the presence of potassium carbonate (K₂CO₃) (1.0 equiv) and acetonitrile (CH₃CN) for 8 h at reflux. Intermediates **3** was obtained in the yield of 39%. Thirdly, intermediate **3** (1.0 equiv) and sodium borohydride (NaBH₄) (2.0 equiv) in an anhydrous ethanol-tetrahydrofuran (THF) solution were reacted at room temperature for 4 h to obtain compound **4** in 80% yield. Finally, compound **4** (1.0 equiv) and various acid (1.5 equiv) were reacted in the presence of *N*,*N*'-dicyclohexylcarbodiimide (DCC) (1.3 equiv), 4-dimethylaminopyridine (DMAP) (1.3 equiv) and anhydrous dichloromethane for overnight at reflux to give final compounds **5a-5q** in 70%-89% yield. The reaction was monitored by TLC, and the reaction mixture was purified by column chromatography.



Chart 1. Synthetic route of compounds **5a-5q**. Reagents and conditions: (i) ClCH₂COCl, AlCl₃, Anhydrous CH₂Cl₂, reflux, 4 h, 80% yield; (ii) Triazole, K₂CO₃, CH₃CN, reflux, 8 h, 39% yield; (iii) NaBH₄, Ethanol-THF, r.t., 4 h, 80% yield; (iii) DCC, DMAP, Anhydrous CH₂Cl₂, reflux, overnight, 70-89% yield.

The structures of all compounds were shown in **Table 1**. All the compounds were elucidated by ¹H NMR, ¹³C NMR and HRMS.

Table 1. Structure, yield, reaction time and melting point of compounds 5a-5q



No.	R	Time (h)	Yield (%)	m.p.(°C)
5a	ζζζ∽_CΙ	13	75	85-87
5b	≻¢ Br	12	70	91-93
5c	Str. Cl	13	83	80-82
5d	چئ Br	14	80	85-87
5e	×~~~_ci	13	81	62-64
5f	¥∕∽∽_Br	12	78	64-65
5g	¥~~~~ _{Cl}	14	85	64-66
5h	XBr	13	79	66-67
5 i	*	13	89	126-128
5j	CH ₃	12	87	85-86
5k	*Q _{CH}	13	87	167-168
51		13	82	90-91
5m	× C	14	80	177-179
5n	× C	14	86	141-142
50	F F	13	78	129-130
5p	² Ar	13	83	175-177
5q	Br	14	80	84-85
3		8	39	113-114

2.2. Biological activity

The antifungal activity of the compounds **5a-5q** against nine human pathogenic fungi was evaluated according to the protocols of the National Committee for Clinical Laboratory Standards. Fluconazole (FCZ), Amphotericin B (Amp B) and Itraconazole (ITZ) were used as positive control drugs. The broth microdilution method was used to determine the minimum

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inhibitory concentrations (MIC₈₀) of the synthesized compounds.²⁰⁾

As shown in **Table 2**, most of the compounds showed weak antifungal activity against *C. par.*, *A. fum.*, *C. kru.* and *C. tro.*, while there was a good antifungal activity against *C. alb.*, *C. gla.* and *C. neo.*. Several compounds, such as compounds **5c**, **5d**, **5e**, **5f**, **5m** and **5n** exhibited superior or comparable antifungal activity to FCZ with MIC₈₀ values in the range of 2-16 μ g/mL, particularly for *C. alb.*(I), *C. alb.*(II), *C. gla.*(I), and *C. gla.*(II). Among them, compounds **5e** and **5n** showed the most potent activity against four important fungal pathogens (MIC₈₀ = 2-8 μ g/mL). Compound **5c** (inhibition of *C. alb.* (II)) could be achieved at 2 μ g/mL and compounds **5d** and **5f** (inhibition of *C. gla.* (II)) could be achieved at 4 μ g/mL. Compound **5m** (inhibition of *C. gla.* (I) and *C. gla.* (II)) could be achieved at 8 μ g/mL. These above-mentioned compounds exhibited superior activity to FCZ. Interestingly, compound **5p** (MIC₈₀ = 2 μ g/mL).

In contrast, the target compounds generally showed improved activity against *C. neo.*, whereas they were less potent against *C. par.*. For example, the inhibitory activity of compounds **5c**, **5d**, **5e**, **5f** and **5g** (MIC₈₀ = 32 µg/mL) against *C. neo.* were comparable to that of FCZ (MIC₈₀ = 32 µg/mL). For *C. kru.*, most compounds showed weak activity except compound **5n** (MIC₈₀ = 32 µg/mL), which was more potent than FCZ (MIC₈₀ = 64 µg/mL). However, all the compounds were inactive against *A. fum.* and *C. tro.*.

As seen from the **Table 2**, compound **3** and intermediate **4** showed weak antifungal activity as compared to the compounds **5a-5q**. The introduction of different ester groups on the intermediate **4** resulted in a better improvement of the antifungal activity.

		MIC ₈₀ (µg/mL)							
Compd	C. alb.	C. alb.	C. kru.	C. par.	C. gla.	C. gla.	C. neo.	A. fum.	C. tro.
•	(I)	(II)			(I)	(II)			
5a	64	16	128	64	128	128	128	64	>128
5b	64	16	128	64	128	128	128	128	>128
5c	8	2	64	32	16	16	32	128	>128
5d	8	4	128	64	16	16	32	64	>128
5e	4	2	128	128	8	8	32	128	>128
5f	8	4	128	128	16	16	32	128	>128
5g	32	16	>128	64	32	32	32	128	>128
5h	64	16	>128	64	64	32	64	128	>128
5i	64	32	>128	128	128	128	128	128	>128
5j	128	64	>128	128	>128	128	128	128	>128
5k	16	32	>128	128	32	128	128	128	>128
51	128	64	>128	128	128	128	128	64	>128
5m	8	8	>128	64	8	8	128	128	>128
5n	2	2	32	64	8	8	128	128	>128
50	128	64	128	128	128	128	128	128	>128
5p	2	32	128	>128	16	32	128	128	>128
5q	128	64	64	>128	128	128	128	128	>128
3	32	32	>128	64	64	64	>128	128	>128
4	16	32	128	16	128	128	64	64	>128
FCZ	8	8	64	16	16	16	32	128	32
ITZ	0.25	0.25	1	0.5	2	2	8	4	1
Amp B	<1	<1	<1	<1	<1	2	1	2	<1

Table 2. In vitro antifungal activity of the target compounds 5a-5q^a

^aAbbreviations: *C. alb.*(I), *Candida albicans (ATCC 90028)*; *C. alb.*(II), *Candida albicans clinical isolate (1802274)*; *C. kru.*, *Candida krusei (ATCC 6528)*; *C. par., Candida parapsilosis clinical isolate (18022019)*; *C. gla.*(I), *Candida glabrata clinical isolate (1802183)*; *C. gla.*(II), *Candida glabrata (ATCC 15126)*; *C. neo., Cryptococcus neoformans (ATCC 34877)*; *A. fum., Aspergillus fumigatus clinical isolate (2423)*; *C. tro., Candida tropicalis clinical isolate (1802372)*; FCZ: Fluconazole; ITZ: Itraconazole; Amp B: Amphotericin B.

2.3. Structure activity relationships study

On the basis of the antifungal activity, preliminary structure-activity relationships (SARs) were obtained. Firstly, compared with the compounds **3** and **4**, the introduction of different acids on the 1-(2,4-dichlorophenyl)-2-(1*H*-1,2,4-triazol-1-yl)ethan-1-one scaffold (**5a-5q**) resulted in a better improvement of the antifungal activity, suggesting that the interactions of the aliphatic and aromatic groups with CYP51 were important for its binding affinity.

Secondly, for the compounds **5a-5h**, the length of halogenated alkyl side chain on ester group was essential to their antifungal activities. The data suggested that the optimal chain

lengths for maximal antifungal activity were C4 and C5 (**5c**, **5d**, **5e**, **5f**), and the general trend for activity versus chain length was C5 > C4 > C6 > C3. Among the compounds **5c**, **5d**, **5e**, **5f**, chloro substituted alkyl side chain on ester group (**5c**, **5e**) was more active than the bromo substituted (**5d**, **5f**) against *C. alb.*, *C. gla.* and *C. neo*.

Thirdly, for the compounds **5i-5q**, electron-withdrawing groups (halogens, such as -F, -Cl and -Br) on benzene ring, enhanced the inhibitory activity against *C. alb.*, *C. gla.* and *C. neo.*. On the contrary, electron-donating groups (such as -CH₃), decreased the inhibitory activity. Therefore, to some extent, we could predict that electron-withdrawing group could affect the charge distribution and hydrophobility of drug molecules to improve the biological activity of compounds. Moreover, 4-substitutions on benzene ring (**5m**, **5n** and **5p**) were more favorable than the 3-substitutions (**5l**, **5o** and **5q**). Among the compounds **5m**, **5n**, **5p**, 4-fluoro substitution (**5n**) was more active than the 4-chloro substitution (**5m**). However, 4-bromo substitution (**5p**) had little effect on the antifungal activity as compared to the compounds **5m** and **5m**. These results gave us a clear conclusion that the halogen position and type.

Finally, the replacement of the halogenated alkyl ester side chains (**5a-5h**) by phenyl groups (**5i-5q**) resulted in a decrease in antifungal activity. These results clearly indicated the importance that halogen substituents have on the antifungal activity of this class of triazole derivatives.

2.4. Molecular docking

In order to rationalize the antifungal activity and provide more straightforward information about ligand-enzyme interactions for further rational drug design, molecular docking was performed for itraconazole (ITZ), compounds **5e** and **5q**. The crystal structure of CYP51²¹⁾ (PDB ID: 5EQB) bound with its active ligand ITZ was served as a useful template for generating proposed binding modes. The docking results were showed in **Fig. 3** and **Table 3**.

The overall conformation of the ITZ in the active site of CYP51 was shown in **Fig. 3A**. The docking method was quite accurate to predict the real interaction of ligands and CYP51, where the re-docking experiment gave a RMSD value of 1.9844 Å. Moreover, the triazole group of the ITZ made a coordination bond with the heme iron and showed carbon hydrogen bond interaction with Gly314. The 2,4-dichlorophenyl group could be put into the hydrophobic pocket formed by Ile139, Met313 and Phe236. In addition, the long tail of ITZ lied within a hydrophobic pocket and formed Van der Waals interactions with the surrounding residues, such as Leu383, Ser382, Ala69 and Phe384. The phenyl group of side chain could form π - π interaction with Phe241 and the piperazine ring of side chain had carbon hydrogen bonds with residues of Ser508 and Met509.

More insights revealed that compound **5e** might bind with the active site cavity of CYP51 (**Fig. 3B**). The -CDOCKER ENERGY value of it, which was 21.4974 kcal/mol, was higher than the natural ligand ITZ (14.2919 kcal/mol). It indicated that compound **5e** might exert better antifungal activity than ITZ. The triazole ring made a coordination bond with the heme iron and formed hydrogen bond interaction with Tyr126. The 2,4-dichlorophenyl group could show π - π interactions with Leu380 and Phe236. Further, the ester side chain was interacting with Met509, Leu383, Phe384, Phe241 and Ser382 through Van der Waals force.

Compared with compound 5e, compound 5q showed weaker antifungal activity. The

calculated -CDOCKER ENERGY value was 3.6624 kcal/mol. The -CDOCKER INTERACTION ENERGY value of it, which was 38.1424 kcal/mol, was lower than the compound **5e** (45.1817 kcal/mol). As shown in **Fig. 3C**, the triazole ring showed π - π interaction with His381 and the ester side chain was interacting with Phe384, Tyr126 and Leu383 through Van der Waals force. Compound **5q** showed weaker interactions with the key amino acid residues of the CYP51 binding pocket than compound **5e**. Thus, the molecular modeling studies suggested that compound **5e** might act as an antifungal agent by occupying the CYP51 binding pocket and showing favorable interactions with the key amino acid residues.





Fig. 3. The binding modes of ITZ (A), 5e (B), and 5q (C) in the active site of CYP51 The heme is shown in blue (Upper) and molecules ITZ, 5e and 5q are shown in purple-red (Lower).

LIGANDS	-CDOCKER ENERGY ^a	-CDOCKER INTERACTION		
	(kcal/mol)	ENERGY ^b (kcal/mol)		
Itraconazole	14.2919	80.1265		
5h	27.1352	48.3584		
5e	21.4974	45.1817		
5 f	21.6036	44.9928		
5g	25.7246	43.1719		
5c	23.5840	40.9958		
5d	19.3272	40.8749		
5m	15.9591	40.5997		
5p	9.7196	40.5280		
5a	18.0549	40.0338		
51	4.0308	39.9109		
5k	17.2615	39.7354		
5j	11.9170	39.5278		
5b	20.2553	39.4440		
5n	14.1954	38.6478		
5q	3.6624	38.1424		
50	11.6228	38.0478		
5 i	12.2871	37.5572		
3	6.8256	32.2895		

Table 3. Ligand-CYP51 interactions of the target compounds 5a-5q

^a -CDOCKER ENERGY: The higher the value of -CDOCKER ENERGY, the more stable the conformation.

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^b -CDOCKER INTERACTION ENERGY: The higher the CDOCKER INTERACTION ENERGY value represents more stable ligand binding.

3. Conclusions

A novel series of triazole derivatives containing different ester skeleton were synthesized and evaluated as antifungal agents. Compounds **5e** and **5n** showed the best antifungal activity *in vitro*, which (inhibition of *Candida albicans* (II) and *Candida glabrata* (I), (II)) could be achieved at the concentrations of 2 μ g/mL and 8 μ g/mL, respectively. In addition, compounds **5c**, **5d**, **5f** and **5m** displayed the remarkable *in vitro* inhibitory activity against *Candida albicans* and *Candida glabrata* strains, which was superior or comparable to that of the reference drug fluconazole. However, all the compounds also showed weak inhibition to other fungi which were *Candida parapsilosis*, *Aspergillus fumigatus*, *Candida krusei* and *Candida tropicalis*. Moreover, molecular docking was performed to position compounds **5e** and **5q** into the CYP51 active site to determine the potential binding mode. In conclusion, these preliminary results are promising and some of these compounds may be potential candidates as antifungal agents.

Experimental

General chemistry methods, synthesis procedures, spectral data, biological assays, molecular docking are given in Supplementary information.

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Conflict of Interest

The authors declare no conflict of interest.

Supplementary Materials

The online version of this article contains supplementary materials.

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