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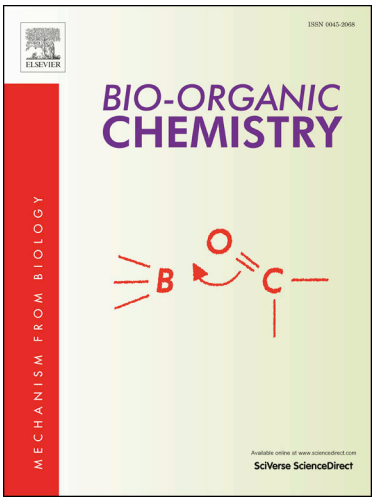

Synthesis, biological properties, and acid dissociation constant of novel naphthoquinone–triazole hybrids

Yahya Nural, Sadin Ozdemir, Osman Doluca, Bunyamin Demir, Mustafa Serkan Yalcin, Hasan Atabey, Beyza Kanat, Selma Erat, Hayati Sari, Zeynel Seferoglu

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naphthoquinone–triazole hybrids

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A series of novel 1,4-naphthoquinone–triazole hybrids, *N*-(3-amino-1,4-dioxo-1,4-dihydronaphthalen-2-yl)-2-(4-R-1*H*-1,2,3-triazol-1-yl)acetamide, was synthesized by click chemistry in the presence of sodium ascorbate and copper(II) sulfate pentahydrate in 81-94% yield. Various biological properties of the synthesized compounds including DNA binding/cleavage, antioxidant, antibacterial and antifungal properties were evaluated. The DNA binding study was performed using dsDNA and G-quadruplex DNA. All of the compounds showed fluorescence increase in the presence of DNA, regardless of the structure. Up to 2.9 and 2.5 times fluorescence increase upon incubation with double stranded or G-quadruplex DNA was detected for **5f** and **5g**, respectively. The docking studies performed on dsDNA and G-quadruplex structures suggested compounds' mode of interactions were populated around the grooves. All of the compounds showed excellent DNA cleavage activity and **5e** was almost degraded all of the plasmid DNA. The highest radical scavenging activity was obtained as 89.9% at 200 mg/L with **5d**. However, the highest ferrous chelating activity was obtained as 68.1% at 200 mg/L with **5g**. The compounds exhibited antimicrobial activity against *Bacillus cereus*, *Legionella pneumophila subsp. pneumophiia*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Escherichia coli* and *Enterococcus hirae* bacteria strains and two microfungus *Candida albicans* and *Candida tropicalis* strains. The compounds exhibited antibacterial and antifungal activity in the range of 4-128 μg/mL and 16-128 μg/mL, respectively. The best antimicrobial activity was obtained with **5d** and **5e** with a MIC value of 4 μg/mL against *Enterococcus hirae*. The acid dissociation constants (pK_a) were determined potentiometrically in 20% (v/v) dimethyl sulfoxide-water hydro-organic solvent at an ionic background of 0.1 mol/L of NaCl, at 25±0.1 °C. Five pK_a values were obtained for each ligand.

***Keywords**:* Naphthoquinone; Triazole; Click chemistry; DNA-binding; DNA-cleavage; Antioxidant; Antibacterial; Antifungal; Acid dissociation constants

Click chemistry, a term introduced by Sharpless in 2001 [1], is one of the most popular strategy for synthesis of pharmacologically active compounds. Because click chemistry offers many advantages such as stereo- and regio-specific product, high yield, mild reaction conditions etc., it has rapidly become a popular tool in drug research. Click chemistry allows the reliable synthesis of a large number of new compounds. Copper-catalyzed azide-alkyne cycloaddition click chemistry is widely used in the synthesis of 1,4-substituted-1,2,3-triazoles as regio- and stereo-specific. Triazoles represent an important group of pharmacophores of drug chemistry and there are many drugs or drug candidates containing 1,2,3-triazole moiety such as Rufinamide, Savolitinib, Seviteronel, Solithromycin, Ticagrelor, Molidustat and Tazobactam in the markets (Fig. 1) [2-5].

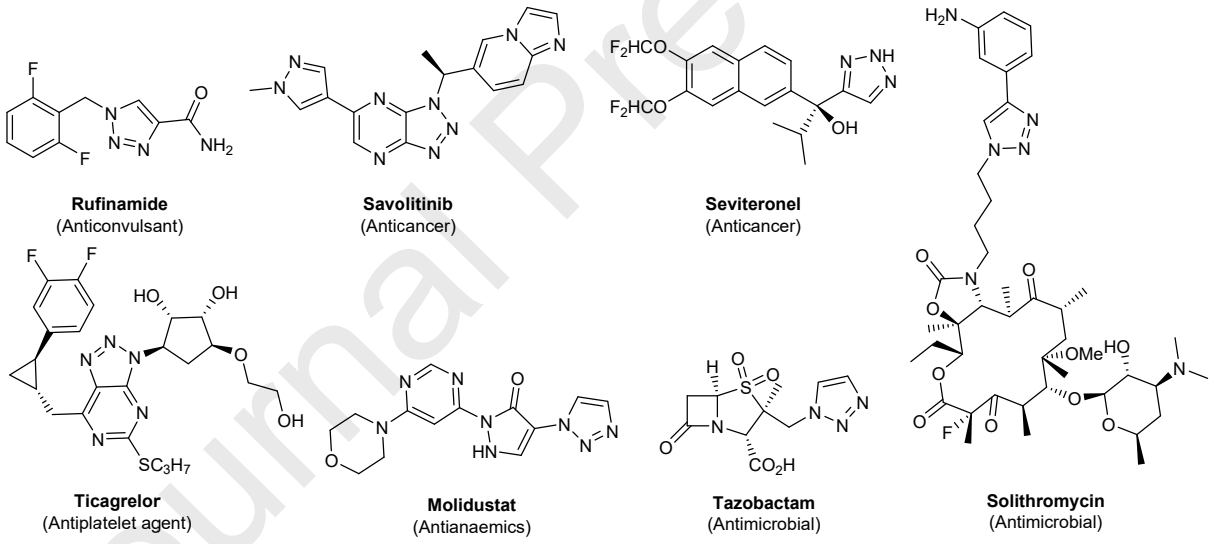


Figure 1. Some pharmaceuticals based on 1,2,3-triazoles

Triazole moiety has many indispensable properties desired in drug researches such as non-toxicity, stability under physiological conditions, H-bonds and π-π stacking interactions and having both acidic and basic character [1-5]. It is known that 1,2,3-triazole-containing hybrids exhibited a wide range of pharmacological activities such as antibacterial [6-9], antimycobacterial [10], antifungal [11-13], antiviral [14], anticancer [15-17], DNA–binding

Alzheimer disease [25].

Naphthoquinone derivatives naturally occur in various fungi, bacteria and plants species and play an active role in various biological processes [26]. The 1,4-naphthoquinone is one of the privileged pharmacophore group in drug research since, a great number of naturally occurring compounds and many synthetic drugs such as atovaquone, buparvaquone and lapachol contain naphthoquinone core in their molecular structures (Fig. 2) [27,28]. The 1,4-naphthoquinone derivatives are known to showed a wide range of pharmacological activities, such as antibacterial [29-33], antifungal [29,34], antimalarial [35], antioxidant [36,37], anticancer [38,39], DNA–binding/cleavage [40,41] activities. The synthesis of hybrids that contain at least two pharmacophore groups is an important strategy for achieving invaluable pharmacological activities. The binding of 1,2,3-triazole core to the molecular structure of pharmacologically active compounds has become very popular in recent years [2-5]. Within this framework, there are numerous studies in medicinal chemistry on the synthesis of compounds that have both triazole and naphthoquinone moieties in their molecular structure. It has been reported that the compounds exhibited a wide range of pharmacological activities such as anticancer [42], anti-inflammatory [43] anti-*T. cruzi* [44], leishmanicidal [45] and antimycobacterial [46] activity. In the light of this information, it was aimed to synthesize potentially bioactive novel hybrid compounds containing naphtoquinone and 1,2,3-triazole pharmacophore groups and to investigate a wide range of biological activities. Although many compounds bearing 1,2,3-triazole and 1,4-naphtoquinone rings have been synthesized in literature, there is no hybrid similar to 1,4-naphthoquinone–triazole hybrids to be reported in present study.

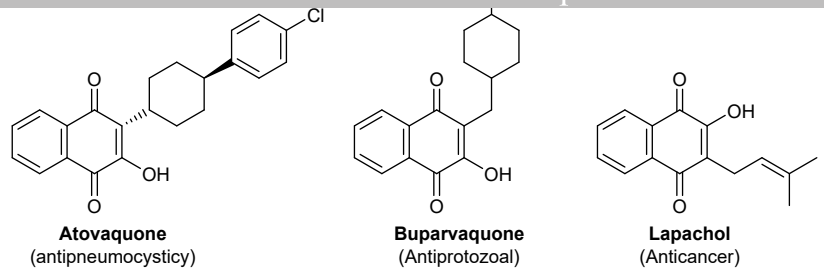


Figure 2. Some pharmaceuticals based on naphthoquinones

In present study, novel 1,4-naphthoquinone–1,2,3-triazole hybrids were synthesized and a wide range pharmacological activities of synthesized hybrids (DNA–binding/cleavage, antioxidant, antibacterial and antifungal properties) were reported. Additionally, acid dissociation constants (pK_a) were determined in 20% (v/v) dimethyl sulfoxide (DMSO) were also reported.

2. Experimental

2.1. Materials and instrumentation

The precursor chemicals purchased from Merck or Aldrich were high-grade and used without further purification. Fourier-transform infrared spectroscopy (FTIR) spectra were recorded by a Mattson 1000 FTIR spectrophotometer. Nuclear magnetic resonance (NMR) spectra and decoupling experiments were recorded with a Bruker Ultrashield Plus Biospin GmbHt at 400 MHz. Chemical shifts were given in parts per million (δ) downfield from TMS as internal standard. Spectra were determined in dimethyl sulfoxide-d_6. The following abbreviations were used; s = singlet, d = doublet, dd = doublet of doublets, q = quartet, t = triplet, td = triplet of doublets and m = multiplet. High resolution mass spectra (HRMS) were recorded in Waters-LCT-Premier-XE-LTOF (TOF-MS) and Agilent 6224 TOF LC-MS instruments using electrospray ionization technique and are reported in m/z (rel. %). Melting points were

a Titroline 7000 automated titrator with SI-Analytics combined with a glass pH electrode, which could be controlled by a computer and had an automatic micro-burette.

2.2. The synthesis of 2,3-diaminonaphthalene-1,4-dione, ***2***

The 2,3-diaminonaphthalene-1,4-dione **2** was prepared from 2,3-dichloronaphthalene-1,4-dione **1** using the method specified in literatures [47,48] and structure of the compound **2** was confirmed by ^{1}H NMR spectroscopy, which were found to be identical with the data described in Ref. [47,48].

2.3. The synthesis of N-(3-amino-1,4-dioxo-1,4-dihydronaphthalen-2-yl)-2-chloroacetamide, ***3***

The stirred solution of 2,3-diaminonaphthalene-1,4-dione (0.190 g, 1 mmol) in chloroform (20 mL) at 0 ºC was supplemented with a solution of pyridine (0.160 g, 2 mmol) in chloroform (10 mL) and the resultant mixture was stirred for 15 min. After then, a solution of chloroacetyl chloride (0.230 g, 2 mmol) in chloroform (20 mL) was added to the reaction medium and was allowed the temperature to come to room temperature. Upon completion of the reaction after 6 h, the solvent was evaporated under reduced pressure and the crude product was first washed with water and then washed several times with diethyl ether and dichloromethane (DCM), respectively, in this way the pure product was obtained as orange powder. Yield, 0.20 g, 75%. m.p.: 198-200 ºC (decomp.). IR (cm^{-1}): υ_{max} 3412, 3259, 3210, 3005, 2954, 1683. ^{1}H NMR (400 MHz, DMSO-d_6): δ 9.36 (s 1H, NH), 7.99-7.95 (m, 2H, Ar-H), 7.83 (td, 1H, J = 7.5 Hz, 1.3 Hz, Ar-H), 7.74 (td, 1H, J = 7.5 Hz, 1.3 Hz, Ar-H), 6.99 (s, 2H, NH_2), 4.29 (s, 2H, CH_2). ^{13}C NMR (100 MHz, DMSO-d_6): δ 181.6 (C=O), 177.5 (C=O), 164.6 (C=O), 144.2, 134.9, 132.5,

265.0380; found 265.0371.

2.4. The synthesis of N-(3-amino-1,4-dioxo-1,4-dihydronaphthalen-2-yl)-2-azidoacetamide, ***4***

The stirred solution of the compound **3** (0.270 g, 1 mmol) in dimethylformamide (DMF) (10 mL) was supplemented with a solution of sodium azide (0.100 g, 1.5 mmol) in DMF (10 mL) and the resultant mixture was stirred at 75 °C for 18 h. After completion of the reaction, the mixture was quenched with saturated aqueous sodium bicarbonate, and extracted with ethyl acetate. The crude mixture was purified by column chromatography (ethyl acetate:hexane / 1:2) to have the pure product **4** (0.220 g, 81%) as brown powder. m.p.: 192-194 ºC (decomp.). IR (cm^{-1}): υ_{max} 3413, 3288, 3204, 3067, 2948, 2114, 1690. ^{1}H NMR (400 MHz, DMSO-d_6): δ 9.24 (s 1H, NH), 7.99-7.96 (m, 2H, Ar-H), 7.85-7.81 (m, 1H, Ar-H), 7.76-7.72 m, 1H, Ar-H), 7.05 (s, 2H, NH_2), 4.04 (s, 2H, CH_2). ^{13}C NMR (100 MHz, DMSO-d_6): δ 181.6 (C=O), 177.6 (C=O), 166.2 (C=O), 144.4, 134.9, 132.4, 132.2, 130.1, 125.8, 125.6, 111.6, 50.8. HRMS (ESI-TOF-MS): calcd. for $C_{12}H_9N_5O_3$ $[MH]^+$ 272.0785; 272.0784.

2.5. General procedure for the synthesis of 1,4-naphthoquinone–triazole hybrids, **5a–h**

The novel 1,4-naphthoquinone–triazole hybrids **5a–h** were synthesized by click chemistry. The stirred solution of the compound **4** (1 mmol) in DMF (20 mL) was supplemented with sodium ascorbate (0.6 mmol), $CuSO_4.5H_2O$ (0.3 mmol), corresponding alkyne compound (1.5 mmol) and 4 mL deionized water, respectively. Resultant mixture was stirred at room temperature for 16 h and then, the mixture was quenched with saturated aqueous sodium bicarbonate, and extracted with ethyl acetate. The crude product was purified by column chromatography (ethyl acetate:hexane / 1:2).

1-yl)acetamide, **5a**

Black powder. Yield, 0.33 g, 89%. m.p.: 172-174 ºC. IR (cm^{-1}): υ_{max} 3424, 3309, 3255, 3066, 2956, 2931, 1689, 1663, 1614. ^{1}H NMR (400 MHz, DMSO-d_6): δ 9.54 (s 1H, NH), 7.99-7.96 (m, 2H, Ar-H), 7.85-7.81 (m, 2H, Ar-H and triazole C-H), 7.74 (td, 1H, J = 7.5 Hz, 0.7 Hz, Ar-H), 7.05 (s, 2H, NH_2), 5.28 (s, 2H, $C(O)CH_2$), 290-2.85 (m, 1H, C2H), 1.66-1.57 (m, 1H, C3H), 1.53-1.44 (m, 1H, C4H), 1.30-1.20 (m, 2H, C3-H', C4H'), 1.21 (d, 3H, J = 7.3 Hz, $C1H_3$), 0.86 (t, 3H, J = 7.3 Hz, $C5H_3$). ^{13}C NMR (100 MHz, DMSO-d_6): δ 181.5 (C=O), 134.8, 132.4, 132.1, 130.0, 125.7, 125.5, 122.2, 51.6, 38.7, 30.0, 20.4, 19.6, 13.8. HRMS (ESI-TOF-MS): calcd. for $C_{19}H_{21}N_5O_3$ $[MH]^+$ 368.1723; found 368.1711.

2.5.2. N-(3-amino-1,4-dioxo-1,4-dihydronaphthalen-2-yl)-2-(4-cyclopropyl-1H-1,2,3-triazol-1-yl)acetamide, **5b**

Orange powder. Yield, 0.31 g, 91%. m.p.: 230-232 ºC (decomp.). IR (cm^{-1}): υ_{max} 3354, 3263, 3211, 3143, 3071, 3005, 2950, 1670, 1613. ^{1}H NMR (400 MHz, DMSO-d_6): δ 9.53 (s 1H, NH), 7.99-7.96 (m, 2H, Ar-H), 7.86-7.80 (m, 2H, Ar-H and triazole C-H), 7.73 (td, 1H, J = 7.5 Hz, 0.8 Hz, Ar-H), 7.04 (s, 2H, NH_2), 5.26 (s, 2H, $C(O)CH_2$), 1.99-1.93 (m, 1H, C1H), 0.92-0.88 (m, 2H, C2H, C3H), 0.74-0.70 (m, 2H, C2H', C3H'). ^{13}C NMR (100 MHz, DMSO-d_6): δ 181.5 (C=O), 177.5 (C=O), 164.2 (C=O), 148.6, 144.3, 134.8, 132.4, 132.1, 130.0, 125.7, 125.5, 122.1, 111.4, 51.6, 7.4 (2 x C), 6.4. HRMS (ESI-TOF-MS): calcd. for $C_{17}H_{15}N_5O_3$ $[MH]^+$ 338.1253; found 338.1245.

2.5.3. N-(3-amino-1,4-dioxo-1,4-dihydronaphthalen-2-yl)-2-(4-cyclopentyl-1H-1,2,3-triazol-1-yl)acetamide, **5c**

Green powder. Yield, 0.32 g, 86%. m.p.: 246-248 ºC (decomp.). IR (cm^{-1}): υ_{max} 3458, 3347, 3280, 3115, 3067, 2950, 2867, 1688, 1666, 1628. ^{1}H NMR (400 MHz, DMSO-d_6): δ 9.55 (s 1H, NH), 7.99-7.94 (m, 2H, Ar-H), 7.85-7.79 (m, 2H, Ar-H and triazole C-H), 7.75-7.72 (m, 1H, Ar-H), 7.07 (s, 2H, NH_2), 5.28 (s, 2H, $C(O)CH_2$), 3.18-3.06 (m, 1H, C1H), 2.06-1.94 (m, 2H, C2H, C5H), 1.70-1.56 (m, 6H, C2H', C3H, C3H', C4H, C4H' C5H'). ^{13}C NMR (100 MHz, DMSO-d_6): δ 181.5 (C=O), 177.5 (C=O), 164.2 (C=O), 144.3, 134.8, 132.4, 132.1, 130.0, 125.7, 125.5, 122.2, 111.5, 51.6, 36.1, 32.7 (2 x C), 24.6 (2 x C). HRMS (ESI-TOF-MS): calcd. for $C_{19}H_{19}N_5O_3$ $[MH]^+$ 366.1566; found 366.1550.

2.5.4. N-(3-amino-1,4-dioxo-1,4-dihydronaphthalen-2-yl)-2-(4-(cyclohexylmethyl)-1H-1,2,3-triazol-1-yl)acetamide, **5d**

Green powder. Yield, 0.37 g, 94%. m.p.: 202-204 ºC (decomp.). IR (cm^{-1}): υ_{max} 3465, 3356, 3281, 3109, 3062, 2925, 2852, 1688, 1669, 1625. ^{1}H NMR (400 MHz, DMSO-d_6): δ 9.55 (s 1H, NH), 8.03-7.93 (m, 2H, Ar-H), 7.85-7.81 (m, 2H, Ar-H and triazole C-H), 7.75-7.72 (m, 1H, Ar-H), 7.06 (s, 2H, NH_2), 5.29 (s, 2H, $C(O)CH_2$), 2.57-2.45 (m, 2H, C1H, C1H'), 1.65-1.53 (m, 6H, C2H, C3H, C4H, C5H, C6H, C7H), 1.23-1.06 (m, 3H, C4H', C5H', C6H'), 0.97-0.86 (m, 2H, C3H', C7H'). ^{13}C NMR (100 MHz, DMSO-d_6): δ 181.6 (C=O), 177.6 (C=O), 164.3 (C=O), 144.3, 134.9, 132.5, 132.2, 130.1, 125.8, 125.6, 123.9, 123.8, 111.5, 51.6, 37.6, 32.7, 32.4 (2 x C), 26.0, 25.6 (2 x C). HRMS (ESI-TOF-MS): calcd. for $C_{21}H_{23}N_5O_3$ $[MH]^+$ 394.1879; found 394.1869.

2.5.5. N-(3-amino-1,4-dioxo-1,4-dihydronaphthalen-2-yl)-2-(4-(2-bromoethyl)-1H-1,2,3-triazol-1-yl)acetamide, **5e**

3168, 3017, 2966, 1686, 1646, 1618. ^{1}H NMR (400 MHz, DMSO-d_6): δ 9.57 (s 1H, NH), 7.99-7.96 (m, 3H, Ar-H and triazole C-H), 7.85-7.81 (m, 1H, Ar-H), 7.75-7.72 (m, 1H, Ar-H), 7.05 (s, 2H, NH_2), 5.33 (s, 2H, C(O)CH_2), 3.75 (t, 2H, J = 6.5 Hz, CH_2), 3.23 (t, 2H, J = 6.5 Hz, CH_2). ^{13}C NMR (100 MHz, DMSO-d_6): δ 181.5 (C=O), 177.6 (C=O), 164.1 (C=O), 144.3, 134.9, 132.5, 132.2, 130.1, 125.8, 125.6, 124.3, 124.2, 111.5, 51.7, 32.6, 29.0. HRMS (ESI-TOF-MS): calcd. for $C_{16}H_{14}BrN_5O_3$ $[MH]^+$ 404.0358; found 404.0344.

2.5.6. Ethyl 1-(2-((3-amino-1,4-dioxo-1,4-dihydronaphthalen-2-yl)amino)-2-oxoethyl)-1H-1,2,3-triazole-4-carboxylate, **5f**

Brown powder. Yield, 0.32 g, 85%. m.p.: 162-164 ºC (decomp.). IR (cm^{-1}): υ_{max} 3459, 3383, 3241, 3149, 3016, 2969, 1712, 1673, 1641, 1621. ^{1}H NMR (400 MHz, DMSO-d_6): δ 9.66 (s 1H, NH), 8.72 (s, 1H, triazole C-H), 7.99-7.97 (m, 2H, Ar-H), 7.85-7.81 (m, 1H, Ar-H), 7.76-7.72 (m, 1H, Ar-H), 7.08 (s, 2H, NH_2), 5.43 (s, 2H, C(O)CH_2), 4.32 (q, 2H, J = 6.9 Hz, CH_2), 1.31 (t, 3H, J = 6.9 Hz, CH_3). ^{13}C NMR (100 MHz, DMSO-d_6): δ 181.5 (C=O), 177.5 (C=O), 163.6 (C=O), 160.2 (C=O), 144.3, 138.6, 134.8, 132.4, 132.1, 130.5, 130.0, 125.7, 125.5, 111.2, 60.4, 51.9, 14.1. HRMS (ESI-TOF-MS): calcd. for $C_{17}H_{15}N_5O_5$ $[MH]^+$ 370.1151; found 370.1143.

2.5.7. N-(3-amino-1,4-dioxo-1,4-dihydronaphthalen-2-yl)-2-(4-phenyl-1H-1,2,3-triazol-1-yl)acetamide, **5g**

Green powder. Yield, 0.30 g, 81%. m.p.: 164-166 ºC (decomp.). IR (cm^{-1}): υ_{max} 3456, 3349, 3274, 3131, 3066, 2956, 1687, 1668, 1624. ^{1}H NMR (400 MHz, DMSO-d_6): δ 9.62 (s 1H, NH), 8.55 (s, 1H, triazole C-H), 7.99-7.97 (m, 2H, Ar-H), 7.88-7.81 (m, 2H, Ar-H), 7.83 (td, 1H, J =

7.32 (m, 1H, Ar-H), 7.09 (s, 2H, NH_2), 5.42 (s, 2H, $C(O)CH_2$). ^{13}C NMR (100 MHz, DMSO-d_6): δ 181.6 (C=O), 177.6 (C=O), 164.1 (C=O), 146.2, 144.4, 134.9, 132.5, 132.2, 130.8, 130.1, 128.9 (2 x C), 127.8, 125.8, 125.6, 125.1 (2 x C), 122.9, 111.5, 51.9. HRMS (ESI-TOF-MS): calcd. for $C_{20}H_{15}N_5O_3$ $[MH]^+$ 374.1253; found 374.1262.

2.5.8. N-(3-amino-1,4-dioxo-1,4-dihydronaphthalen-2-yl)-2-(4-phenethyl-1H-1,2,3-triazol-1-yl)acetamide, **5h**

Brown powder. Yield, 0.33 g, 82%. m.p.: 177-179 ºC (decomp.). IR (cm^{-1}): υ_{max} 3365, 3207, 3154, 3025, 2953, 1695, 1668, 1639, 1615. ^1H NMR (400 MHz, DMSO-d_6): δ 9.54 (s 1H, NH), 7.99-7.97 (m, 2H, Ar-H), 7.85-7.81 (m, 2H, Ar-H and triazole C-H), 7.75-7.72 (m, 1H, Ar-H), 7.30-7.24 (m, 4H, Ar-H), 7.20-7.16 (m, 1H, Ar-H), 7.05 (s, 2H, NH_2), 5.29 (s, 2H, $C(O)CH_2$), 2.94 (s, 4H, 2 x CH_2). ^{13}C NMR (100 MHz, DMSO-d_6): δ 181.6 (C=O), 177.6 (C=O), 164.3 (C=O), 144.3, 141.2, 134.9, 132.5, 132.2, 130.1, 128.30 (2 x C), 128.27 (2 x C), 125.9, 125.8, 125.6, 123.6, 123.5, 111.6, 51.7, 34.9, 27.0 HRMS (ESI-TOF-MS): calcd. for $C_{22}H_{19}N_5O_3$ $[MH]^+$ 402.1566; found 402.1559.

2.6. DNA Binding

Double stranded DNA structure (dsDNA) was prepared by mixing 5′-GACGTGTCGAAAGAGCTCCGATTA-3′ and 5′-TAATCGGAGCTCTTTCGACACGTC-3′ oligonucleotides in Tris HCl buffer (pH 7.4). The mixture was heated up to 50 °C and left to cool down to room temperature. For G-quadruplexes, 5′-AGGGTTAGGGTTAGGGTTAGGG-3′ oligonucleotide was incubated in Tris HCl buffer containing 50 mM KCl. G-quadruplexes were heated up to 95 °C and left to

prepared at 50 μM.

The samples for fluorescence measurement were prepared for each compound at 10 μM in Tris HCl (pH 7.4) containing 5% DMSO. The DNA stock solutions were added to the sample to achieve respective equivalent concentrations. For all titrations, up to three consecutive fluorescence measurements were performed with 15 minutes intervals. Only the measurements after the fluorescence reached equilibrium were taken into account.

All fluorescence measurements were performed at 360 nm excitation and emissions between 380 and 700 nm with 10 nm excitation and emission slits using Jasco FP8300 in quartz cuvettes.

Docking studies of the compounds with DNA structures associated topologies were performed using Autodock Vina and in-house Python script to automate the process. To represent the dsDNA and the G-quadruplex, PDB files with accession numbers, 1BNA and 1XAV were used, respectively. After removal of water molecules, removal of non-polar hydrogens and calculation of Kollman charges, the PDB files were saved as PDBQT. Docking studies were performed with a grid size of 26 Å for all axes and centred at coordinates 0,0,0 for 1BNA and with a grid of 18, 18 and 40 Å for x, y and z axes and centred at coordinates 14.78, 20.976 and 8.807 for 1XAV. The exhaustiveness was set to 64. Docking studies were performed on a workstation with 32 cores and 128 GB ECC Ram.

2.7. DNA cleavage activity

The DNA cleavage activity was tested by agarose gel electrophoresis, which was studied by incubation at 37 °C as follows: pBR322DNA (0.1 μg/μL) in Tris–HCl 50 mM and NaCl buffer (18 mM; pH:7.2) was reacted with 1,4-naphthoquinone–triazole hybrids **5a–h** and then the mixture was incubated for 2 h. After that the samples were electrophoresed for 120 min at 80

were monitored under UV-A light and photographed [49].

2.8. DPPH Scavenging Activity

DPPH radical scavenging activities of the 1,4-naphthoquinone–triazole hybrids **5a–h** were performed according to the Blois method [50]. For this test, 2 mL of 0.004% DPPH solution was taken to each tube and then 0.5 mL of **5a–h** was added into each tube. The tubes were shaken quickly and kept at room temperature for 30 minutes in the dark for incubation. After incubation, absorbances were then measured at 517 nm. The ability to scavenge DPPH free radical was calculated by the following equation:

$$\%\text{Inhibition Activity} = [A_0\text{-}A_1)/A_0] \times 100$$

A_0 = Control absorbance and A_1 = the absorbance value of the solution containing compounds and DPPH after 30 min. The IC50 values were calculated using linear regression analysis and used to indicate antioxidant capacity.

2.9. Chelating Activity

The iron chelating activities of the 1,4-naphthoquinone–triazole hybrid compounds **5a–h** were determined with the method specified by Dinis et al. [51]. Different concentrations of **5a–h** solutions were separately put into the test tubes and $FeCl_2$ (0.05 mL of 2 mM) solution was added to each tubes. The reaction was started by adding 0.1 mL of 5 mM ferrozine ($C_{20}H_{13}N_4NaO_6S_2$). After the total volume was completed to 2.5 mL with the solvent used, the solution was mixed quickly and kept for 10 minutes at room temperature. Then absorbance values were read at 562 nm. Without adding **5a–h**, 50 µL $FeCl_2$ (2 mM) and 2.35 mL solvent were added to 100 µL of ferrozine (5 mM) and measured spectrophotometrically at 562 nm.

The same procedures were applied for the standard (EDTA). The chelating activity of the compounds for Fe^{2+} was calculated using the following equation:

$$\text{Chelating activity (\%)} = (A_{control} - A_{sample})/A_{control} \times 100;$$

where; $A_{control}$ is the absorbance of the control reaction, and A_{sample} represents the absorbance obtained in the presence of compounds or EDTA. In addition, IC50 values were calculated using linear regression analysis to determine iron chelating ability.

2.10. Antimicrobial activity

Six different bacteria strains (*Bacillus cereus*, *Legionella pneumophila* subsp. *pneumophiia* (ATCC 33152), *Staphylococcus aureus* (ATCC 6538), *Pseudomonas aeruginosa* (ATCC 9027), *Escherichia coli* (ATCC 10536) and *Enterococcus hirae* (ATCC 10541)) and two microfungus (*Candida albicans* and *Candida tropicalis* (ATCC 750)) were used to investigate antimicrobial activity. The minimum inhibition concentrations (MICs) of the tested compounds were evaluated by two-fold serial dilution method. Microorganisms (about 10^8-10^9 colony forming units (CFU/mL) were inoculated in medium containing different concentrations of **5a–h**. Culture media were then incubated at 37 °C and 120 rpm for 24 hours in a shaker [52].

2.11. Determination of Acid Dissociation Constants

The pH values were measured by model TitroLine® 7000 automatic titrator using a combined glass electrode. The glass electrode was calibrated using standard buffer solutions of pH 4.01, pH 7.00 and pH 10.01 according to the procedure described elsewhere [53,54] and determination of pK_a values was performed by modification of a literature method [55,56]. To maintain the ionic strength at a desired value a high concentrated solution of NaCl was used for

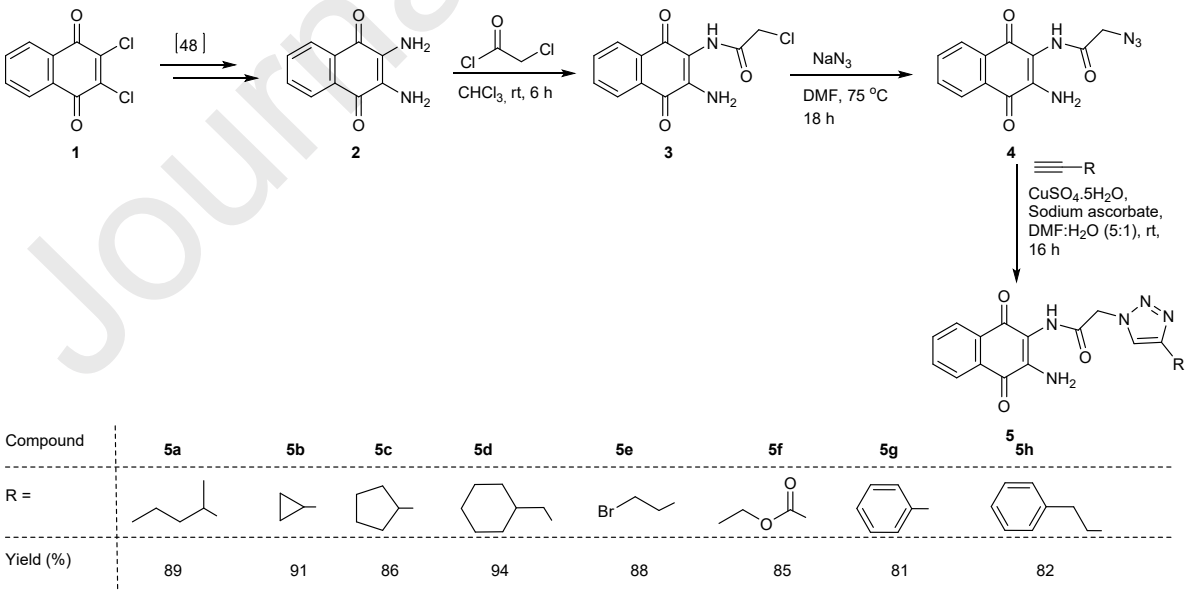
25.0±0.1 °C using a thermostat and the titration cell was stirred at a constant rate throughout the titration using a magnetic stirrer. The ligand **5a–h** solutions were prepared as 1.10^{-3} mol/L in DMSO, and 0.025 mol/L NaOH, 0.1 mol/L HCl and 1.0 mol/L NaCl stock solutions were prepared in deionized water. To determine the pK_a values of **5a–h** in 20% (v/v) DMSO-water hydro-organic solvent, titration cell was supplemented with 10 mL of the **5a–h** ligand solutions, 1 mL of the HCl solution and 5 mL of the NaCl solution from previously prepared stock solutions. After that, the titration cell was filled to 50.00 mL with deionized water, The pK_w value, which is defined as $-\log[H^+][OH^-]$ for the aqueous system, was obtained as 14.49±0.07 at the ionic strength employed. The pK_a values of **5a–h** were calculated from the potentiometric data using HYPERQUAD.

3. Results and discussion

3.1. Synthesis and characterization

The synthesis of 1,4-naphthoquinone–triazole hybrids **5a–h** was performed by click chemistry in the presence of copper(II) sulfate pentahydrate and sodium ascorbate as catalyst in 81-94% yield. The 2,3-diaminonaphthalene-1,4-dione **2** was prepared from 2,3-dichloronaphthalene-1,4-dione **1** according to a literature method [47,48]. The 2,3-diaminonaphthalene-1,4-dione **2** was reacted with chloroacetyl chloride, then the resultant product **3** was reacted with sodium azide to prepare the desired intermediate **4**, *N*-(3-amino-1,4-dioxo-1,4-dihydronaphthalen-2-yl)-2-azidoacetamide. To obtain the targeted products **5a–h**, the *N*-(3-amino-1,4-dioxo-1,4-dihydronaphthalen-2-yl)-2-azidoacetamide **4** was reacted with various alkyne compounds in the presence of sodium ascorbate (0.6 equivalent) and $CuSO_4.5H_2O$ (0.3 equivalent) in DMF / water (5:1 v/v) at room temperature [17]. **5a–h** were obtained with a good to excellent yield

fully characterized by various analytical techniques such as ^{1}H NMR, ^{13}C NMR, DEPT, COSY, HMQC, FT-IR and HRMS (See supplementary part). In the FT-IR spectra of **5a–h,** after formation of the triazole ring in the 1,4-naphthoquinone–triazole hybrids **5a–h**, the characteristic strong band of the azide group of the intermediate compound **3**, which appeared at 2114 cm^{-1}, disappeared. In the ^{1}H NMR spectra of **5a–h**, proton of the triazole ring were observed as a singlet in the range of 8.72 –7.79 ppm depending on inductive effect. In the ^{1}H NMR spectra of compound **4**, the signal observed as singlet at 4.04 ppm was assigned to the protons of C(O)CH_2 group. After formation of the triazole ring, in the ^{1}H NMR spectra of **5a–h**, the corresponding signal was observed as singlet in the range of 5.43–5.26 ppm. In the ^{13}C NMR spectra of **5a–h**, although the NMR solution was prepared as very concentrated and the NMR measurement time was extended, the carbon peak belonging to the quaternary carbon atom in the triazole ring was sometimes observed in the NMR spectrum and sometimes not observed. In the ^{13}C NMR spectra of **5a**, some of the C=O peaks were by no means observed. The shift values of the all carbon peaks originating from alkyne compounds have been found to be appropriate.



Scheme 1. Synthesis of the naphthoquinone–triazole hybrids **5a–h**

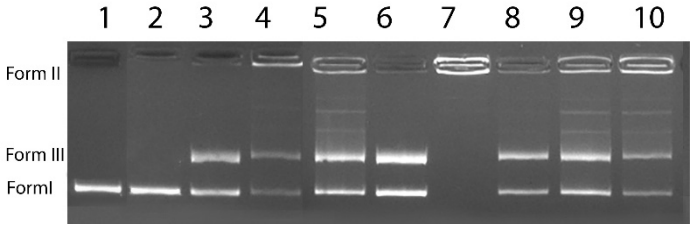
The potential of **5a–h** as DNA-binding fluorescent agents were analyzed through fluorescence change in the presence of dsDNA and G-quadruplex DNA. While all components showed absorbance at low wavelengths, the emission maxima varied between 400 and 440 depending on the compound. While all compounds showed very low fluorescence, an increase in fluorescence was apparent for all compounds, except for **5b** (Fig. S65). The fluorescence increase was most dramatic for **5f**, **5g** and **5h**. For these compounds, the titrations of dsDNA and G-quadruplex resulted in an increase in fluorescence (Fig. S66). Surprisingly, a shift in the emission maxima was apparent only for **5g** in the presence of dsDNA with a new maximum at 450 nm. The increase in intensity was tested up to 1:10 equivalent for dsDNA and 3:40 equivalent for G-quadruplex. The most dramatic increases in fluorescence emission in the presence of dsDNA was detected for **5f**, and in the presence of G-quadruplex it was detected for **5g** with up to 2.9 and 2.5 times increase, respectively. While all three compounds, showed fluorescence increase for the both DNA topologies, **5g** showed the highest difference in fluorescence emission in the presence of difference DNA-topologies as it showed a fluorescence increase of only 1.3 times increase for equal amount of dsDNA in comparison to G-quadruplex. This could be interpreted as **5g** having higher potential as a topology specific fluorescent probe among the compounds tested.

The docking studies have showed that affinity of the compounds varied between -7.0 and -8.3 kcal/mol for G-quadruplex topology, 1XAV (Table S1). For all fluorescently active compounds, **5f–h**, grooves of the G-quadruplex remained as the main target (Figure S67.d-f). Docking positions also revealed formation of hydrogen bonds by all compounds and it may be associated to change in fluorescence properties in the presence of DNA structures (Table S2). In the case of duplex DNA, higher affinities were observed in general and minor groove was determined as the only target by the docking studies (Figure S67.a-c). For all compounds, the

cases, the highest affinities for both DNA structures belonged to **5g**. Subsequently, a dramatic increase in fluorescence was detected in both dsDNA and G-quadruplex of **5g**.

3.3. DNA Cleavage Activity

The DNA cleavage activity of **5a–h** was performed by agarose gel electrophoresis. DNA cleavage was examined by relaxation of super coiled circular form into nicked circular form and linear form. When circular DNA is run to electrophoresis, relatively fast move will be monitored for the uncleaved super coiled form. If one strand cleavage is observed, the super coiled form relax to generate slower moving open circular form. If double strand cleavage is occurred, linear form produces and moves between super coiled and open circular forms. The results of DNA cleavage activity of **5a–h** are presented in Fig. 3. After the gel electrophoresis, it can clearly be seen in Fig. 3 that all tested compounds **5a–h** exhibited nuclease activity, whereas DNA+DMSO did not exhibit cleavage activity in Lane 2. The compound **5a** showed single strand cleaved DNA activity, whereas **5b**, **5f** and **5g** demonstrated double strand cleaved DNA activities in Lane 4 Lane 6 and Lane 8, respectively. Among the test compounds, especially **5c**, **5d**, **5e** and **5h** showed excellent cleaved DNA activities in Lane 5, Lane 7, Lane 9 and Lane 10, respectively. The plasmid DNA was cleaved into the unidentified small DNA fragments with **5c**, **5d** and **5h**. And also, **5e** was nearly degraded the plasmid DNA into the indistinguishable particles. As a result, the 1,4-naphthoquinone-triazole hybrids **5a–h** may be applied in medicine industries after further studies.



pBR 322 DNA + DMSO; Lane 3, pBR 322 DNA + 250 μg/mL of **5a**; Lane 4, pBR 322 DNA + 250 μg/mL of **5b**; Lane 5, pBR 322 DNA + 250 μg/mL of **5c**; Lane 6, pBR 322 DNA + 250 μg/mL of **5g**; Lane 7 pBR 322 DNA + 250μg/mL of **5e**; Lane 8, pBR 322 DNA + 250 μg/mL of **5f**; Lane 9, pBR 322 DNA + 250 μg/mL of **5d**; Lane 10, pBR 322 DNA + 250 μg/mL of **5h**

3.4 DPPH Scavenging and Chelating Activity

The antioxidant activities of 1,4-naphthoquinone-triazole hybrids **5a–h** were determined using 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical. The DPPH process is one of the most powerful procedures for measuring the quantity of radical scavenging compounds active by chain-breaking operations [57]. DPPH scavenging activity measurement is also a fast, easy, economic and suitable method to check the antioxidant activity of the compounds. Fig. 4 is depicted the radical scavenging ability of **5a–h**, compared with Trolox and Ascorbic acids. All tested compounds exhibited well scavenging radical ability. As seen in Fig. 4, the radical scavenging activities were concentration dependent. When the concentration of **5a**, **5b**, **5c**, **5d**, **5e**, **5f**, **5g** and **5h** was increased from 25 mg/L to 50 mg/L, the scavenging activities were increased from 27.1% to 30.1%, from 31.4% to 59.9%, from 30.6% to 36.8%, from 41.8% to 53.4%, from 29.6% to 52.9%, from 33.5% to 47.6%, from 26.2% to 39.1%, and from 32.9% to 37.6%, respectively. The DPPH scavenging activities at 100 mg/L of **5b**, **5d**, **5e**, **5f**, and **5h** were obtained as 75.6%, 83.7%, 61.1%, 59.4%, and 49.8%, respectively. The highest scavenging activity was obtained as 89.9% at 200 mg/L with **5d**. Additionally, IC50 values of **5a**, **5b**, **5c**, **5d**, **5e**, **5f**, **5g** and **5h** were determined as 223.6 mg/L, 48.9 mg/L, 89.2 mg/L, 40.4 mg/L, 47.9 mg/L, 77.9 mg/L, 125.8 mg/L and 107.2 mg/L, respectively. The antioxidant abilities of the compounds **5a–h** may be due to their redox characteristics, which give them to treat as reducing agents or H^+ donor and free radical scavengers. These results showed that the synthesized compounds may be better candidate for the future studies to develop new antioxidant agents.

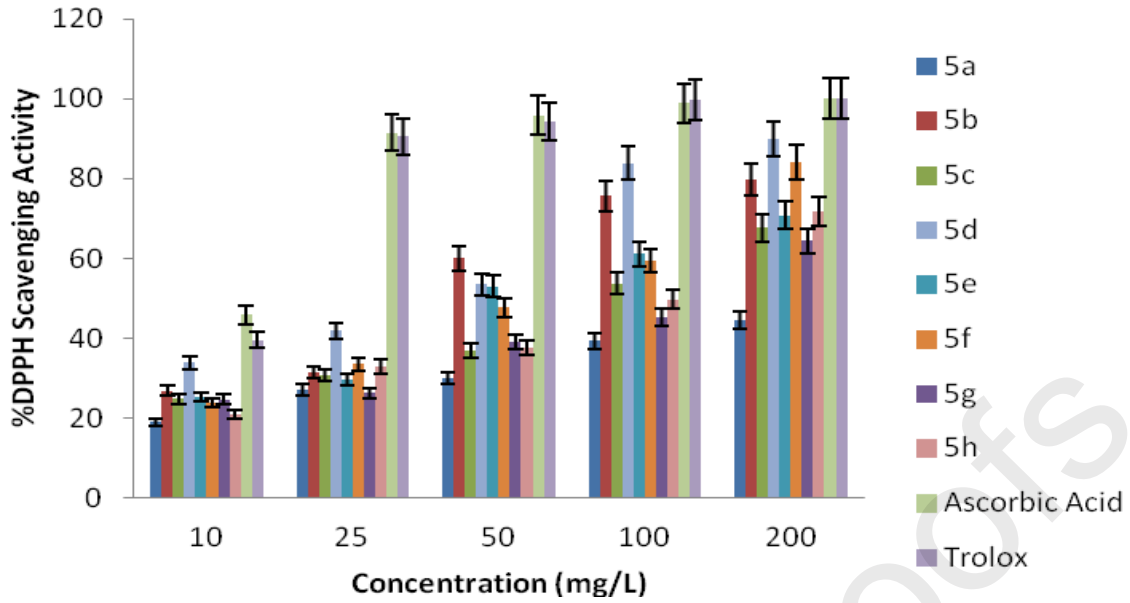


Figure 4. % Radical scavenging activity of the 1,4-naphthoquinone-triazole hybrids **5a–h**

Ferrous Chelating Activity: Most of the reactive oxygen species (ROS) are created as by-product throughout the electron transport system and other metabolic activities and ROS are also generated by metal catalyzed oxidation reactions. The transition metal Fe(II) ions have the ability to maintain the generation of free radicals by loss or gain of electrons. So, the reducing of the generation of ROS can be achieved by the chelation of toxic metal ions with chelating agents [58]. The **5a–h** have been tested for their ferrous chelating activities. The ferrous chelating assay of the compounds **5a–h** indicated that all compounds **5a–h** exhibited chelating activities and as presented in Fig. 5, chelating activities were concentration dependent. Chelating activity of **5a–h** was compared with the standard (EDTA) material. The ferrous chelating activities at 25 mg/L were obtained as 26.1%, 18.8%, 24.3%, 24.7%, 22.6%, 30.1%, 26.4%, and 21.1% for **5a**, **5b**, **5c**, **5d**, **5e**, **5f**, **5g** and **5h**, respectively. The ferrous chelating method displayed that the compounds **5a–h** also exhibited chelating ability in the order of **5g** > **5f** > **5a** > **5c** > **5e** > **5b** > **5d** > **5h** at concentration of 100 mg/L. The highest ferrous chelating activity was obtained as 68.1% with **5g** at concentration of 200 mg/L. Moreover, the IC50 values of **5a**, **5b**, **5c**, **5d**, **5e**, **5f**, **5g** and **5h** were found as 153.2 mg/L, 163.7 mg/L, 169.6 mg/L,

agents are used for neutralizing iron overload in the body in chelation therapy for treatment of several diseases such as Alzheimer, Parkinson, Thalassemia [59,60]. According to present findings, especially **5e**, **5f** and **5g** can be used in chelation therapy after further investigations.

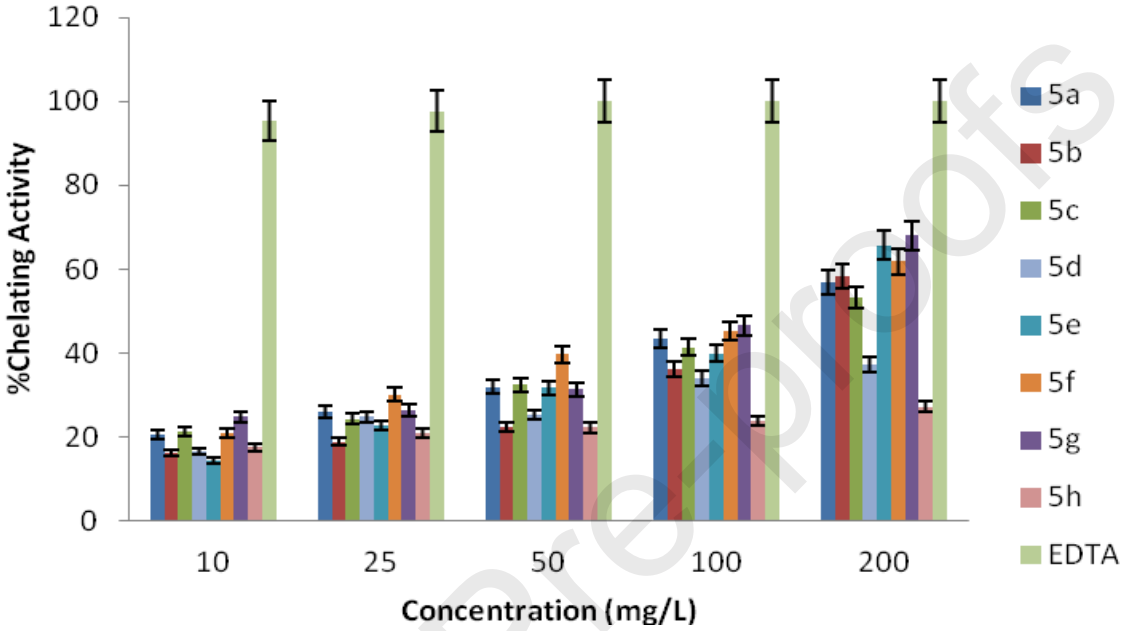


Figure 5. Chelating activity of the 1,4-naphthoquinone-triazole hybrids **5a–h**

3.5. Antimicrobial activities

The increase of antibiotic tolerance in microorganisms especially bacteria has become a significant attention for successful treatment and diagnosis of infectious illnesses. Over the past few decades, major investigations have been realized on the synthesis and development of new medicines for combating multi-drug tolerance in microorganisms [61]. The results of the antimicrobial activity of **5a–h** and standard antibiotics are demonstrated in Table 1. All compounds tested **5a–h** exhibited antimicrobial activity against all tested microorganisms with MICs ranging between 4 and 128 µg/mL. Only one substituent has been changed in the molecular structure of the 1,4-naphthoquinone-triazole hybrids **5a–h**. Compound **5b**, which contains the cyclopropyl group as a substituent, exhibited the best activity among the others

Compared to ampicillin (MIC value = 0.5 μg/mL and 1 μg/mL, respectively), all compounds showed moderate antibacterial activity against *E. coli* strain but **5b** showed noteworthy antibacterial activity against *S. aureus* strains. The **5d**, which contains the cyclohexylmethyl group as a substituent, and **5b** exhibited the best activity among the others against *B. cereus* and *P. aeruginosa* strains with a MIC value of 8 μg/mL. Compared to ampicillin (MIC value = 1 μg/mL, 0.5 μg/mL, respectively), **5b** and **5d** exhibited remarkable antibacterial activity against *B. cereus* strain. All compounds exhibited antibacterial activity against *L. pneumophila subsp. Pneumophiia* strain with a MIC value in the range of 16-64 μg/mL. The **5e**, which contains the 2-bromoethyl as a substituent, and **5d** exhibited better antibacterial activity against *E. hirae* strain with a MIC value of 4 μg/mL and *E. hirae* was the most sensitive microorganism among the test compounds. Compound **5d** exhibited the best antifungal activity among the others against *C. albicans* and *C. tropicalis* strains with a MIC value of 32 μg/mL and 16 μg/mL, respectively. Compared to Fluconazole (MIC value = 0.5 μg/mL), **5a–h** exhibited moderate antifungal activity against the tested fungus strains. As a result of antimicrobial activity study, it was observed that especially **5d** could be an important antimicrobial agent after some modifications to its molecular structure.

Table 1. The MIC values (μg/mL) of **5a–h** against the microbial strains (MIC: The minimal inhibitory concentrations)

Compound	*E. coli*	*B. cereus*	*S. aureus*	*P. aeruginosa*	*E. hirae*	*L. pneumophila subsp. pneumophiia*	*C. albicans*	*C. tropicalis*
5a	128	32	64	32	32	32	64	32
5b	16	8	8	8	16	16	128	64
5c	64	16	16	16	16	16	128	64
5d	32	8	16	8	4	16	32	16
5e	32	16	16	16	4	16	64	32
5f	128	32	32	32	32	16	128	64
5g	128	64	32	32	16	32	64	32
5h	128	16	16	16	16	64	128	64
Ampicillin	0.5	1	1	0.5	0.5	0.5	-	-
Fluconazole	-	-	-	-	-	-	0.5	0.5

3.6. Acid Dissociation Constants

In order to continue drug research studies on a molecule, pK_a value(s) which is one of the most important physicochemical parameters must be determined because it provides critical data related to acidity, degree of ionization, solubility and hydrogen bonding capacity of the compounds [62,63]. The pK_a values of **5a–h** were determined potentiometrically at 25.0±0.1 °C, 0.1 mol / L ionic strength of NaCl in 20% (v/v) DMSO-water hydro-organic solvent system using a literature method [54,55]. The calculated pK_a values are given in Table 2, and titration curves of the ligands **5a–h** and distribution curve of **5a** for symbolized all ligands **5a–h** are presented in Fig. 6. All obtained logβ values and distribution curves were submitted as supporting data (Fig. S68).

Table 2. pK_a values of **5a–h** (20% (*v*/*v*) DMSO-water, 25.0 ±0.1 ºC, *I*= 0.1 mol/L by NaCl)

Ligand	pK_{a1}	pK_{a2}	pK_{a3}	pK_{a4}	pK_{a5}
5a	2.85±0.01	6.25±0.02	10.43±0.02	10.86±0.01	11.39±0.03
5b	3.06±0.03	5.25±0.02	10.81±0.03	10.99±0.05	11.36±0.07
5c	2.13±0.02	6.66±0.02	10.47±0.02	11.05±0.03	11.46±0.06
5d	2.85±0.01	6.25±0.02	10.43±0.03	10.89±0.03	11.36±0.06
5e	2.60±0.03	6.69±0.03	9.83±0.05	10.38±0.05	11.21±0.08
5f	2.93±0.02	7.07±0.03	10.29±0.03	10.67±0.05	11.17±0.08
5g	3.07±0.01	5.72±0.01	10.39±0.03	10.92±0.02	11.31±0.06
5h	2.71±0.01	7.27±0.02	10.04±0.02	10.68±0.03	11.29±0.047

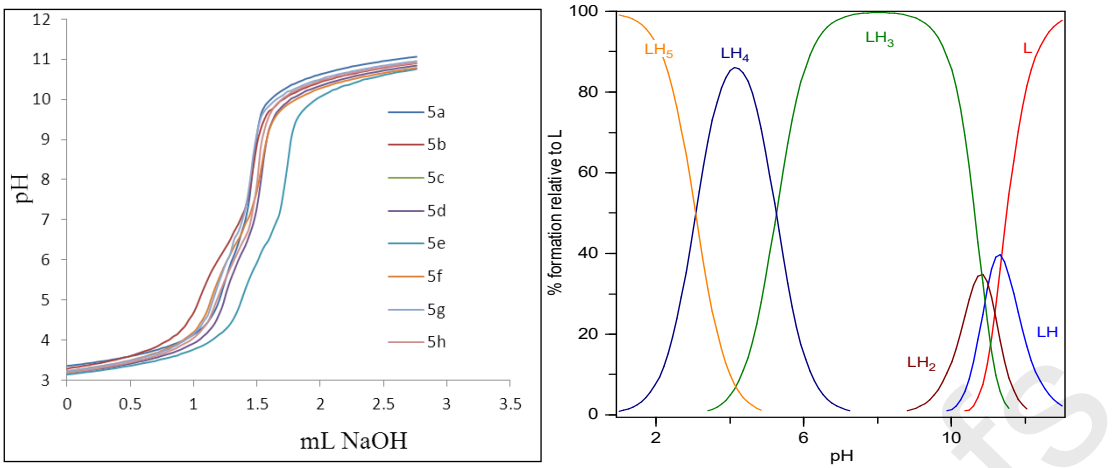


Figure 6. Titration curves of **5a–h** and distribution curve of **5a** for symbolized all ligands. (25.0±0.1 °C, 0.1 mol/L ionic strength of NaCl in 20% (v/v) DMSO-water)

According to the results, five protonated species were obtained as LH, LH_2, LH_3, LH_4 and LH_5 calculated five pK_a values. Full protonated form of the ligands is showed in Fig. 7 and four protonated species are related nitrogen atoms and a protonated species is related oxygen in the ligands.

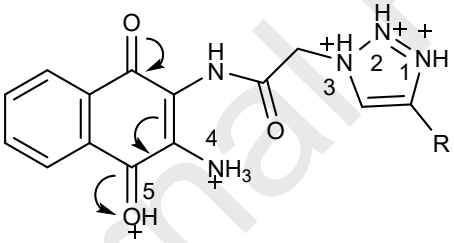


Figure 7. Full protonated form of the ligands **5a–h** (LH_5^{5+})

In our previous study [17], pyrrolidine linked to 1,2,3-triazole derivatives were examined and dissociation properties of 1,2,3-triazole moieties were explained clearly. The 1,2,3-triazole includes diazinyl group (-N=N-) and tertiary imine nitrogen. While one of two nitrogen atoms in diazinyl groups is very acidic, the other is very basic. Thus, calculated pK_{a1} values (between 2.85-3.07) in present study are related to acidic diazinyl nitrogen. Additionally, other diazinyl nitrogen is very basic and it is related to pK_{a3} values (between 10.61 – 10.97). The third nitrogen

atom in 1,2,3-triazole ring is the most basic atoms due to electronic effects and related pK_{a5} values varied between 11.01 – 11.54. In present study, proton affinity of nitrogen atoms in the ligands were calculated theoretically using semi-empirical methods such as modified neglect of diatomic overlap (MNDO). The formation heats (H*f*) and the total energies (TE) of the ligands and mono-protonated species were calculated. In addition, the proton affinity of each nitrogen atom (PA) in the ligands was found using formation heats in the following equation and given in Table 3.

$$PA = 367.2 + \Delta H_f(B) - \Delta H_f(BH^+)$$

where; PA is the proton affinity of B types; ΔH_f(B) is the formation heat of B molecule; ΔH_f(BH^+) is the formation heat of BH^+ molecule, and 367.2 is the formation heat of H^+ [64].

Table 3. The calculated formation heat (H_f), total energy (*TE*) and proton affinity (*PA*) values with MNDO methods for **5a** for symbolized all of the ligands and their mono-protonated forms.

	MNDO		
Species	T.E. (kcal/mol)	Hf (kcal/mol)	PA
5a	-94591.29	-6.50	-
1 N-H^+	-94717.18	194.28	166.42
2 N-H_2^+	-94756.19	155.27	205.45
3 N-H_2^+	-94769.26	142.21	218.49
4 N-H_2^+	-94747.62	163.84	196.86

According to Table 3, the proton affinity of nitrogen in position 3 was higher than the other nitrogen atoms and proton affinity of the nitrogen in position 1 was lower than the others. The nitrogen atom in position 3 was the most basic atom and the most acidic atom was the nitrogen in position 1. Therefore, the first protonated atom was the nitrogen atom in position 3 which

present in the molecular structures of the ligands. According to the calculation results, the protonation order for nitrogen atoms in the ligand was as 3N, 2N, 4N and 1N. Quinoline molecules behave as an aromatic cycle due to delocalization of π electrons. Thus, other two pK_a values (pK_{a2} and pK_{a4}) related to phenolic oxygen and aniline nitrogen atoms. Therefore, attention was paid in this study to explain the pK_a values of 1,4-naphthoquinone-triazole hybrid ligands because of their pharmacological importance.

4. Conclusion

The synthesis of the novel 1,4-naphthoquinone-triazole hybrid compounds **5a–h** containing two very valuable pharmacophore groups was demonstrated and their various biological properties including DNA binding / cleavage, antioxidant and antimicrobial activities were evaluated in present study. The **5a–h** exhibited high DPPH scavenging and ferrous chelating activities. A form of DNA interaction was apparent due to fluorescence increase for all compounds in the presence of the duplex DNA. While the level of increase was limited up to 2.9 times, this was measured in presence of as low as 1:10 equivalent DNA. Moreover, in spite of the toplogy of the DNA used, dsDNA or G-quadruplex, the fluorescence was enhanced for all tested compounds, **5f**, **5g**, **5h**. This result indicating the potential of these compounds as fluorescent probes; however, fluorescence enhancement may be overshadowed by the DNA cleavage activity. Especially **5c**, **5d**, **5e** and **5h** showed high DNA cleavage activity. All tested compounds displayed antibacterial and antifungal activities. Among all the compounds, **5d** demonstrated the most efficient antimicrobial activity. Present findings on biological activities of the compounds should pass through further pharmacological and toxicological tests and probably will conveniently be used in further antioxidant, antimicrobial and anticancer medicine development. Further new drug researches are recommended to be performed on such

1,4-naphthoquinone-triazole hybrids. The pK_a values containing critical information about the compounds will provide significant contributions to further pharmacological studies on these types of compounds.

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Conflict of Interest: The authors declare no conflict of interest.

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CAPTIONS

Scheme Captions

Scheme 1. Synthesis of the naphthoquinone–triazole hybrids **5a–h**

Figure Captions

Figure 1. Some pharmaceuticals based on 1,2,3-triazoles

Figure 2. Figure 2. Some pharmaceuticals based on naphthoquinones

Figure 3. DNA cleavage activities of the compounds **5a–h**. Lane 1, pBR 322 DNA; Lane 2, pBR 322 DNA + DMSO; Lane 3, pBR 322 DNA + 250 µg/mL of **5a**; Lane 4, pBR 322 DNA + 250 µg/mL of **5b**; Lane 5, pBR 322 DNA + 250 µg/mL of **5c**; Lane 6, pBR 322 DNA + 250 µg/mL of **5g**; Lane 7 pBR 322 DNA + 250µg/mL of **5e**; Lane 8, pBR 322 DNA + 250 µg/mL of **5f**; Lane 9, pBR 322 DNA + 250 µg/mL of **5d**; Lane 10, pBR 322 DNA + 250 µg/mL of **5h**

Figure 4. % Radical scavenging activity of the 1,4-naphthoquinone-triazole hybrid **5a–h**

Figure 5. Chelating activity of the 1,4-naphthoquinone-triazole hybrids **5a–h**

Figure 6. Titration curves of **5a–h** and distribution curve of **5a** for symbolized all ligands. (25.0±0.1 °C, 0.1 mol/L ionic strength of NaCl in 20% (v/v) DMSO-water)

Figure 7. Full protonated form of the ligands **5a–h** (LH_5^{5+})

Table Captions

Table 1. The MIC values (μg/mL) of **5a–h** against the microbial strains (MIC: The minimal inhibitory concentrations)

Table 2. pK_a values of **5a–h** (20% (*v*/*v*) DMSO-water, 25.0 ±0.1 ºC, *I*= 0.1 mol/L by NaCl)

Table 3. The calculated formation heat (H_f), total energy (*TE*) and proton affinity (*PA*) values with MNDO methods for **5a** for symbolized all of the ligands and their mono-protonated forms.

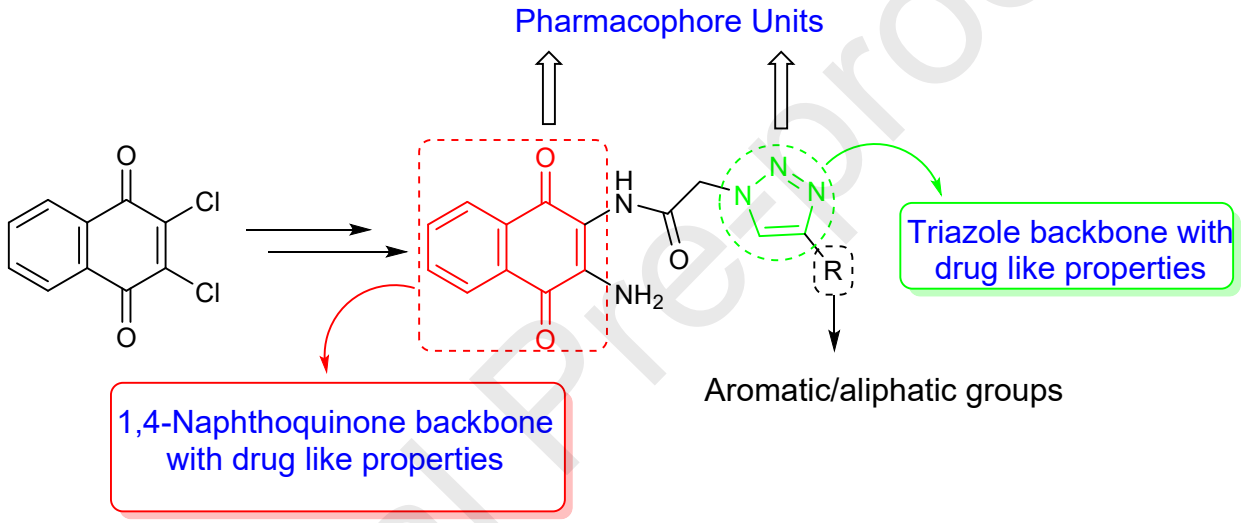
Highlights

- Synthesis of novel naphthoquinone–triazole hybrids was reported.
- The compounds were shown to have fluorescence increase upon DNA.
- The fluorescence enhancement was shown to be indifferent towards G-quadruplexes or double stranded DNA.
- *In vitro* DNA cleavage properties were observed.
- Compounds were found to show antioxidant activity.
- Compounds were screened for antibacterial and antifungal activities.
- Acid dissociation constants were determined

Graphical Abstract

Synthesis, biological properties, and acid dissociation constant of novel naphthoquinone–triazole hybrids

Yahya Nural[1,2*], Sadin Ozdemir[3,4], Osman Doluca[5], Bunyamin Demir[2,6], Mustafa Serkan Yalcin[4,5], Hasan Atabey[7], Beyza Kanat[4], Selma Erat[2,8], Hayati Sari[9], Zeynel Seferoglu[10**]



Screening a wide range of pharmacological properties

- DNA binding property	→ All of the compounds
- DNA cleavage property	→ All of the compounds
- Antioxidant property	→ All of the compounds
- Chelating activity	→ All of the compounds
- Antibacterial activity	→ MIC values of 4-128 µg/mL
- Antifungal activity	→ MIC values of 16-128 µg/mL
- Acid dissociation constant	→ Ok

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Conflict of Interest: The authors declare no conflict of interest.