

Journal Pre-proofs

Synthesis and biological screening of thiosemicarbazones of substituted 3-acetylcoumarins having d-glucose moiety

Vu Ngoc Toan, Nguyen Dinh Thanh, Nguyen Minh Tri

PII: S0960-894X(20)30775-7
DOI: <https://doi.org/10.1016/j.bmcl.2020.127664>
Reference: BMCL 127664

To appear in: *Bioorganic & Medicinal Chemistry Letters*

Received Date: 12 August 2020
Revised Date: 24 October 2020
Accepted Date: 28 October 2020



Please cite this article as: Ngoc Toan, V., Dinh Thanh, N., Minh Tri, N., Synthesis and biological screening of thiosemicarbazones of substituted 3-acetylcoumarins having d-glucose moiety, *Bioorganic & Medicinal Chemistry Letters* (2020), doi: <https://doi.org/10.1016/j.bmcl.2020.127664>

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Synthesis and biological screening of thiosemicarbazones of substituted 3-acetylcoumarins having D-glucose moiety

Vu Ngoc Toan^{a,b}, Nguyen Dinh Thanh^{*b}, Nguyen Minh Tri^{a,b}

^a Department of Toxicological Chemistry and Radiation, Institute for Advanced Technology

(Vietnam Academy of Military Science and Technology), 17 Hoang Sam, Cau Giay, Ha Noi, Viet Nam

^b Faculty of Chemistry, VNU University of Science (Vietnam National University, Ha Noi), 19 Le Thanh Tong, Hoan Kiem, Ha Noi, Viet Nam

Abstract

Thiosemicarbazones **5a-j** were synthesized with yields of 45–68% by condensation of 3-acetylcoumarins **3a-j** and tetra-*O*-acetyl- β -D-thiosemicarbazide **4**. All obtained thiosemicarbazones were screened for anti-microorganic activities against bacteria (*B. subtilis*, *S. aureus*, *S. epidermidis*, *E. coli*, *P. aeruginosa*, *K. pneumoniae*, *S. typhimurium*) and fungi (*A. niger*, *C. albicans*, *S. cerevisiae*, and *A. flavus*). Some compounds had significant inhibitory activity with MICs of 0.78–3.125 μ M in comparison with **5a**, including **5e,h,i** for *S. aureus*, and **5c,f,i** for *S. epidermidis* (Gram-(+) bacteria), **5c,f,g** for *E. coli*, **5f** for *K. pneumoniae*, **5b,c,g** for *P. aeruginosa*, and **5i** for *S. typhimurium* (Gram-(–) bacteria), **5d,h,i** for *A. niger*, **5i** for *A. flavus*, **5b,d,e,h** for *C. albicans*, and **5i** for *S. cerevisiae*. Compounds exhibited excellent activity against tested microorganism with MIC = 0.78 μ M, including **5h,i** (against *S. aureus*), **5h** (against *C. albicans*), and **5i** (against *S. cerevisiae*).

Keywords: 3-Acetylcoumarins; Antibacterial; Antifungal; D-Glucose; Thiosemicarbazides; Thiosemicarbazones

Thiosemicarbazides and thiosemicarbazone derivatives had occupied important roles in organic and medicinal chemistry. These classes of compounds presented chelating properties by using C=S and NH groups for coordination with metal center.¹⁻³ They could be used as versatile intermediates for preparing various heterocyclic compounds.^{4, 5} They exhibited a broad spectrum of potential pharmacological activities, such as antibacterial,⁶ antifungal,⁷ anticancer,⁸ antimycobacterial,⁹ and antidiabetic¹⁰ activity. For example, nopinone-based thiosemicarbazone **A** displayed potent *in vitro* antitumor activity against three human cancer cell lines (MDA-MB-231, SMMC-7721 and Hela) with the IC₅₀ values of 2.79 ± 0.38 , 2.64 ± 0.17 and 3.64 ± 0.13 μM , respectively.⁸

Thiosemicarbazone **B** exhibited antimycobacterial activity against *Mycobacterium tuberculosis* H37Ra with MIC value of 0.031 $\mu\text{g/mL}$ and MBC (minimum bactericidal concentration) value of 0.063 $\mu\text{g/mL}$.⁹ Compounds **C** were found to be the most active ALR2 inhibitors with IC₅₀ values of 0.52 ± 0.04 μM (with R = Br) and 0.19 ± 0.03 μM (with R = F), respectively, and these compounds were more effective inhibitors as compared to the ALR2 (standard aldose reductase 2) inhibitor, sorbinil, which has an IC₅₀ value of 3.14 ± 0.02 μM .¹⁰

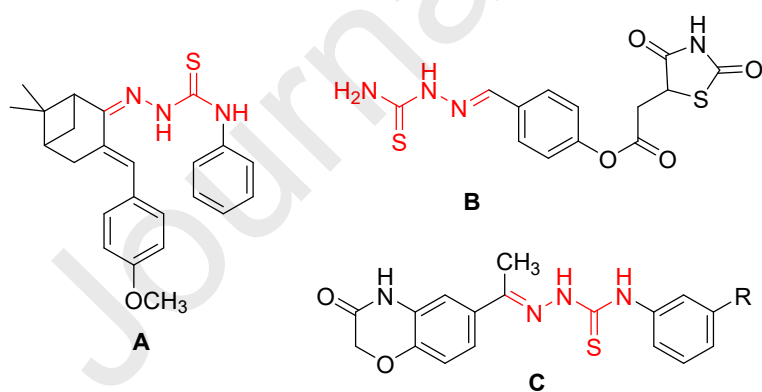


Figure 1. Various bioactive thiosemicarbazones.

The synthesis of substituted aromatic or heterocyclic aromatic aldehyde/ketone thiosemicarbazones with sugar moieties were reported.^{1, 4, 11-13} The series of aldehyde 4-(β -D-

glucopyranosyl)thiosemicarbazones¹²⁻¹⁴ were prepared from per-*O*-acetylated β -D-glucopyranosyl isothiocyanate and several of them showed micromolar inhibitions against typical Gram-positive (*Bacillus subtilis*, *Staphylococcus aureus*, *Staphylococcus epidermidis*), Gram-negative bacteria (*Escherichia coli*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Salmonella typhimurium*) and fungi (*Aspergillus niger*, *Candida albicans*, *Aspergillus flavus*). These such compounds also had shown various biological properties, such as antibacterial,¹³ antifungal,¹³ antioxidant,¹¹ anti-dyslipidemic,¹¹ and anticancer activities.¹⁵ Figure 2 displayed some examples of bioactive thiosemicarbazones bearing sugar moiety. Compound **D** of D-glucose significantly decreased cancer cell growth with IC₅₀ value of 1.60 μ M on breast cancer cell line MDA-MB-231 and IC₅₀ value of 2.8 μ M on prostate cancer cell line PC-3, which was lower than that of cells treated with the well-known anticancer drug 5-fluorouracil with IC₅₀ = 4.75 \pm 1.41 and 2.88 \pm 1.39 μ M, respectively).¹⁵ Sugar-based thiosemicarbazone of peracetylated galactoside (**E**) was discovered to be a potent rhodesain inhibitor with IC₅₀ = 1.2 \pm 1.0 μ M.¹⁶ Thiosemicarbazone **F** with *para*-F group had inhibitory efficiency for rabbit muscle glycogen phosphorylase b with IC₅₀ = 5.7 μ M.¹⁷ Compound **G** containing simultaneously monosaccharide and isatin moieties exhibited *in vitro* antibacterial and *in vivo* antioxidant activities. When R' = Br, R = H the compound showed selective cytotoxic effects against some cancer (LU-1, HepG2, MCF7, P338, SW480, KB) cell lines and normal fibroblast cell line NIH/3T3. This compound had good inhibitory activity with MIC of 1.56 μ M against *B. subtilis*, *S. aureus*, and *S. epidermidis*.¹³

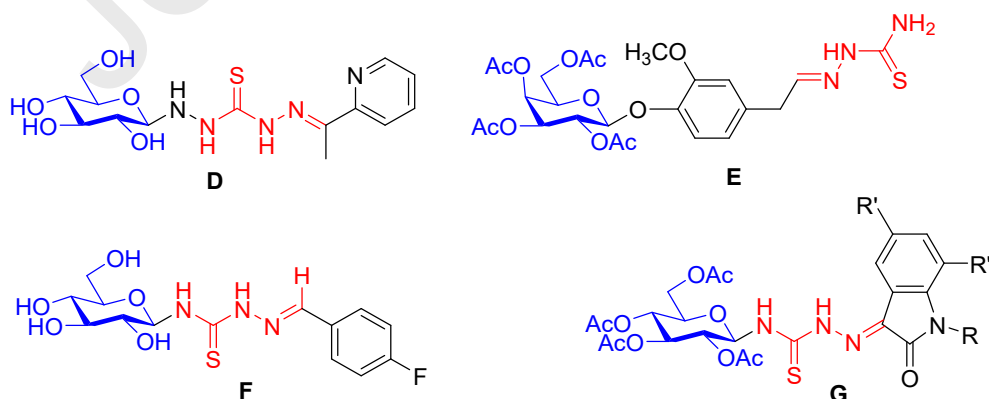


Figure 2. Some bioactive thiosemicarbazones bearing sugar moiety.

Coumarin (2*H*-chromen-2-one) was a compound of cinnamon. This heterocycle possessed remarkably pharmacological activities, such as antibacterial,¹⁸ antioxidant,¹⁹ anti-inflammatory,²⁰ anticancer²¹ activity, and widely distributed in nature in many plants.^{22, 23} Compound **H** and **I** (Fig. 3) were potential anti-tubercular agents with MIC of 0.09 $\mu\text{g/mL}$ and 3.9 $\mu\text{mol/mL}$, respectively.²⁴ ²⁵ Hydroxycoumarin **J** was used as selective MAO-B inhibitor with $\text{IC}_{50} = 2.79 \pm 0.19 \mu\text{M}$.²⁶

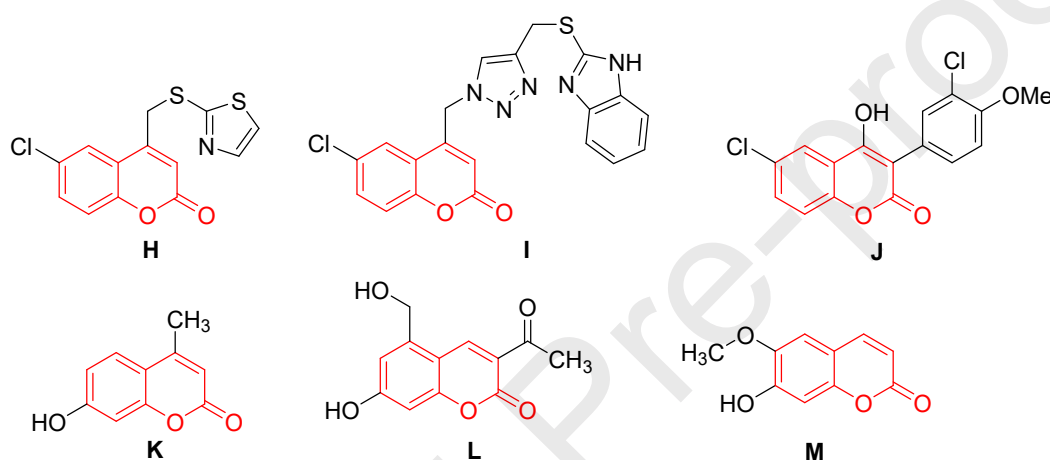
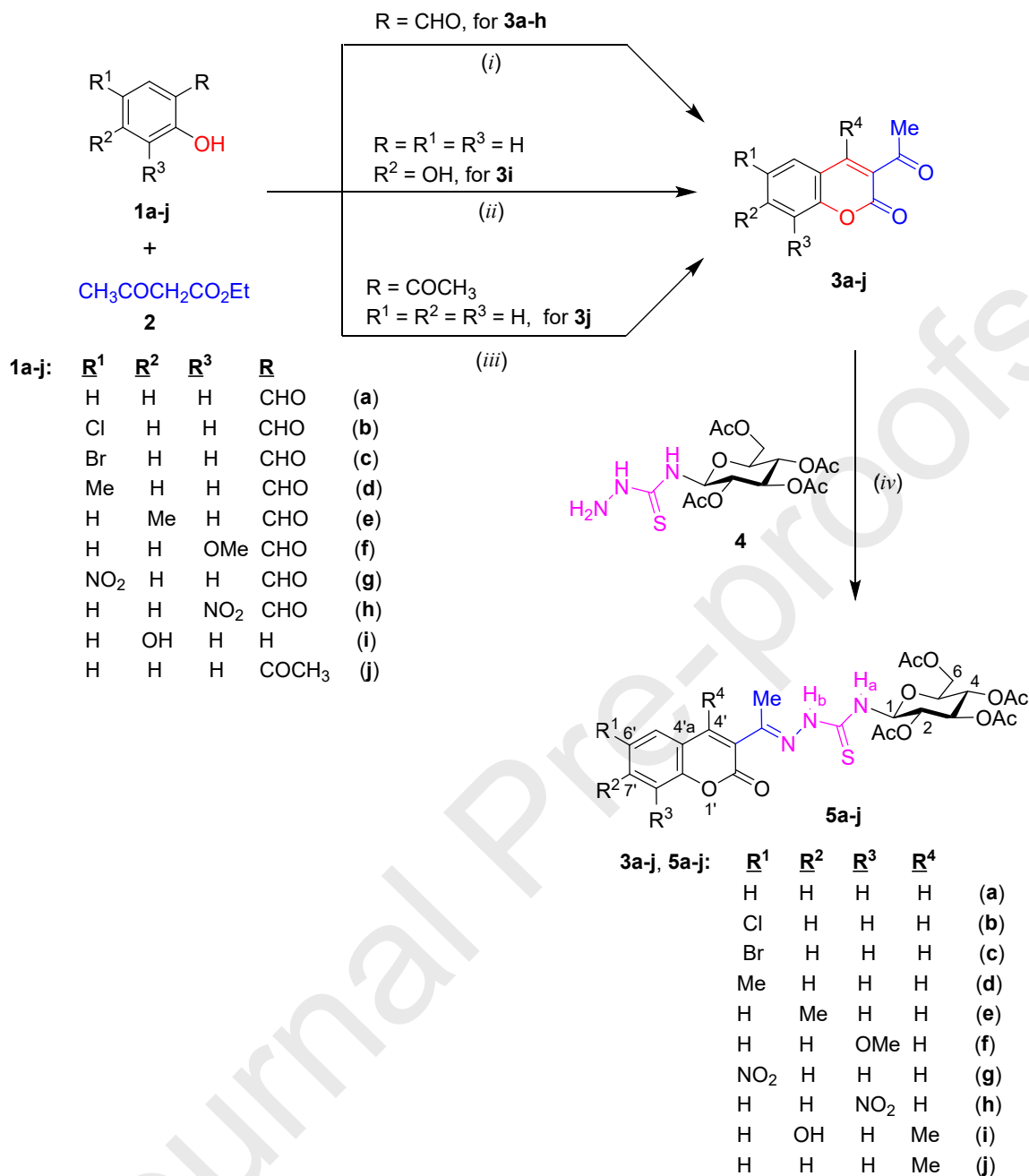


Figure 3. Some biologically active agents containing coumarin ring.

Some natural and synthetic coumarin-containing derivatives were used as biologically active agents. For example, hymecromone (4-methylumbelliferone or 4-methyl-7-hydroxycoumarin, **K**) was used as choleric and antispasmodic agent.²⁷ Armillarisin A (3-acetyl-5-hydroxymethyl-7-hydroxycoumarin, **L**), extracted from the fungus *Armillariella tabescens* (Scop. ex Fr.) Sing.),²⁸ was used as an antibiotic agent. Scopoletin (6-methoxy-7-hydroxycoumarin, **M**), isolated from several plant species, exhibited antioxidant, hepatoprotective, anti-inflammatory and antifungal activity and inhibited *in vitro* against α -glucosidase enzyme.²⁹

Based on the above-mentioned reviews on biological properties of coumarin- and sugar-containing compounds and continuing our previous studies on the synthesis and reactivity of *N*-(per-*O*-acetyl-

D-glycopyranosyl)thiosemicarbazides,^{13, 30-32} we reported herein the synthesis and evaluation of biological activities of a series of substituted 3-acetylcoumarins *N*-(2,3,4,6-tetra-*O*-acetyl- β -D-glycopyranosyl)thiosemicarbazones having D-glucose moiety. The connection of monosaccharide moiety to coumarin ring through thiosemicarbazone linkage could induce novel activities. They were then evaluated by their antibacterial and antifungal activities against some Gram-positive bacteria (*S aureus*, *B subtilis*, *S epidermidis*), Gram-negative bacteria (*P aeruginosa*, *K pneumonia*, *S typhimurium*, and *E coli*), and some fungi (*A niger*, *C albicans*, *S cerevisiae*, and *A flavus*). Synthetic pathway for target thiosemicarbazones was displayed in Scheme 1. Substituted coumarins (**3a-j**) were prepared according to modified literature procedures by reaction of corresponding substituted salicylaldehydes (**1a-h**) with resorcinol (**1i**) or *o*-hydroxyacetophenone (**1j**), respectively (see Schema 1S-4S in Supplementary data in the online version of this article). Novel substituted 3'-acetylcoumarin *N*-(2,3,4,6-tetra-*O*-acetyl- β -D-glucopyranosyl)thiosemicarbazones (**5a-j**) were obtained by condensation of corresponding substituted 3-acetylcoumarins (**3a-j**) with *N*-(2,3,4,6-tetra-*O*-acetyl- β -D-glucopyranosyl)thiosemicarbazide (Scheme 1). Reactions were performed in absolute methanol in the presence of glacial acetic acid as catalyst using microwave-assisted heating method. We found that these thiosemicarbazides could be soluble in methanol at room temperature, and substituted 3-acetylcoumarins were dissolved with difficulty in this solvent with the same conditions, but they were dissoluble in this hot solvent. At the initial stage of reaction, the initial materials began dissolving when heated under the microwave-assisted conditions. Subsequently, the appearance of precipitate was used to identify the product formation.



Scheme 1. Synthetic route for final substituted 3-acetylcoumarin tetra-*O*-acetyl-β-*D*-glucopyranosylthiosemicarbazones **5a-j**. Reaction conditions: (i) Piperidine as catalyst, **Method A**: MW Irradiation; **Method B**: Ultrasound Irradiation; **Method C**: Solvent-free Ultrasound Irradiation. (ii) a) 70% Sulfuric acid, 0 °C then 25 °C, 24 h; b) Glacial acetic acid, POCl₃, under reflux, 90 min. (iii) Ethyl acetoacetate, CH₃COONa, under microwave-assisted conditions, 90 min. (iv) Glacial acetic acid (cat), MeOH (solvent), 30–45 min under MW-assisted conditions at 400W.

The end of reaction was monitored by thin chromatography. Irradiation times were 30–45 min at microwave power of 400 W. Glacial acetic acid as catalyst was used in order to activate the carbonyl group of 3-acetylcoumarins and to facilitate nucleophilic attack of thiosemicarbazide, and protonation of hydroxyl group in the dehydration stage. Strong inorganic acids, such as HCl, H₂SO₄, HClO₄, did not favor for this reaction because so the nucleophilic amino group of thiosemicarbazide was protonated, which loose unshared electron pair. Weak organic acids, such as acetic acid, was sufficient to activate the carbonyl group without protonation of amino-thiosemicarbazide group. The formation of thiosemicarbazone products **5a-j** could be preliminary confirmed by IR spectroscopy and compared to the IR spectra of corresponding substituted 3-acetylcoumarins **3a-j** and thiosemicarbazide **4**. On IR spectrum of thiosemicarbazones, characteristic absorption bands appeared for both components, coumarin and per-acetylated D-glucose, and stretching band for imine group >C=N–, whereas specific bands for amino group in thiosemicarbazide and for carbonyl group in substituted 3-acetylcoumarin were not present. These evidences initially indicated that the reaction had occurred, and we expected thiosemicarbazone was formed. IR spectra showed characteristic absorptions in the range of 3354–3447 and 3328–3234 cm⁻¹ belonged stretching vibrations of N–H groups of thiosemicarbazone linkage group (–NHCSNHN=C<). Chemical shifts that appeared at 11.15–9.28 (singlet) and 9.05–8.66 ppm (doublet) in ¹H NMR spectra confirmed the presence of N–H groups in molecular structure of thiosemicarbazone **5a-j**. The former chemical shift was assigned to proton H_b and the latter one specified to proton H_a; this proton had coupling interaction to proton H-1 on pyranose ring with coupling constant $J = 9.25\text{--}8.50$ Hz. These values of coupling constants agreed with *trans*-axial H–H disposition and β-anomeric configuration. Resonance of carbon atom C-1 on pyranose ring was easily recognized at δ= 81.5 ppm, which was the signal lying in the weakest field in resonance region of pyranose's carbon. The cause of this phenomenon was that the carbon atom C-1 was influenced by electronegative oxygen atom of pyranose ring and of the adjacent nitrogen atom of thiosemicarbazone linkage. ¹³C NMR spectra of compound **5a-j** showed resonance signals at δ=181.6–180.0 ppm (carbon atom in C=S group),

δ =170.5–168.8 ppm (carbon atoms in C=O bond of acetyl groups). Carbon atoms in coumarin ring had chemical shifts at δ =155.4–110.1 ppm, whereas resonance signals at δ =82.0–61.1 ppm belonged carbon atoms on D-glucopyranose ring, and chemical shifts were at δ =21.5–21.1 ppm belonged methyl carbons in acetyl groups. Carbon atom in lactone carbonyl group on coumarin ring had chemical shift at δ =159.6–158.7 ppm.

All the synthesized thiosemicarbazones **5a-j** were screened for their *in vitro* antibacterial activity against three representative Gram-positive bacteria, which were *B. subtilis* (ATCC 11774), *S. aureus* (ATCC 11632), *S. epidermidis* (ATCC 12228), and four representative Gram-negative bacteria, which were *E. coli* (ATCC 25922), *P. aeruginosa* (ATCC 25923), *K. pneumoniae* (ATCC 4352), *S. typhimurium* (ATCC 14028). Some antibiotics were used as positive references for antibacterial activity, including ciprofloxacin and vancomycin. Vancomycin and ciprofloxacin are used to treat a number of bacterial infections, with the former used to treat serious, life-threatening infections by Gram-positive bacteria that are unresponsive to other antibiotics. The latter treated serious infections caused by Gram-negative bacteria, including *P. aeruginosa*. All the experiments were performed three times. Evaluated results for compounds **5a-j** were given in Table 1.

The results showed that almost all novel molecules exhibited antibacterial activity against the tested bacteria at their different concentrations. In general, it has been observed that almost all tested compounds showed mild to moderate activity against the tested bacteria in comparison with the MIC values of the reference compounds. MIC values of these reference drugs are as follows: Ciprofloxacin, 3.125 μ M (for Gram-positive bacteria), 1.56 μ M (for Gram-negative bacteria); Vancomycin, 0.78–3.125 μ M (for Gram-positive bacteria). Some thiosemicarbazones **5a-j** had higher ability to inhibit to both Gram-positive bacteria (*B. subtilis*, *C. difficile*, *S. aureus*, and *S. epidermidis*) and Gram-negative bacteria (*E. Coli*, *K. pneumoniae*, *P. aeruginosa*, and *S. Typhimurium*), with MIC values of 1.56–3.125 μ M. From obtained results of inhibitory activities in Table 1 for tested bacteria, some comments were discussed as follows. Almost all thiosemicarbazones **5a-j** series had remarkable inhibitory activity against Gram-positive bacteria *S.*

aureus, *S. epidermidis* (especially, compounds **5h** and **5i** with MIC = 0.78 and 1.56 μ M), Gram-negative bacteria *E. coli* (especially, compound **5f** with 8-methoxy group, MIC = 1.56 μ M) and *P. aeruginosa* (**5g**, MIC = 1.56 μ M). Thiosemicarbazones **5a-j** exhibited similar inhibitory activities for both Gram-negative and Gram-positive bacteria. Almost all these compounds displayed significant inhibitory activity against all tested bacteria (with MIC were equal 3.125 μ M or smaller), especially for *S. aureus* and *S. epidermidis*. In ranges of MIC = 0.78–3.125 μ M, thiosemicarbazones **5a-j** series displayed better inhibitory activity against Gram-negative bacteria than against Gram-positive bacteria. In ranges of MIC = 0.78–3.125 μ M. Compound **5i** and **5h** had strongest inhibitory activity against bacterium *S. aureus*. Both compounds had MIC values of 0.78 μ M.

Based on the obtain results of the inhibitory activities in Table 1 for the tested bacteria, some SAR results were deduced as follows:

1. Thiosemicarbazone **5a** from unsubstituted 3-acetylcoumarin (**3a**) had unnoticeable inhibitory activity against all tested bacteria, with MIC values in the range from 50 μ M to 400 μ M, except against bacterium *S. epidermidis* with MIC = 12.5 μ M
2. Electron-donating groups on benzene ring of coumarin (such as methyl, hydroxy, and methoxy): Compounds that had strong inhibitory activity with MIC values of 0.78–3.125 μ M against Gram-positive bacteria were **5e** and **5i** (for *S. aureus*), **5f** and **5i** (for *S. epidermidis*). Compounds that exhibited notable inhibition against Gram-negative bacteria included **5f** (for *E. coli* and *K. pneumoniae*), **5i** (for *S. typhimurium*). The remaining compounds had unremarkably inhibitory activity for both tested bacterial types. Additionally, all of the tested compounds **5d,e,f,i,j** had unremarkably inhibitory activity against *B. subtilis*. The methyl group on position 4 led to decrease inhibition for almost all tested bacteria. The presence of 7-hydroxy group in compound **5i** strengthened inhibition against several tested bacteria, such as *S. aureus*, *S. epidermidis*, and *S. typhimurium*.
3. Electron-withdrawing groups on benzene ring of coumarin (including 6-chloro, 6-bromo, 6-nitro, and 8-nitro): In general, thiosemicarbazones **5b,c,g,h** had unremarkable inhibitory

activity against tested Gram-positive bacteria, except **5h** (for *S. aureus*) and **5c** (for *S. epidermidis*). For Gram-negative bacteria, compounds **5c** (bromo substituent) and **5g** (nitro substituent) had strong inhibition against *E. coli*, and *P. aeruginosa* with MICs of 1.56–3.125 μ M. The presence of chloro substituent (compound **5b**) increased activity against only for *P. aeruginosa*.

We also evaluated the antifungal activities of the above thiosemicarbazones **5a-j** against some fungi, such as *Aspergillus niger* (ATCC 439), *Aspergillus flavus* (ATCC 204304), *Candida albicans* (ATCC 7754), and *Saccharomyces cerevisiae* (SH 20). Miconazole and fluconazole were used as references. Miconazole is mainly used externally for the treatment and prophylactic treatment of *Candida* infection in the oral cavity and the digestive tract. Fluconazole is a first-generation triazole antifungal medication. It has a spectrum of activity, which includes most *Candida* species but not *C. krusei* or *C. glabrata*, *Cryptococcus neoformans*, some dimorphic fungi, and dermatophytes. MIC values of miconazole were 1.56, 1.56, 3.125 and 3.125 μ M for each fungus, respectively, and of fluconazole were 1.56, 0.78, 0.78 and 0.78 μ M for each fungus, respectively. All the experiments were performed three times. The obtained results were given in Table 2. Some comments could deduce as follows. In range of MIC values of 0.78–3.125 μ M, almost all the tested thiosemicarbazones **5a-j** were remarkably active against fungus *A. niger* and *C. albicans*, and were more resistant to *A. niger*, *A. flavus*, and *S. cerevisiae* than miconazole and fluconazole. Remarkably, compounds **5h** (against *C. albicans*) and **5i** (against *S. cerevisiae*) were more active (all MIC = 0.78 μ M) than miconazole and comparably active with miconazole. Almost all compounds **5a-j** did not exhibit inhibitory activity against fungus *S. cerevisiae*, except compound **5i** (bearing 4-methyl and 7-hydroxy) that expressed the best inhibition with MIC = 0.78 μ M that could be comparable with reference drug fluconazole (MIC = 0.78 μ M). This compound had higher inhibition than miconazole (MIC = 3.125 μ M) against *S. cerevisiae*.

Table 1. Antibacterial activity of thiosemicarbazones **5a-j**

Compounds	Micro-organisms/ MIC (μM)						
	Gram-positive bacteria			Gram-negative bacteria			
	<i>B. subtilis</i>	<i>S. aureus</i>	<i>S. epidermidis</i>	<i>E. coli</i>	<i>K. pneumoniae</i>	<i>P. aeruginosa</i>	<i>S. typhimurium</i>
5a	>100	N.E.	12.5 ± 0.65	50 ± 4.23	100 ± 12.35	50 ± 3.56	100 ± 12.35
5b	N.E.	>100	>100	25 ± 1.45	N.E.	3.125 ± 0.48	50 ± 3.75
5c	N.E.	100 ± 12.35	3.125 ± 0.32	3.125 ± 0.37	25 ± 1.45	3.125 ± 0.51	25 ± 1.12
5d	N.E.	12.5 ± 0.78	50 ± 3.51	50 ± 3.54	100 ± 12.35	12.5 ± 0.95	25 ± 1.18
5e	100 ± 11.45	1.56 ± 0.14	N.E.	25 ± 1.15	25 ± 1.14	25 ± 1.15	N.E.
5f	50 ± 3.43	50 ± 3.39	3.125 ± 0.36	1.56 ± 0.75	1.56 ± 0.71	N.E.	N.E.
5g	>100	12.5 ± 0.86	12.5 ± 0.75	3.125 ± 0.41	50 ± 3.46	1.56 ± 0.21	100 ± 12.35
5h	12.5 ± 0.79	0.78 ± 0.08	12.5 ± 0.72	25 ± 1.15	12.5 ± 0.71	N.E.	>100
5i	100 ± 11.35	0.78 ± 0.07	1.56 ± 0.15	100 ± 12.35	100 ± 12.35	12.5 ± 0.69	3.125 ± 0.32
5j	>100	25 ± 1.21	100 ± 12.35	N.E.	50 ± 4.35	N.E.	N.E.
Ciprofloxacin	3.125 ± 0.34	3.125 ± 0.36	3.125 ± 0.35	1.56 ± 0.23	1.56 ± 0.15	1.56 ± 0.16	1.56 ± 0.15
Vancomycin	1.56 ± 0.12	3.125 ± 0.25	0.78 ± 0.06	–	–	–	–

Note: N.E. = not evaluated

Table 2. Antifungal activity of thiosemicarbazones **5a-j**

Compounds	Fungi/ MIC (μ M)			
	<i>A. niger</i>	<i>A. flavus</i>	<i>C. albicans</i>	<i>S. cerevisiae</i>
5a	>100	25 \pm 1.65	100 \pm 15.09	50 \pm 3.54
5b	100 \pm 12.12	12.5 \pm 1.10	1.56 \pm 0.06	50 \pm 3.35
5c	N.E.	6.25 \pm 0.78	50 \pm 3.34	25 \pm 1.34
5d	1.56 \pm 0.27	N.E.	3.125 \pm 0.89	25 \pm 1.65
5e	25 \pm 1.64	>100	3.125 \pm 0.96	>100
5f	6.25 \pm 0.84	6.25 \pm 0.82	25 \pm 1.26	50 \pm 3.46
5g	12.5 \pm 0.89	100 \pm 12.17	25 \pm 1.52	25 \pm 1.54
5h	3.125 \pm 0.34	12.5 \pm 1.04	0.78 \pm 0.06	12.5 \pm 0.96
5i	3.125 \pm 0.23	1.56 \pm 0.34	>100	0.78 \pm 0.09
5j	12.5 \pm 0.97	25 \pm 1.54	12.5 \pm 0.89	12.5 \pm 0.89
Miconazole	1.56 \pm 0.09	1.56 \pm 0.21	3.125 \pm 0.24	3.125 \pm 0.22
Fluconazole	1.56 \pm 0.12	0.78 \pm 0.02	0.78 \pm 0.03	0.78 \pm 0.02

Some SAR results were deduced as follows, based on obtained results of inhibitory activities for all tested fungi:

1. Thiosemicarbazones from unsubstituted 3-acetylcoumarin (**5a**) had no activity against all tested fungi.
2. The electron-donating groups on benzene ring of coumarin, such as methyl, hydroxy, and methoxy: In general, these groups created strong to medium inhibitory activity against tested fungi in thiosemicarbazones **5d,e,f,i,j** with MIC = 0.78–12.5 μ M when compared with thiosemicarbazone of unsubstituted 3-acetylcoumarin. Compound **5i** had remarkable inhibitory activity against three fungi, *A. niger*, *A. flavus* and *S. cerevisiae*.

3. The electron-withdrawing groups on benzene ring of coumarin (such as 6-chloro, 6-bromo, 6-nitro, and 8-nitro): These substituents also decreased the inhibitory activity against *A. niger* and *S. cerevisiae* of thiosemicarbazones **5b,c,g,h**, but greatly increased the inhibitory activity against *A. flavus* (**5g,h**) and *C. albicans* (**5b,h**). Compound **5h** exhibited strong activity against *A. niger* and *C. albicans*.

In summary, microwave-assisted heating method was an efficient and convenient method for synthesis of *N*-(2,3,4,6-tetra-*O*-acetyl- β -D-glucopyranosyl)thiosemicarbazones of substituted 3-acetylcoumarins. All synthesized thiosemicarbazones except for **5a** displayed significant inhibition in vitro against bacteria (*B. subtilis*, *S. aureus*, *S. epidermidis*, *E. coli*, *P. aeruginosa*, *K. pneumoniae*, *S. typhimurium*) and fungi (*A. niger*, *C. albicans*, *S. cerevisiae*, and *A. flavus*).

Appendix A. Supplementary data

Supplementary data (experimental section, IR, ^1H NMR, ^{13}C NMR, mass spectral data and elemental analysis for the synthesized compounds) can be found, in the online version, at <http://dx.doi.org/10.1016/j.bmcl...>

References

1. Bal-Demirci T, Şahin M, Kondakçı E, Özyürek M, Ülküseven B, Apak R. Synthesis and antioxidant activities of transition metal complexes based 3-hydroxysalicylaldehyde-*S*-methylthiosemicarbazone. *Spectrochim Acta A: Mol Biomol Spectr.* 2015;138:866-872. <https://doi.org/10.1016/j.saa.2014.10.088>.
2. Gou Y, Wang J, Chen S, et al. α -*N*-heterocyclic thiosemicarbazone Fe(III) complex: Characterization of its antitumor activity and identification of anticancer mechanism. *Eur J Med Chem.* 2016;123:354-364. <http://doi.org/10.1016/j.ejmech.2016.07.041>.

3. Lobana TS, Sharma R, Bawa G, Khanna S. Bonding and structure trends of thiosemicarbazone derivatives of metals—An overview. *Coord Chem Rev.* 2009;253(7):977-1055. <https://doi.org/10.1016/j.ccr.2008.07.004>.
4. Martins Alho MA, D'Accorso NB. Behavior of free sugar thiosemicarbazones toward heterocyclization reactions. *Carbohydr Res.* 2000;328(4):481-488. [https://doi.org/10.1016/S0008-6215\(00\)00127-0](https://doi.org/10.1016/S0008-6215(00)00127-0).
5. Ashry ESHE. Heterocycles from Carbohydrate Precursors. In: Gupta RR, ed. *Topics in Heterocyclic Chemistry*. Vol 7. Springer-Verlag Berlin Heidelberg; 2007.
6. El-Sharief MAMS, Abbas SY, El-Bayouki KAM, El-Gammal EW. Synthesis of thiosemicarbazones derived from *N*-(4-hippuric acid) thiosemicarbazide and different carbonyl compounds as antimicrobial agents. *Eur J Med Chem.* 2013;67:263-268. <http://doi.org/10.1016/j.ejmech.2013.06.031>.
7. Rogolino D, Gatti A, Carcelli M, et al. Thiosemicarbazone scaffold for the design of antifungal and antiaflatoxic agents: evaluation of ligands and related copper complexes. *Sci Rep.* 2017;7(1):11214. <https://doi.org/10.1038/s41598-017-11716-w>.
8. Wang Y, Gu W, Shan Y, et al. Design, synthesis and anticancer activity of novel nopinone-based thiosemicarbazone derivatives. *Bioorg Med Chem Lett.* 2017;27(11):2360-2363. <https://doi.org/10.1016/j.bmcl.2017.04.024>.
9. Trotsko N, Golus J, Kazimierzak P, et al. Design, synthesis and antimycobacterial activity of thiazolidine-2,4-dione-based thiosemicarbazone derivatives. *Bioorg Chem.* 2020;97:103676. <https://doi.org/10.1016/j.bioorg.2020.103676>.
10. Shehzad MT, Imran A, Njateng GSS, et al. Benzoxazinone-thiosemicarbazones as antidiabetic leads via aldose reductase inhibition: Synthesis, biological screening and molecular docking study. *Bioorg Chem.* 2019;87:857-866. <https://doi.org/10.1016/j.bioorg.2018.12.006>.

11. Ghosh S, Misra AK, Bhatia G, Khan MM, Khanna AK. Syntheses and evaluation of glucosyl aryl thiosemicarbazide and glucosyl thiosemicarbazone derivatives as antioxidant and anti-dyslipidemic agents. *Bioorg Med Chem Lett*. 2009;19(2):386-389. <https://doi.org/10.1016/j.bmcl.2008.11.070>.
12. Tenchiu A-C, Kostas ID, Kovala-Demertzi D, Terzis A. Synthesis and characterization of new aromatic aldehyde/ketone 4-(β -D-glucopyranosyl)thiosemicarbazones. *Carbohydr Res*. 2009;344(11):1352-1364. <https://doi.org/10.1016/j.carres.2009.05.010>.
13. Thanh ND, Giang NTK, Quyen TH, Huong DT, Toan VN. Synthesis and evaluation of in vivo antioxidant, in vitro antibacterial, MRSA and antifungal activity of novel substituted isatin *N*-(2,3,4,6-tetra-*O*-acetyl- β -D-glucopyranosyl)thiosemicarbazones. *Eur J Med Chem*. 2016;123:532-543. <https://doi.org/10.1016/j.ejmech.2016.07.074>.
14. Bognár R, Somogyi L, Szilágyi L, Györgydeák Z. *N*-Glykosyl-Derivate: Teil XIII. Der nachträgliche ausbau des aglykons. Synthese von *N*-Glykosyl-Derivaten des 2-amino-thiazols, 2-amino-1,3,4-thiadiazols und 5-amino-1,2,3,4-thiatriazols. *Carbohydr Res*. 1967;5(3):320-328. [https://doi.org/10.1016/S0008-6215\(00\)80506-6](https://doi.org/10.1016/S0008-6215(00)80506-6).
15. Bonaccorso C, Grasso G, Musso N, et al. Water soluble glucose derivative of thiocarbohydrazone acts as ionophore with cytotoxic effects on tumor cells. *J Inorg Biochem*. 2018;182:92-102. <https://doi.org/10.1016/j.jinorgbio.2018.01.019>.
16. Fonseca NC, da Cruz LF, da Silva Villela F, et al. Synthesis of a Sugar-Based Thiosemicarbazone Series and Structure-Activity Relationship versus the Parasite Cysteine Proteases Rhodesain, Cruzain, and *Schistosoma mansoni* Cathepsin B1. *Antimicrob Agents Chemother*. 2015;59(5):2666-2677. <http://doi.org/10.1128/aac.04601-14>.
17. Alexacou K-M, Tenchiu A-C, Chrysina ED, et al. The binding of β -d-glucopyranosyl-thiosemicarbazone derivatives to glycogen phosphorylase: A new class of inhibitors. *Bioorg Med Chem*. 2010;18(22):7911-7922. <https://doi.org/10.1016/j.bmc.2010.09.039>.

18. Souza Simone Md, Monache Franco D, Smânia A. Antibacterial Activity of Coumarins. *Z Naturforsch, C: Chem Sci.* 2005;60(9-10):693. <http://doi.org/10.1515/znc-2005-9-1006>.
19. Guilherme Borges B, Damiana da Rocha V, Alexander M-R, et al. The Antioxidant Activity of Coumarins and Flavonoids. *Mini-Reviews in Medicinal Chemistry.* 2013;13(3):318-334. <http://dx.doi.org/10.2174/1389557511313030002>.
20. Bansal Y, Sethi P, Bansal G. Coumarin: a potential nucleus for anti-inflammatory molecules. *Med Chem Res.* 2013;22(7):3049-3060. <http://doi.org/10.1007/s00044-012-0321-6>.
21. Yang F, Zhao N, Song J, et al. Design, Synthesis and Biological Evaluation of Novel Coumarin-Based Hydroxamate Derivatives as Histone Deacetylase (Hdac) Inhibitors with Antitumor Activities. *Molecules.* 2019;24(14):2569. <http://doi.org/10.3390/molecules24142569>.
22. Ito Y, Kitagawa H, Tamaoki B. Coumarin derivatives for medical purpose.1. Anthelmintic action. *Yakugaku Zasshi.* 1950;70(12):730-733. http://doi.org/10.1248/yakushi1947.70.12_730.
23. Soine TO. Naturally Occurring Coumarins and Related Physiological Activities. *J Pharm Sci.* 1964;53(3):231-264. <http://doi.org/10.1002/jps.2600530302>.
24. Reddy DS, Hosamani KM, Devarajegowda HC, Kurjogi MM. A facile synthesis and evaluation of new biomolecule-based coumarin–thiazoline hybrids as potent anti-tubercular agents with cytotoxicity, DNA cleavage and X-ray studies. *RSC Adv.* 2015;5(79):64566-64581. <https://doi.org/10.1039/C5RA09508E>.
25. Anand A, Kulkarni MV, Joshi SD, Dixit SR. One pot Click chemistry: A three component reaction for the synthesis of 2-mercaptobenzimidazole linked coumarinyl triazoles as anti-tubercular agents. *Bioorg Med Chem Lett.* 2016;26(19):4709-4713. <https://doi.org/10.1016/j.bmcl.2016.08.045>.

26. Serra S, Ferino G, Matos MJ, et al. Hydroxycoumarins as selective MAO-B inhibitors. *Bioorg Med Chem Lett*. 2012;22(1):258-261. <https://doi.org/10.1016/j.bmcl.2011.11.020>.
27. Nagy N, Kuipers HF, Frymoyer AR, et al. 4-Methylumbelliferone Treatment and Hyaluronan Inhibition as a Therapeutic Strategy in Inflammation, Autoimmunity, and Cancer. *Front Immunol*. 2015;6(123). <https://doi.org/10.3389/fimmu.2015.00123>.
28. Wang Y, Wang Y, Li P, Tang Y, Fawcett JP, Gu J. Quantitation of Armillarisin A in human plasma by liquid chromatography–electrospray tandem mass spectrometry. *J Pharm Biomed Anal*. 2007;43(5):1860-1863. <https://doi.org/10.1016/j.jpba.2006.12.023>.
29. Jang JH, Park JE, Han JS. Scopoletin inhibits α -glucosidase in vitro and alleviates postprandial hyperglycemia in mice with diabetes. *Eur J Pharmacol*. 2018;834:152-156. <https://doi.org/10.1016/j.ejphar.2018.07.032>.
30. Nguyen DT, Le TH, Bui TTT. Antioxidant activities of thiosemicarbazones from substituted benzaldehydes and *N*-(tetra-*O*-acetyl- β -D-galactopyranosyl)thiosemicarbazide. *Eur J Med Chem*. 2013;60:199-207. <https://doi.org/10.1016/j.ejmech.2012.10.004>.
31. Thanh ND, Kim Van HT, Thu TT. Synthesis and Characterization of Some Novel Thiosemicarbazones of Substituted Benzaldehydes and *N*-(Hepta-*O*-Acetyl- β -D-Lactosyl)Thiosemicarbazide. *J Carbohydr Chem*. 2015;34(9):514-544. <http://doi.org/10.1080/07328303.2015.1114119>.
32. Thanh ND, Duc HT, Duyen VT, Tuong PM, Van Quoc N. Synthesis and antibacterial and antifungal activities of *N*-(tetra-*O*-acetyl- β -D-glucopyranosyl)thiosemicarbazones of substituted 4-formylsydnones. *Chem Cent J*. 2015;9(1):60. <http://doi.org/10.1186/s13065-015-0138-8>.

Highlights

- Ten new thiosemicarbazones of substituted 3-acetylcoumarins with D-glucose moiety were synthesized in 45–68% yields.
- Several thiosemicarbazones were active against Gram-(+) and Gram-(–) bacteria with MICs of 1.56–3.125 μ M.
- Some thiosemicarbazones had activity against fungi with MICs of 1.56–3.125 μ M.

- Compounds **5h** and **5i** exhibited excellent activity against *S. aureus* with MIC of 0.78 μ M.
- Compounds exhibited excellent activity with MIC of 0.78 μ M, including **5h** (for *C. albicans*), and **5i** (for *S. cerevisiae*).