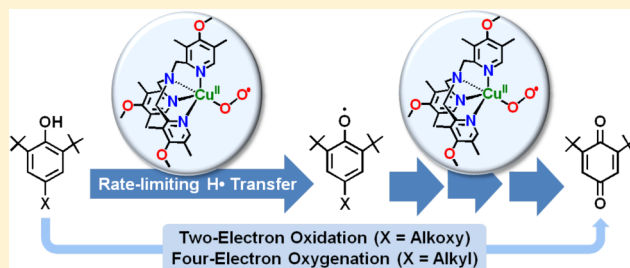


Mechanistic Insights into the Oxidation of Substituted Phenols via Hydrogen Atom Abstraction by a Cupric–Superoxo Complex

Jung Yoon Lee,[†] Ryan L. Peterson,[†] Kei Ohkubo,[‡] Isaac Garcia-Bosch,[†] Richard A. Himes,[†] Julia Woertink,[§] Cathy D. Moore,[†] Edward I. Solomon,^{*,§} Shunichi Fukuzumi,^{*,‡} and Kenneth D. Karlin^{*,†}[†]Department of Chemistry, Johns Hopkins University, Baltimore, Maryland 21218, United States[‡]Department of Material and Life Science, Graduate School of Engineering, and ALCA, Japan Science and Technology Agency (JST), Osaka University, Suita, Osaka 565-0871, Japan[§]Department of Chemistry, Stanford University, Stanford, California 94305, United States

S Supporting Information

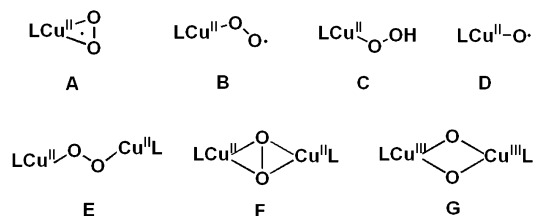
ABSTRACT: To obtain mechanistic insights into the inherent reactivity patterns for copper(I)–O₂ adducts, a new cupric–superoxo complex [(DMM-tmpa)Cu^{II}(O₂^{•−})]⁺ (**2**) [DMM-tmpa = tris((4-methoxy-3,5-dimethylpyridin-2-yl)methyl)-amine] has been synthesized and studied in phenol oxidation–oxygenation reactions. Compound **2** is characterized by UV–vis, resonance Raman, and EPR spectroscopies. Its reactions with a series of *para*-substituted 2,6-di-*tert*-butylphenols (*p*-X-DTBPs) afford 2,6-di-*tert*-butyl-1,4-benzoquinone (DTBQ) in up to 50% yields. Significant deuterium kinetic isotope effects and a positive correlation of second-order rate constants (*k*₂) compared to rate constants for *p*-X-DTBPs plus cumylperoxyl radical reactions indicate a mechanism that involves rate-limiting hydrogen atom transfer (HAT). A weak correlation of (*k*_B*T*/*e*) ln *k*₂ versus *E*_{ox} of *p*-X-DTBP indicates that the HAT reactions proceed via a partial transfer of charge rather than a complete transfer of charge in the electron transfer/proton transfer pathway. Product analyses, ¹⁸O-labeling experiments, and separate reactivity employing the 2,4,6-tri-*tert*-butylphenoxyl radical provide further mechanistic insights. After initial HAT, a second molar equiv of **2** couples to the phenoxyl radical initially formed, giving a Cu^{II}–OO–(ArO′) intermediate, which proceeds in the case of *p*-OR-DTBP substrates via a two-electron oxidation reaction involving hydrolysis steps which liberate H₂O₂ and the corresponding alcohol. By contrast, four-electron oxygenation (O–O cleavage) mainly occurs for *p*-R-DTBP which gives ¹⁸O-labeled DTBQ and elimination of the R group.



INTRODUCTION

Investigation of the structure and reactivity of various Cu^I/O₂ adducts (Chart 1), those derived from the reaction of reduced

Chart 1



ligand–copper(I) complexes with molecular oxygen (dioxygen), occurs in large part due to their critical roles in metalloenzymes.¹ Monocopper complexes A–C or dinuclear species E–G, or their analogues, have been discovered, and insights into their electronic structural characteristics and 3-D structures have been obtained.¹ Formally high-valent species D

has been detected in the gas phase,² hinted at in coordination chemistry studies³ or discussed computationally.⁴ Among them, the mononuclear species such as cupric–superoxide (A and B), cupric hydroperoxide (C), and copper oxyl (D) species all have been considered as highly reactive intermediates which likely participate in overall reaction sequences occurring in copper enzyme biotransformations.⁵

They may hydroxylate the substrate C–H bond in copper enzymes such as peptidylglycine- α -hydroxylating monooxygenase (PHM) and dopamine- β -monooxygenase (D β M)⁶ or initiate O–H oxidation in copper oxidases such as galactose oxidase (GO)^{6a,7} or copper amine oxidase (CAO).^{6a,8} From relatively recent experimental and computational studies, the [Cu^{II}(O₂^{•−})]⁺ (A or B) moiety has been declared as the reactive intermediate which effects C–H^{9,10} or O–H^{7,8} hydrogen atom transfer (HAT) reactions. Relevant to this is an X-ray crystallographic study on a PHM derivative, which

Received: August 27, 2013

reveals a dioxygen-derived species assigned as an end-on bound cupric–superoxo species (**B**) (also discussion below).¹¹ As is relevant to part of the GO catalytic cycle, recent experimental studies support a mechanism wherein O₂ binding to this fully reduced copper ion affords a cupric–superoxo species (Figure 1), and that this mediates the oxidation of the ligated tyrosine

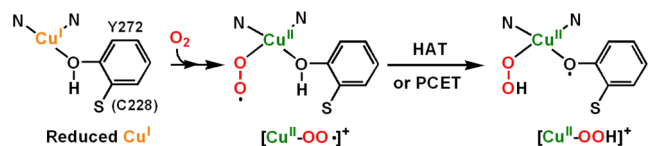


Figure 1. Reduction of molecular oxygen in galactose oxidase (GO). See Supporting Information (Figure S1a) for fuller details.

residue resulting in the formation of a cupric hydroperoxide species (**C**) plus phenoxyl radical.^{7c} Hydrolysis leads to the release of hydrogen peroxide, the observed stoichiometric O₂ reduction product in the overall mechanism.

With this background, one of the research goals that we have recently been emphasizing is on the chemistry of cupric–superoxo complexes. It is critical to elucidate chemical/physical properties, spectroscopic characteristics, and reactivity toward substrates possessing C–H or O–H bonds. An understanding of such aspects is critical in order to fully elucidate fundamentals involved in oxidative processes and the reduction of dioxygen by copper centers in chemical systems or at the active sites of metalloenzymes.

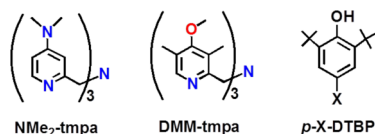
In this report, we describe the chemistry of a new [Cu^{II}(O₂^{•−})]⁺ complex and a detailed investigation into its reactions with phenolic substrates. As discussed above, the net hydrogen atom abstraction reaction from a phenol derivative is directly relevant to the enzyme chemistry in GO. Similar reactivity occurs in CAO, where a cupric–superoxo species is thought to undergo a proton-coupled electron transfer (PCET) process converting TPQ_{SQ} (2,4,5-trihydroxyphenylalanine semiquinone) to TPQ_{IMQ}, the one-electron oxidized iminoquinone form (Supporting Information Figure S1b).⁸ As mentioned, phenols can undergo oxidation by two pathways, both resulting in formation of the neutral phenoxyl radical and transfer of a hydrogen atom to the oxidant (eq 1). Mechanistically, the process can occur by PCET¹² or hydrogen atom transfer, and these are fundamentally important reactions occurring in many biological systems.



Osako et al.¹³ previously investigated phenol (and C–H) oxidation chemistry using binuclear complexes of the type (μ-η²:η²-peroxo)dicopper(II) (**F**) and/or bis(μ-oxo)dicopper(III) (**G**). For a series of phenols, kinetic/mechanistic studies demonstrated that these oxidations proceeded via PCET rather than HAT. However, studies on the reactivity of mononuclear Cu^I/O₂ species are scarce, and in fact, no detailed mechanistic investigations have been described. Only recently have ligand design and synthetic methodologies better allowed for the formation of discrete [Cu^{II}(O₂^{•−})]⁺ adducts, making it possible for the systematic investigation into their inherent chemical and physical properties, along with substrate reactivity.¹⁴

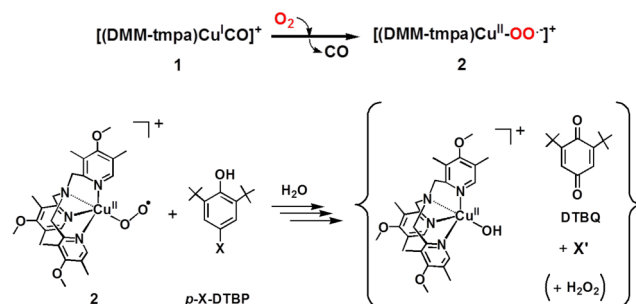
Furthermore, the investigation into exogenous phenolic substrate O–H oxidation by [Cu^{II}(O₂^{•−})]⁺ complexes has been limited to the report of product yields for a few phenol or catechol substrates. Thus, utilizing the tripodal tetradentate ligand NMe₂-tmpa (Chart 2), which forms a low-temperature

Chart 2



stable end-on bound superoxo–copper(II) complex, we found that 2,6-di-*tert*-butylphenol derivatives (*p*-X-DTBP; X = OMe, ^tBu, H) were oxidized to 2,6-di-*tert*-butyl-1,4-benzoquinone (DTBQ) (Scheme 1).

Scheme 1



We herein describe a detailed study employing a new electron-rich ligand supporting the cupric–superoxo complex, [(DMM-tmpa)Cu^{II}(O₂^{•−})]⁺ (**2**) (Scheme 1), which is capable of mediating this chemistry for a large series of *p*-X-DTBP (X = alkoxy/alkyl) substrates. We provide decisive kinetic evidence supporting cupric–superoxo complex-mediated HAT as the rate-limiting step in phenol O–H bond activation and two- and/or four-electron reduction of molecular oxygen (vide infra). There are three possible reaction pathways in the overall hydrogen atom transfer from *p*-X-DTBP to [Cu^{II}(O₂^{•−})]⁺: (1) electron transfer (ET) followed by proton transfer (PT); (2) PT followed by ET; and (3) concerted ET and PT. The former two pathways correspond to PCET processes, whereas the third one (i.e., concerted ET and PT) corresponds to a direct HAT process. The mechanistic conclusions are drawn by several corroborating studies employing physical measurements leading to (i) kinetic isotope effect (KIE) determination, (ii) correlation of the relative reactivity for hydrogen atom abstraction using cumylperoxyl radical (**4**) as a mechanistic benchmark, (iii) comparison of the rate dependences in the phenol oxidations by **2** and **4** on the one-electron oxidation potentials of the *p*-X-DTBPs, and (iv) activation parameters determined from the reaction kinetics. We also discuss the experimental results from product analysis showing that either overall two-electron oxidation or four-electron oxygenation of the *p*-X-DTBPs can occur; the results depend on the identity of the substituent X. Detailed pathways leading to the products observed are proposed.

EXPERIMENTAL SECTION

General Considerations. All materials used were of commercially available reagent quality unless otherwise stated. Acetone was distilled from Drierite under argon atmosphere. Tetrahydrofuran (THF) and 2-methyltetrahydrofuran (MeTHF) inhibitor-free were purchased from Sigma-Aldrich and distilled under argon from sodium/benzophenone prior to use. Pentane was distilled under argon over CaH₂. Acetonitrile was purified via passage through a double alumina column solvent purification system from Innovative Technologies, Inc. Di-*tert*-butyl

peroxide was purchased from Nacalai Tesque Co., Ltd. and purified by chromatography through alumina which removes traces of the hydroperoxide. Cumene was purchased from Tokyo Chemical Industry Co., Ltd. Air-sensitive compounds were synthesized and transferred under an argon atmosphere using standard Schlenk techniques and stored in an MBraun glovebox filled with N₂. [Cu^I(CH₃CN)₄]B(C₆F₅)₄ was synthesized as previously reported.¹⁵ 2,4,6-Tri-*tert*-butylphenoxyl radical ([•]Bu₃ArO[•]) was synthesized following a procedure reported in the literature¹⁶ and characterized with UV–vis absorption band at 384 and 402 nm and sharp EPR signal at *g* = 2.00. Elemental analyses were performed by Desert Analytics, Tucson, AZ. The ¹H NMR spectrum was measured on a Bruker 300 MHz or a Bruker 400 MHz spectrometer. ²H NMR was recorded on the broad-band coil on a 300 MHz instrument with ²H resonance at 46 MHz. Chemical shifts are reported in parts per million downfield against TMS or residual solvent signals unless otherwise specified. Benchtop low-temperature UV–vis experiments were carried out on a Cary bio-50 spectrophotometer equipped with a Unisoku USP-203A cryostat using a 1 cm modified Schlenk cuvette. EPR measurements were performed on a Bruker X-band EPR 5 mm quartz EPR tubes (Willmad). Electrospray ionization (ESI) mass spectra were acquired using a Finnigan LCQDeca ion-trap mass spectrometer equipped with an electrospray ionization source (Thermo Finnigan, San Jose, CA). Sulfur dioxide (SO₂, gas) was prepared by mixing sodium metabisulfite-saturated water and diluted sulfuric acid solution (Figure S2). 2,4,6-Tri-*tert*-butyl-4-hydroperoxycyclohexa-2,5-dienone was prepared as in the literature¹⁷ and characterized by ¹H NMR spectroscopy.

Ligand Synthesis. DMM-*tmpa* [tris((4-methoxy-3,5-dimethylpyridin-2-yl)methyl)amine] ligand utilized in this report was synthesized following a procedure described in the literature.¹⁸

2-Phthalimidomethyl-4-methoxy-3,5-dimethylpyridine: TLC on alumina (1:1 = EtOAc/hexane), *R_f* = 0.63; ¹H NMR (CDCl₃) δ 8.05 (s, 1H), 7.89–7.86 (dd, 2H, *J* = 4 Hz), 7.72–7.70 (dd, 2H, *J* = 4 Hz), 4.93 (s, 2H), 3.75 (s, 3 H), 2.31 (s, 3H), 2.17 (s, 3H); FAB-MS calcd MH⁺ = 297.12392, found 297.12352.

(4-Methoxy-3,5-dimethylpyridin-2-yl)methanamine: TLC on alumina (95:5 = DCM/MeOH), *R_f* = 0; ¹H NMR (CDCl₃) δ 8.16 (s, 1 H), 3.87 (s, 2H), 3.71 (s, 3H), 2.20 (s, 3H), 2.16 (s, 3H), 1.98 (br s, 2H).

DMM-*tmpa*: TLC on alumina (95:5 = DCM/MeOH), *R_f* = 0.29; ¹H NMR CDCl₃ δ 8.18 (s, 3H), 3.74 (s, 6H), 3.65 (s, 9H), 2.22 (s, 9H), 1.61 (s, 9H); ¹H NMR (acetone-*d*₆) δ 8.16 (s, 3H), 3.68 (s, 9H), 3.67 (s, 6H), 2.21 (s, 9H), 1.62 (s, 9H); FAB-MS calcd MH⁺ = 465.28657, found 465.28582.

4-Bromo-2,6-di-*tert*-butyl-2,5-cyclohexadienone: The preparation of 4-bromo-2,6-di-*tert*-butyl-2,5-cyclohexadienone was accomplished using a modified published procedure:¹⁹ ¹H NMR (CDCl₃) δ 6.77–6.75 (d, 2H), 5.38–5.36 (t, 1H), 1.24 (s, 9H); ¹³C NMR (CDCl₃) δ 185.05, 143.74, 136.60, 77.31.

Synthesis of *para*-Alkoxy-2,6-di-*tert*-butylphenols (*p*-OR-DTBPs). Selected phenols were synthesized from adapted published procedures.²⁰ A 22 mL vial was charged with 810.7 mg of silver triflate and a magnetic stir bar. To this vial was added 10 mL of the corresponding alcohol (RO-H). If needed, a minimal amount of dimethylethanol (DME) was used to dissolve the silver salt. To this solution was rapidly added 900 mg of 4-bromo-2,6-di-*tert*-butyl-2,5-cyclohexadienone dissolved in 2.5 mL of DME, resulting in the immediate formation of a precipitate and the solution to turn yellow. After 3 min, the solution was poured into a 50 mL solution containing ~1 g of NaSH. The organics were separated by extraction four times with 50 mL portions of pentane. This pentane solution was dried over sodium sulfate and filtered using a 0.45 μm filter. The pentane was removed by vacuum, and the phenol was purified by flash chromatography using silica gel eluting with 0–2.5% gradient of EtOAc/petroleum ether. The phenols were isolated as pale yellow to white solids, and the purity was checked by NMR and GC–MS. If necessary, phenols were crystallized from petroleum ether. Phenol purity for kinetic analysis was >95%. The yields of phenols varied according to substituent but were typically 25–70%.

***p*-OCH₂CH₃-DTBP:** ¹H NMR (CDCl₃) δ 6.758 (s, 2H), 4.757 (s, 1H), 4.02–3.95 (q, 2H, *J* = 6.9 Hz), 1.43 (s, 18H), 1.419–1.372 (t, 3H, *J* = 6.9 Hz); ¹³C NMR (CDCl₃) δ 151.83, 147.66, 137.17, 111.262, 63.80, 34.576, 30.22, 15.10; FAB-MS calcd M⁺ = 250.19328, found 250.19312.

***p*-OCD₂CD₃-DTBP:** ¹H NMR (CDCl₃) δ 6.758 (s, 2H), 4.757 (s, 1H), 4.02–3.95 (q, 2H, *J* = 6.9 Hz), 1.43 (s, 18H), 1.419–1.372 (t, 3H, *J* = 6.9 Hz); ¹³C NMR (CDCl₃) δ 151.83, 147.66, 137.17, 111.262, 63.80, 34.576, 30.22, 15.10; FAB-MS calcd M⁺ = 255.22466, found 255.22453.

***p*-OCD₃-DTBP:** ¹H NMR (CDCl₃) δ 6.766 (s, 2H), 4.773 (s, 1H), 1.43 (s, 18H); ²H NMR (CDCl₃) δ 3.759 (s); ¹³C NMR (CDCl₃) δ 152.41, 147.68, 110.53, 34.57, 30.18; FAB-MS calcd M⁺ = 239.19696, found 239.19629.

***p*-OCH₂CF₃-DTBP:** ¹H NMR (CDCl₃) δ 6.86 (s, 2H), 4.975 (s, 1H), 4.40–4.316 (q, 2H, *J* = 8.4 Hz), 1.49 (s, 18H); ¹³C NMR (CDCl₃) δ 150.64, 149.17, 137.49, 129.14–118.082 (q, *J* = 278 Hz) 112.19, 67.71–66.316 (q, *J* = 35.3 Hz), 34.59, 30.121; ¹⁹F NMR δ 74.61–74.67 (t, *J* = 8.5 Hz); FAB-MS calcd M⁺ = 304.16501, found 304.16607.

***p*-OMPP-DTBP (2-Methyl-1-phenylpropan-2-yloxy):** ¹H NMR (CDCl₃) δ 7.332–7.31 (m, 5H), 6.778 (s, 2H), 4.931 (s, 1H), 2.989 (s, 2H), 1.431 (s, 18H), 1.235 (s, 6H); ¹³C NMR (CDCl₃) δ 149.82, 147.18, 138.50, 136.15, 130.77, 127.83, 126.19, 120.88, 79.83, 48.85, 34.27, 30.28, 30.25, 26.23; FAB-MS calcd M⁺ = 354.25588, found 354.25570.

Preparation of ²H-O-2,6-Di-*tert*-butyl-4-methoxyphenol. A Schlenk flask containing 1.25 g of *p*-OMe-DTBP and a stir bar was dissolved in 20 mL of freshly distilled THF. The sample was then chilled to –78 °C and 1.1 equiv of *n*-butyllithium dissolved in pentane was slowly added to the solution. The solution was allowed to react at low temperature for 30 min, after which 1.4 mL of D₂O was added to the solution in ~150 μL portions. The solution was slowly allowed to warm to room temperature and the solvent removed in vacuo, yielding a white precipitate. The desired phenol was extracted from the flask by the addition of ~20 mL of freshly distilled pentane, and the resulting pentane solution was filtered through a plug of Celite. The pentane was removed in vacuo, yielding the desired product as a white solid. The sample was stored in the glovebox, and the ²H content was assessed by the absence of the RO–H proton resonance at ~4.8 ppm. ²H content was determined to be >98%.

Redox Potentials of *para*-Substituted 2,6-Di-*tert*-butylphenols (*p*-X-DTBPs). The redox potentials were measured by second harmonic AC voltammetry (SHACV) in CH₃CN containing 0.10 M Bu₄N⁺PF₆[–] as a supporting electrolyte using an ALS-630B electrochemistry analyzer with a three-electrode setup consisting of a platinum disk working electrode, platinum wire counter electrode, and a Ag/AgNO₃ reference electrode. The voltammograms are plotted against the [Fe(Cp)₂]^{+/0} potential which was measured as an external standard. The *E*⁰ values (vs Ag/AgNO₃) were converted to those versus the [Fe(Cp)₂]^{+/0} (Fc/Fc⁺) potential which was measured as an external standard. All electrochemical measurements were carried out at 25 °C under argon atmosphere. Scans were run at 4 mV s^{–1}.

Synthesis of [(DMM-*tmpa*)Cu^I(CO)]B(C₆F₅)₄ (1). This complex was synthesized in a manner very similar to that for the previously reported complex [(NMe₂-*tmpa*)Cu^I(CO)]B(C₆F₅)₄.²¹ A 100 mL Schlenk flask containing a stir bar, 100 mg of DMM-*tmpa*, and 195 mg of [Cu^I(CH₃CN)₄]B(C₆F₅)₄ was evacuated and the flask purged with argon on the vacuum line. To this reaction flask was attached a Claisen adapter fitted with two air-free addition funnels consisting of ~50 mL of THF and ~125 mL of pentane, respectively. The argon line was replaced with a carbon monoxide (CO) line, and the resulting solutions were deaerated with briskly flowing CO(g) for 20 min. Approximately, 10 mL of the CO-saturated THF solution was used to dissolve the ligand and copper salt. The yellow solution was allowed to stir for ~5 min under a positive pressure of CO. The CO-saturated pentane was added to the THF solution, resulting in the solution turning cloudy and an oil settling to the bottom of the flask. Excess solvent was decanted off under a CO(g) atmosphere and the resulting oil dried under vacuum, yielding a white solid (70% yield): ¹H NMR

(acetone- d_6) δ 8.39 (s, 3H), 4.13 (s, 6H), 3.82 (s, 9H), 2.29 (s, 9H), 2.24 (s, 9H). Cu–CO stretch 2085 cm^{-1} in CH_3CN . Anal. Calcd: C, 50.56; H, 2.94; N, 4.54. Found: C, 50.96; H, 2.95; N, 4.52). Note: The use of the CO adduct of the copper(I) complex is required in order to prevent disproportionation. The counteranion, $\text{B}(\text{C}_6\text{F}_5)_4$, is utilized for better solubility of the superoxo–copper(II) complex and products derived from its reactions at very low temperatures.

Generation of $[(\text{DMM-tmpa})\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]\text{B}(\text{C}_6\text{F}_5)_4$ (2) and Reaction with p -X-DTBP Derivatives. *Kinetic Measurements.* In the glovebox, 0.27 mM solution of $[(\text{DMM-tmpa})\text{Cu}^{\text{I}}(\text{CO})]\text{B}(\text{C}_6\text{F}_5)_4$ (1) was prepared in a 2.5 mL acetone solvent mixture (10% MeTHF), and the 1 cm Schlenk cuvette was sealed with a rubber septum. Out of the glovebox and at room temperature, the solution was immediately purged for 15 s with $\text{CO}_{(\text{g})}$ using a long syringe needle. The solution was then cooled to the appropriate temperature (-100 to -85 °C), and dioxygen was gently bubbled through the solution. Clean isosbestic conversion to 2 was obtained within 15 min as monitored by UV–vis spectroscopy.²¹ This green intermediate was stable for hours at -90 °C. Phenol oxidation reactions were initiated by the addition of a stock solution of phenol to the fully generated 2 after three Ar/vacuum purge cycles. Pseudo-first-order rate plots were performed by observing the disappearance of a 409 nm band to obtain plots of $\ln[(A - A_t)/(A_i - A_t)]$ versus time (seconds), which were found to be linear for three or more half-lives. {General notes: (i) Complex 2 is stable for an hour at -90 °C in acetone, as judged by any absorbance loss at $\lambda_{\text{max}} = 409$ nm. This “lifetime” is well beyond times needed for kinetic studies. (ii) The superoxo complex 2 can also be generated in other solvents such as THF or MeTHF, but it is less stable than in acetone. (iii) If $\text{CO}_{(\text{g})}$ is not present in excess in the initial solution of 1 (in 10% MeTHF/acetone at -90 °C), the binuclear peroxodicopper(II) species $\{[(\text{DMM-tmpa})\text{Cu}^{\text{II}}]_2(\mu-1,2-\text{O}_2^{2-})\}^{2+}$ readily forms upon addition of O_2 . (iv) Further, this dicopper species is formed when higher concentrations of 1 (>1 mM) are employed in the generation of the 2, thus confining the range of concentrations used in all of the studies described. (v) Superoxo complex 2 does slowly decay with warming to above -85 °C; however, $\{[(\text{DMM-tmpa})\text{Cu}^{\text{II}}]_2(\mu-1,2-\text{O}_2^{2-})\}^{2+}$ is not the product.}

Quantification of 2,6-Di-*tert*-butyl-1,4-benzoquinone with Gas Chromatography–Mass Spectrometry (GC–MS). A Schlenk flask was charged with 10 mL of 0.25 mM acetone solution of $[(\text{DMM-tmpa})\text{Cu}^{\text{I}}(\text{CO})]\text{B}(\text{C}_6\text{F}_5)_4$ (1) in the glovebox, and out of the glovebox, the solution was immediately purged with $\text{CO}_{(\text{g})}$ at room temperature. The solution was then cooled to the -90 °C acetone/liquid nitrogen cooling bath, and dioxygen was gently bubbled through the solution to generate $[(\text{DMM-tmpa})\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]\text{B}(\text{C}_6\text{F}_5)_4$ (2). After 2 was fully formed, three Ar/vacuum purge cycles were applied to remove excess dioxygen, and a stock solution of substrates (1–50 equiv) was added to the solution. When the reaction ended, the Schlenk flask was warmed to room temperature. (Note: we also analyzed solutions quenched at low temperature by addition of SO_2 , but the results and yields of reactions were not affected). The solvent was removed in vacuo, redissolved in 300 μL of the solvent, and transferred in a GC–MS vial with addition of 0.8 μmol of naphthalene as a standard. Then 1 μL was injected into the GC–MS. The area ratio was converted to mole ratio to quantify the yield of DTBQ by using a standard curve. All GC–MS experiments were carried out and recorded using a Hewlett-Packard 6890 series gas chromatograph system equipped with 5973N mass selective detector. The GC–MS conditions for the product analysis were as follows: injector port temperature = 250 °C; column temperature = initial temperature = 80 °C; initial time = 2 min; final temperature = 280 °C; final time = 2 min; gradient rate = 10 °C/min; flow rate = 14.2 mL/min; ionization voltage = 1.3 kV.

^{18}O -Labeling Experiments. A Schlenk cuvette was charged with 2.5 mL of 1 mM acetone solution of $[(\text{DMM-tmpa})\text{Cu}^{\text{I}}(\text{CO})]\text{B}(\text{C}_6\text{F}_5)_4$ (1) in the glovebox, and out of the glovebox, the solution was immediately purged with $\text{CO}_{(\text{g})}$ at room temperature. The solution was then cooled to the -90 °C, and $^{18}\text{O}_2$ was gently bubbled through the solution to generate $[(\text{DMM-tmpa})\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]\text{B}(\text{C}_6\text{F}_5)_4$ (2) monitored by UV–vis spectroscopy. $^{18}\text{O}_2$ (Icon 6393) was prepared in

100 mL of a Hamilton gastight syringe equipped with a three-way valve and needle outlet. After 2 was fully formed, three Ar/vacuum purge cycles were applied to remove excess dioxygen and a stock solution of substrates was added to the solution. When the reaction ended, the Schlenk cuvette was taken out to be warmed at room temperature. The solvent was removed in vacuo, redissolved in 300 μL of the solvent, and transferred in a GC–MS vial. Then 1 μL was injected into the GC–MS.

OR Product Analysis Using ^2H NMR. Reactions were performed using a 0.75 mM solution of $[(\text{DMM-tmpa})\text{Cu}^{\text{I}}(\text{CO})]\text{B}(\text{C}_6\text{F}_5)_4$ (1) in acetone. The $[(\text{DMM-tmpa})\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]\text{B}(\text{C}_6\text{F}_5)_4$ (2) was generated in a similar fashion described above at -95 °C. After oxygenation, a solution containing 5 equiv of p -OR-DTBP (OR = OCD_3 or OCD_2CD_3) and benzene- d_6 (as internal reference) was added to the NMR tube. The sample was loaded to the spectrophotometer at low temperature and the spectrum recorded. After which, the sample was removed from the instrument, allowed to be warmed to room temperature, and the spectrum of the solution recorded.

Oxidation of Phenols by the Cumylperoxyl Radical (4). Kinetic measurements were performed on a JEOL X-band spectrometer (JES-ME-LX) at 183 K. Typically, photoirradiation of an oxygen-saturated acetone solution containing di-*tert*-butyl peroxide (1.0 M) and cumene (1.0 M) with a 1000 W mercury lamp resulted in formation of cumylperoxyl radical ($g = 2.0156$) which could be detected at low temperatures. The g values were calibrated by using a Mn^{2+} marker. Upon cutting off the light, the decay of the EPR intensity was recorded with time. The decay rate was accelerated by the presence of p -X-DTBPs (1.0×10^{-2} M). Rates of hydrogen transfer from p -X-DTBPs to $\text{PhCMe}_2\text{OO}\bullet$ were monitored by measuring the decay of the EPR signal of $\text{PhCMe}_2\text{OO}\bullet$ in the presence of various concentrations of p -X-DTBPs in acetone at 183 K. Pseudo-first-order rate constants were determined by a least-squares curve fit using a personal computer. The first-order plots of $\ln(I - I_\infty)$ versus time (I and I_∞ are the EPR intensity at time t and the final intensity, respectively) were linear for three or more half-lives with the correlation coefficient, $\rho > 0.99$. In each case, it was confirmed that the rate constants derived from at least five independent measurements agreed within an experimental error of $\pm 5\%$.

Hydrogen Peroxide Quantification. Detection of H_2O_2 as a product has been performed with CH_3CN -saturated NaI solution as in a recent report.²² First, 2.5 mL of a 0.6 mM solution of $[(\text{DMM-tmpa})\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]\text{B}(\text{C}_6\text{F}_5)_4$ (2) was generated in a Schlenk cuvette in a typical way. After the reaction with p -X-DTBP was ended, the solution was taken out and warmed to room temperature. Then, 70 μL of solution was added into 2.0 mL of a CH_3CN -saturated NaI solution at room temperature in the darkness. The UV–vis spectrum of this solution displayed the formation of triiodide (I_3^-) at 362 nm, and the yield was calculated by comparing with a standard H_2O_2 solution of known concentrations. 2,4,6-Tri-*tert*-butyl-4-hydroperoxycyclohexa-2,5-dienone was also found to oxidize the iodide ion.

Attempt To Detect Formaldehyde. The detection of the formaldehyde was accomplished spectrophotometrically via an aqueous-based Nash assay.²³ The Nash reagent was prepared by dissolving 7.5 g of ammonium acetate, 100 μL of 2,4-pentadione, and 150 μL of acetic anhydride in 50 mL of water. Eight milligrams of $[(\text{DMM-tmpa})\text{Cu}^{\text{I}}(\text{CO})]\text{B}(\text{C}_6\text{F}_5)_4$ (1) was dissolved in 5 mL of acetone and transferred to a Schlenk flask, capped with a rubber septum, and bubbled with $\text{CO}_{(\text{g})}$ for ~ 30 s. This reaction vesicle was put in a -95 °C acetone bath. The formation of $[(\text{DMM-tmpa})\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]\text{B}(\text{C}_6\text{F}_5)_4$ (2) was achieved by O_2 displacement for ~ 10 min at which time 7.4 mg of *para*-methoxy-2,6-di-*tert*-butylphenol dissolved in 200 μL of acetone was added to the solution. The reaction was allowed to react at -95 °C for 45 min at which time the reaction was quenched at low temperature with $\text{SO}_{2(\text{g})}$. The solution was allowed to be warmed to room temperature, and a 250 μL aliquot of the reaction solution was added to a vial charged with 2 mL of the Nash reagent cocktail. The reaction mixture was capped, sealed, and heated to 60 °C for 90 min at which time the absorbance at 413 nm was recorded.

Attempt To Detect Isobutylene. The attempt for detecting isobutylene (gas) as one of the products was performed by using GC–MS. An isobutylene-saturated acetone solution was injected as an authentic standard, and it could be easily detected by GC–MS. However, when the reaction solution was tested, there was no evidence of isobutylene. We attribute this to the low concentrations (maximum 1 mM) used for reaction.²⁴

Resonance Raman Spectroscopy. Resonance Raman spectra were recorded on a Princeton Instruments ST-135 back-illuminated CCD detector and on a Spex 1877 CP triple monochromator with 1200, 1800, and 2400 grooves/mm holographic spectrograph gratings. Excitation was provided by a Coherent 190C-K Kr⁺ ion laser ($\lambda_{\text{ex}} = 407$ nm). The spectral resolution was <2 cm⁻¹. Spectra were recorded at 5 mW power at the sample, and the samples were cooled to 77 K in a quartz liquid nitrogen finger Dewar (Wilmad). Baseline spectra were collected using ground, activated charcoal. Isotopic substitution was achieved by oxygenating with ¹⁸O₂ (Icon, Summit, NJ).

RESULTS AND DISCUSSION

Spectroscopic Characterization of [(DMM-tpmpa)-Cu^{II}(O₂^{•-})]⁺ (2). There exist three structurally characterized cupric–superoxo complexes. One is a synthetic complex from Fujisawa–Kitajima with a side-on η^2 -bound O₂^{•-} fragment. It is a ground-state singlet species ($S = 0$) and from resonance Raman spectroscopy $\nu_{\text{O-O}} = 1043$ cm⁻¹.²⁵ There is also the already mentioned case of the PHM protein X-ray structure, considered to have the Cu^{II}(O₂^{•-}) formulation and end-on bound O₂ fragment.¹¹ The physical properties of this species within the protein are still lacking. Closely related to our own case here, 2, is the now very well-known complex from Sundermeyer and Schindler,²⁶ [(TMG₃tren)Cu^{II}(O₂^{•-})]⁺ (TMG₃tren = (1,1,1-tris[2-[N²-(1,1,3,3-tetramethylguanidino)]ethyl]amine), with analogous tripodal tetradentate ligand, and end-on superoxo binding ($\angle\text{Cu-O-O} = 123^\circ$) (X-ray structure, Figure 2) with $\nu_{\text{O-O}} = 1120$ cm⁻¹ ($\Delta^{18}\text{O}_2 = -63$



Figure 2. Structural representation of [(TMG₃tren)Cu^{II}(O₂^{•-})]⁺.²⁶

cm⁻¹). The electronic structure of this molecule has been delineated by Solomon and co-workers,²⁷ and the molecule possesses a triplet ground state with ferromagnetically coupled spins on both the Cu(II) ion and superoxo fragment. In fact, the finding of a triplet ground state for cupric–superoxo complexes possessing tripodal tetradentate ligands is general.²⁸

Based on the physical properties of [(DMM-tpmpa)-Cu^{II}(O₂^{•-})]⁺ (2), as described here, it has a physical and electronic structure very similar to that of [(TMG₃tren)-Cu^{II}(O₂^{•-})]⁺. Complex 2 was generated in acetone and/or MeTHF at -90°C via displacement of CO(g) by bubbling O₂ gas through a solution of [(DMM-tpmpa)Cu^I(CO)]⁺ (1); a further discussion of why/how this procedure is employed, with references, is given in the Experimental Section. Thus, 2 has been presently characterized by UV–vis and resonance Raman spectroscopies. The absorption spectrum of this green colored complex is shown in Figure 3a, and three primary absorption bands are observed: 409, 587, and 743 nm with $\epsilon = 4250$, 1100,

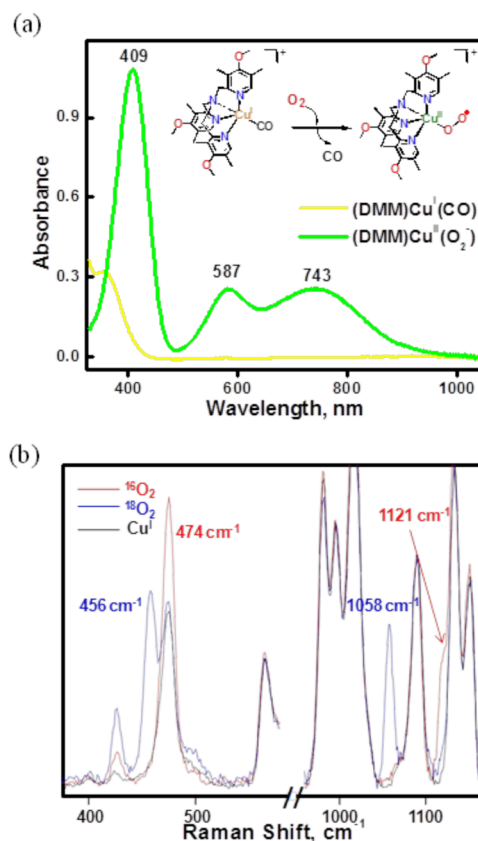


Figure 3. (a) Absorption spectra of 1 (0.27 mM) and 2 after addition of O_{2(g)} in acetone (10% MeTHF) at 183 K. (b) Resonance Raman spectra of 2 (0.7 mM) measured in MeTHF ($\lambda_{\text{ex}} = 407$ nm). Red, ¹⁶O₂; blue, ¹⁸O₂.

and 1030 M⁻¹ cm⁻¹, respectively. This complex is EPR silent (Figure S3). Complex 2 is stable enough ($t_{1/2} > \sim 3$ h; -90°C) to investigate its reactivity toward external substrates. Compared to the parent tmpa ligand (without any pyridyl substituents) Cu^{II}–superoxo complex, which can only be observed as a fleeting intermediate at -128°C in MeTHF,²⁹ the electron-donating groups on the DMM-tpmpa ligand (as also for NMe₂-tpmpa; Chart 2)²¹ provide significant electron density to the copper center, resulting in stabilization of [(ligand)-Cu^{II}(O₂^{•-})]⁺ species.

Resonance Raman spectra of [(DMM-tpmpa)Cu^{II}(O₂^{•-})]⁺ (2) ($\lambda_{\text{ex}} = 407$ nm) reveal two dioxygen isotope-sensitive vibrations (Figure 3b). An O–O vibration is observed at 1121 cm⁻¹, which shifts to 1058 cm⁻¹ ($\Delta^{18}\text{O}_2 = -63$ cm⁻¹) upon ¹⁸O₂ substitution. An additional isotope-sensitive vibration attributed to the Cu–O stretch occurs at 474 cm⁻¹ ($\Delta^{18}\text{O}_2 = -18$ cm⁻¹). These parameters closely match those for [(TMG₃tren)Cu^{II}(O₂^{•-})]⁺ and other cupric–superoxo complexes with tripodal tetradentate N₄ ligands.^{21,27,30}

Reactivity of [(DMM-tpmpa)Cu^{II}(O₂^{•-})]⁺ (2) toward *para*-Substituted 2,6-Di-*tert*-butylphenols. As mentioned in the Introduction, the broader perspective for why the present study was undertaken is that our general knowledge of [Cu^{II}(O₂^{•-})]⁺ reactivity is very limited. There are a few highly interesting examples of C–H oxidation reaction (*vide infra*), and as also indicated, we provided initial descriptions of phenol oxygenation (giving benzoquinones) with both [(NMe₂-tpmpa)Cu^{II}(O₂^{•-})]⁺ (Chart 2)²¹ and [(TMG₃tren)Cu^{II}(O₂^{•-})]⁺

Table 1. Phenol BDE's, Redox Potentials (E_{ox}), Second-Order Rate Constants (183 K) for *p*-X-DTBP Phenol Oxidations by 2 and 4 and Reaction Yields (See Figure S10 for Kinetics Details)

substituent (X)		BDE ^a (kcal mol ⁻¹)	<i>E</i> _{ox} V (vs Fc/Fc ⁺) ^c	<i>k</i> ₂ of 2 ^d (M ⁻¹ s ⁻¹)	<i>k</i> ₂ ^C of 4 ^d (M ⁻¹ s ⁻¹)	DTBQ yield (%)
OR	OCH ₂ CH ₃	79.6	0.532	24	714	49
	OCH ₃		0.526	23	520	
	OCD ₃		0.496	21	513	
	OCH ₃ , -OD	80.1	0.585	2.1	58	44
	OMPP ^b		0.614	0.84	350	
	OCH ₂ CF ₃		0.805	0.81	329	
R	CH ₃	80.1	0.81	0.042	185	38
	CH ₂ CH ₃	80.0	0.875	0.027	160	
	<i>sec</i> -butyl		0.884	0.023	152	
	CH ₃ , -OD	82.3	0.896	0.010		38
	<i>tert</i> -butyl		0.927	0.008	106	
	H	82.7	1.074	NR		

^aBond dissociation energy in DMSO.³¹ ^bOMPP = 2-methyl-1-phenylpropan-2-yl oxy. ^cThese were determined from SHACV measurements; see Experimental Section. The experimental error is ± 0.01 V. ^dThe experimental error is $\pm 5\%$.

(Figure 2).³² At first, we planned to employ either or both of these complexes for detailed phenol oxidation/oxygenation reactivity studies but found them to have a very narrow range of substrates that could be oxidized (in terms of O–H BDE), and even where oxidation occurred, the reactions were exceptionally slow (Figure S4), thus not amenable to kinetic investigations, as compared to what we find here for reactions of complex 2. A very simple analysis and conclusion would be that the superoxo moiety in [(NMe₂-tmpa)Cu^{II}(O₂^{•-})]⁺, with its very electron-releasing pyridyl *p*-NMe₂ substituents, is less electrophilic than the superoxo complex in 2, with its pyridyl *p*-OMe substituents. As to how this translates to reactivity of phenols with varying O–H BDE, we can judge from the BDE values of substrates in Table 1 that the BDE of the Cu–OOH complex must be larger than 79.6 kcal mol⁻¹ but smaller than 82.7 kcal mol⁻¹. Initial survey of reaction of 2 with a variety of phenols indicated that the kinetic studies and product analyses were viable for a good range of phenols, both *p*-alkoxy-2,6-di-*tert*-butylphenols (*p*-OR-DTBP) and *p*-alkyl-2,6-di-*tert*-butylphenols (*p*-R-DTBP) (Table 1).

Kinetic and Thermodynamic Studies. Demonstration of Hydrogen Atom Transfer Chemistry. Kinetic isotope labeling studies on the oxidation of the two phenol substrates *p*-OMe-DTBP and *p*-Me-DTBP by [(DMM-tmpa)-Cu^{II}(O₂^{•-})]⁺ (2) were performed under pseudo-first-order conditions at 183 K by following the disappearance of the 409 nm band, as shown in Figure 4a.³³ In both cases, the decay behavior observed fits to a first-order kinetics and yields an observed rate constant (k_{obs}) which was linear with respect to the substrate concentration, as shown in Figure 4b. A second-order rate constant (k_2) of 23 M⁻¹ s⁻¹ was obtained for the oxidation of proteo *p*-OMe-DTBP. Using a substrate which had been subjected to deuterium (²H–O) exchange (see Experimental Section), a significant deceleration of the reaction rate was observed, yielding a second-order rate constant of 2.1 M⁻¹ s⁻¹. Thus, a primary kinetic isotope effect (KIE) of 11 was obtained. Using *p*-Me-DTBP, a lower but significant KIE of 4.2 was observed based on the determination of $k_2 = 4.2 \times 10^{-2}$ and 1.0×10^{-2} M⁻¹ s⁻¹ obtained for the oxidation of the proteo- and deuterium-labeled phenols, respectively. Thus, it appears that the oxidations of both the *p*-alkoxy-DTBP and *p*-alkyl-DTBP by 2 occur via a rate-limiting O–H activation event.

Comparison of KIEs to Other Metal–Superoxo-Mediated Oxidations. There have been no prior reports of O–H KIEs by cupric–superoxo complexes. However, primary KIEs of 10.6 and 10.9 are reported for the C–H activation of hippuric acid and dopamine by PHM and DBM, respectively,³⁴ thought to be effected by a protein cupric–superoxide moiety.^{6b,10} In a bioinspired synthetic system, a value of 12.1 was reported for the oxidation of BNAH (1-benzyl-1,4-dihydronicotinamide) for the PV-tmpa [bis(pyrid-2-ylmethyl) ([6-(pivalamido)pyrid-2-yl)methyl]amine] cupric–superoxo complex.³⁰ A lower magnitude of 4.1 was reported for the intramolecular benzylic C–H oxidation of a phenethyl ligand arm in a chelated copper(II)–superoxo complex reported by Itoh and co-workers.³⁵ In addition, this value (KIE = 12) is approximately equal to what is observed for the oxidation of a water-soluble trisubstituted phenol by a chromium–superoxo described by Bakac and co-workers.³⁶ All of these results are consistent with the substrate activation via homolytic O–H bond cleavage.

Cumylperoxyl Radical (4) Plus Phenol Substrate HAT Reactions for Comparison. To provide further evidence that hydrogen atom transfer is the rate-determining step in these reactions, we studied the same substrates with cumylperoxyl radical (4); the latter is known to effect “pure” HAT chemistry with phenols (and *N,N*-dimethylanilines).^{13,37} This hydrogen atom acceptor reacts with phenols to yield cumene hydroperoxide and phenoxyl radical; here, we wished to compare the behavior of 4 with phenols to that of the reactions of [(DMM-tmpa)Cu^{II}(O₂^{•-})]⁺ (2) and phenols (Scheme 2). Thus, to understand the relationship between second-order rate constants for 2 (k_2) and 4 (k_2^{C}) (Table 1), the reaction of the whole series of substrates with 2 and 4 has been explored monitoring the chemistry with the use of UV–vis and EPR spectroscopies, respectively (see Experimental Section). A large KIE value (KIE = 9.0) was observed for hydrogen atom transfer from *p*-OMe-DTBP to 4, as shown in Figure S5. The k_2 values display a linear correlation with k_2^{C} , which further indicates that HAT is involved in the rate-limiting step of phenol oxidations by 2 (Figure 5). If the rate-determining step was to be pure HAT, as in case of 4, the slope would be 1. The larger slope observed (=4.85) from the plot obtained indicates that some degree of contribution of partial transfer of charge is also involved in the rate-determining step for 2 plus phenol reactions.

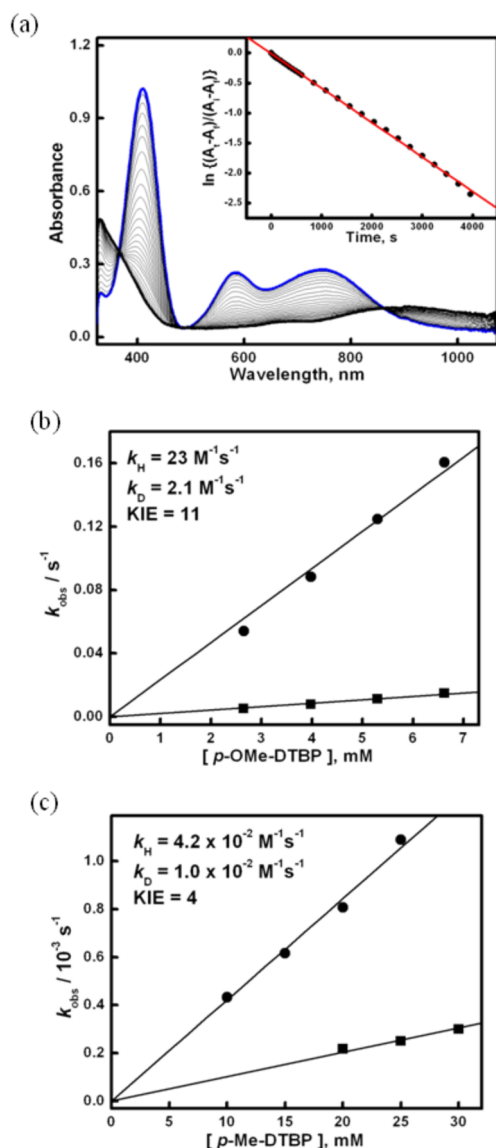


Figure 4. (a) UV-vis spectral changes observed by addition of *p*-Et-DTBP (20 mM) to **2** (0.27 mM) in acetone (10% MeTHF) at 183 K; $k_{\text{obs}} = 5.73 \times 10^{-4} \text{ s}^{-1}$. (b,c) Plots of k_{obs} values against the concentrations of substrates (circles) and deuterated ^2H -O substrates (squares) for *p*-OMe-DTBP (b) and *p*-Me-DTBP (c) to determine second-order rate constants and KIEs.

Scheme 2

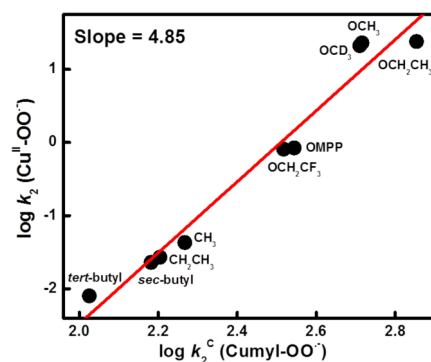
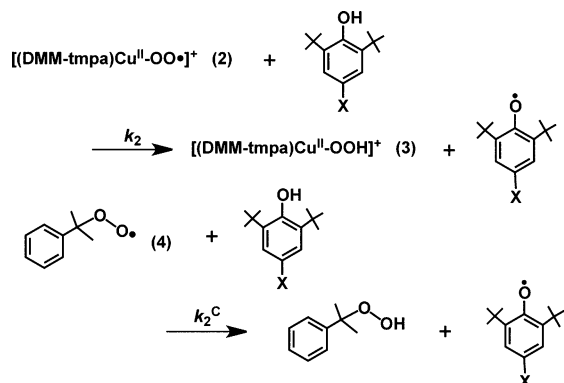


Figure 5. Correlation between $\log k_2$ of **2** and $\log k_2^{\text{C}}$ of **4** with *p*-X-DTBPs. Slope is 4.85.

Rate Constant (k_2) Correlation to Phenol Redox Potentials
 ? We also considered possible relationships between the second-order rate constants for the reaction of $[(\text{DMM-tmpa})\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]^+$ (**2**) with phenols (k_2) and substrate redox potentials. In fact, k_2 values for the reactions increase with decreasing the redox potentials, that is, with increasing driving force of electron transfer from phenols to **2**. The plot of $(k_{\text{B}}T/e) \ln k_2$ versus E_{ox} exhibits a linear correlation with a negative slope = -0.29 , as shown in Figure 6. By contrast, for the case of

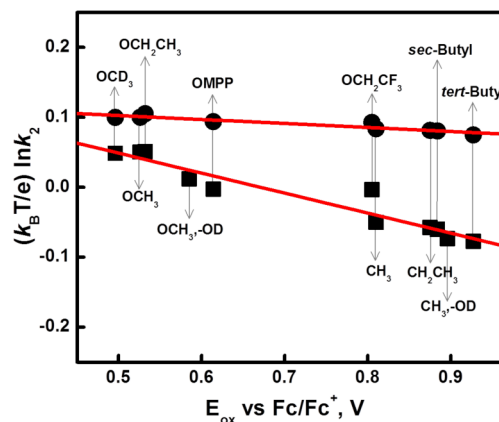


Figure 6. Plots of $(k_{\text{B}}T/e) \ln k_2$ for the reactions of *p*-X-DTBPs with **2** (squares) and **4** (circles) against the one-electron oxidation potentials (E_{ox}) of substrates. The slopes are -0.29 and -0.05 , respectively.

cumylperoxyl radical (**4**), the plot of $(k_{\text{B}}T/e) \ln k_2^{\text{C}}$ versus E_{ox} exhibits a much weaker dependence on the E_{ox} values, and the slope is only -0.05 . Although the bond dissociation energy (BDE) of *p*-X-DTBP is known to decrease by electron-donating substituents,³⁸ the difference in BDE between OCH_3 and *tert*-butyl group is only $2.7 \text{ kcal mol}^{-1}$, which is equivalent to 0.12 eV , whereas the difference in E_{ox} between OCH_3 and *tert*-butyl group is 0.40 eV (Table 1). Based on thermochemical cycles,³⁷ the difference in BDE (ΔBDE) is given by eq 2.

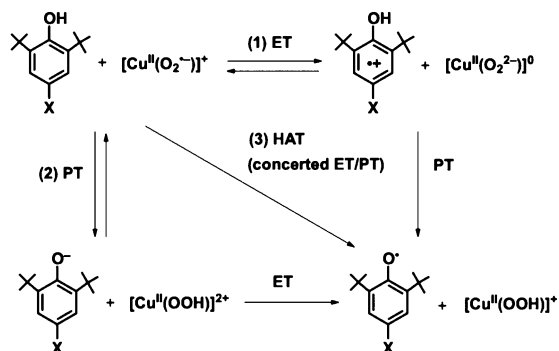
$$\Delta\text{BDE} = e\Delta E_{\text{ox}} + k_{\text{B}}T\Delta pK_{\text{a}} \quad (2)$$

where ΔpK_{a} is the difference in the pK_{a} values of *p*-X-DTBP⁺⁺ and entropy changes are neglected. The pK_{a} value becomes smaller with increased E_{ox} value, and the constant BDE value (eq 2) indicates that an increase in the E_{ox} value is partially canceled by a decrease in the pK_{a} value. Such cancellation may be the reason why the BDE value is much less sensitive to

electron-donating substituents in Table 1 as compared to the E_{ox} value.

Mechanistic Considerations. There are three possible reaction pathways in the apparent hydrogen atom transfer from p -X-DTBP to $[\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]^+$: (1) electron transfer followed by proton transfer, (2) PT followed by ET; and (3) concerted ET and PT (Scheme 3). The observation of large

Scheme 3



KIEs in Figure 4 indicates that PT should be involved in the rate-determining step. In such a case, PT should be the rate-determining step following the ET equilibrium, which is endergonic, in the electron transfer/proton transfer (ET/PT) pathway (1).³⁹ In this case, the observed second-order rate constant (k_2) is given by eq 3.

$$k_2 = k_p K_{\text{et}} \quad (3)$$

where k_p is the rate constant of proton transfer from p -X-DTBP^{•+} to $[\text{Cu}^{\text{II}}(\text{O}_2^{2-})]^0$ and K_{et} is the ET equilibrium constant between p -X-DTBP and $[\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]^+$ ($K_{\text{et}} \ll 1$). Because the Brønsted slope of k_p is between 0 and 0.5 for an exergonic proton transfer reaction,⁴⁰ and the slope of the plot of $(k_B T/e) \ln K_{\text{et}}$ versus E_{ox} is -1.0 , the predicted slope of a plot of $(k_B T/e) \ln k_2$ versus E_{ox} is between -0.5 and -1.0 .

In the case of the PT/ET pathway (2), PT should also be the rate-determining step followed by fast ET. In this case, the slope of the plot of $(k_B T/e) \ln k_2$ versus E_{ox} is positive because an increase in the E_{ox} value with electron-withdrawing substituents results in an decrease in the $\text{p}K_{\text{a}}$ value (more acidic) when the proton transfer from p -X-DTBP to $[\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]^+$ becomes thermodynamically more favorable. The observed negative slope (-0.29) clearly rules out the PT/ET pathway. Although the ET/PT pathway affords the negative slope of a plot of $(k_B T/e) \ln k_2$ versus E_{ox} is between -0.5 and -1.0 , the observed slope (-0.29) is less negative than -0.5 . In the case of the concerted ET/PT pathway (one-step hydrogen atom transfer), the slope is -0.05 as observed for hydrogen atom transfer from p -X-DTBP to **4**. The smaller slope than expected for transfer of a full unit of charge has been reported to result from only partial transfer of charge in hydrogen atom transfer reactions from hydrogen donors to the triplet excited state of benzophenone.⁴¹ Thus, it is most likely that hydrogen transfer from p -X-DTBP proceeds via a partial transfer of charge rather than an ET/PT pathway in which a full unit of charge is transferred. Thus, more electron-deficient ligands may be required in order to increase the hydrogen transfer reactivity of $[\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]^+$ by facilitating charge transfer from p -X-DTBP to $[\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]^+$.

Activation Parameters and Comparisons. The rates of the $[(\text{DMM-tmpa})\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]^+$ (**2**) + p -X-DTBP reactions were of course temperature-dependent. However, we could only examine a narrow temperature range, as colder (than 173 K) conditions involved solution freezing, while self-decomposition of **2** was too extensive when warming above 188 K. Also, the slow self-decomposition precluded the generation of good quality data except for the fastest reaction, that being with the p -OMe-DTBP substrate. Nevertheless, an Eyring plot developed from data between 173 and 188 K for this substrate yielded activation parameters as follows: $\Delta H^\ddagger = 3.6 \pm 0.6 \text{ kcal mol}^{-1}$ and $\Delta S^\ddagger = -32 \pm 3 \text{ cal mol}^{-1} \text{ K}^{-1}$ (Figure S6). The large negative activation entropy suggests that the rate-determining step involves a well-ordered transition state.

Although no comparable activation parameters for p -X-DTBP oxidation by cupric-superoxide or other Cu-oxygen complexes have been reported, there are a few values measured for oxidation of phenol derivatives by various other metal ion complexes, including Cu^{III} , Ru^{VI} , Mn^{V} , and Ru^{III} (Table 2).⁴²

Table 2. Kinetic Parameters in Various Metal Complexes Plus Phenol Oxidation Reactions

reactants		$\Delta H^\ddagger$ (kcal mol ⁻¹)	$\Delta S^\ddagger$ (cal mol ⁻¹ K ⁻¹)
$\text{Cu}^{\text{II}}-\text{O}_2^{\bullet-}$ ^a	p -OMe-DTBP	3.6 ± 0.6	-32 ± 3
$\text{Cu}(\text{III})$ ^{42a}	2,4-DTBP	8.3 ± 1.1	-27 ± 3
$\text{Ru}(\text{VI})$ ^{42b}	phenol	11.3 ± 0.8	-14 ± 2
$\text{Mn}(\text{V})$ ^{42c}	2,4-DTBP	6.3 ± 1.4	-35.6 ± 2.3
$\text{Ru}(\text{III})$ ^{42d}	p - ^t Bu-DTBP	1.6 ± 0.2	-36 ± 2

^aThis work.

Except for the case of the reaction of a Ru^{III} -pterin complex, which was concluded to involve PCET, all other cases are stated as involving HAT in the rate-limiting step. In particular, the HAT reaction of a manganese(V)-oxo corrolazine complex plus 2,4-di-*tert*-butylphenol (2,4-DTBP) affording $\text{Mn}^{\text{IV}}(\text{OH})$ plus 2,4-di-*tert*-butylphenoxyl radical gives both $\Delta H^\ddagger$ and $\Delta S^\ddagger$ values close to those obtained from our reaction of **2** and p -OMe-DTBP.

KIE Temperature Dependence? We also investigated the temperature dependence of the KIE values for the reaction with deuterated p -OMe-DTBP (Table 3, Figure S6). While the

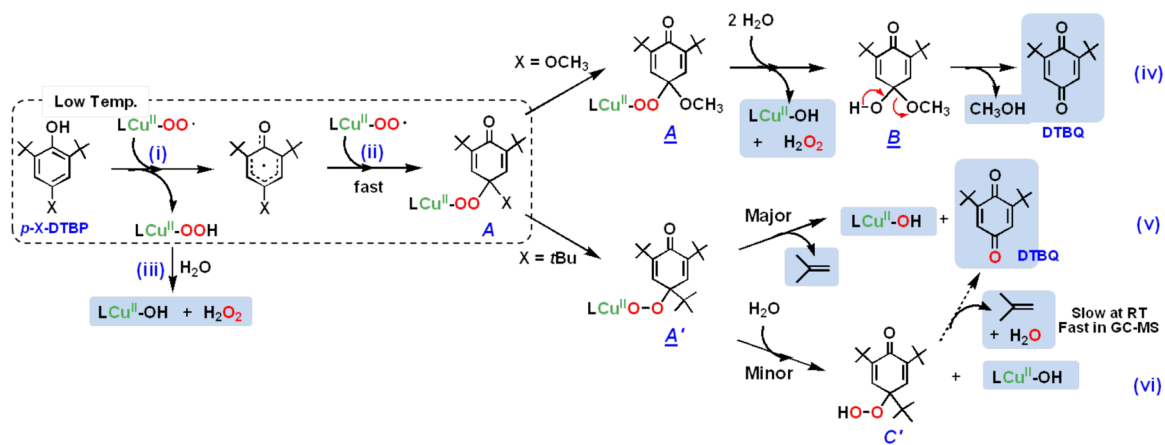
Table 3. Temperature Dependence on the KIEs of the HAT Reactions from p -OMe-DTBP to $[(\text{DMM-tmpa})\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]^+$ (**2**)

	k_{H} (M ⁻¹ s ⁻¹)	k_{D} (M ⁻¹ s ⁻¹)	$k_{\text{H}}/k_{\text{D}}$
173 K	11	1.0	11
178 K	14	1.2	12
183 K	23	2.1	11
188 K	26	2.7	10

temperature range is again limited, the results suggest there is no temperature dependence on the magnitude of the primary KIEs. Thus, the Arrhenius pre-exponential factor for the phenol O-H and O-D isotopically labeled analogues remains essentially constant as a function of temperature. This behavior is consistent with a substrate oxidation mechanism involving classical rate-limiting H atom abstraction.⁴³

Summary of the Kinetic-Thermodynamic Studies. As detailed above, our in-depth studies are consistent with the

Scheme 4. Proposed Pathways for $[\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]^+$ Reactivity with *p*-OMe-DTBP or *p*-*t*Bu-DTBP Substrates Leading to Observed Products^a



^aSee text for detailed descriptions of direct/indirect evidence or literature support for the individual reaction steps or intermediates described. EPR spectra observed for (A $\equiv$ A') (from mixing 2 and *p*-X-DTBP at -90°C), for $\text{LCu}^{\text{II}}\text{--OH(H)}$ (observable at room temperature and refreezing to record EPR spectra) are indistinguishable; see Figure S7 and caption.

conclusion that the cupric-superoxo complex $[(\text{DMM-tmpa})\text{--Cu}^{\text{II}}(\text{O}_2^{\bullet-})]^+$ (2), as a kind of prototype for the initial adducts formed in chemical or biological $\text{Cu}^{\text{I}}/\text{O}_2$ interactions, reacts with *p*-OMe-DTBP (and the other phenols) via a HAT oxidative mechanism. Likewise, as mentioned above, rate-limiting HAT occurs for the galactose oxidase active site special Tyr, but PCET in CAO has been suggested by Klinman^{7c} and Roth.⁸ In relevant C–H bond activation by $[\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]^+$ species, Klinman and Blackburn reported C–H bond cleavage in *N*-benzoylglycine by PHM having large intrinsic isotope effects.^{34a} They also demonstrated hydrogen tunneling as occurring in PHM with a lack of a temperature dependence of an intrinsic isotope effect on the C–H bond cleavage of the substrate hippuric acid.⁴⁴ As also mentioned, Itoh and co-workers reported intramolecular C–H hydroxylation (HAT) by a cupric-superoxo species supported by tridentate ligand possessing a distorted tetrahedral geometry.^{35a}

Product Analyses and Further Mechanistic Insights.

Phenol Oxidations Lead to Benzoquinone Products. As described above in detail, the initial products generated after rate-limiting hydrogen atom transfer occurs are presumed to be a $[(\text{DMM-tmpa})\text{Cu}^{\text{II}}(\text{OOH})]^+$ (3) and a *para*-X-2,6-di-*tert*-butylphenoxyl radical (Scheme 2). Such a phenoxyl radical would possess distinctive $\lambda_{\text{max}} = 384$ and 402 nm absorptions and exhibit a sharp $g \sim 2.0$ EPR spectroscopically detectable signal. However, neither of these spectroscopic signals could be observed. Thus, qualitative and quantitative product analyses were carried out, allowing us to understand and build up the proposed overall mechanism and stoichiometry of the reactions observed. In fact, for all phenolic substrates, benzoquinones are produced. As mentioned above, reactions of 2 with *p*-OR-DTBP and *p*-R-DTBP proceed via different pathways, two-electron oxidation and four-electron oxygenation, respectively, and these further aspects of the overall chemistry are now detailed.

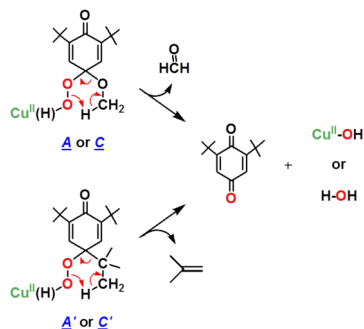
Reaction of $[(\text{DMM-tmpa})\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]^+$ (2) with *p*-Alkoxy-2,6-di-*tert*-butylphenols (*p*-OR-DTBPs). All synthetic reactions were carried out by mixing excess phenol substrate with 2 at -90°C . The cryogenic solutions were interrogated by UV–vis, NMR, and EPR spectroscopies. As described for the initial kinetics of the system (vide supra), 2 rapidly disappears. Low-

temperature UV–vis (new d–d bands) and EPR (EPR silent to EPR active) spectroscopies reveal that a new mononuclear Cu^{II} complexes has formed, but NMR spectroscopy suggests that no quinone has yet formed and only starting extra phenol is present. However, warming of these initial reaction solutions to room temperature followed by NMR and GC–MS analyses indicates that up to 50% yields of 2,6-di-*tert*-butyl-1,4-benzoquinones form (Scheme 1). An ^{18}O -labeling experiment (see Experimental Section) where 2 was generated using $^{18}\text{O}_2$ gas revealed that no ^{18}O was incorporated into the DTBQ. This is explained below; only the overall two-electron oxidation of the substrate phenol takes place, while hydrogen peroxide and alcohol products are also formed. The fate of the initial *para*-OR fragment, after warming, was identified by $^2\text{H(D)}$ -labeling experiments using *p*- $\text{OCD}_3/\text{OCD}_2\text{CD}_3$ -DTBP substrates (Figure S9). In each case, the product detected was the corresponding alcohol (methanol and ethanol), not the oxidized form (formaldehyde and acetaldehyde). The inorganic product is a mononuclear copper(II) complex $[(\text{DMM-tmpa})\text{Cu}^{\text{II}}\text{--OH(H)}]^{+/2+}$, and H_2O_2 is produced ($\sim 50\%$ yield), as determined by EPR/UV–vis spectroscopy (see Figures S7, S8, and S13) and iodometric titration, respectively.⁴⁵

Proposed Pathway for $[\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]^+$ (2) Reaction with *p*-OR-DTBP Substrates (Scheme 4). Initially, a hydrogen atom is transferred from the phenol to $[\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]^+$ as the (low-temperature) rate-determining step, where $[\text{Cu}^{\text{II}}(\text{OOH})]^+$ and the corresponding phenoxyl radical form (Scheme 4, step i). The $[\text{Cu}^{\text{II}}(\text{OOH})]^+$ intermediate would “hydrolyze”, giving H_2O_2 and $[\text{Cu}^{\text{II}}(\text{OH})]^+$ (Scheme 4, step iii) (upon warming to room temperature). Strong support for this claim comes from our independent generation of authentic complex $[\text{Cu}^{\text{II}}(\text{OOH})]^+$ (3) and demonstration that it releases H_2O_2 (which was directly identified) and forms the hydroxide complex, $[\text{Cu}^{\text{II}}(\text{OH})]^+$. This was identified as the Cu product by matching EPR and UV–vis spectroscopic data with authentically generated compounds (see Supporting Information). The formation of the eventual benzoquinone and other products is explained by (a) fast radical–radical coupling reaction of the newly generated $\text{ArO}^\bullet$ moiety and a second equiv $[\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]^+$, affording a peroxy $\text{Cu}^{\text{II}}\text{--OO--(ArO')}$

species **A** (Scheme 4, step ii); (b) then this reacts with solvent water to give **B**, which eliminates the alcohol (RO–H) derived from the *p*-OR-DTBP substrate (step iv). {Note: if this hydrolysis step involved Cu^{II}–OOH rather than Cu^{II}–OO–(ArO') bond cleavage, then the corresponding hydroperoxo intermediate **C** (HOO–(ArO')) (Scheme 5) forms, rather than

Scheme 5



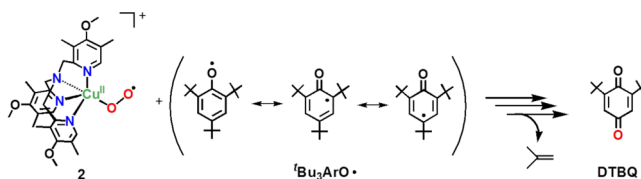
our postulated **B** with HO–(ArO') formulation. However, **C** would be incapable of producing methoxide/methanol plus benzoquinone, our observed products. If **C** or Cu^{II}–peroxo derivative **A** were to form, CH₂=O would be produced (Scheme 5), but none was observed (Experimental Section). Kinetic aspects for quinone formation were not investigated in detail, as this step occurs later, with warming of the reaction solutions (also see Experimental Section).

Reaction of [(DMM-*tmpa*)Cu^{II}(O₂^{•−})]⁺ (2**) with *p*-Alkyl-2,6-di-*tert*-butylphenols (*p*-R-DTBPs).** For the case where the *para*-substituent is an alkyl group like *tert*-butyl, the reaction of *p*-R-DTBP plus [(DMM-*tmpa*)Cu^{II}(O₂^{•−})]⁺ (**2**) proceeds differently, via an overall four-electron substrate oxygenation. In the initial reaction, steps i and ii proceed analogously, producing the Cu^{II}–OO–(ArO') species **A'** (Scheme 4). This, however, can undergo a *tert*-butyl substituent oxidation via elimination of isobutylene, then directly giving the DTBQ and Cu(II) products observed (Scheme 4, step v).⁴⁶ Isobutylene or *t*BuOH (=isobutylene + H₂O) elimination from *tert*-butyl-substituted phenols under oxidative conditions is well precedented.⁴⁷ We, however, were not able to detect isobutylene as a product.^{24,48}

As can be seen from Scheme 4 (step v), the product DTBQ in the [(DMM-*tmpa*)Cu^{II}(O₂^{•−})]⁺ (**2**) plus *p*-R-DTBP substrate reaction should have one of its O atoms derived from molecular oxygen (derived from **2**). Indeed, this appears to be the case. The reaction of *p*-^tBu-DTBP with **2** provides for a quantitative yield of DTBQ (based on the stoichiometry of two molecules of **2** per one mole of phenol substrate). For ¹⁸O-labeled **2** (derived from **1** plus ¹⁸O₂), reaction with *p*-^tBu-DTBP affords a 40% of ¹⁸O-incorporated DTBQ (GC–MS). The smaller isotope incorporation yield than expected may result from the ¹⁸O–¹⁶O exchange reaction with water because carbonyl compounds are known to undergo oxygen exchange reactions with water, catalyzed by acids or bases.⁴⁹

[Cu^{II}(O₂^{•−})]⁺ (2**) Coupling with 2,4,6-Tri-*tert*-butylphenoxyl Radical.** A set of experiments that provides further and strong support for our mechanistic picture of the chemistry comes from reacting [(DMM-*tmpa*)Cu^{II}(O₂^{•−})]⁺ (**2**) with isolated ^tBu₃ArO• (Scheme 6). The addition of 1 equiv of ^tBu₃ArO• to **2** was monitored by UV–vis and EPR spectroscopies. A fast reaction occurs;⁵⁰ the intermediate **2** quickly disappears (UV–

Scheme 6



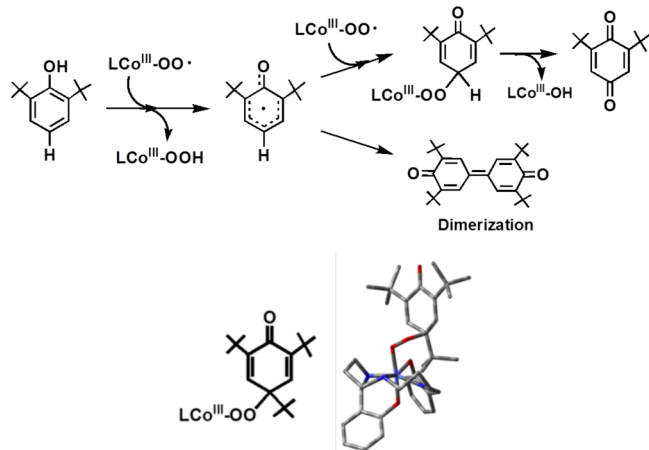
vis monitoring), and the Cu(II) EPR spectrum previously observed for the [(DMM-*tmpa*)Cu^{II}(OH(H))]^{+/2+} product appears. The yield of DTBQ was 98%, and a 70–80% ¹⁸O incorporation into the DTBQ product was observed. Thus, the **2** + ^tBu₃ArO• occurs with a 1:1 stoichiometry (Scheme 4, step ii), and this then supports the [Cu^{II}(O₂^{•−})]⁺/phenol = 2:1 overall stoichiometry. In contrast to the case of *p*-OR-DTBP, the oxygen incorporated into the oxidized product (DTBQ) obtained from the reaction of *p*-R-DTBP with **2** originates from **2** via O–O bond cleavage followed by the elimination of the oxidized R group (Scheme 4, step v).

Minor Reaction Pathway Contributing to *p*-R-DTBP Oxidation. If the reaction of [(DMM-*tmpa*)Cu^{II}(O₂^{•−})]⁺ (**2**) with *p*-R-DTBP (or that of **2** + ^tBu₃ArO•) occurred only via step v (Scheme 4), the product solution should not oxidize iodide since no H₂O₂ would have been generated. However, we observed 20–30% of triiodide absorption band formation when testing for peroxide. We explain this by invoking the “minor” step vi, where hydrolysis of **A'** occurs giving 2,4,6-tri-*tert*-butyl-4-hydroperoxycyclohexa-2,5-dienone (**C'**), which (i) does oxidize iodide ion (see Experimental Section) and (ii) slowly converts to DTBQ (at room temperature) but (iii) fully converts to DTBQ in the GC–MS experiment.

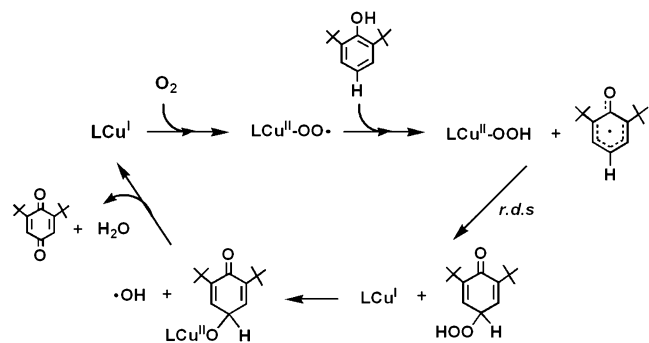
Precedent and Comparison with Cobalt and Nickel–Superoxide Phenol Oxidations. In the overall process involving reaction of the *p*-X-DTBP, steps i and ii (top line; Scheme 4), a second molar equiv of [(DMM-*tmpa*)Cu^{II}(OH)]⁺ and H₂O₂ is produced, all consistent with the identity and yields of products obtained and thus the reaction stoichiometry. Nishinaga and co-workers, in classical studies of the reactions of [Co^{III}(O₂^{•−})]⁺ (formed from Co^{II} and O₂), also observed this same reaction stoichiometry, involving two [Co^{III}(O₂^{•−})]⁺ moieties with one phenol substrate. Further, Nishinaga's work provides precedent for the proposed (Scheme 4) reaction of [(DMM-*tmpa*)Cu^{II}(O₂^{•−})]⁺ (**2**) with the initially generated phenoxyl radical intermediate;⁵¹ Nishinaga^{51b} also obtained a cobalt peroxy species Co^{III}–OO–(ArO'), and its X-ray structure for a *para*-^tBu phenol substrate (Scheme 7). Further, in order to explain the products observed in very recent investigations by Driess and co-workers,⁵² involving phenol reactions with a new [Ni^{II}(O₂^{•−})]⁺ complex, an intermediate analogous to **A'** was proposed to form from *p*-R-DTBP (R = H, Me, ^tBu) substrates.

Prior Computational Study of [Cu^{II}(O₂^{•−})]⁺ Reaction with 2,6-Di-*tert*-butylphenol (*p*-H-DTBP) Leading to a 2,6-Di-*tert*-butyl-1,4-benzoquinone. In 2009, based on DFT calculations, Güell et al.⁵³ proposed a mechanism (Scheme 8) of *p*-H-DTBP oxidation by a cupric–superoxide complex close analogue of **2**, [(NMe₂-*tmpa*)Cu^{II}(O₂^{•−})]⁺ (see Introduction), which differs considerably from what we have found and proposed in the present study (Scheme 4). They suggest HAT occurs at the first step (as we observe), but then it is suggested that the LCu^{II}–OOH species formed undergoes Cu–OOH homolytic cleavage and the hydroperoxyl radical thus formed attacks the

Scheme 7



Scheme 8



phenoxyl radical, overall leading to copper(I) and hydroperoxy organic compound. This C–O bond formation process is described as the rate-determining step. Then, the resulting copper(I) facilitates O–O homolytic cleavage, forming a hydroxyl radical plus a copper(II)–O–ArO' complex. The last step involves HAT (or PCET) from the ring to hydroxyl radical, leading to the formation of copper(I), a water molecule, and DTBQ. Thus, their overall chemistry is formally a catalytic reaction (regenerating copper(I), to react again with more O₂ to give the cupric–superoxo species), with different stoichiometry than we observed and a different rate-determining step as mentioned. However, we should point out that the substrate used in their calculations was *p*-H-DTBP, a perhaps rather special case compared to the series of substrates we examined (Table 1). In fact, we find that *p*-H-DTBP is unreactive toward [(DMM-tmpa)Cu^{II}(O₂^{•−})]⁺ (2).

CONCLUSION

In this report, we have obtained detailed mechanistic insights into the oxidation of a wide series of *para*-substituted 2,6-di-*tert*-butylphenols (*p*-X-DTBPs) by a newly synthesized cupric–superoxo complex supported by an electron-rich ligand, [(DMM-tmpa)Cu^{II}(O₂^{•−})]⁺ (2). With detailed kinetic investigations, we proved that hydrogen atom abstraction is the first step and the rate-determining-step for the oxidation of *p*-X-DTBPs. The key observations supporting this were the finding of a large deuterium kinetic isotope effect, a good correlation with reactivity of the cumylperoxyl radical toward the same substrates, comparison of the rate dependences in the phenol oxidations on the one-electron oxidation potentials of the *p*-X-

DTBP substrates, and the observed activation parameters for the reaction. The hydrogen atom transfer from *p*-X-DTBPs proceeds via a partial transfer of charge rather than a complete transfer of charge in the ET/PT pathway. The qualitative and quantitative product analyses and reactivity study carried out with the 2,4,6-tri-*tert*-butylphenoxyl radical (^tBu₃ArO[•]) reacting with 2 allowed us to build the case for the proposed overall stoichiometry of reaction and, furthermore, the detailed mechanism. Two moles of copper(II)–superoxo species are required to oxidize *para*-substituted 2,6-di-*tert*-butylphenols to 2,6-di-*tert*-butyl-1,4-benzoquinones. It is found that the reaction of 2 toward *para*-alkoxy-2,6-di-*tert*-butylphenol proceeds via two-electron oxidation with hydrolysis, while reaction with *para*-alkyl-2,6-di-*tert*-butylphenol proceeds via a process involving four-electron oxygenation chemistry.

This study contributes significantly to our understanding of the fundamental chemistry and oxidative capabilities of initial copper(I)–dioxygen adducts, such as cupric–superoxide complexes. As described in the Introduction, Cu^{II}(O₂^{•−}) species are implicated in reactions with both C–H and O–H containing substrates, and the latter biomimetic chemistry was the focus of attention here. The supporting ligand is well-known to modulate the structural, electronic structure/bonding, as well as the reactivity nature of copper complexes, here with O₂-derived fragments bound to the copper ion. Thus, future efforts will include the generation of Cu^{II}(O₂^{•−}) complexes with rather differing supporting ligands to explore the range of reactivity possible for such primary copper–dioxygen adducts.

ASSOCIATED CONTENT

Supporting Information

Reaction schemes for copper proteins, synthetic details, EPR spectra, additional kinetic data and NMR spectra. This material is available free of charge via the Internet at <http://pubs.acs.org>.

AUTHOR INFORMATION

Corresponding Authors

edward.solomon@stanford.edu
fukuzumi@chem.eng.osaka-u.ac.jp
karlin@jhu.edu

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

We are grateful to the NIH (E.I.S., DK31450; K.D.K., GM28962), JSPS (S.F., 20108010; K.O., 23750014, 26620154, 26288037), and JST (S.F., ALCA) for financial support of this research.

REFERENCES

- (1) (a) Mirica, L. M.; Ottenwaelter, X.; Stack, T. D. P. *Chem. Rev.* **2004**, *104*, 1013–1045. (b) Lewis, E. A.; Tolman, W. B. *Chem. Rev.* **2004**, *104*, 1047–1076. (c) Hatcher, L. Q.; Karlin, K. D. *J. Biol. Inorg. Chem.* **2004**, *9*, 669–683. (d) Peterson, R. L.; Kim, S.; Karlin, K. D. In *Comprehensive Inorganic Chemistry II*, 2nd ed.; Elsevier: Amsterdam, 2013; pp 149–177.
- (2) (a) Schröder, D.; Holthausen, M. C.; Schwarz, H. *J. Phys. Chem. B* **2004**, *108*, 14407–14416. (b) Dietl, N.; van der Linde, C.; Schlagen, M.; Beyer, M. K.; Schwarz, H. *Angew. Chem., Int. Ed.* **2011**, *50*, 4966–4969.
- (3) (a) Hong, S.; Huber, S. M.; Gagliardi, L.; Cramer, C. C.; Tolman, W. B. *J. Am. Chem. Soc.* **2007**, *129*, 14190–14192. (b) Donoghue, P. J.;

- Tehranchi, J.; Cramer, C. J.; Sarangi, R.; Solomon, E. I.; Tolman, W. B. *J. Am. Chem. Soc.* **2011**, *133*, 17602–17605. (c) Kunishita, A.; Ishimaru, H.; Nakashima, S.; Ogura, T.; Itoh, S. *J. Am. Chem. Soc.* **2008**, *130*, 4244–4245. (d) Maiti, D.; Narducci Sarjeant, A. A.; Karlin, K. D. *Inorg. Chem.* **2008**, *47*, 8736–8747.
- (4) (a) Nakao, Y.; Hirao, K.; Taketsugu, T. *J. Chem. Phys.* **2001**, *114*, 7935–7940. (b) Decker, A.; Solomon, E. I. *Curr. Opin. Chem. Biol.* **2005**, *9*, 152–163. (c) Yoshizawa, K.; Kihara, N.; Kamachi, T.; Shiota, Y. *Inorg. Chem.* **2006**, *45*, 3034–3041.
- (5) Itoh, S. *Curr. Opin. Chem. Biol.* **2006**, *10*, 115–122.
- (6) (a) Klinman, J. P. *Chem. Rev.* **1996**, *96*, 2541–2561. (b) Klinman, J. P. *J. Biol. Chem.* **2006**, *281*, 3013–3016.
- (7) (a) McGuirl, M. A.; Dooley, D. M. *Curr. Opin. Chem. Biol.* **1999**, *3*, 138–144. (b) Rokhsana, D.; Shepard, E. M.; Brown, D. E.; Dooley, D. M. Amine Oxidase and Galactose Oxidase. In *Copper–Oxygen Chemistry*; Itoh, S.; Karlin, K. D., Eds.; John Wiley & Sons, Inc.: Hoboken, NJ, 2011; Vol. 4; pp 53–106. (c) Humphreys, K. J.; Mirica, L. M.; Wang, Y.; Klinman, J. P. *J. Am. Chem. Soc.* **2009**, *131*, 4657–4663.
- (8) Liu, Y.; Mukherjee, A.; Nahumi, N.; Ozbil, M.; Brown, D.; Angeles-Boza, A. M.; Dooley, D. M.; Prabhakar, R.; Roth, J. P. *J. Phys. Chem. B* **2012**, *117*, 218–229.
- (9) Evans, J. P.; Ahn, K.; Klinman, J. P. *J. Biol. Chem.* **2003**, *278*, 49691–49698.
- (10) (a) Chen, P.; Solomon, E. I. *J. Am. Chem. Soc.* **2004**, *126*, 4991–5000. (b) Chen, P.; Bell, J.; Eipper, B. A.; Solomon, E. I. *Biochemistry* **2004**, *43*, 5735–5747.
- (11) Prigge, S. T.; Eipper, B.; Mains, R.; Amzel, L. M. *Science* **2004**, *304*, 864–867.
- (12) Warren, J. J.; Tronic, T. A.; Mayer, J. M. *Chem. Rev.* **2010**, *110*, 6961–7001.
- (13) Osako, T.; Ohkubo, K.; Taki, M.; Tachi, Y.; Fukuzumi, S.; Itoh, S. *J. Am. Chem. Soc.* **2003**, *125*, 11027–11033.
- (14) During the review of this present manuscript, Itoh and co-workers reported on the “redox reactivity” of a $[\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]^+$ complex first described in 2009.³⁵ The complex with tridentate N_3 ligand does not oxidize phenols but only reacts with them in an acid–base fashion.
- (15) Liang, H.-C.; Kim, E.; Incarvito, C. D.; Rheingold, A. L.; Karlin, K. D. *Inorg. Chem.* **2002**, *41*, 2209–2212.
- (16) Manner, V. W.; Markle, T. F.; Freudenthal, J. H.; Roth, J. P.; Mayer, J. M. *Chem. Commun.* **2008**, 256–258.
- (17) Nishinaga, A.; Shimizu, T.; Matsuura, T. *J. Org. Chem.* **1979**, *44*, 2983–2988.
- (18) Zhang, C. X.; Kaderli, S.; Costas, M.; Kim, E.-i.; Neuhold, Y.-M.; Karlin, K. D.; Zuberbühler, A. D. *Inorg. Chem.* **2003**, *42*, 1807–1824.
- (19) Fyfe, C. A.; Vanveen, L. J. *Am. Chem. Soc.* **1977**, *99*, 3366–3371.
- (20) Omura, K. *J. Org. Chem.* **1996**, *61*, 7156–7161.
- (21) Maiti, D.; Fry, H. C.; Woertink, J. S.; Vance, M. A.; Solomon, E. I.; Karlin, K. D. *J. Am. Chem. Soc.* **2007**, *129*, 264–265.
- (22) Kim, S.; Saracini, C.; Siegler, M. A.; Drichko, N.; Karlin, K. D. *Inorg. Chem.* **2012**, *51*, 12603–12605.
- (23) (a) Nash, T. *Biochem. J.* **1953**, *55*, 416–421. (b) Zhang, C. X. Ph.D. Thesis, Synthetic analogues for copper metalloenzymes: effects of ligand electronic and structural variations on the formation, stability and reactivity of biomimetic copper-dioxygen adducts. The Johns Hopkins University, Baltimore MD, 2001.
- (24) If higher concentrations of $[(\text{DMM-tmpa})\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]^+$ (**2**) are employed, **2** reacts with $[(\text{DMM-tmpa})\text{Cu}^{\text{I}}(\text{CO})]^+$ (**1**), forming dicopper(II) species; see Introduction (structure type E) and the Experimental Section concerning the synthesis of **1** and the generation of **2**.
- (25) (a) Fujisawa, K.; Tanaka, M.; Morooka, Y.; Kitajima, N. *J. Am. Chem. Soc.* **1994**, *116*, 12079–12080. (b) Chen, P.; Root, D. E.; Campochiaro, C.; Fujisawa, K.; Solomon, E. I. *J. Am. Chem. Soc.* **2003**, *125*, 466–474.
- (26) Würtele, C.; Gaoutchenova, E.; Harms, K.; Holthausen, M. C.; Sundermeyer, J.; Schindler, S. *Angew. Chem., Int. Ed.* **2006**, *45*, 3867–3869.
- (27) Woertink, J. S.; Tian, L.; Maiti, D.; Lucas, H. R.; Himes, R. A.; Karlin, K. D.; Neese, F.; Würtele, C.; Holthausen, M. C.; Bill, E.; Sundermeyer, J.; Schindler, S.; Solomon, E. I. *Inorg. Chem.* **2010**, *49*, 9450–9459.
- (28) Ginsbach, J. W.; Peterson, R. L.; Cowley, R. E.; Karlin, K. D.; Solomon, E. I. *Inorg. Chem.* **2013**, *52*, 12872–12874.
- (29) Lucas, H. R.; Meyer, G. J.; Karlin, K. D. *J. Am. Chem. Soc.* **2010**, *132*, 12927–12940.
- (30) Peterson, R. L.; Himes, R. A.; Kotani, H.; Suenobu, T.; Tian, L.; Siegler, M. A.; Solomon, E. I.; Fukuzumi, S.; Karlin, K. D. *J. Am. Chem. Soc.* **2011**, *133*, 1702–1705.
- (31) Bordwell, F. G.; Zhang, X.-M. *J. Phys. Org. Chem.* **1995**, *8*, 529–535.
- (32) Maiti, D.; Lee, D.-H.; Gaoutchenova, K.; Würtele, C.; Holthausen, M. C.; Sarjeant, A. A. N.; Sundermeyer, J.; Schindler, S.; Karlin, K. D. *Angew. Chem., Int. Ed.* **2008**, *47*, 82–85.
- (33) It is only for the *p*-alkyl-DTBP cases that it was convenient to take a large number of full spectra, as in Figure 3a, to follow the kinetics. For all other cases, reactions which were much faster; only the 409 nm absorption value changes were monitored to deduce pseudo-first-order rate constants.
- (34) (a) Francisco, W. A.; Merkler, D. J.; Blackburn, N. J.; Klinman, J. P. *Biochemistry* **1998**, *37*, 8244–8252. (b) Miller, S. M.; Klinman, J. P. *Biochemistry* **1983**, *22*, 3091–3096.
- (35) (a) Kunishita, A.; Kubo, M.; Sugimoto, H.; Ogura, T.; Sato, K.; Takui, T.; Itoh, S. *J. Am. Chem. Soc.* **2009**, *131*, 2788–2789. (b) Kunishita, A.; Ertem, M. Z.; Okubo, Y.; Tano, T.; Sugimoto, H.; Ohkubo, K.; Fujieda, N.; Fukuzumi, S.; Cramer, C. J.; Itoh, S. *Inorg. Chem.* **2012**, *51*, 9465–9480.
- (36) Nemes, A.; Bakac, A. *Inorg. Chem.* **2001**, *40*, 746–749.
- (37) Fukuzumi, S.; Shimoosako, K.; Suenobu, T.; Watanabe, Y. *J. Am. Chem. Soc.* **2003**, *125*, 9074–9082.
- (38) Pratt, D. A.; Dilabio, G. A.; Mulder, P.; Ingold, K. U. *Acc. Chem. Res.* **2004**, *37*, 334–340.
- (39) If the ET step is exergonic, this should occur first, followed by rate-determining PT. In such a case, phenol radical cation derivatives should be observed, but they were not. If an exergonic ET were the rate-determining step, followed by rapid PT, no KIE would be observed in contrast to our experimental observations. We further note that $[\text{Cu}^{\text{II}}(\text{O}_2^{\bullet-})]^+$ complex **2** is unreactive to the strong reductant decamethylferrocene.
- (40) (a) Matsumoto, T.; Ohkubo, K.; Honda, K.; Yazawa, A.; Furutachi, H.; Fujinami, S.; Fukuzumi, S.; Suzuki, M. *J. Am. Chem. Soc.* **2009**, *131*, 9258–9267. (b) Marcus, R. A. *J. Phys. Chem.* **1966**, *72*, 891.
- (41) (a) Guttenplan, J. B.; Cohen, S. G. *J. Am. Chem. Soc.* **1972**, *94*, 4040–4042. (b) Wagner, P. J.; Lam, H. M. H. *J. Am. Chem. Soc.* **1980**, *102*, 4167–4172.
- (42) (a) Lockwood, M. A.; Blubaugh, T. J.; Collier, A. M.; Lovell, S.; Mayer, J. M. *Angew. Chem., Int. Ed.* **1999**, *38*, 225–227. (b) Yiu, D. T. Y.; Lee, M. F. W.; Lam, W. W. Y.; Lau, T.-C. *Inorg. Chem.* **2003**, *42*, 1225–1232. (c) Lansky, D. E.; Goldberg, D. P. *Inorg. Chem.* **2006**, *45*, 5119–5125. (d) Miyazaki, S.; Kojima, T.; Mayer, J. M.; Fukuzumi, S. *J. Am. Chem. Soc.* **2009**, *131*, 11615–11624.
- (43) (a) Kohen, A.; Klinman, J. P. *Acc. Chem. Res.* **1998**, *31*, 397–404. (b) Kwart, H. *Acc. Chem. Res.* **1982**, *15*, 401–408.
- (44) Francisco, W. A.; Knapp, M. J.; Blackburn, N. J.; Klinman, J. P. *J. Am. Chem. Soc.* **2002**, *124*, 8194–8195.
- (45) (a) The smaller yield of H_2O_2 than expected from Scheme 4 may result from partial decomposition during the reaction. (b) The DMM-tmpa ligand in Cu(II) reaction products remains intact (based on direct TLC and MS analyses); no ligand oxidation occurred.
- (46) In fact, we were not able to detect isobutylene as a product; see the Experimental Section for details and explanation.
- (47) (a) Wali, A.; Das, J.; Pillai, S. M.; Ravindranathan, M. *Green Chem.* **2002**, *4*, 587–591. (b) Mandal, S.; Macikenas, D.; Protasiewicz, J. D.; Sayre, L. M. *J. Org. Chem.* **2000**, *65*, 4804–4809. (c) Nishinaga, A.; Itahara, T.; Shimizu, T.; Matsuura, T. *J. Am. Chem. Soc.* **1978**, *100*, 1820–1825. (d) Peters, A.; Trumm, C.; Reinmuth, M.; Emeljanenko, D.; Kaifer, E.; Himmel, H.-J. *Eur. J. Inorg. Chem.* **2009**, *2009*, 3791–

3800. (e) Gupta, R.; Mukherjee, R. *Tetrahedron Lett.* **2000**, *41*, 7763–7767. (f) Cvetkovic, M.; Batten, S. R.; Moubaraki, B.; Murray, K. S.; Spiccia, L. *Inorg. Chim. Acta* **2001**, *324*, 131–140. (g) Gupta, M.; Upadhyay, S. K.; Sridhar, M. A.; Mathur, P. *Inorg. Chim. Acta* **2006**, *359*, 4360–4366. (h) Singla, M.; Mathur, P.; Gupta, M.; Hundal, M. S. *Transition Met. Chem.* **2008**, *33*, 175–182. (i) Mukherjee, C.; Weyhermüller, T.; Bothe, E.; Chaudhuri, P. *C. R. Chimie* **2007**, *10*, 313–325. (j) Singh, A. P.; Gupta, R. *Eur. J. Inorg. Chem.* **2010**, 4546–4554. (k) Bakshi, R.; Mathur, P. *Inorg. Chim. Acta* **2010**, *363*, 3477–3488. (l) Ahuja, G.; Mathur, P. *Inorg. Chem. Commun.* **2012**, *17*, 42–48.

(48) For the substrate *p*-Me-DTBP, which was studied for its kinetics (Table 1), there is a question as to what the final organic product could be. The mechanism applied to the *p*-^tBu-DTBP substrate which would eliminate isobutylene, cannot apply to *p*-Me-DTBP; methanol or a Cu(II)–OMe species might be formed, as suggested by one reviewer, but a detailed analysis was not performed.

(49) (a) Damuel, D.; Silver, B. L. *Adv. Phys. Org. Chem.* **1965**, *3*, 123–186. (b) Kawanishi, Y.; Suzuki, Y.; Miyazawa, A. *Chem. Eng. J.* **2011**, *167*, 531–535.

(50) Upon benchtop mixing of **2** and ^tBu₃ArO[•] (less than 1 min, with 1:1 ratios of reagents), the reaction is already over by the time the first UV–vis spectrum could be recorded, as indicated by the complete loss of the UV–vis signals ascribed to **2** (Figure 3) and an expected 402 nm peak for ^tBu₃ArO[•]. It is outside the scope of this study to attempt stopped-flow (triple mixing) experiments which would be required.

(51) (a) Nishinaga, A.; Tomita, H.; Matsuura, T. *Tetrahedron Lett.* **1980**, *21*, 3407–3408. (b) Nishinaga, A.; Tomita, H.; Nishizawa, K.; Matsuura, T.; Ooi, S.; Hirotsu, K. *J. Chem. Soc., Dalton Trans.* **1981**, 1504–1514.

(52) Company, A.; Yao, S.; Ray, K.; Driess, M. *Chem.—Eur. J.* **2010**, *16*, 9669–9675.

(53) Güell, M.; Luis, J.; Solà, M.; Siegbahn, P. *J. Biol. Inorg. Chem.* **2009**, *14*, 229–242.