by CC on silica gel (30 g) and elution with  $C_6H_6$ -EtOAc (3:1, 2:1). Appropriate column fractions were separated by prep. TLC using silica gel and  $C_6H_6$ -EtOAc (3:1). Subsequent purification on silica gel (20 g) and recrystallization from the same solvent mixture yielded 23 mg of pure 1.

Compound 1. Thin needles with blue fluorescence (360 nm). Mp 262–263° ( $C_6H_6$ –EtOAc 3:1).  $C_{15}H_{10}O_4$  requires [M]<sup>+</sup> 254.0579; found 254.0585. EI-MS: m/z (rel. int.); 254 [M]<sup>+</sup> (100), 225 [M – CHO]<sup>+</sup> (90), 139 (40); CI-MS: 255 [MH]<sup>+</sup>; UV  $\lambda_{max}^{McOH}$  nm: 230, 257, 266, 289sh, 300, 315, 330sh, 345; IR  $\nu_{max}^{Nujol}$  cm<sup>-1</sup>: 1725 (conjugated ester); <sup>1</sup>H NMR (DMSO- $d_6$ , ppm): 3.493 (2H, t, J = 6.1 Hz, CH<sub>2</sub>-4), 4.678 (2H, t, J = 6.1 Hz, CH<sub>2</sub>-3), 5.851 (2H, d, J = 1.1 Hz, CH<sub>2</sub>-6), 7.875 (1H, dd, J = 8.5, 7.3 Hz, H-10), 7.964 (1H, br s, H-5), 8.376 (1H, dd, J = 7.3, 1.1 Hz, H-9), 8.517 (1H, dd, J = 8.5, 1.1 Hz, H-11); <sup>13</sup>C NMR (DMSO- $d_6$ , ppm, multiplets from DEPT): 23.06 (t, C-4), 66.20 (t, C-3), 68.73 (t, C-6), 120.23 (d, C-10), 127.30 (d, C-5), 130.29 (d, C-9), 130.64 (d, C-11), 120.52 s, 122.32 s, 126.65 s, 128.33 s, 129.61 s, 138.04 s (C-4a, C-5a, C-5b, C-8a, C-11a, C-11b), 162.49 s and 163.61 s (C-1 and C-8).

## REFERENCES

- 1. Stojanov, N. (1973) Our Medicinal Plants Vol. 2, pp. 13-18. Nauka i Iskustvo, Sofia.
- 2. Goetz, M., Hostettmann, K. and Jacot-Guillarmod, A. (1976) *Phytochemistry* 15, 2014.
- 3. Goetz, M. and Jacot-Guillarmod, A. (1977) *Helv. Chim. Acta* 60, 1322.
- 4. Goetz, M. and Jacot-Guillarmod, A. (1977) Helv. Chim. Acta 60, 2104.
- 5. Goetz, M. and Jacot-Guillarmod, A. (1977) *Helv. Chim. Acta* **61**, 1373.
- 6. Marekov, N. L. and Popov, S. S. (1968) Tetrahedron 24, 1323.
- Rulko, F. and Nadler, K. (1970) Diss. Pharm. Pharmacol. 22, 329.
- 8. Mpondo, E. M. and Chulia, A. J. (1988) Planta Med. 54, 185.
- 9. Kitanov, G. M. (1992) Khim. Prir. Soedin. (in press).
- 10. Dombrowicz, E. and Swiatek, I. (1987) Farm. Pol. 43, 639.
- Pretsch, E., Seibl, J., Simon, W. and Clerc, T. (1983) Tables of Spectral Data for Structure Determination of Organic Compounds, p. H245. Springer, Berlin.

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# ISOSALVIANOLIC ACID C, A DEPSIDE POSSESSING A DIBENZOOXEPIN SKELETON

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Key Word Index--Salvia chinensis; Labiatae; depsides; isosalvianolic acid C.

Abstract—Isosalvianolic acid C, a new depside possessing a dibenzooxepin skeleton, was isolated from the aqueous extract of Salvia chinensis along with salvianolic acid B and D, lithospermic acid, rosmarinic acid,  $(R-(+)-\beta-(3,4-dihydroxyphenyl))$  lactic acid, caffeic acid and protocatechualdehyde.

# INTRODUCTION

Salvia chinensis Benth. is a herbal medicinal plant distributed in the southern part of the Yangtze River in China. It is used in Chinese folk medicine for the treatment of hepatitis, nephritis, dysmenorrhea and several kinds of cancer [1]. No chemical investigation of this plant has been reported previously. During our studies on the biologically active components of the traditional Chinese medicine Danshen (Salvia miltiorrhiza), we isolated several depsides [2-4], some of which possessed significant antiplatelet [5] and anti-oxidant activities [6]. These observations prompted us to extend our studies to other species of the genus Salvia. Chemical studies on the aqueous extract of S. chinensis led to the isolation of five depsides, among which one is a new compound named isosalvianolic acid C (1a), together with two phenolic acids and protocatechualdehyde.

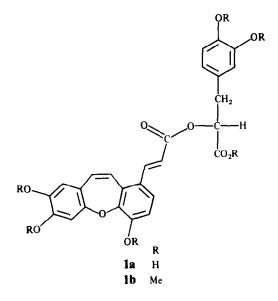
## **RESULTS AND DISCUSSION**

Systematic isolation of the aqueous extract of S. chinensis by Sephadex LH-20 CC, silica gel LPLC and preparative TLC yielded a new depside named isosalvianolic acid C (1a), along with four known depsides, salvianolic acid B [3] and D [4], lithospermic acid and rosmarinic acid, two phenolic acids,  $R-(+)-\beta-(3,4-dihydroxy$ phenyl)lactic acid and caffeic acid, and protocatechualdehyde.

Isosalvianolic acid C (1a) was recognized as a phenolic acid from a positive test with ferric ferricyanide and bromocresol green. UV maxima at 202, 222, 288, 326 and 340 nm suggested the presence of a highly conjugated system. Its FD mass spectrum displayed  $[M]^+$  at m/z 492, similar to that of salvianolic acid C [3].

The HR mass spectrum of the methylated compound **1b** showed [M]<sup>+</sup> at m/z 576.2000 (calc. for C<sub>32</sub>H<sub>32</sub>O<sub>10</sub>, 576.1996) and diagnostic fragments of a  $\beta$ -(3,4-dimethoxyphenyl)-lactic ester (m/z 222, 191, 181, 163) [2].

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Hydrolysis of 1b with 10% KOH-methanol yielded  $R-(+)-\beta-(3,4-dimethoxyphenyl)$  lactic acid [2] and a yellowish crystalline compound 1c with  $[M]^+$  at m/z 354. Its <sup>1</sup>H and <sup>13</sup>C NMR spectra revealed the presence of 16 olefinic carbons, eight olefinic protons, three methoxyls and one carboxyl group (Table 1), giving a composition  $C_{20}H_{18}O_5$  of 338. The molecular ion of 1c indicated the presence of an additional oxygen, linked between two olefinic carbons, thus 1c should have a molecular formula of  $C_{20}H_{18}O_6$ . Consideration of the <sup>1</sup>H and <sup>13</sup>C NMR data indicated that the 12 degrees of unsaturation inherent in the molecule consisted of one carboxylic carbonyl, eight olefinic groups and a tricyclic carbon skeleton. A dibenzooxepin skeleton for 1c was deduced from the fact that the two olefinic proton signals at  $\delta 6.94$  and 7.00 possessed a coupling constant of J = 11.5 Hz similar to that of pacharin, a dibenzooxepin derivative isolated from the heartwood of Bauhinia racemosa [7]. A pair of singlets at  $\delta 6.80$  and 6.90 and a pair of doublets with J = 8at  $\delta$ 7.11 and 7.53 indicated the presence of two para protons and two ortho protons in the two separated benzo groups. The chemical structure of 1c was finally verified by H,C-COLOC analysis as depicted in Fig. 1. Isosalvianolic acid C is thus a depside having structure **1a.** Following the isolation of salvianolic acid G from S. miltiorrhiza [8], this is the second dibenzooxepin derivative isolated from the genus Salvia.

#### **EXPERIMENTAL**

General. Sephadex LH-20 CC, solvent MeOH; LPLC, 300 g silica gel, sample 4g, solvent A,  $CHCl_3$ -MeOH-HCO<sub>2</sub>H (85:15:1) 2l, **B**,  $CHCl_3$ -MeOH (17:3) 1.51, **C**, (4:1) 800 ml, **D**, (1:1) 800 ml; TLC, silica gel-GF<sub>254</sub>, solvent CHCl<sub>3</sub>-MeOH-HCO<sub>2</sub>H (85:15:1). Mps: uncorr. <sup>1</sup>H and <sup>13</sup>C NMR (90 or 400 MHz) with TMS as int. standard.

*Plant material.* The whole plants of *S. chinensis* were collected in Jiangxi province, China, and identified by Prof. Zhang Haidao of the Jiangxi Institute of Materia Medica. A voucher specimen is deposited in the Institute of Materia Medica in Beijing.

Extraction and isolation of the depsides. The dried plant material (4.25 kg) was extracted with 95% EtOH and the residue boiled with H<sub>2</sub>O. The aq. extract was concd under red. pres. and

EtOH added to the concentrate until the EtOH content was 70%. After filtering, the filtrate was concd to 21 and successively extracted with CHCl<sub>3</sub> and EtOAc. Evapn of the EtOAc extract yielded 4.6 g of fr. 1. The aq. portion was acidified with 10% HCl to pH 3-4 and successively extracted with EtOAc and n-BuOH. After removal of the solvent, 38.5 g of fr. 2 and 23.94 g of fr. 3 were respectively obtained from the EtOAc and n-BuOH extracts. Chromatography of fr. 1 (3.9 g) over Sephadex LH-20 afforded six fractions. Purification of frs 1-2 and 1-3 by prep. TLC yielded 99 mg of protocatechualdehyde and 40 mg of caffeic acid. Fr. 2 (34g) was sepd by LPLC and 12 frs were collected. Rechromatography over Sephadex LH-20 yielded rosmarinic acid (347 mg) from fr. 2-4, isosalvianolic acid C (1a) (180 mg) from fr. 2-5, and salvianolic acid D (30 mg) and lithospermic acid (167 mg) from fr. 2-6. Further fractionation of fr. 2-7 by prep. TLC furnished 1.07 g of salvianolic acid B. Separation of fr. 3 (4 g) by Sephadex LH-20 CC, followed by prep. TLC led to the isolation of  $R-(+)-\beta-(3,4-dihydroxy$ phenyl)lactic acid (52 mg).

Isosalvianolic acid C (1a). Amorphous yellowish compound,  $[\alpha]_D^{12} + 39^\circ$  (EtOH; c 0.06). UV $\lambda_{max}^{\text{EtOH}}$  nm (log  $\epsilon$ ): 202 (4.77), 222 (4.46 sh), 288 (4.21), 326 (4.30), 340 (4.24); FD-MS m/z: 492 [M]<sup>+</sup> <sup>1</sup>H NMR (Me<sub>2</sub>CO-d<sub>6</sub>):  $\delta$  5.20 (1H, dd, J = 6, 8 Hz), 6.28 (1H, d, J = 16 Hz), 7.92 (1H, d, J = 16 Hz), 6.72–7.41 (9H, m).

Methylation of isosalvianolic acid C. Dry  $K_2CO_3$  (200 mg) was suspended in a soln of 1a (80 mg) in dry  $Me_2CO$  (10 ml) under an atmosphere of  $N_2$ .  $Me_2SO_4$  (1.5 ml) was added, and after stirring for 6 hr a second portion (0.5 ml) added and stirring was continued for 26 hr. The mixture was filtered and concd under red. pres. The residue was purified by silica gel VLC [9] using a mixture of petrol and EtOAc with increasing polarity. Methylated 1b (72 mg) was obtained as a yellowish gum.  $[\alpha]_{\rm D}^{12} + 21^{\circ}$ (EtOH; c 0.06). UV $\lambda_{\rm max}^{\rm EtOH}$  nm (log  $\varepsilon$ ): 203 (4.72), 232 (4.40 sh), 288 (4.07), 326 (4.22), 340 (4.15); HRMS m/z (rel. int.): 576.2000 [M] <sup>+</sup> (100) (C<sub>32</sub>H<sub>32</sub>O<sub>6</sub> requires 576.1996), 354 (39), 337 (24), 309 (72),

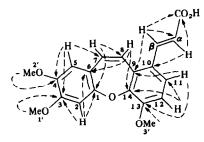


Fig. 1. H, C-COLOC of 1c

Table 1. <sup>13</sup>C and <sup>1</sup>HNMR spectral data of compound 1c (DMSO- $d_6$ )

С	<sup>13</sup> C	<sup>1</sup> H	С	<sup>13</sup> C
2	105.3	6.80 (1H, s)	1	150.7
5	111.6	6.90 (1H, s)	3	146.0
7	131.2	6.94 (1H, d, J = 11.5)	4	150.4
8	124.4	7.00 (1H, $d$ , $J = 11.5$ )	6	121.9
11	124.2	7.53 (1H, $d, J = 8$ )	9	130.1
12	112.7	7.11 (1H, $d, J = 8$ )	10	124.5
1′	55.9	3.72 (3H, s)	13	152.5
2′	55.7	3.77 (3H, s)	14	145.7
3′	56.1	3.91 (3H, s)	C = 0	167.2
α	119.6	6.30 (1H, d, J = 16)		
β	134.0	7.80 (1H, $d, J = 16$ )		

## Short Reports

278 (37), 222 (98), 191 (42), 181 (19), 163 (13), 151 (77); <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$ 3.11 (1H, d, J = 6 Hz), 3.12 (1H, d, J = 8 Hz), 5.28 (1H, dd, J = 6, 8 Hz) 3.72, 3.86, 3.92 (each 3H, s) 3.84 (9H, s), 6.20, 7.91 (each 1H, d, J = 16 Hz), 6.61–7.32 (9H, m).

Hydrolysis of compound **1b**. Compound **1b** (40 mg) was refluxed with 5 ml 10% KOH-MeOH for 1 hr. After acidification and work-up with EtOAc and H<sub>2</sub>O, the organic layer was concd yielding 19 mg of yellow crystals **1c**. The mother liquor was purified by TLC using  $C_6H_6$ -EtOH-HCO<sub>2</sub>H (80:20:1) as solvent, 5 mg of **1c** along with *R*-(+)- $\beta$ -(3, 4-dimethoxyphenyl)lactic acid were obtained.

Compound 1c. Yellow crystals, mp 178–180°. EI-MS m/z (rel. int.): 354 ([M]<sup>+</sup>, 100), 339 (28), 309 (55), 278 (75).

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# REFERENCES

- 1. Compilation of Chinese Herb Medicine (1975) Vol. 1, p. 242. People's Publishing House, Beijing.
- 2. Li, L. N., Tan, R. and Chen, W. M. (1984) Planta Med. 227.
- 3. Ai, C. B. and Li, L. N. (1988) J. Nat. Prod. (Lloydia) 51, 145.
- 4. Ai, C. B. and Li, L. N. (1992) Planta Med. (in press).
- Xu, L. N., Wang, J. P., Tian, Q. Y. and Han, Y. H. (1990) Proceedings of the 3rd National Symposium of Activating Blood Circulation and Removing Blood Stasis Research, p. 198. Lushan, China.
- Xu, L. N., Li, D. Y. and Tian, Q. Y. (1990) Proceedings of the 3rd National Symposium of Activating Blood Circulation and Removing Blood Stasis Research, p. 198. Lushan, China.
- 7. Anjaneyulu, A. S. R., Reddy, A. V. R. and Reddy, D. S. K. (1984) Tetrahedron 40, 4245
- 8. AI, C. B. and Li, L. N. (1991) Chinese Chem. Letters 1, 17.
- Pelletier, S. W., Chokshi, H. P. and Desai, H. K. (1986) J. Nat. Prod. 49, 892.

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# CHROMENES FROM ARNICA SACHALINENSIS AND A. AMPLEXICAULIS

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Key Word Index—Arnica sachalinensis; A. amplexicaulis; Asteraceae; chromenes (benzopyrans); 12-acetoxy-demethoxyencecalin; 12-isobutyryloxy-demethoxyencecalin.

Abstract—Flowerheads of Arnica sachalinensis yielded, in addition to the known chromenes acetovanillochromene, demethylencecalin and demethoxyencecalin, the new compounds 12-acetoxy-, 12-isobutyryloxy- and 12-isovaleryloxy-demethoxyencecalin. Demethoxy-and demethylencecalin are also constituents of A. amplexicaulis. The assignment of the <sup>13</sup>C NMR signals of demethoxyencecalin is revised by  $2D-^{13}C$ , <sup>1</sup>H NMR experiments. Chemotaxonomic aspects are discussed.

### INTRODUCTION

Chromenes (benzopyrans) and benzofurans are prominent natural products of many genera of the Asteraceae and have become useful taxonomic markers within this family [1-3]. In a previous paper [4], we reported on the isolation of demethoxyencecalin (1) from the flowers of *Arnica sachalinensis* (Regl.) A. Gray, which was the first chromene found in the genus *Arnica*. The tribal position of the genus *Arnica*, traditionally within the tribe Senecioneae, has been the subject of considerable discussion [5, 6]. On morphological, serological and chemical grounds, it has been argued that *Arnica* is more related to the Heliantheae than to the Senecioneae, thus Robinson [7] includes *Arnica* in this tribe, subtribe Chaenactidinae. In Bohlmann's opinion, however, *Arnica* has more resemblance to the closely related subtribe Gaillardiinae

