

Synthesis and Fluorescence Properties of new Monastrol Analogs Conjugated Fluorescent Coumarin Scaffolds

Najim A. Al-Masoudi^{1,2} · Niran J. Al-Salihi¹ · Yossra A. Marich¹ · Timo Markus³

Received: 3 July 2015 / Accepted: 14 October 2015
© Springer Science+Business Media New York 2015

Abstract A mild and efficient method has been used for the synthesis of ethyl 4-(3-hydroxyphenyl)-6-methyl-2-thioxo-1,3-dihydroprimidine-5-carboxylate (monastrol) (**2**), via Biginelli reaction. Alkylation of **2** with the fluorescent coumarin **3** afforded the new thioether analog **4**. Similarly, ethyl 4-(6,8-dichloro-2-oxo-2 H-chromen-3-yl)-6-methyl-2-thioxo-1,2,3,4-tetrahydro-pyrimidine-5-carboxylate (**6**) was prepared. The synthesized compounds are fluorescent active and show wavelength of maximum absorption (λ_{max}) in UV or visible region in MeOH at room temperature.

Keywords Biginelli reaction · Coumarins · MPA supported on Y zeolite · Monastrol · Fluorescence properties

Introduction

A common strategy for cancer therapy is the development of drugs that interrupt the cell cycle during the stage of mitosis. Compounds that perturb microtubule shortening (depolymerization) or lengthening (polymerization) cause arrest of the cell cycle in mitosis due to perturbation of the normal microtubule dynamics necessary for chromosome

movement [1]. Therefore, microtubule dynamics is an important target for the developing anticancer drugs [2] but differ in their mode of action from other drugs because they target the mitotic spindle and not the DNA. A variety of such drugs that bind to tubulin and thus inhibit spindle assembly are currently under investigation in cancer therapy, however, they have been classified on the basis of their mode of action and binding site [3–5]. Among these drug is monastrol (ethyl 4-(3-hydroxyphenyl)-6-methyl-2-sulfanylidene-3,4-dihydro-1 H-pyrimidine-5-carboxylate) (**2**) [6–9]. Coumarin core moieties have wide biological application, in particular for the imaging of living cells [10, 11], since exhibited spectral range and high emission quantum yields [12]. In this communication, two new monastrol analogs have been prepared with study of their fluorescence properties.

Experimental

Physical Measurements: See ref. [13]

Ethyl 4-(3-hydroxyphenyl)-6-methyl-2-thioxo-1,3-dihydroprimidine-5-carboxylate (Monastrol) (2)

Method A. A mixture of **1** (122 mg, 1.0 mmol), ethyl acetoacetate (130 mg, 1.0 mmol), and thiourea (190 mg, 2.5 mmol) in MeCN (15 ml) was mixed with HPA supported on Y zeolite (8 wt% NaY + 0.5 mM HPA) and refluxed for 7 h. After cooling, the heteropoly acid (HPA) supported on HY filtered off and washed with hot water and ethanol to remove thiourea from the surface of the catalyst. Then, the catalyst dried and was maintained for new runs. The filtrate was evaporated to dryness and the residue was recrystallized

✉ Najim A. Al-Masoudi
najim.al-masoudi@gmx.de

¹ Department of Chemistry, College of Science, University of Basrah, Basrah 61001, Iraq

² Present address: Am Tannenhof 8, 78464 Constance, Germany

³ Department of Chemistry, University of Konstanz, P.O. Box 5560, 78457 Constance, Germany

from EtOH to afford **2** (254 mg, 87 %), m.p. 181–185 °C (Lit. [6, 7] 184–186 °C), $R_f = 0.71$. ^1H NMR (DMSO- d_6): δ 10.29 (s, 1 H, NH), 9.60 (s, 1 H, NH), 9.44 (s, 1 H, OH), 7.10 (t, $J = 7.8$ Hz, H-5), 6.65 (d, 1 H, $J_{2,4'} = 2.4$ Hz, $\text{H}_{\text{arom.}-2'}$), 6.64 (m, 2 H, $\text{H}_{\text{arom.}-4} + \text{H}_{\text{arom.}-6}$), 5.09 (d, 1 H, $J_{\text{NH},4} = 5.5$ Hz, H-4), 4.02 (q, 2 H, $J = 7.8$ Hz, CH_2CH_3), 2.28 (s, 3 H, $\text{C}_6\text{-Me}$), 1.12 (t, 3 H, $J = 7.8$ Hz, CH_2CH_3). ^{13}C NMR (DMSO- d_6): δ 174.6 (C = S), 165.7 (CO_2Et), 157.9 (C-6), 145.3 ($\text{C}_3\text{'-OH} + \text{C}_{\text{arom.}-1'}$), 130.0 ($\text{C}_{\text{arom.}-5'}$), 117.7 ($\text{C}_{\text{arom.}-2'}$ + $\text{C}_{\text{arom.}-6'}$), 133.6 ($\text{C}_{\text{arom.}-4'}$), 101.2 (C-5), 60.1 (CH_2CH_3), 54.5 (C-4), 17.7, 17.6 ($\text{C}_6\text{-Me}$), 14.5 (CH_2CH_3). EI-MS: m/z (%) = 292 [$\text{M}]^+$. Anal. Calcd. for $\text{C}_{14}\text{H}_{16}\text{N}_2\text{O}_3\text{S}$ (292.35): C, 57.52; H, 5.52; N, 9.58. Found: C, 57.32; H, 5.39; N, 9.32 %.

Method B. A mixture of **1** (244 mg, 2.0 mmol), ethyl acetoacetate (300 mg, 2.30 mmol), and thiourea (380 mg, 5.0 mmol) in EtOH (15 ml) in the presence conc. Hydrochloric acid (1 ml) was heated under reflux for 3 h. After cooling, the mixture was poured onto ice (20 g). The precipitate was filtered and dried and recrystallized from EtOH to give **2** (292 mg, 50 %). The NMR spectra, m.p. and mixed m.p. were almost similar for those of **2** prepared in method A.

Ethyl 4-(3-hydroxyphenyl)-2-(((7-methoxy-2-oxo-2 H-chromen-4-yl)methyl)thio)-6-methyl-1,4-dihydropyrimidine-5-carboxylate (4)

A mixture of **2** (100 mg, 0.34 mmol) and 4-bromomethyl-7-methoxycoumarin (**3**) (97 mg, 0.36 mmol) in DMF (10 ml) containing K_2CO_3 (50 mg, 0.36 mmol) was heated 100 °C for 5 h. After cooling, the solution was evaporated to dryness and the residue was partitioned between CHCl_3 (20 ml) and water (3×20 ml). The combined organic extracts were dried (Na_2SO_4), filtered and evaporated to dryness. The residue was poured onto a SiO_2 column (10 g) and eluted, in gradient, with MeOH (0–5 %) and CHCl_3 as eluent to give **4** (60 mg, 35 %), m.p. 94–97 °C, $R_f = 0.84$. ^1H NMR (DMSO- d_6): δ 9.88 (d, 1 H, $J_{\text{NH},4} = 4.5$ Hz, NH), 7.93–7.84 (m, 8 H, $\text{H}_{\text{arom.}} + \text{OH}$), 6.69 (s, 1 H, $\text{H}_{\text{coum.}-3'}$), 5.18 (d, 1 H, $J_{\text{NH},4} = 4.5$ Hz, $\text{H}_{\text{pyrimid.}-4}$), 4.06 (q, 2 H, $J = 7.2$ Hz, CH_2CH_3), 2.23 (s, 3 H, $\text{C}_6\text{-Me}$), 1.16 (t, 3 H, $J = 7.2$ Hz, CH_2CH_3). ^{13}C NMR (DMSO- d_6): δ = 166.7 (CO_2Et), 160.3 ($\text{C}_{\text{pyrimid.}-2} + \text{C}_{\text{coum.}-2'}$), 160.2 ($\text{C}_{\text{coum.}-7'}$), 156.9 ($\text{C}_3\text{'-OH}$), 155.7 (C-4 + $\text{C}_{\text{coum.}-4'}$), 153.8 ($\text{C}_{\text{coum.}-8\text{a}'}$), 139.4 ($\text{C}_{\text{arom.}-1}$), 132.4 ($\text{C}_{\text{arom.}-5}$), 128.3 ($\text{C}_{\text{coum.}-5'}$), 116.9 ($\text{C}_{\text{arom.}-6}$), 115.5 ($\text{C}_{\text{coum.}-4\text{a}'}$), 114.3 ($\text{C}_{\text{arom.}-2}$), 112.8 ($\text{C}_{\text{coum.}-3'}$ + $\text{C}_{\text{arom.}-4}$), 112.0 ($\text{C}_{\text{coum.}-6'}$), 107.9 ($\text{C}_{\text{coum.}-8}$), 102.0 ($\text{C}_{\text{arom.}-6}$), 59.9 ($\text{CH}_2\text{-CH}_3$), 59.6 (OMe), 56.5 (C-4), 34.8 (CH_2S), 22.4 ($\text{C}_6\text{-Me}$), 14.6 (CH_2CH_3). EI-MS: m/z (%) = 480 [$\text{M}]^+$. Anal. Calcd. for $\text{C}_{25}\text{H}_{24}\text{N}_2\text{O}_6\text{S}$ (480.53): C, 62.49; H, 5.03; N, 5.83. Found: C, 62.28; H, 4.89; N 5.59 %.

Ethyl 4-(6,8-dichloro-2-oxo-2 H-chromen-3-yl)-6-methyl-2-thioxo-1,2,3,4-tetrahydro-pyrimidine-5-carboxylate (6)

Method A. This compound was prepared following the same procedure of preparation **2** (method B) using aldehyde **5** (243 mg, 1.0 mmol), ethyl acetoacetate (130 mg, 1.0 mmol), and thiourea (190 mg, 2.5 mmol), and HPA supported on Y zeolite (8 wt% NaY + 0.5 mM HPA) in MeCN (15 ml). The mixture was heated under reflux for 9 h to give, after working up, **6** (343 mg, 83 %); m.p. 256–257 °C, $R_f = 0.81$. ^1H NMR (DMSO- d_6): δ 10.36 (s, 1 H, NH-1), 9.91 (d, 1 H, $J_{\text{NH},4} = 5.0$ Hz, NH-3), 8.37 (s, 1 H, $\text{H}_{\text{coum.}-4'}$), 9.97 (d, 1 H, $J_{5,7} = 2.3$ Hz, $\text{H}_{\text{coum.}-7'}$), 8.18 (d, 1 H, $J_{5,7'} = 2.3$ Hz, $\text{H}_{\text{coum.}-5'}$), 4.08, 3.98 (2xq, 2 H, $J = 7.5$ Hz, CH_2CH_3 [$R + S$]), 2.28 (s, 3 H, $\text{C}_6\text{-Me}$), 1.17, 1.08 (2xt, 3 H, $J = 7.5$ Hz, CH_2CH_3 [$R + S$]). ^{13}C NMR (DMSO- d_6): δ 174.6 (C = S), 166.5 (CO_2Et), 163.5 ($\text{C}_{\text{coum.}-2'}$), 144.4 ($\text{C}_{\text{coum.}-8\text{a}'}$), 136.2 ($\text{C}_{\text{coum.}-4'}$), 130.1 ($\text{C}_{\text{coum.}-8'}$), 129.8 ($\text{C}_{\text{coum.}-6'}$ + $\text{C}_{\text{coum.}-7'}$), 126.4 (C-4a' + $\text{C}_{\text{coum.}-3'}$), 122.3 ($\text{C}_{\text{coum.}-5'}$), 104.6 (C-5), 60.1 (CH_2CH_3), 53.1 (C-4), 19.8 ($\text{C}_6\text{-Me}$), 14.2 (CH_2CH_3). EI-MS: m/z (%) = 413 [$\text{M}]^+$. Anal. Calcd. for $\text{C}_{17}\text{H}_{14}\text{Cl}_2\text{N}_2\text{O}_4\text{S}$ (413.28): C, 49.41; H, 3.41; N, 6.78. Found: C, 49.20; H, 3.33; N, 6.52 %.

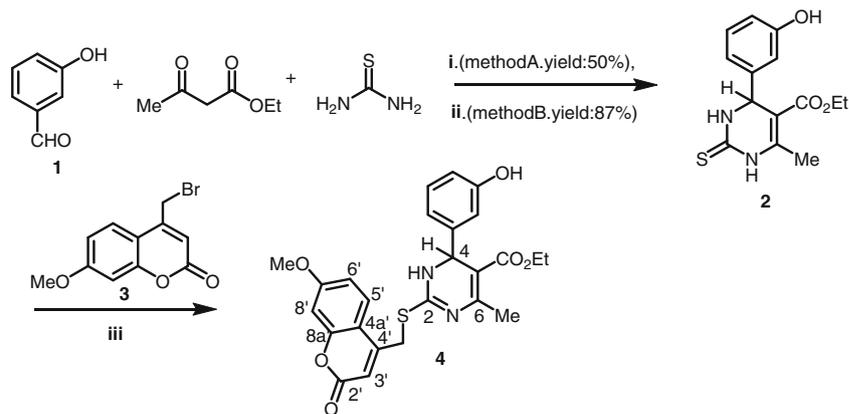
Method B. A mixture of ethyl acetoacetate (42 mg, 0.32 mmol), 6,8-dichloro-2-oxo-2 H-chromene-3-carbaldehyde (**5**) (100 mg, 0.29 mmol) and thiourea (55 mg, 0.73 mmol) in EtOH (10 ml) in the presence conc. Hydrochloric acid (0.5 ml) was heated under reflux for 3 h. After cooling, the mixture was poured onto ice (10 g). The precipitate was filtered and dried and recrystallized from EtOH to give **6** (50 mg, 42 %). The NMR spectra, m.p. and mixed m.p. were identical for those of **6** prepared in method A.

Results and Discussion

Chemistry

Our attention was focused on the synthesis of a new fluorescent coumarin conjugated monastrol via alkylation of thio group of the later scaffold, aiming to study their application as a tracer for detection of hypermetabolic circulating tumor cells (CTC) by fluorescence imaging. Thus, **2** has been prepared (50 %), via Biginelli reaction [14] in a one-pot three-component cyclocondensation reaction of 3-hydroxybenzaldehyde **1**, ethyl acetoacetate and thiourea in the presence of catalytic amount of HCl. The literatures reported various Lewis acids as a catalyst in an attempt to optimize the yield percentage due to the formation of monastrol in *R* and *S* isomers. However, we have optimized the yield of **2** (87 %) by using molybdophosphoric acid (MPA) supported on Y zeolite [15] as an efficient and reusable catalyst in boiling MeCN.

Scheme 1 Reagents and condition: (i) method A: HCl, EtOH, reflux, 3 h; (ii) method B: molybdophosphoric acid (MPA) supported on Y zeolite, MeCN, reflux 7 h; (iii) 3, DMF, K₂CO₃, 100 °C, 5 h

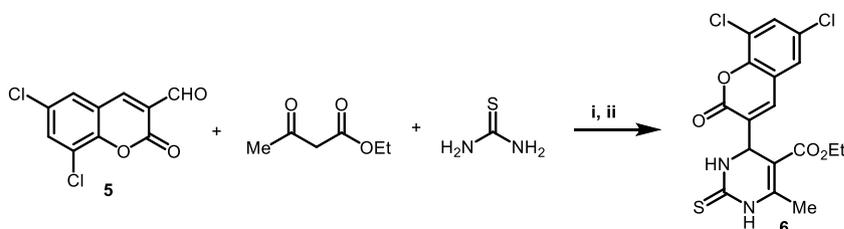


Alkylation of **2** with the commercially available coumarin fluorescent, 4-(bromomethyl)-7-methoxy-2 *H*-chromon-2-one (**3**) in DMF as a solvent and K₂CO₃ as a catalyst at 100 °C to give, after chromatographic purification, **4** (35 %) (Scheme 1).

In the ¹H NMR spectrum of **4**, NH-3 and H-4 appeared as doublets at δ 9.88 and 5.18 ppm ($J_{\text{NH},4} = 4.5$ Hz), respectively, whereas the aromatic protons and OH resonated as a multiplet at the region δ 7.93–7.84 ppm. In the ¹³C NMR spectrum of **6**, the lower field signals at δ 166.7 ppm were assigned to C-6 and CO₂Et of the pyrimidine ring, while the resonance at δ 160.3 ppm was attributed to C-S (C_{pyrimidin.-2}) together with C-2 of coumarin ring. The resonances at δ 160.2 and 155.7 ppm were assigned to C_{coum.-7'} and C_{pyrimid.-4} together with C-4' of coumarin, respectively, meanwhile carbon atoms 4a', 5', 6' and 8' of coumarin appeared at δ 115.5, 128.3, 112.0 and 107.9 ppm, respectively. Carbon atom of the same moiety 3' appeared together with C_{arom.-4} at δ 112.8 ppm, while CH₂S and C-4 of pyrimidine resonated at δ 34.8 and 56.5 ppm, respectively. The difference in the chemical shifts value (~ 14 ppm) between C_{pyrimidin.-2} (C-S) of **4** and C_{pyrimid.-2} (C = S) of **2**, is indicative for the alkylation of **2** by the fluorescent coumarin **3**.

By following the same procedure of preparation of **2**, via Biginelli reaction, **6** was obtained in 42 % yield via a one-pot three-component cyclocondensation reaction of 6,8-dichlorocoumarin 3-carboaldehyde (**5**), ethyl acetoacetate and thiourea in the presence of catalytic amount of hydrochloric acid.

Scheme 2 Reagents and condition: (i) method A: HCl, EtOH, reflux, 3h, 42 % yield; (ii) method B: molybdophosphoric acid (MPA) supported on Y zeolite, MeCN, reflux, 9 h, 83 % yield



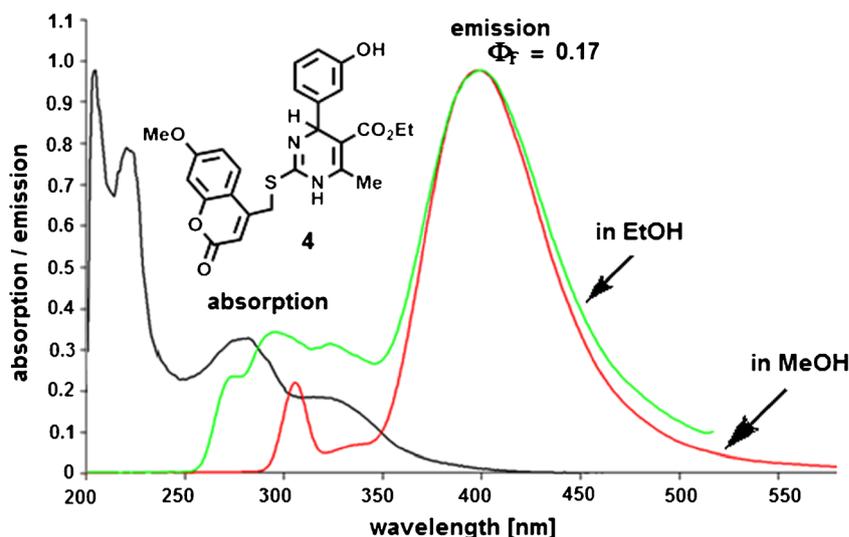
Alternatively, **6** was obtained in 83 % yield, using MPA in boiling MeCN (Scheme 2).

The structure of **6** was assigned from the ¹H and ¹³C NMR spectra, where the singlets at δ 10.36 and 8.37 ppm were assigned to NH-1 and H-4' of the coumarin moiety. The doublets at δ 9.91 and 8.81 ppm ($J_{\text{NH},4} = 5.0$ Hz, $J_{5',7'} = 2.3$ Hz) were attributed to NH-3, and H-5' of the coumarin, whereas the doublet and singlet at δ 9.97 and 2.28 ppm were assigned to H-7 of coumarin and methyl protons at C-6 of pyrimidine ring, respectively. The methylene and methyl protons of CO₂Et group were appeared as two doublets and triplets (as *R* + *S* isomers) at δ 4.08, 3.98 ppm and δ 1.17, 1.08 ppm ($J = 7.5$ Hz), respectively. In the ¹³C NMR spectrum of **6**, the resonance at δ 174.6 ppm was attributed to C = S (C_{pyrimid.-2}), meanwhile the signals at δ 163.5, 136.2, 126.4 ppm were assigned to carbon atoms 2', 4', 4'a and 3' of coumarin scaffold. The carbon atoms 5'-8a' of coumarin were appeared at δ 122.3, 129.8, 130.1 and 144.4 ppm, respectively, in addition to resonance of carbonyl carbon atom (CO₂Et) at δ 166.5 ppm. C-5 and C-4 of pyrimidine ring together with methylene and methyl carbon atoms of ester group were oriented at δ 104.6, 60.1, 53.1 and 14.2 ppm, respectively. The structures of **4** and **6** were further identified from the HSQC [16] and HMBC [17] NMR experiments.

Fluorescence and Photophysical Properties

Derivatization procedures in fluorescence enhancement are especially attractive because they introduce an additional

Fig. 1 UV-visible and fluorescence spectra of compounds **4** in MeOH and EtOH



dimension of selectivity than can simplify analyses in a predictable and reproducible manner. The importance of **3** as an active fluorescent label in biological system, especially for fatty acids [18–21] as well as alkylating agent prompted us for measurement the fluorescence property of coumarinyl-monastrol **4**, with the aim to develop a new tracer for detection of hypermetabolic circulating tumor cells (CTC) by fluorescence imaging. An effective fluorescent agent for biological application has to present a good fluorescent intensity, high quantum yield and high photostability.

The absorption and emission spectra of **4** (Fig. 1) and **6** were measured in MeOH and EtOH. Table 1 illustrated the absorption (λ_{ex}), emission (λ_{em}) and quantum yields (Φ_F) of these analogs, using Rhodamin 6G as standard. Compounds **4** and **6** exhibited λ_{em} = 398 and 339 nm, with quantum yields (Φ_F) = 0.17 and 0.09, respectively, and remarkable stoke's effects (76 and 28, respectively) in MeOH. Unexpectedly, **4** showed a low quantum yield although **3** possesses a spectral property of (λ_{ex} 322 nm; λ_{em} = 395 nm, stoke's effect 73) [21] on derivatization. It is well known that the electron donating groups substituted on coumarin will increase the intermolecular electron transfer and thus enhance the fluorescence of coumarin derivatives, especially at position 7 [22]. Although **4** having coumarin moiety with methoxy group at C-7, but exhibited a low quantum yield. Such data may due to two

factors: distortion of molecule in its own plane or twisted out of plane on intramolecular charge transfer caused by solvent, whereas the other factor may due to collisional quenching occurs when the excited state fluorophore (coumarin-methylthio moiety) is deactivated upon joining with other molecule (monastrol) via the thioether linkage in solution [23].

Regarding compound **6**, the presence of two electron withdrawing groups (3,5-dichloro residues) at coumarin moiety might decrease the conjugation therefore would reduce the fluorescence property of such molecules instead of its optimization. The other reason of low quantum yield of **6** may due to distortion of the planarity of the molecule as in compound **4**, as well as the decrease in π -conjugation because of the saturated carbon atom (C-4) of monastrol. Furthermore, the emission spectrum of **4** has been measured in EtOH which revealed almost a similar pattern of emission as in MeOH (Fig. 1). The fluorescence of compounds **4** and **6** in CH_2Cl_2 showed a red chromic shift (λ_{em} = 386 and 322 nm, respectively), therefore, it is obvious that λ_{em} of both compounds are shifted to longer wavelengths with increasing solvent polarity. Such result indicated that solvent polarity had obvious effect on the emission spectrum [24, 25].

The quantum yield (Φ_F) has been calculated according to the equation reported by C. Vielsack, h.D. Thesis, University of Konstanz, 1999.

Table 1 Absorption and emission data of some new monastrol analogs

| Compd. | Mass [μ g] | Molar Mass [μ g/ μ mol] | λ_{em} [nm] ^a | λ_{em} [nm] ^b | Φ_F ^a | λ_{ex} [nm] ^a | λ_{ex} [nm] ^a |
|-----------------------|-----------------|----------------------------------|----------------------------------|----------------------------------|-----------------------|----------------------------------|----------------------------------|
| 4 | 176 | 480.45 | 398 | 386 | 0.17 | 278 | 322 |
| 6 | 271 | 413.28 | 339 | 322 | 0.09 | 224 | 311 |
| 3 ^c | – | 269.09 | 395 | – | – | 322 | – |

^a in MeOH; ^b in CH_2Cl_2 ; ^c after derivatization [21]; Volume: 25 ml; Ref.-Subst. Rh6G in EtOH (Φ_F : 0.95) at RT; n_D^{20} (MeOH): 1.77; n_D^{20} (EtOH): 1.86/ Equipment and software: Perkin Elmer LS 50, Cary UV/vis 50; SpekWin 1.71.3, FL WinLab.

Acknowledgments We thank Basrah University for a scholarship to Miss Y.A. Marich. Miss A. Friemel of Chemistry Department, Konstanz University, Germany is highly acknowledged for the 2D-NMR experiments.

References

- Bhalla KN (2003) Microtubule-targeted anticancer agents and apoptosis. *Oncogene* 8:9075–9786
- Jordan M (2012) Mechanism of action of antitumor drugs that interact with microtubules and tubulin. *Curr Med Chem Anti-Cancer Agents* 2:1–17
- Perez EA (2009) Microtubule inhibitors: differentiating tubulin-inhibiting agents based on mechanisms of action, clinical activity, and resistance. *Mol Cancer Ther* 8:2086–2095
- Hamel E (1996) Antimitotic natural products and their interactions with tubulin. *Med Res Rev* 16:207–231
- Kingston DGI (2009) Tubulin-interactive natural products as anticancer agents (1). *J Nat Prod* 72:507–517
- Mayer TU, Kapoor T, Haggarty SJ, King RW, Schreiber SL, Mitchison TJ (1999) Small molecule inhibitor of mitotic spindle bipolarity identified in a phenotype-based screen. *Science* 286:971–974
- Kappe CO (2000) Enantioseparation, and determination of absolute configuration of the mitotic kinesin Eg5 inhibitor monastrol. *Tetrahedron* 56:1859–1862
- Haque SA, Hasaka TP, Brooks AD, Lobanov PV, Baas PW (2004) Monastrol, a prototype anti-cancer drug that inhibits a mitotic kinesin, induces rapid bursts of axonal outgrowth from cultured postmitotic neurons. *Cell Motil Cytoskeleton* 58:10–16
- Bose D, Sudharshan M, Chavhan SW (2005) New protocol for biginelli reaction—a practical synthesis of monastrol. *Arxivoc* iii 228–236
- Signore G, Nifosi R, Albertazzi L, Storti B, Bizzarri R (2010) Polarity-sensitive coumarins tailored to live cell imaging. *J Am Chem Soc* 132:1276–1288
- Sun YF, Xu SH, Wu RT, Wang ZY, Zheng ZB, Li JK, Cui YP (2010) The synthesis, structure and photoluminescence of coumarin-based chromophores. *Dyes Pigments* 87:109–118
- Ivana K, Andrea J, Pavol K (2006) Synthesis of coumarin or ferrocene labeled nucleosides via Staudinger ligation. *Beilstein J Org Chem* 2:23–26
- Al-Masoudi NA, Maricha YA, Al-Salihi NJ, Saeed B (2014) Synthesis and modeling study of some potential pyrimidine derivatives as HIV inhibitors. *Z Naturforsch* 69b:913–923
- Biginelli P (1893) Derivati aldeiduredici degli eteri acetyl-e dossal-acetico. *Gaza Chim Ital* 23:360–416
- Moosavifar M (2012) An appropriate one-pot synthesis of dihydropyrimidinones catalyzed by heteropoly acid supported on zeolite: An efficient and reusable catalyst for the Biginelli reaction. *C R Chimie* 15:444–447 references therein cited
- Willker W, Leibfritz KR, Bermel W (1993) Gradient selection in inverse heteronuclear correlation spectroscopy. *Magn Reson Chem* 31:287–292
- Davis AL, Keele J, Laue MD (1992) Experiments for recording pure-absorption heteronuclear correlation spectra using pulsed field gradients. *J Magn Reson* 98:207–216
- Duenges W (1977) 4-bromomethyl-7-methoxycoumarin. *Anal Chem* 49:442–445
- Zhenming X, Wen LL, Qinying D (2007) Determination of sodium mono-fluoroacetate (1080) in biological samples as its bromo-7-methoxycoumarin derivatives by RP-HPLC. *J Chromatogr Sci* 45:405–408
- Güldütuna S, You T, Kurts W, Leuschner U (1993) High performance liquid chromatographic determination of free and conjugated bile acids in serum, liver biopsies, bile, gastric juice and feces by fluorescence labelling. *Clin Chim Acta* 214:195–207
- Xie Z, Shi W, Deng Q (2007) Determination of sodium monofluoroacetate (1080) in biological samples as its 4-bromomethyl-7-methoxycoumarin derivative by RP-HPLC. *J Chromatogr Sci* 45:405–408
- Harishumar N, Mahadevan KM, Masagalli JN, Chandrashekarappa KKH (2012) *Org Synth* 5:196–208
- Tully E, O’Kennedy R (2014) Fluorescent labelling. In: Li D (ed) *Encyclopedia of microfluidics and nanofluidics*. Springer, New York, pp. 1–17
- Chun-feng Y, He-ping Z (2011) Synthesis and spectroscopic study of coumarin derivatives. *Chem Res Chin Univ* 27:599–603
- Donovalová J, Cigán M, Stankovičová H, Gašpar J, Danko M, Gáplovský A, Hrdlovič P (2012) Spectral properties of substituted coumarins in solution and polymer matrices. *Molecules* 17:3259–3276