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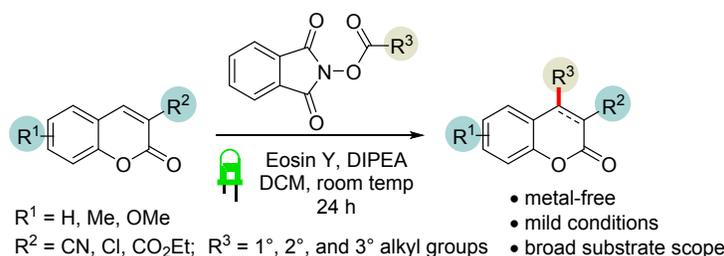
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Organo Photoinduced Decarboxylative Alkylation of Coumarins with N-(Acyloxy)phthalimide

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Abstract: A metal free and mild, photo-induced decarboxylative 4-position alkylation of coumarins has been reported. Photo-induced single electron transfer has been initiated by utilizing the visible-light absorptivity of Eosin Y for a reductive generation of alkyl radicals from N-(acyloxy)phthalimide esters. Depending on the nature of N-(acyloxy)phthalimide esters (primary, secondary, and tertiary carboxylic acid derived) several saturated and unsaturated C-4 alkylated coumarins were synthesized. Both control experiments and photophysical studies supported a radical based mechanism for the selective alkylation.

Visible light catalyzed selective construction of carbon-carbon bond has started to garner attention from chemists recently.¹ The attractiveness of such selective photoreactions are attributed to the synthetic utility of a photo active catalyst, which on excitation by appropriate wavelength functions as single electron or energy transfer source to yield free radical intermediates.² Many metals based homogeneous photo redox catalysts such as ruthenium and iridium complexes, and heterogeneous semiconductor catalyst have been

extensively explored for various synthetic transformations.³ On the contrary, while many organic dyes have demonstrated photocatalytic activities, their use as catalyst for reactions that enables previously inaccessible synthetic transformations are scarce in selective organic synthesis.⁴ For example, it has been reported that aliphatic carboxylic acids and their derivatives can generate a corresponding alkyl radical by photo-redox mediated single electron transfer and CO₂ extrusion mechanism. Typically, the alkyl radical can be generated either by esterification of the acid followed by reductive cleavage of the esters^{5a-c} or by direct oxidative cleavage of the acid.^{5d-f} Similar reactions that uses the low energy visible light has been utilized to achieve photocatalytic decarboxylative reactions such as arylations,^{6a} fluorinations,^{6b} vinylations,^{6c} allylations^{6d} and alkynylations.^{6e} It has been demonstrated that *in situ* generated radicals can then react with the activated alkene to yield corresponding analogues⁷. Recently, Eosin Y has been extensively used as a catalyst to achieve various photochemical transformations.

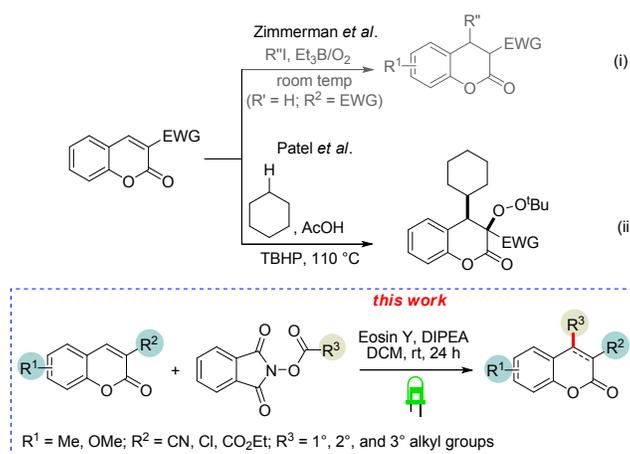
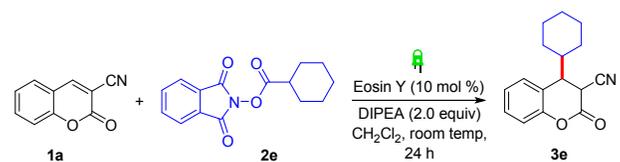


Fig. 1. Previous Contributions and Current Approach for Alkylation of Coumarins.

König and coworkers have demonstrated, Eosin Y catalyzed decarboxylative alkylation of acrylates from N-(acyloxy)phtalimide esters.^{5a} Interestingly, while one can find many examples of alkylation of monosubstituted and disubstituted alkene, the trisubstituted alkene alkylations are limited. Addressing the

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2
3 lacuna, in our present protocol, we have attempted to explore the reactivity of the radical intermediates
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5 generated by photocatalytic decarboxylation of N-(acyloxy)phthalimide esters with 3-substituted
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7 coumarins and its analogues. Coumarins are naturally occurring compounds and have utility in organic
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9 materials for optical property^{8a-c} and in pharmaceuticals for anti-inflammatory, anti-oxidant, antimicrobial
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11 and anti-cancer bio-activities.^{8d-g} Various approaches such as nucleophilic addition to Michael acceptor,
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13 direct C-H functionalization through oxidative cross coupling reaction with metal or without metal have
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15 been explored for the synthesis of coumarin derivatives.⁹ However, the alkylation of coumarins yielding
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17 chain alkylated derivatives and other highly functionalized scaffold installation through alkylation *via*
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19 oxidative coupling reaction is still an arduous task. To the best of our knowledge, thus far there is no
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21 report on such alkylation of coumarin at C-4 position except Zimmerman's report,^{10a} which could provide
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23 the alkylation of coumarins by a metal free approach (Fig. 1, eq. i). Recently, Patel^{10b} demonstrated a
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25 similar alkylation method of coumarin using TBHP as an oxidant ended in di-functionalization (an
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27 alkylation at C-4 and peroxidation at C-3) (Fig. 1, eq. ii). In all the reports while it's commendable that
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29 the reactions were direct and without any side products the limited substrate scopes and high temperature
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31 reactions limits them for post synthesis functional group manipulations. While preparing our manuscript,
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33 Sun¹¹ reported visible-light induced Ir catalyzed regioselective, decarboxylative C-3 alkylation of
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35 coumarin, although with limited substrate scope. In the present work we are reporting a photocatalyzed
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37 approach to access a library of alkylated coumarins where several phthalimide esters have been used as
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39 source of alkyl radicals. Interestingly, we could isolate the saturated and unsaturated products depending
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41 on the nature of the phthalimide esters used.
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Table 1.^a Selected Optimization Reaction Condition Results.


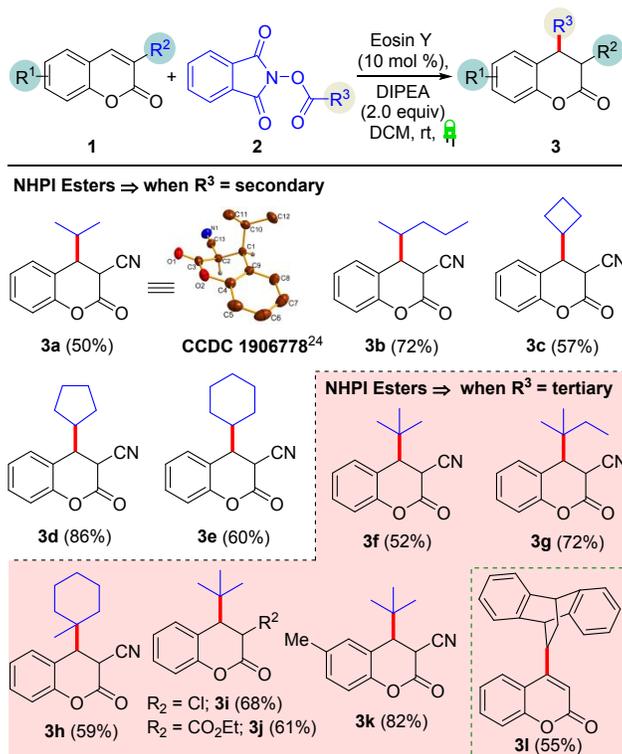
entry	deviation from above	yield (%) ^b
1	none	60
2	in the absence of Eosin Y	ND
3	in the absence of DIPEA	ND
4	in the absence of light	ND
5	5 mol % Eosin Y	42
6	5 mol % of Ru(bpy) ₂ PF ₆ instead of Eosin Y	30
7	10 mol % of Mes-Acr ⁺ instead of Eosin Y	ND
8	10 mol % of Rose Bengal instead of Eosin Y	20
9		35
10	CH ₃ CN instead of CH ₂ Cl ₂	ND
11	DMF instead of CH ₂ Cl ₂	22
12	CHCl ₃ instead of CH ₂ Cl ₂	45
13	C ₂ H ₄ Cl ₂ instead of CH ₂ Cl ₂	ND
	Et ₃ N instead of DIPEA	ND

^aReaction Conditions **1a:2e** = 1.2:1, for 0.2 mmol scale, solvent 3mL. ^bIsolated yield. DIPEA = N,N-Diisopropylethylamine, ND = not detected, Mes-Acr⁺ = 9-Mesityl-10-methylacridinium tetrafluoroborate.

Our study was initiated by an investigation of the proposed alkylation of 3-cyanocoumarins (**1**) and N-(acyloxy) phthalimide of cyclohexyl carboxylic acid (**2e**) as model substrates for the decarboxylative alkylation in the presence of photoredox catalyst under a green LED irradiation. After extensive screening we were delighted to established that a combination of Eosin Y (10 mol %), DIPEA (2.0 equiv.) in dichloromethane (0.08 M) under photo-irradiation at room temperature was optimum, affording the desired product **3e** in 60% isolated yield (Table 1, entry 1). Control experiments indicated that deviation from the optimal condition led to either no product or significant drop in the yield of **3e**. Notably, in the absence of any reaction component such as Eosin Y or DIPEA or visible-light irradiation, no alkylated product was observed (Table 1, entry 2-4). Lowering of catalyst loading from 10 mol % to 5 mol % yielded lower conversion (Table 1, entry 5). Eosin Y was the only effective photocatalyst and with other photocatalyst no improvement in yield of the product was observed. The common Ru metallic photocatalyst (Table 1, entry 6) or other organo-photocatalysts such as Mes-Acr⁺ and Rose Bengal were

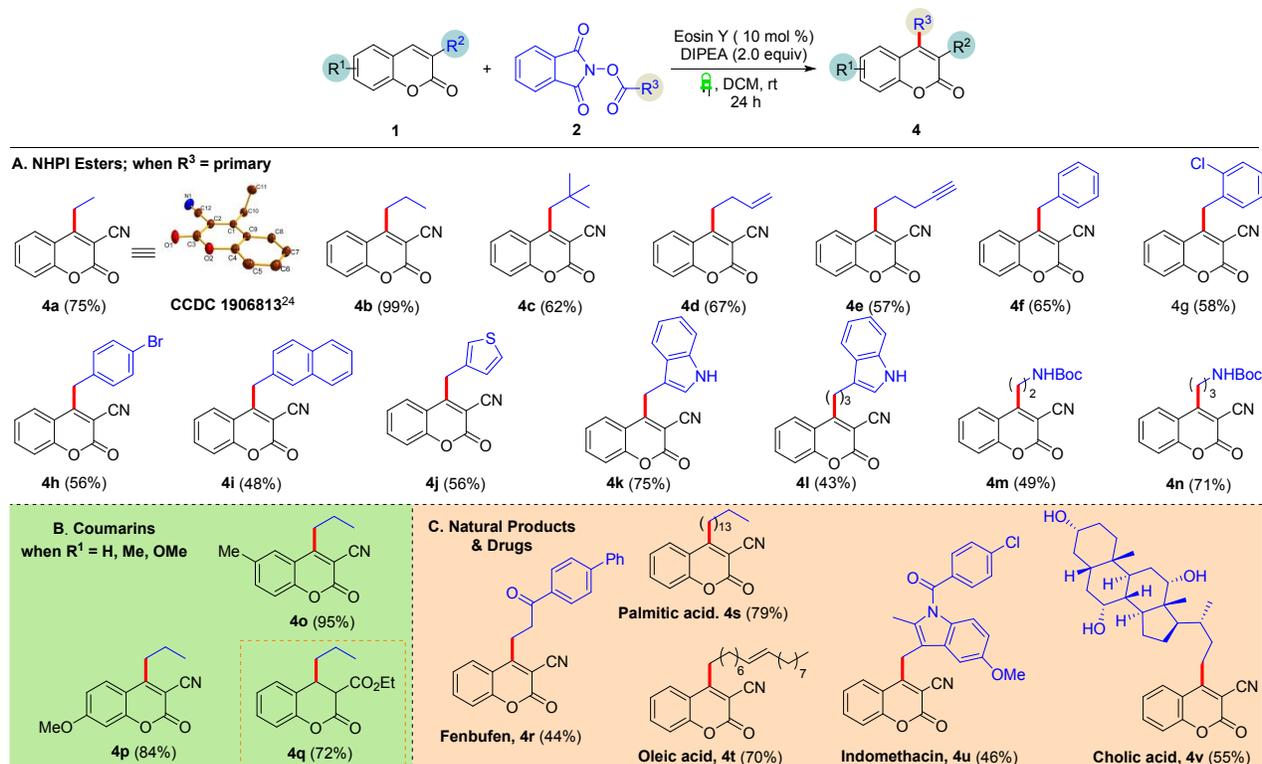
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3 least effective (Table 1, entry 7, 8). The choices of solvents played critical role in optimizing the reaction
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5 outcome. Switching reaction solvent from DCM to acetonitrile or CHCl_3 or DMF or DCE produced
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7 alkylated product in much lower yield (Table 1, entry 9-12). A survey of organic bases revealed that the
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9 DIPEA was the best choice. Replacing DIPEA with Et_3N yielded no product (Table 1, entry 13). The
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11 substrate scope, which ensued from the above optimized reaction conditions is summarized in Table 2
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13 and Table 3. Unexpectedly, secondary carboxylic acids derived NHPI esters when coupled with 3-
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15 substituted coumarins under optimized reaction conditions provided 3,4-substituted 2-chromanone
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17 instead of 3,4-substituted 2-chromenones. NHPI esters of isobutyric and 2,2-dimethylbutanoic acids
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19 provided 4-(2-propyl) and 4-(2-pentyl) substituted 3-cyano-2-chromanones **3a** and **3b** in 50% and 72%
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21 of isolated yields, respectively (Table 2). Structure of **3a** was unambiguously confirmed by single crystal
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23 XRD (CCDC no. 1906778). Secondary carboxylic acids appended to acyclic chains (cyclobutyl,
24
25 cyclopentyl and cyclohexyl) were converted into corresponding 4-alkylated 2-chromanones products with
26
27 57%-86% of isolated yields (Table 2, entries **3c-3e**). In lieu of above results, we tried pivalic, 2,2-
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29 dimethylbutanoic and 1-methyl-1-cyclohexane tertiary aliphatic carboxylic acids derived NHPI esters.
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31 Coherently, we obtained 52%-82% of isolated yield of 4-alkylated products when pivalic acid derived
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33 NHPI ester has been used with various coumarins {Table 2, $\text{R}^2 = \text{CN}$ (**3f**), Cl (**3i**) and CO_2Et (**3j**)}. NHPI
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35 esters of 2,2-dimethylbutanoic and 1-methyl-1-cyclohexane tertiary aliphatic carboxylic acids gave 72%
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37 and 59% of isolated yield of 4-alkylated 3-cyano-2-chromanones (Table 2, entry **3g**, **3h**). NHPI ester
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39 derived from 9,10-dihydro-9,10-ethanoanthracene-11-carboxylic acid gave anomalous result in which we
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41 observed unsaturation with elimination of nitrile group (Table 2, entry **3l**, yield 55%).
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Table 2.^a Scope of decarboxylative 4-position alkylation of coumarins with various secondary and tertiary carboxylic acid derived NHPI esters.



^aReaction conditions: All the reactions were carried out with 0.24 mmol of **1** and 0.2 mmol of **2** in the presence of 10 mol % of Eosin Y in 3 mL of DCM at room temperature under green light irradiation.

Table 3.^a Scope of decarboxylative 4-position alkylation of coumarins with various primary carboxylic acid derived NHPI esters.

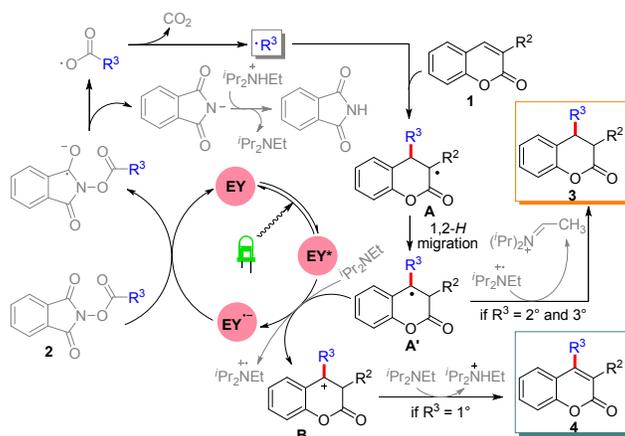


^aReaction conditions: All the reactions were carried out with 0.24 mmol of **1** and 0.2 mmol of **2** in presence of 10 mol% of Eosin Y in 3 mL of DCM at room temperature under green light irradiation.

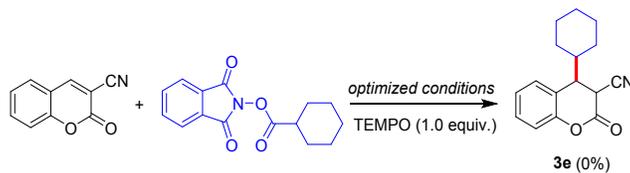
Primary carboxylic acid derived NHPI esters produced surprising results when used with 3-substituted coumarins under this visible-light mediated decarboxylative alkylation strategy (Table 3). In this process, primary carboxylic acids of alkyl, branched alkyl, β -positioned alkene, γ -positioned alkyne, benzylic substrates, substrates bearing extended aromatics, heterocycles (thiophene, indole) and amino acids (β -alanine and γ -alanine) were converted into the corresponding 4-alkylated-3-cyanocoumarins in good to high yields *via* their phthalimide esters (Table 3, Part A, entry **4a-4n**). Functional variations on coumarins itself with electron donating 6-Me and 7-OMe groups supported this alkylation well with yields up to 95%

(Table 3, Part B, entry **4o**, **4p**). In case when we made an attempt to alkylate ethyl 3-coumarincarboxylate, we observed saturated instead of unsaturated coumarin **4q** in 72% yield (Table 3, Part B).

Scheme 1. Plausible mechanism of alkylation.



Scheme 2. Control Experiment.



To showcase the utility of the transformation for diversifying natural-products and drugs carboxylic acids, we prepared a diverse collection of *N*-(acyloxy)phthalimide esters and subjected them to the optimized reaction conditions (Table 3, Part C, entry **4r-4v**, 44%-79% yield). The proposed mechanism illustrated in Scheme 1 can be rationalized by spectroscopic and electrochemical studies performed on control reactions (Supporting Information) and based on similar observations in the literature.^{4b,5a} Photo-irradiation *via* the green LED, excites Eosin Y leading to single electron transfer (SET) from DIPEA (0.72 V vs SCE in CH₃CN) to excited Eosin Y* (Eosin Y*/Eosin Y^{•-} = 0.83 V vs SCE in CH₃CN-H₂O, 1:1) (Scheme 1). Following this, regeneration of Eosin Y take place by single electron reduction of NHPI ester (-1.20 V vs SCE in CH₃CN-H₂O, 1:1). The reduced ester produces [•]R³ after breaking of the N–O bond

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2 and subsequent elimination of CO₂. To confirm the formation of radical $\cdot R^3$, we performed radical
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4 trapping experiment with 1.0 equiv. of TEMPO. Presence of radical trapper inhibited the reaction and no
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6 product formation was observed (Scheme 2), thus, confirming the radical pathway. The $\cdot R^3$ is then added
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8 on coumarin and generates an open radical **A**. Further, intramolecular 1,2-hydrogen migration took place
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10 which facilitated the generation of 4-positioned benzylic radical **A'**. We hypothesized two possible
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12 pathways of terminations. First, if $\cdot R^3$ is secondary or tertiary then oxidized DIPEA will transfer H \cdot to **A'**
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14 and consequently formation of **3** will be observed. Secondly, if $\cdot R^3$ is primary then **A'** will get oxidized
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16 by Eosin Y* and **B** will be formed, which will produce **4** after abstraction of H⁺ by DIPEA. Since Eosin
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18 Y has a fluorescence emission ($\lambda_{em} = 550\text{nm}$, $\lambda_{ex} = 520\text{nm}$) it can be used to study the structural changes
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20 in Eosin Y by observing the changes in the fluorescence emission. When the Fluorescence spectra of
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22 Eosin Y and NHPI were recorded no significant difference in the fluorescence peak (Supporting
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24 Information, Figure S1) was observed suggesting very poor quenching due to any electron transfer.
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26 Similarly addition of coumarin to Eosin Y and NHPI mixture did not alter the fluoresce emission
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28 (Supporting Information, Figure S1). However, when the same reaction was monitored with addition of
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30 DIPEA, a significant quenching of the fluorescence emission was observed over time with almost no
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32 emission after 8 h (Supporting Information, Figure S2), thus, suggesting an electron transfer with DIPEA.
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39 In conclusion, we have developed a mild, metal-free, low energy (visible-light) mediated organo-
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41 photocatalyzed strategy for 4-alkylation of 3-substituted coumarins. Primary carboxylic acids derived
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43 NHPI esters produced 4-alkylated 3-substituted saturated coumarins while those of secondary and tertiary
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45 carboxylic acids produced 4-alkylated 3-substituted unsaturated coumarins. We believe that developed
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47 methodology offers a powerful post-synthetic modification route for generating various new classes of
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49 highly functionalized small molecules.
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52 53 **Experimental Section** 54 55 56 57 58 59 60

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2 **General Information.** The *N*-(acyloxy)phthalimide esters have been prepared from the commercially
3 available corresponding carboxylic acids and *N*-hydroxyphthalimides. Nuclear magnetic resonance
4 (NMR) spectra were recorded in deuterated solvents with residual protonated solvent signal as internal
5 reference on a BrückerAva-300, BrückerAva-400 and BrückerAva-500. Chemical shifts are reported in
6 parts per million using the solvent resonance internal standard (chloroform, 7.26 and 77.0 ppm or DMSO,
7 2.50 and 40.0 ppm). Data is reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t =
8 triplet, q = quartet, m = multiplet, bs = broad singlet), coupling constant, and integration. Electrospray
9 and electron impact high resolution mass spectrometry was performed by Brücker mass spectrometer.
10 The data is recorded as the ionization method followed by the calculated and measured masses. Solvents
11 for starting material preparation and coupling reactions were dried before use. Green LEDs (2.50 W, λ =
12 530 nm) Rebel LED, mounted on a 25 mm cool base was purchased from commercial supplier Luxeon
13 Star LEDs Quadica Developments Inc. 10-3447 30 Ave N. Lethbridge, Alberta T1H 7B5 Canada. No filter
14 has been used. Glass material of round-bottom flask is of borosilicate. Distance of RB from LED was
15 kept fixed at 5cm (see Supporting Information for photoreaction set-up).
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35 **General Method for the Synthesis of Starting Materials Coumarins (1) and N-(Acyloxy)phthalimide** 36 **Esters (2)**

37 **2.1. Preparation of Coumarins**

38 **2.1.1. Preparation of 3-Cyanocoumarins (1a-c)**

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42 3-Cyanocoumarins were synthesized using known literature procedure.¹² To a solution of salicylaldehyde
43 (10.0 mmol) and malononitrile (660 mg, 10.0 mmol) in EtOH (10.0 mL) was added NH₄OAc (462 mg,
44 6.0 mmol) slowly. The reaction mixture was stirred at room temperature for 4h to form the precipitations.
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2 The resulting reaction mixture was vigorously stirred at 75 °C in oil bath for 30 min. Subsequently, it was
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4 poured in ice water. A solid precipitate was obtained. The resulting solid was filtered, washed with cold
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6 water and recrystallized in EtOH, yielded corresponding 3-cyanocoumarins.
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10 **2.1.2. Synthesis of ethyl coumarin-3-carboxylate (1d)**

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12 Ethyl coumarin-3-carboxylate **1d** were synthesized following known literature procedure.¹³ To a solution
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14 of salicylaldehyde (1.83 g, 15 mmol) in 30 mL of ethanol was added diethyl malonate (2.88 g, 18 mmol),
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16 2 drops of acetic acid and piperidine (0.15 mL). The reaction mixture was stirred under reflux for 8 h, and
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18 dilution was done with ice-cold water (50 mL). After completion of the reaction, the cake was filtered and
19
20 washed with cold water. The product was dried under vacuum to give a white solid (3.07 g, 94% yield).
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24 **2.1.3. Synthesis of 3-Chlorocoumarin (1e)**

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26 3-Chlorocoumarin (**1e**) was synthesized following known literature procedure.¹⁴ To a solution of
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28 coumarin (10 mmol) in 1M HCl-DMF (15 mL) solution was added *m*-CPBA (15 mmol). The reaction
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30 mixture was stirred at rt for 30 min. After completion, reaction was quenched with 1M NaHCO₃ and
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32 extracted with Et₂O (3 x 50 mL) after removal of solvent in vacuo, the product was purified by silica gel
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34 chromatography using EtOAc-hexane (1:4) as eluent.
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38 **2.2. Preparation of *N*-(acyloxy)phthalimides (2)**

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40 All *N*-(Acyloxy)phthalimides (**2a-2o**, **2q-2t**, **2w-2z**, and **2za**) were synthesized by following known
41
42 literature procedures.^{5a,15-23} The NHPI esters **2p**, **2u**, **2v**, and **2zb** were synthesized by following the known
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44 method of Koenig^{5a} in which we took respective carboxylic acids (4.0 mmol, 1.0 equiv.), *N*-
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46 hydroxyphthalimide (721 mg, 4.40 mmol, 1.1 equiv.), *N,N'*-dicyclohexylcarbodiimide (988 mg, 4.8
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48 mmol, 1.2 equiv.) and 4-dimethylaminopyridine (48 mg, 0.80 mmol, 0.1 equiv.) in a round bottom flask.
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50 Dry THF (20 mL) was added and the orange reaction mixture was stirred for 15 h at room temperature.
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52 After completion of reaction the white precipitate was filtered off and the solution was concentrated by
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2 evaporation of the solvent. Purification by column chromatography on silica gel (EtOAc: hexane = 3:7)
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4 gave a solid **2**. All characterization data for **2p**, **2u**, **2v**, and **2zb** are reported.
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7 **3. General Methods of Preparations for the Synthesis of 3 and 4**

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9
10 0.2 mmol of *N*-(acyloxy)phthalimides (1.0 equiv.), coumarin (0.24 mmol, 1.2 equiv.) were taken in a long
11 neck round bottom flask and 10 mol % of Eosin Y was added into it, the RB was capped with septum.
12 After that 2.0 equiv. of DIPEA and then 3 ml of dry DCM were added into the reaction mixture via
13 syringe. The mixture was degassed and filled with N₂ (three times). The reaction mixture was irradiated
14 with green LED for 24 h. After completion (monitored through TLC), reaction was quenched with
15 saturated NaHCO₃ solution and extracted with DCM (3 x 10 mL), washed with brine solution. After
16 removal of solvent in vacuo, the product was purified by silica gel chromatography using EtOAc-hexane
17 (1:9 to 4:6) as eluent to provide the desired product.
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28 **1,3-Dioxoisindolin-2-yl 2-(2-chlorophenyl)acetate (2p)**. Physical state: White solid; Yield: 856.8 mg
29 (68%). ¹H NMR (300 MHz, CDCl₃) δ 7.89 – 7.87 (m, 2H), 7.80 – 7.77 (dd, *J* = 5.4, 2.8 Hz, 2H), 7.42 (s,
30 2H), 7.30 – 7.25 (m, 2H), 4.15 (d, *J* = 2.6 Hz, 2H). ¹³C{¹H} NMR (75 MHz, CDCl₃) δ 166.9, 161.8,
31 134.9, 129.8, 129.5, 129.0, 127.3, 124.1, 35.8. HRMS (ESI/QTOF), *m/z*: [M+Na]⁺ Calcd for
32 C₁₆H₁₀ClNO₄Na, 338.0190; Found: 338.0180.
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41 **1,3-Dioxoisindolin-2-yl 4-(1H-indol-3-yl)butanoate (2t)**. Physical state: yellow solid; Yield: 1.08 g,
42 (78%). ¹H NMR (300 MHz, CDCl₃) δ 8.05 (s, 1H), 7.90 – 7.87 (m, 2H), 7.79 – 7.77 (m, 2H), 7.63 (d, *J*
43 = 7.7 Hz, 1H), 7.37 (d, *J* = 7.9 Hz, 1H), 7.26 – 7.07 (m, 3H), 2.94 (t, *J* = 7.2 Hz, 2H), 2.70 (t, *J* = 7.1 Hz,
44 2H), 2.31 – 2.04 (m, 2H). ¹³C{¹H} NMR (75 MHz, CDCl₃) δ 169.8, 162.1, 136.5, 134.8, 129.0, 127.4,
45 124.0, 122.1, 119.4, 114.7, 111.3, 30.3, 25.1, 24.0. HRMS (ESI/QTOF), *m/z*: [M+Na]⁺ Calcd for
46 C₂₀H₁₆N₂O₄Na, 371.1002; Found: 371.0990.
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2 **1,3-Dioxoisindolin-2-yl 3-(tert-butoxycarbonylamino)propanoate (2v).** Physical state: white solid;
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4 **Yield:** 1.01 g (76%). ¹H NMR (300 MHz, CDCl₃) δ 7.90- 7.71 (m, 4H), 5.15 (s, 1H), 3.55 (d, *J* = 5.8
5
6 Hz, 2H), 2.90 (t, *J* = 5.9 Hz, 2H), 1.44 (s, 9H). ¹³C{¹H} NMR (75 MHz, CDCl₃) δ 168.6, 161.8, 155.8,
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8 134.9, 128.8, 124.0, 79.8, 36.1, 32.1, 28.3. HRMS (ESI/QTOF), *m/z*: [M+Na]⁺ Calcd for
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10 C₁₆H₁₈N₂O₆Na, 357.1057; Found: 357.1045.

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14 **2-Oxo-4-((3*S*)-3-((3*S*,5*R*,7*S*,8*S*,10*R*,12*R*,13*S*,14*R*,17*S*)-3,7,12-trihydroxy-10,13-**
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16 **dimethylhexadecahydro-1*H*-cyclopenta[*a*]phenanthren-17-yl)butyl)-2*H*-chromene-3-carbonitrile**
17
18 **(2zb).** Physical state: white solid; **Yield:** 1.15 g (52%). ¹H NMR (300 MHz, CDCl₃) 7.88 (s, 2H), 7.79
19
20 (s, 2H), 4.00 (s, 1H), 3.85 (s, 1H), 3.45 (s, 1H), 2.64 (t, *J* = 16.4 Hz, 2H), 2.26 – 2.22 (m, 3H), 1.92 - 1.81
21
22 (m, 7H), 1.81 – 1.67 (m, 6H), 1.56 (s, 6H), 1.42 -1.31 (m, 3H), 1.24 (d, *J* = 7.1 Hz, 1H), 1.13 (d, *J* = 13.3
23
24 Hz, 2H), 0.89 (s, 3H), 0.72 (s, 3H). ¹³C{¹H} NMR (75 MHz, CDCl₃) δ 170.2, 162.0, 134.7, 128.9, 123.9,
25
26 73.1, 71.9, 68.4, 60.4, 46.8, 41.5, 39.5, 35.4,35.1, 34.8, 30.7, 28.1, 27.5, 26.3, 22.3, 22.4, 21.0, 17.3, 14.2,
27
28 12.5. HRMS (ESI/QTOF), *m/z*: [M+Na]⁺ Calcd for C₃₂H₄₃NO₇Na, 576.2931; Found: 576.2916.
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33
34 **4-Isopropyl-2-oxochromane-3-carbonitrile (3a).** Physical state: White solid; **Yield:** 21.50 mg (50%).
35
36 ¹H NMR (300 MHz, CDCl₃) δ 7.36 – 7.32 (dd, *J* = 10.4, 4.2 Hz, 1H), 7.23 – 7.11 (m, 3H), 4.08 (d, *J* =
37
38 6.3 Hz, 1H), 3.30 (dd, *J* = 6.1, 4.0 Hz, 1H), 2.49-2.39 (m, 1H), 1.09 (d, *J* = 6.9 Hz, 3H), 0.80(d, *J* = 6.8
39
40 Hz, 3H). ¹³C{¹H} NMR (75 MHz, CDCl₃) δ 161.9, 151.1, 129.8, 129.8, 125.0, 120.2, 117.5, 114.0, 44.6,
41
42 38.1, 30.6, 21.5, 16.8. HRMS (ESI/QTOF), *m/z*: [M+Na]⁺ Calcd for C₁₃H₁₃NO₂Na, 238.0838; Found:
43
44 238.0832.
45
46
47

48 **2-Oxo-4-(pentan-2-yl)chromane-3-carbonitrile (3b).** Physical state: White solid; **Yield:** 35 mg (72%).
49
50 ¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.32 (m, 1H), 7.25 – 7.15 (m, 2H), 7.15 – 7.09 (m, 1H), 4.06 (t, *J*
51
52 = 5.2 Hz, 1H), 4.01 (s, 1H), 3.41 (dd, *J* = 6.4, 3.2 Hz, 1H), 3.36 – 3.31 (m, 0.21H), 3.13 (d, *J* = 7.4 Hz,
53
54
55

0.50H), 2.27 (s, 1H), 1.67 (s, 3H), 1.49 – 1.35 (m, 6H), 1.21 (m, 4H), 1.08 (d, $J=6.9$ Hz, 1H), 0.99 – 0.92 (m, 3H), 0.87 (m, 5H), 0.76 (d, $J=6.8$ Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 161.9, 151.2, 150.7, 129.6, 125.1, 120.3, 117.5, 111.3, 46.3, 43.2, 38.2, 37.4, 35.4, 20.4, 14.1, 14.0. HRMS (ESI/QTOF), m/z : $[\text{M}+\text{Na}]^+$ Calcd for $\text{C}_{15}\text{H}_{17}\text{NO}_2\text{Na}$, 266.1151; Found: 266.1148.

4-Cyclobutyl-2-oxochromane-3-carbonitrile (3c). Physical state: Brown gum; Yield: 25.88 mg (57%).

^1H NMR (300 MHz, CDCl_3) δ 7.39 – 7.33 (m, 1H), 7.24-7.19 (m, 2H), 7.16-7.10 (m, 1H), 3.98 (d, $J=5.2$ Hz, 1H), , 3.27 (dd, $J=7.9, 5.2$ Hz, 1H), , 2.79-2.74 (m, 1H), 2.26 - 2.17 (m, 1H), 1.99-1.82 (m, 3H), 1.71 (bs, 1H), 0.98 – 0.80 (m, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 161.1, 150.5, 129.7, 128.8, 125.3, 122.2, 117.5, 113.9, 43.8, 37.5, 37.4, 27.3, 26.1, 18.7. HRMS (ESI/QTOF), m/z : $[\text{M}+\text{Na}]^+$ Calcd for $\text{C}_{14}\text{H}_{13}\text{NO}_2\text{Na}$, 250.0839; Found: 250.0843.

4-Cyclopentyl-2-oxochromane-3-carbonitrile (3d). Physical state: Brown gum; Yield: 41.45 mg (86%).

^1H NMR (500 MHz, CDCl_3) δ 7.39 – 7.36 (m, 1H), 7.29 – 7.21 (m, 2H), 7.18 - 7.13(m, 1H), 4.09 (d, $J=5.5$ Hz, 1H), 3.43 (t, $J=5.7$, Hz, 1H), 2.32-2.30 (m, 1H), 2.06-2.04 (m, 1H), 1.86 – 1.75 (m, 1H), 1.60 – 1.51 (m, 4H), 1.45 – 1.33 (m, 1H), 1.01 – 0.83 (m, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 101 MHz) δ 161.7, 159.8, 1508, 129.7, 129.4, 125.5, 117.5, 114.2, 42.1, 42.5, 39.0, 31.1, 29.4, 24.3. HRMS (ESI/QTOF), m/z : $[\text{M}+\text{Na}]^+$ Calcd for $\text{C}_{15}\text{H}_{15}\text{NO}_2\text{Na}$, 264.0995; Found: 264.0997.

4-Cyclohexyl-2-oxochromane-3-carbonitrile (3e). Physical state: White solid; Yield: 35.70 mg (70%).

^1H NMR (300 MHz, CDCl_3) δ 7.38 – 7.32 (m, 1H), 7.22 – 7.09 (m, 3H), 4.04 (dd, $J=4.4, 3.7$ Hz, 1H), 3.26 (dd, $J=6.0, 3.7$ Hz, 1H), 2.08 – 1.99 (m, 1H), 1.78 (t, $J=13.4$ Hz, 2H), 1.63 (s, 2H), 1.39 – 1.07 (m, 4H), 1.04 – 0.82 (m, 1H), 0.79 – 0.65 (m, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 75 MHz) δ 162.0, 151.0, 129.8, 129.7, 125.0, 121.7, 117.4, 114.0, 44.6, 40.4, 37.8, 31.5, 27.2, 26.2, 25.8, 25.7. HRMS (ESI/QTOF), m/z : $[\text{M}+\text{Na}]^+$ Calcd For $\text{C}_{16}\text{H}_{17}\text{NO}_2\text{Na}$, 278.1151; Found: 278.1160.

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2 **4-(tert-Butyl)-2-oxochromane-3-carbonitrile (3f)**. Physical state: White solid; Yield: 23.82 mg (52%).
3
4 ¹H NMR (300 MHz, CDCl₃) δ 7.43 – 7.33 (m, 1H), 7.26-7.23 (m, 2H), 7.25- 7.16 (m, 1H), 4.12 (s, 1H),
5
6 3.05 (s, 1H), 0.98 (s, 9H). ¹³C{¹H} NMR (75 MHz, CDCl₃) δ 160.6, 151.8, 131.4, 130.4, 125.9, 119.7,
7
8 117.7, 114.2, 51.8, 35.6, 33.4, 27.8. HRMS (ESI/QTOF), m/z: [M+Na]⁺ Calcd For C₁₄H₁₅NO₂Na,
9
10 252.0995; Found: 252.1004.
11
12

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14 **2-Oxo-4-(tert-pentyl)chromane-3-carbonitrile (3g)**. Physical state: White solid; Yield: 34.99 mg (72%).
15
16 ¹H NMR (300 MHz, CDCl₃) δ 7.42 – 7.37 (m, 1H), 7.25 (d, *J* = 5.4 Hz, 2H), 7.22 – 7.15 (m, 1H), 4.09
17
18 (s, 1H), 3.15 (s, 1H), 1.37 (dd, *J* = 15.0, 7.5 Hz, 2H), 0.94-0.87 (m, 9H). ¹³C{¹H} NMR (CDCl₃, 300
19
20 MHz) δ 160.7, 151.2, 131.4, 129.9, 125.1, 119.7, 117.6, 114.7, 49.6, 37.7, 33.5, 31.9, 24.2, 23.3, 8.2.
21
22 HRMS (ESI/QTOF), m/z: [M+Na]⁺ Calcd for C₁₅H₁₇NO₂Na, 266.1151; Found: 266.1148.
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27 **4-(1-Methylcyclohexyl)-2-oxochromane-3-carbonitrile (3h)**. Physical state: White solid; Yield: 34.45
28
29 mg (59%). ¹H NMR (500 MHz, CDCl₃) δ 7.41 – 7.38 (m, 1H), 7.26 – 7.20 (m, 2H), 7.16 (d, *J* = 8.2 Hz,
30
31 1H), 4.16 (s, 1H), 3.12 (s, 1H), 1.57-1.25 (m, 10H), 0.85 (s, 3H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ
32
33 160.8, 151.2, 131.6, 129.9, 125.0, 119.4, 117.6, 114.9, 37.5, 35.4, 35.0, 33.0, 25.7, 21.7, 21.4, 19.5.
34
35 HRMS (ESI/QTOF), m/z: [M+Na]⁺ Calcd for C₁₇H₁₉NO₂Na, 292.1308; Found: 292.1316.
36
37
38

39 **4-(tert-Butyl)-3-chlorochroman-2-one (3i)**. Physical state: White solid; Yield: 32.37 mg (68%). ¹H
40
41 NMR (300 MHz, CDCl₃) δ 7.32 (dd, *J* = 9.0, 8.2, 1H), 7.16-7.08 (m, 3H), 5.03 (d, *J* = 5.8 Hz, 1H), 3.18
42
43 (d, *J* = 5.7 Hz, 1H), 1.08 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 165.8, 150.6, 130.6, 129.4, 125.1,
44
45 124.3, 117.4, 56.8, 52.7, 35.9, 29.5. HRMS (ESI/QTOF), m/z: [M+Na]⁺ Calcd for C₁₃H₁₅ClO₂Na,
46
47 261.0653; Found: 261.0655.
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51 **Ethyl 4-(tert-butyl)-2-oxochromane-3-carboxylate (3j)**. Physical state: Brown gum; Yield: 36.47 mg
52
53 (61%). ¹H NMR (300 MHz, CDCl₃) δ 7.40 – 7.33 (m, 1H), 7.26 – 7.15 (m, 3H), 4.18 - 4.06 (m, 2H),
54
55

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2 4.05 (s, 1H), 3.22 (s, 1H), 1.06 (s, 12H). $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 168.0, 165.9, 151.6, 131.2,
3
4 129.0, 124.1, 121.4, 117.0, 62.3, 50.7, 48.7, 34.7, 27.4, 13.9. HRMS (ESI/QTOF), m/z : $[\text{M}+\text{Na}]^+$ Calcd
5 for $\text{C}_{16}\text{H}_{20}\text{O}_4\text{Na}$, 299.1254; Found: 299.1259.
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9
10 **4-(tert-Butyl)-6-methyl-2-oxochromane-3-carbonitrile (3k)**. Physical state: White solid; Yield: 39.85
11 mg (82%). ^1H NMR (300 MHz, CDCl_3) δ 7.17 (d, $J = 8.4$ Hz, 1H), 7.05- 6.96 (m, 2H), 4.09 (s, 1H), 2.99
12 (s, 1H), 2.36 (s, 3H), 0.97 (s, 9H). $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) δ 160.8, 149.0, 134.8, 131.8, 130.6,
13
14 (s, 1H), 2.36 (s, 3H), 0.97 (s, 9H). $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) δ 160.8, 149.0, 134.8, 131.8, 130.6,
15
16 119.4, 117.3, 114.8, 51.5, 35.0, 34.0, 27.30, 20.9. HRMS (ESI/QTOF), m/z : $[\text{M}+\text{Na}]^+$ Calcd for
17 $\text{C}_{15}\text{H}_{17}\text{NO}_2\text{Na}$, 266.1151; Found: 266.1151.
18
19

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21
22 **4-((9S,10S)-9,10-Dihydro-9,10-ethanoanthracen-11-yl)-2H-chromen-2-one (3l)**. Physical state: White
23 solid; Yield: 41.0 mg (55%). ^1H NMR (400 MHz, CDCl_3) δ 7.40-7.31 (m, 5H), 7.24 (s, 2H), 7.17 – 7.13
24 (m, 3H), 6.97 (dd, $J = 7.2, 1.8$ Hz, 2H), 5.78 (s, 1H), 4.51 (d, $J = 2.5$ Hz, 1H), 4.31 (d, $J = 1.7$ Hz, 1H),
25
26 3.63 – 3.59 (m, 1H), 2.94 – 2.89 (m, 1H), 2.07 – 2.01 (m, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ
27
28 160.61, 148.5, 144.5, 144.1, 143.7, 140.2, 138.0, 128.2, 126.3, 125.9, 125.8, 125.1, 125.0, 123.9, 123.8,
29
30 123.4, 123.1, 121.2, 117.3, 49.6, 44.6, 38.4, 28.7. HRMS (ESI/QTOF), m/z : $[\text{M}+\text{Na}]^+$ Calcd for
31
32 $\text{C}_{25}\text{H}_{18}\text{O}_2\text{Na}$, 373.1199; Found: 373.1223.
33
34
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38 **4-Ethyl-2-oxo-2H-chromene-3-carbonitrile (4a)**. Physical state: White solid; Yield: 29.85 mg (75%).
39
40 ^1H NMR (300 MHz, CDCl_3) δ 7.88 – 7.68 (m, 2H), 7.43 (t, $J = 7.8$ Hz, 1H), 3.14 (q, $J = 7.7$ Hz, 2H),
41
42 1.42 (t, $J = 7.7$ Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 168.1, 157.0, 154.0, 135.2, 125.9, 125.6,
43
44 118.1, 117.2, 113.3, 101.5, 25.4, 14.1. HRMS (ESI/QTOF), m/z : $[\text{M}+\text{Na}]^+$ Calcd for $\text{C}_{12}\text{H}_{10}\text{NO}_2$,
45
46 200.0706; Found: 200.0717.
47
48

49
50 **2-Oxo-4-propyl-2H-chromene-3-carbonitrile (4b)**. Physical state: White solid; Yield: 42.17 mg (99%).
51
52 ^1H NMR (400 MHz, CDCl_3) δ 7.76-7.68 (m, 2H), 7.43 – 7.40 (m, 2H), 3.10 (t, $J = 8.0$ Hz, 2H), 1.85-
53
54 1.79 (m, 2H), 1.16-1.12 (t, $J = 8.0$ Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 166.5, 157.0, 153.9,
55
56

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2 135.2, 126.05, 125.5, 118.1, 117.5, 113.5, 102.3, 33.8, 23.4, 14.3. **HRMS (ESI/QTOF), m/z:** [M+Na]⁺
3
4 Calcd for C₁₃H₁₁NO₂Na, 236.0682; Found: 236.0678.
5
6

7 **4-Neopentyl-2-oxo-2H-chromene-3-carbonitrile (4c).** **Physical state:** White solid; **Yield:** 29.88 mg
8
9 (62%). **¹H NMR (300 MHz, CDCl₃)** δ 7.80 (d, *J* = 7.3 Hz, 1H), 7.71 - 7.66 (m, 1H), 7.41-7.36 (m, 2H),
10
11 3.11 (s, 2H), 1.12 (s, 9H). **¹³C{¹H} NMR (101 MHz, CDCl₃)** δ 164.5, 157.0, 153.8, 135.0, 127.7, 124.9,
12
13 118.8, 117.9, 114.6, 103.9, 43.65, 35.7, 30.09. **HRMS (ESI/QTOF), m/z:** [M+Na]⁺ Calcd for
14
15 C₁₅H₁₅NO₂Na, 264.0995; Found: 264.1020.
16
17
18

19 **4-(But-3-en-1-yl)-2-oxo-2H-chromene-3-carbonitrile (4d).** **Physical state:** White solid; **Yield:** 33.23 mg
20
21 (67%). **¹H NMR (500 MHz, CDCl₃)** δ 7.76 – 7.69 (m, 2H), 7.45 – 7.26 (m, 2H), 5.90 (ddt, *J* = 17.0, 10.5,
22
23 6.8 Hz, 1H), 5.11 – 5.07 (m, 2H), 3.21 (t, *J* = 7.8 Hz, 2H), 2.52 (dd, *J* = 15.0, 7.0 Hz, 2H). **¹³C{¹H} NMR**
24
25 **(126 MHz, CDCl₃)** δ 165.6, 156.9, 153.8, 135.2, 135.1, 126.0, 125.6, 118.1, 117.6, 117.3, 113.4, 102.2,
26
27 33.6, 31.4. **HRMS (ESI/QTOF), m/z:** [M+Na]⁺ Calcd for C₁₄H₁₁NO₂Na, 248.0681; Found: 248.0682.
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32 **2-Oxo-4-(pent-4-yn-1-yl)-2H-chromene-3-carbonitrile (4e).** **Physical state:** White solid; **Yield:** 29.64
33
34 mg (57%). **¹H NMR (300 MHz, CDCl₃)** δ 7.84 (d, *J* = 8.0 Hz, 1H), 7.75-7.68 (m, 1H), 7.45-7.39 (m, 2
35
36 H), 3.25 (dd, *J* = 9.2, 7.2 Hz, 2H), 2.44 (td, *J* = 6.6, 2.5 Hz, 2H), 2.14 (dd, *J* = 3.3, 1.8 Hz, 1H), 1.96 (dt,
37
38 *J* = 16.1, 7.2 Hz, 2H). **¹³C{¹H} NMR (75 MHz, CDCl₃)** δ 166.9, 156.5, 153.8, 135.4, 126.0, 125.7, 118.0,
39
40 117.3, 113.3, 102.1, 82.5, 70.5, 30.8, 28.3, 18.8. **HRMS (ESI/QTOF), m/z:** [M+Na]⁺ Calcd for
41
42 C₁₅H₁₁NO₂Na, 260.0681; Found: 260.0667.
43
44
45

46 **4-Benzyl-2-oxo-2H-chromene-3-carbonitrile (4f).** **Physical state:** White solid; **Yield:** 33.93 mg (65%).
47
48 **¹H NMR (400 MHz, CDCl₃)** δ 7.75 (d, *J* = 8.0 Hz, 1H), 7.65 (t, *J* = 7.8, 1H), 7.40 (t, *J* = 8.2, 1H), 7.36-
49
50 7.24 (m, 6H), 4.48 (s, 2H). **¹³C{¹H} NMR (101 MHz, CDCl₃)** δ 163.9, 156.9, 154.0, 135.2, 135.0 129.4,
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2 128.3, 127.9, 127.0, 125.5, 118.0, 117.6, 113.7, 103.4, 37.7. **HRMS (ESI/QTOF), m/z:** [M+H]⁺ Calcd
3 for C₁₇H₁₂NO₂, 262.0863; Found: 262.0880.
4
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7 **4-(2-Chlorobenzyl)-2-oxo-2H-chromene-3-carbonitrile (4g).** Physical state: White solid; Yield: 33.93
8 mg (58%) ¹H NMR (500 MHz, CDCl₃) δ 7.69 – 7.64 (m, 1H), 7.53 (dd, *J* = 8.1, 1.3 Hz, 1H), 7.49 (dd, *J*
9 = 8.0, 1.0 Hz, 1H), 7.41 (dd, *J* = 8.3, 0.6 Hz, 1H), 7.30 – 7.27 (m, 1H), 7.24 (dd, *J* = 7.7, 1.1 Hz, 1H),
10 7.16 (td, *J* = 7.6, 1.1 Hz, 1H), 6.93 (d, *J* = 7.4 Hz, 1H), 4.60 (s, 2H). ¹³C{¹H} NMR (126 MHz, CDCl₃)
11 δ 163.1, 156.7, 153.8, 135.3, 133.6, 132.9, 130.3, 129.2, 128.9, 127.7, 126.9, 125.8, 117.9, 117.5, 113.3,
12 104.5, 35.0. **HRMS (ESI/QTOF), m/z:** [M+Na]⁺ Calcd for C₁₇H₁₀ClNO₂Na, 318.0292; Found:
13 318.0295.
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24 **4-(4-Bromobenzyl)-2-oxo-2H-chromene-3-carbonitrile (4h).** Physical state: White solid; Yield: 38.08
25 mg (56%). ¹H NMR (300 MHz, CDCl₃) δ 7.70 – 7.64 (m, 2H), 7.48 – 7.45 (m, 2H), 7.42 (dd, *J* = 9.0,
26 1.2 Hz, 1H), 7.36 – 7.31 (m, 1H), 7.16 (d, *J* = 8.5 Hz, 2H), 4.42 (s, 2H). ¹³C{¹H} NMR (126 MHz, CDCl₃)
27 δ 162.8, 156.6, 154.1, 135.4, 134.0, 132.6, 130.0, 126.7, 125.6, 122.0, 118.1, 117.4, 113.6, 103.6, 37.1.
28
29 **HRMS (ESI/QTOF), m/z:** [M+Na]⁺ Calcd for C₁₇H₁₀BrNO₂Na, 361.9787; Found: 361.9802.
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31
32
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35 **4-(Naphthalen-2-ylmethyl)-2-oxo-2H-chromene-3-carbonitrile (4i).** Physical state: White solid; Yield:
36 29.86 mg (48%). ¹H NMR (300 MHz, CDCl₃) δ 7.84 – 7.75 (m, 4H), 7.68-7.61 (m, 2H), 7.51 – 7.45 (m,
37 2H), 7.41 - 7.38 (m, 2H), 7.30 (d, *J* = 9.0 Hz, 1H), 4.64 (s, 2H). ¹³C{¹H} NMR (75 MHz, CDCl₃) δ 163.4,
38 156.9, 154.0, 135.2, 133.5, 132.6, 129.3, 127.8, 127.8, 127.1, 127.0, 126.8, 126.5, 126.0, 125.6, 118.0,
39 117.6, 113.8, 103.51, 37.9. **HRMS (ESI/QTOF), m/z:** [M+K]⁺ Calcd for C₂₁H₁₃NO₂K, 350.0578;
40 Found: 350.0591.
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51 **2-Oxo-4-(thiophen-3-ylmethyl)-2H-chromene-3-carbonitrile (4j).** Physical state: White solid; Yield:
52 29.90 mg (56%). ¹H NMR (300 MHz, CDCl₃) δ 7.82 (d, *J* = 7.9 Hz, 1H), 7.68 (t, *J* = 7.4 Hz, 1H), 7.42-
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2 7.37 (m, 2H), 7.35 – 7.31 (m, 1H), 7.15 (s, 1H), 7.04 (d, $J = 6.0$ Hz, 1H), 4.46 (s, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (75
3
4 MHz, CDCl_3) δ 163.3, 156.9, 154.0, 135.3, 134.3, 127.4, 127.4, 127.1, 126.7, 125.6, 123.2, 118.0, 117.4,
5
6 113.6, 102.7, 32.5. HRMS (ESI/QTOF), m/z : $[\text{M}+\text{Na}]^+$ Calcd for $\text{C}_{15}\text{H}_9\text{NO}_2\text{SNa}$, 290.0246; Found:
7
8 290.0245.
9

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12 **4-((1H-Indol-3-yl)methyl)-2-oxo-2H-chromene-3-carbonitrile (4k)**. Physical state: White solid; Yield:
13
14 45.0 mg (75%). ^1H NMR (300 MHz, DMSO) δ 11.05 (s, 1H), 7.92 (d, $J = 8.0$ Hz, 1H), 7.73 (t, $J = 7.9$
15
16 Hz, 2H), 7.50 (d, $J = 8.3$ Hz, 1H), 7.35 (t, $J = 7.1$ Hz, 2H), 7.20 (d, $J = 1.5$ Hz, 1H), 7.15 – 7.04 (m, 2H),
17
18 4.53 (s, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, DMSO) δ 164.3, 157.1, 153.44, 136.1, 135.1, 127.3, 126.3, 125.2,
19
20 124.2, 121.5, 119.0, 118.26, 117.5, 117.2, 114.5, 111.6, 108.9, 102.1, 33.4. HRMS (ESI/QTOF), m/z :
21
22 $[\text{M}+\text{Na}]^+$ Calcd for $\text{C}_{19}\text{H}_{12}\text{N}_2\text{O}_2\text{Na}$, 323.0791; Found: 323.0795.
23
24

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26
27 **4-(3-(1H-Indol-3-yl)propyl)-2-oxo-2H-chromene-3-carbonitrile (4l)**. Physical state: White solid;
28
29 Yield: 28.21 mg (43%). ^1H NMR (300 MHz, CDCl_3) δ 11.05 (s, 1H), 7.63-7.59 (m, 1H), 7.40 (d, $J =$
30
31 8.0 Hz, 1H), 7.39- 7.35 (M, 1H) 7.34 (dd, $J = 4.8, 3.6$ Hz, 2H), 7.22 (t, $J = 7.3$ Hz, 1H), 7.16 (d, $J = 8.1$
32
33 Hz, 1H) 7.15-7.06 (m, 2H), 3.15-3.10 (m, 2H), 3.03 (t, $J = 7.0$ Hz, 2H), 2.21-2.17 (m, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR
34
35 (75 MHz, CDCl_3) δ 167.0, 156.9, 156.7, 153.7, 131.5, 136.0, 127.1, 126.0, 125.4, 122.3, 122.2, 119.5,
36
37 118.8, 117.9, 117.3, 114.5, 113.6, 111.4, 101.7, 31.6, 30.6, 25.4. HRMS (ESI/QTOF), m/z : $[\text{M}+\text{Na}]^+$
38
39 Calcd for $\text{C}_{21}\text{H}_{16}\text{N}_2\text{O}_2\text{Na}$, 351.1104; Found: 351.1107.
40
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44 **tert-Butyl (2-(3-cyano-2-oxo-2H-chromen-4-yl)ethyl)carbamate (4m)**. Physical state: White solid;
45
46 Yield: 30.77 mg (49%). ^1H NMR (300 MHz, CDCl_3) δ 7.99 (d, $J = 7.7$ Hz, 1H), 7.70 (t, $J = 7.7$ Hz, 1H),
47
48 7.43 (dd, $J = 18.4, 8.1$ Hz, 2H), 4.96 (s, 1H), 3.52 (q, $J = 6.6$ Hz, 2H), 3.35 (t, $J = 6.7$ Hz, 2H), 1.42 (s,
49
50 9H). $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 163.7, 156.8, 156.1, 153.8, 135.4, 126.5, 125.8, 117.9, 113.6,
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2 102.7, 80.2, 40.2, 32.9, 33.07, 28.4. **HRMS (ESI/QTOF), m/z:** [M+Na]⁺ Calcd for C₁₇H₁₈N₂O₄Na,
3
4 337.1159; Found: 337.1172.
5

6
7 **tert-Butyl (3-(3-cyano-2-oxo-2H-chromen-4-yl)propyl)carbamate (4n).** **Physical state:** White solid;
8
9 **Yield:** 46.58 mg (71%). **¹H NMR (300 MHz, CDCl₃)** δ 7.76-7.66 (m, 2H), 7.40 (t, *J* = 1.0 Hz, 2H), 4.94
10
11 (s, 1H), 3.31 (d, *J* = 6.2 Hz, 2H), 3.31 (d, *J* = 6.2 Hz, 2H) 3.15 – 3.10 (m, 2H), 1.43 (s, 9H). **¹³C{¹H} NMR**
12
13 **(75 MHz, CDCl₃)** δ 166.2, 156.8, 156.2, 153.9, 135.9, 126.0, 125.7, 118.1, 117.3, 113.5, 101.9, 79.7,
14
15 40.2, 33.8, 30.0, 28.4. **HRMS (ESI/QTOF), m/z:** [M+Na]⁺ Calcd for C₁₈H₂₀N₂O₄Na, 351.1315; Found:
16
17 351.1321.
18
19

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21
22 **6-Methyl-2-oxo-4-propyl-2H-chromene-3-carbonitrile (4o).** **Physical state:** White solid; **Yield:** 43.13
23
24 mg (95%). **¹H NMR (300 MHz, CDCl₃)** δ 7.49 (d, *J* = 9.0 Hz, 2H), 7.30 (d, *J* = 9.0 Hz, 1H), 3.10 – 3.05
25
26 (m, 2H), 2.47 (s, 3H), 1.85 – 1.78 (m, 2H), 1.14 (t, *J* = 7.4 Hz, 3H). **¹³C{¹H} NMR (75 MHz, CDCl₃)** δ
27
28 166.4, 157.2, 152.1, 136.3, 135.4, 125.6, 117.8, 117.2, 113.7, 101.6, 33.7, 23.7, 21.2, 14.3. **HRMS**
29
30 **(ESI/QTOF), m/z:** [M+Na]⁺ Calcd for C₁₄H₁₃NO₂Na, 250.0838; Found: 250.0840.
31
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35 **7-methoxy-2-oxo-4-propyl-2H-chromene-3-carbonitrile (4p).** **Physical state:** White solid; **Yield:** 40.33
36
37 mg (84%). **¹H NMR (300 MHz, CDCl₃)** δ 7.63 (d, *J* = 9.0 Hz, 1H), 6.95 (dd, *J* = 9.0, 2.5 Hz, 1H), 6.83
38
39 (s, 1H), 3.92 (s, 3H), 3.10-2.93 (m, 2H), 1.77 (dt, *J* = 14.9, 7.5 Hz, 2H), 1.11 (t, *J* = 7.3 Hz, 3H). **¹³C{¹H}**
40
41 **NMR (75 MHz, CDCl₃)** δ 166.4, 165.4, 157.7, 156.2, 127.3, 114.2, 114.0, 111.2, 101.36, 98.1, 56.2,
42
43 33.7, 23.5, 14.2. **HRMS (ESI/QTOF), m/z:** [M+Na]⁺ Calcd for C₁₄H₁₃NO₃Na, 266.0787, Found:
44
45 266.0790.
46
47

48
49 **Ethyl 2-oxo-4-propylchromane-3-carboxylate (4q).** **Physical state:** White solid; **Yield:** 41 mg (72%). **¹H**
50
51 **NMR (300 MHz, CDCl₃)** δ 7.31-7.26 (m, 1H), 7.11 – 7.07 (m, 3H), 4.16 – 3.97 (m, 2H), 3.78 (d, *J* = 2.4
52
53 Hz, 1H), 3.41 – 3.36 (m, 1H), 1.61 – 1.55 (m, 2H), 1.42 – 1.34 (m, 2H), 1.03 (t, *J* = 7.1 Hz, 3H). 0.94 (t,
54
55

$J = 7.2$ Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 167.3, 164.8, 150.8, 128.8, 128.7, 124.7, 124.4, 117.1, 62.2, 52.2, 39.8, 36.6, 19.9, 13.9, 13.8. HRMS (ESI/QTOF), m/z : $[\text{M}+\text{Na}]^+$ Calcd for $\text{C}_{15}\text{H}_{18}\text{O}_4\text{Na}$, 285.1097; Found: 285.1107.

4-(3-(Biphenyl-4-yl)-3-oxopropyl)-2-oxo-2H-chromene-3-carbonitrile (4r). Physical state: White solid; Yield: 35.37 mg (44%). ^1H NMR (400 MHz, CDCl_3) δ 8.03 (d, $J = 8.4$ Hz, 2H), 7.85 (dd, $J = 8.3$, 1.2 Hz, 1H), 7.22 - 7.69 (m, 3H), 7.64 - 7.61 (m, 2H), 7.49 - 7.41 (m, 5H), 3.59 (dd, $J = 8.4$, 6.4 Hz, 2H), 3.47 (t, $J = 8.0$ Hz, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) δ 195.9, 166.1, 154.0, 146.7, 135.5, 129.1, 128.9, 128.6, 127.6, 127.2, 126.0, 125.8, 118.2, 117.3, 113.41, 102.6, 37.5, 25.9. HRMS (ESI/QTOF), m/z : $[\text{M}+\text{Na}]^+$ Calcd for $\text{C}_{25}\text{H}_{17}\text{NO}_3\text{Na}$, 402.1100; Found: 402.1080.

2-Oxo-4-pentadecyl-2H-chromene-3-carbonitrile (4s). Physical state: White solid; Yield: 60.12 mg (79%). ^1H NMR (CDCl_3 , 300 MHz) δ 7.75 - 7.67 (m, 2H), 7.42 (t, $J = 9$ Hz, 2H), 3.12 - 3.07 (m, 2H), 1.80 - 1.70 (m, 2H), 1.52 - 1.47 (m, 2H), 1.25 (m, 22H), 0.87 (t, $J = 6.6$ Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) δ 166.9, 157.0, 154.0, 135.2, 126.0, 125.6, 118.1, 117.6, 113.5, 101.9, 77.5, 77.2, 76.9, 32.1, 30.1, 29.8, 29.5, 29.4, 22.8, 14.3. HRMS (ESI/QTOF), m/z : $[\text{M}+\text{Na}]^+$ Calcd for $\text{C}_{25}\text{H}_{35}\text{NO}_2\text{Na}$, 404.2560; Found: 404.2546.

(Z)-4-(Heptadec-8-en-1-yl)-2-oxo-2H-chromene-3-carbonitrile (4t). Physical state: White solid; Yield: 56.98 mg (70%). ^1H NMR (300 MHz, CDCl_3) δ 7.71 (dd, $J = 15.5$, 7.9 Hz, 2H), 7.42 (t, $J = 6.8$ Hz, 2H), 5.35 - 5.34 (m, 2 H), 3.13 - 3.08 (m, 2 H), 2.01 (s, 4 H), 1.77 (dd, $J = 14.6$, 7.1 Hz, 2H), 1.58 - 1.53 (m, 2H), 1.34 (s, 9H), 1.26 (s, 9H), 0.87 (t, $J = 5.8$ Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) δ 166.8, 157.0, 153.9, 135.1, 130.2, 129.7, 126.0, 125.5, 118.1, 117.5, 113.5, 101.9, 32.1, 32.0, 30.0, 29.8, 29.7, 29.6, 29.4, 29.2, 29.1, 27.3, 27.2, 22.8, 14.2. HRMS (ESI/QTOF), m/z : $[\text{M}+\text{Na}]^+$ Calcd for $\text{C}_{27}\text{H}_{37}\text{NO}_2\text{Na}$, 430.2717; Found: 430.2723.

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2 **4-((1-(4-Chlorobenzoyl)-5-methoxy-2-methyl-1H-indol-3-yl)methyl)-2-oxo-2H-chromene-3-**
3
4 **carbonitrile (4u).** Physical state: Brown solid; Yield: 38.0 mg (44%). ¹H NMR (300 MHz, CDCl₃) δ
5 7.86 (d, *J* = 7.9 Hz, 1H), 7.70 – 7.64 (m, 3H) 7.43 (dd, *J* = 14.0, 8.4 Hz, 3H), 7.33 (t, *J* = 7.6 Hz, 1H),
6 6.84 (d, *J* = 9.0 Hz, 1H), 6.75 (s, 1H), 6.66 (d, *J* = 8.8 Hz, 1H), 5.47 (s, 2H), 3.71 (s, 3H), 2.46 (s, 3H).
7
8 ¹³C{¹H} NMR (75 MHz, CDCl₃) δ 168.4, 163.6, 153.5, 139.8, 136.3, 135.4, 133.4, 131.5, 130.9, 129.5,
9 129.3, 126.1, 125.6, 118.1, 115.2, 130.0, 112.0, 100.4, 100.8, 55.7, 33.9, 29.1. HRMS (ESI/QTOF), *m/z*:
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11 [M+Na]⁺ Calcd for C₂₈H₁₉ClN₂O₄Na, 505.0925, Found: 505.0921.
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19 **2-Oxo-4-((3S)-3-((3S,5R,7S,8S,10R,12R,13S,14R,17S)-3,7,12-trihydroxy-10,13-**
20 **dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)butyl)-2H-chromene-3-carbonitrile**
21 **(4v).** Physical state: Brown solid; Yield: 57.56 mg (54%). ¹H NMR (400 MHz, CDCl₃) δ 7.69 – 7.60
22 (m, 2H), 7.38 – 7.20 (m, 2H), 3.89 (s, 1H), 3.79 (s, 1H), 3.48- 3.33 (m, 4H), 3.11 – 3.07 (m, 1H), 2.98 (s,
23 1H), 2.22 – 2.10 (m, 1H), 1.97 – 1.82 (m, 3H), 1.66 – 1.46 (m, 10H), 1.21 – 1.18 (m, 9H), 0.92 – 0.87 (m,
24 2H), 0.79 (s, 3H), 0.69 (s, 3H). ¹³C{¹H} NMR (75 MHz, CDCl₃) δ 168.5, 157.2, 153.9, 135.3, 126.1,
25 125.7, 118.0, 117.5, 113.6, 101.3, 73.2, 72.0, 68.6, 46.6, 45.8, 41.7, 39.8, 39.4, 36.6, 36.4, 35.5, 34.9,
26 30.5, 29.8, 28.5, 28.2, 27.9, 26.3, 23.5, 22.5, 17.7, 14.3, 12.5. HRMS (ESI/QTOF), *m/z*: [M+K]⁺ Calcd
27 for C₃₃H₄₃NO₅K, 572.2773; Found: 572.2772.
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40 ASSOCIATED CONTENT

41
42
43 The Supporting Information is available free of charge on the ACS Publications website at DOI:

44
45 Mechanistic studies, X-ray crystallography data, CIF files, ¹H and ¹³C NMR spectra
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43
44 **DEDICATION**
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46
47 This work is dedicated to Professor Elias J. Corey on the occasion of his 90th birthday.
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