ISSN 1070-3632, Russian Journal of General Chemistry, 2016, Vol. 86, No. 6, pp. 1292–1299. © Pleiades Publishing, Ltd., 2016. Original Russian Text © N.M. Chernov, P.V. Filippova, I.P. Yakovlev, V.E. Zakhs, A.V. Belyakov, 2016, published in Zhurnal Obshchei Khimii, 2016, Vol. 86, No. 6, pp. 962–969.

Synthesis and Reactivity of 4-Hydroxy-5-methyl-2-(2-oxo-2*H*-chromen-3-yl)-6*H*-1,3-oxazin-6-ones

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Received February 18, 2016

Abstract—4-Hydroxy-6*H*-1,3-oxazin-6-ones with a coumarin fragment in position 2 of the oxazine ring were synthesized. Their hydrolysis, alcoholysis, and hydrazinolysis afforded a number of new coumarin derivatives containing malonamic acid and 1,2,4-triazole residues. The low stability of the title compounds toward oxygencentered nucleophiles was interpreted by quantum chemical calculations.

Keywords: 4-Hydroxy-6*H*-1,3-oxazin-6-one, coumarin, malonamic acid, 1,2,4-triazole, quantum chemical calculations

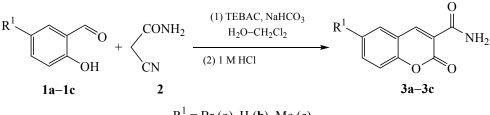
DOI: 10.1134/S1070363216060128

Coumarin derivatives constitute an abundant group of natural and synthetic compounds, many of which exhibit important pharmacological activities. In particular, a number of indirect anticoagulants (e.g., warfarin) are 4-hydroxycoumarin derivatives. Nowadays, extensive search for biologically active substances is performed among coumarin derivatives containing an amide or hydrazide moiety, as well as nitrogen heterocycle residues, in the 3-position [1–4]. Such compounds were found to possess antitubercular [1], antitumor [2, 3], and many other kinds of biological activity [4].

On the other hand, 4-hydroxy-6*H*-1,3-oxazin-6ones are known to react with nucleophiles to give various acyclic (malonamic acid derivatives [5]) and heterocyclic systems (azoles [6], azines [7]). Herein, we report the synthesis of 1,3-oxazines containing a coumarin fragment and their reactions with nucleophiles with the goal of obtaining new biologically active compounds.

A convenient procedure for the synthesis of 4hydroxy-6H-1,3-oxazin-6-ones is based on the well studied reaction of the corresponding carboxylic acid amides with malonyl chlorides [5, 7]. The required coumarin-3-carboxamides **3a**-**3c** were prepared by a modified known procedure [8, 9] (Scheme 1). According to published data, the Knoevenagel condensation of salicylaldehydes 1a-1c with cyanoacetamide (2) is carried out in a two-phase system water-methylene chloride (9:1) in the presence of a phase-transfer catalyst. The subsequent acidification of the reaction mixture leads to closure of pyran ring with formation of coumarin-3-carboxamides 3a-3c in 84-90% yield. Instead of cetyl(trimethyl)ammonium bromide [8] or Katamin AB [9], as phase-transfer catalyst we used more accessible benzyl(triethyl)ammonium chloride (BTEAC). In this case, the yields of the target products did not decrease, and the experimental procedure was not complicated. Addition of methylene chloride to the reaction mixture made it possible to accomplish the reaction with solid 5-bromoand 5-methylsalicylaldehydes 1a and 1c. The melting point and ¹H NMR spectrum of 3b coincided with published data for 2-oxo-2H-chromene-3-carboxamide [9]. Apart from aromatic proton signals, the ¹H NMR spectrum of **3b** characteristically showed two broadened singlets due to protons of the NH₂ group (δ 7.91 and 8.08 ppm) and 4-H signal at δ 8.86 ppm. The structure of 3a-3c was also confirmed by ¹³C NMR spectra and elemental analyses (Tables 1, 2).

Amides **3a–3c** were reacted with methylmalonyl chloride (4) in aprotic solvents such as carbon



 $R^{1} = Br(a), H(b), Me(c).$

tetrachloride, benzene, 1,2-dichloroethane, and acetonitrile at the boiling point. Acetonitrile turned out to be the best solvent; it ensured reaction completion in 2-3 h, and the yield of 1,3-oxazines 5a-5c was 76-84% (Scheme 2). 1,2-Dichloroethane was less efficient: the reactions were complete in 4-6 h, and the yields were slightly lower (70–78%). Insofar as initial amides 3a– 3c are poorly soluble in carbon tetrachloride and benzene, the reaction time in these solvents increased to 8-10 h, and the yield of 5a-5c decreased to 20-24%, which may be due to polymerization of methylmalonyl chloride on prolonged heating.

The ¹H NMR spectra of 5a-5c contained signals from aromatic protons of the coumarin fragment (\delta 7.43–8.86 ppm), a singlet at δ 1.82–1.83 ppm from the

Comp.	\mathbf{R}^1	D ²	R ³	Yield,	mp,	Found, %			Calculated, %			
no.	K.	\mathbb{R}^2	R	%	°C	С	Н	Ν	Formula	С	Н	N
3a	Br	_	_	86	294–296	44.03	1.89	5.17	C ₁₀ H ₆ BrNO ₃	44.81	2.26	5.23
3b	Н	_	_	90	263–264	63.31	3.51	7.04	C ₁₀ H ₇ NO ₃	63.49	3.73	7.40
3c	Me	_	_	84	218–220	64.77	4.12	6.76	C ₁₁ H ₉ NO ₃	65.02	4.46	6.89
5a	Br	_	_	84	251-252	47.93	2.20	3.89	C14H8BrNO5	48.03	2.30	4.00
5b	Н	_	_	76	197–200	61.85	3.29	5.00	C ₁₄ H ₉ NO ₅	62.00	3.34	5.16
5c	Me	_	_	79	205–207	63.01	3.65	4.72	C ₁₄ H ₁₁ NO ₅	63.16	3.89	4.91
6a	Br	Н	_	95	209–210	45.46	2.61	3.77	C14H10BrNO6	45.68	2.74	3.80
6b	Н	Н	_	97	191–193	57.92	3.64	4.80	C ₁₄ H ₁₁ NO ₆	58.14	3.83	4.84
6c	Me	Н	_	95	194–195	59.33	4.12	4.49	C ₁₅ H ₁₃ NO ₆	59.41	4.32	4.62
7a	Br	Et	-	87	173–174	48.44	3.51	3.47	C ₁₆ H ₁₄ BrNO ₆	48.51	3.56	3.54
7b	Н	Et	-	91	144–145	60.46	4.52	4.35	C ₁₆ H ₁₅ NO ₆	60.57	4.77	4.41
7c	Me	Et	_	94	153–153	61.43	4.79	4.07	C ₁₇ H ₁₇ NO ₆	61.63	5.17	4.23
8a	Br	_	Н	69	288–290	41.98	2.35	6.91	$C_{14}H_{10}Br_2N_2O_2$	42.24	2.53	7.04
9a	Br	_	Н	81	270–271	46.07	2.53	11.21	$C_{14}H_{10}BrN_3O_4$	46.18	2.77	11.54
9b	Н	_	Н	72	212–214	58.68	3.82	14.42	$C_{14}H_{11}N_3O_4$	58.95	3.89	14.73
9c	Me	_	Н	85	210–211	59.93	4.17	13.86	$C_{15}H_{13}N_3O_4$	60.20	4.38	14.04
10a	Br	_	Ph	89	172–174	54.40	2.98	9.33	$C_{20}H_{14}BrN_3O_4$	54.56	3.21	9.54
10b	Н	_	Ph	81	153–155	66.04	4.12	11.39	$C_{20}H_{15}N_3O_4$	66.48	4.18	11.63
10c	Me	_	Ph	83	158–159	66.97	4.36	10.97	$C_{21}H_{17}N_3O_4$	67.19	4.56	11.19

Table 1. Yields, melting points, and elemental analyses of compounds 3 and 5-10

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Table 2. ¹ H and ¹³ C	NMR spectra (DMSO-d ₆) of compounds 3 and 5–10
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14010 20	H and C NMR spectra (DMSO- a_6) of compounds 5 and 5–10	
Comp. no.	¹ H NMR spectrum, δ , ppm (<i>J</i> , Hz)	¹³ C NMR spectrum, δ_C , ppm
3 a	7.44 d (1H, 8-H, <i>J</i> = 8.8), 7.86 d.d (1H, 7-H, <i>J</i> = 2.4, 8.8), 7.97 br. s (1H, NH), 8.04 br.s (1H, NH), 8.20 d (1H, 5-H, <i>J</i> = 2.4), 8.81 s (1H, 4-H)	
3b	7.43 t (1H, 6-H, J = 7.8), 7.48 d (1H, 8-H, J = 7.8), 7.74 t (1H, 7-H, J = 7.8), 7.91 br.s (1H, NH), 7.96 d (1H, 5-H, J = 7.8), 8.08 br.s (1H, NH), 8.86 s (1H, 4-H)	
3c	2.36 s (3H, CH ₃), 7.34 d (1H, 8-H, $J = 8.5$), 7.52 d.d (1H, 7-H, $J = 1.3$, 8.5), 7.68 d (1H, 5-H, $J = 1.3$), 7.90 br.s (1H, NH), 8.08 br.s (1H, NH), 8.74 s (1H, 4-H)	
5a	1.83 s (3H, CH ₃), 7.45 d (1H, 8-H, <i>J</i> = 8.8), 7.90 d (1H, 7-H, <i>J</i> = 8.8), 8.25 s (1H, 5-H), 8.74 s (1H, 4-H), 12.66 br.s (1H, OH)	8.6, 91.6, 117.0, 118.9, 119.0, 120.3, 132.6, 137.2, 146.8, 153.8, 155.6, 158.0, 162.0, 165.5
5b	1.82 s (3H, CH ₃), 7.46 m (2H, 6-H, 8-H), 7.76 t (1H, 7-H, $J = 8.0$), 7.97 d (1H, 5-H, $J = 8.0$), 8.78 s (1H, S ⁴ N), 12.65 br. s (1H, OH)	8.5, 91.2, 116.6, 117.8, 118.4, 118.6, 125.5, 130.8, 135.1, 148.2, 154.5, 156.1, 158.4, 165.6
5c	1.83 s (3H, CH ₃), 7.37 d (1H, 8-H, <i>J</i> = 8.5), 7.59 d (1H, 7-H, <i>J</i> = 8.5), 7.75 s (1H, 5-H), 8.69 s (1H, 4-H), 12.65 br.s (1H, OH)	8.4, 20.9, 90.5, 116.4, 118.7, 119.3, 130.4, 134.7, 135.6, 148.3, 152.5, 160.5, 162.2, 163.0, 165.6
6a	1.29 d (3H, <i>J</i> = 7.0, CH ₃), 3.97 q (1H, <i>J</i> = 7.0, CH), 7.51 d (1H, <i>J</i> = 8.8, 8-H), 7.93 d.d (1H, 7-H, <i>J</i> = 2.4, 8.8), 8.23 d (1H, 5-H, <i>J</i> = 2.4), 8.71 s (1H, 4-H), 11.30 s (1H, NH), 12.81 br.s (1H, COOH)	13.6, 48.0, 117.3, 119.1, 120.5, 122.1, 132.6, 137.2, 146.3, 153.5, 159.4, 162.3, 171.1, 171.7
6b	1.31 d (3H, CH ₃ , $J = 7.2$), 4.01 q (1H, CH, $J = 7.2$), 7.48 t (1H, 6-H, $J = 7.5$), 7.54 d (1H, 8-H, $J = 8.3$), 7.80 t (1H, 7-H, $J = 8.3$), 7.98 d (1H, 5-H, $J = 8.3$), 8.83 s (1H, 4-H), 11.29 s (1H, NH), 12.73 br.s (1H, COOH)	13.6, 48.1, 116.8, 118.6, 120.1, 125.9, 130.9, 135.3, 148.5, 154.4, 160.2, 162.1, 171.2, 171.8.
6c	1.31 d (3H, CH ₃ , <i>J</i> = 7.3), 4.03 q (1H, CH, <i>J</i> = 7.3), 7.40 d (1H, 8- H, <i>J</i> = 8.4), 7.58 d (1H, 7-H, <i>J</i> = 8.4), 7.71 s (1H, 5-H), 8.74 s (1H, 4-H), 11.28 s (1H, NH), 12.71 br.s (1H, COOH)	13.6, 20.6, 48.2, 116.5, 118.4, 119.7, 130.3, 135.3, 136.2, 148.7, 152.7, 160.3, 162.0, 171.1, 171.8.
7a	1.19 t (3H, CH ₃ CH ₂ , $J = 7.0$), 1.32 d (3H, CH ₃ , $J = 7.3$), 4.06 m (3H, CH ₃ CH ₂ , CH), 7.50 d (1H, 8-H, $J = 8.8$), 7.91 d (1H, 7-H, $J = 8.8$), 8.22 s (1H, 5-H), 8.71 s (1H, 4-H), 11.33 s (1H, NH)	13.5, 14.4, 48.0, 61.3, 117.3, 119.1, 120.5, 121.9, 132.7, 137.2, 146.6, 153.5, 159.4, 162.3, 170.2, 170.7
7b	1.14 t (3H, CH ₃ CH ₂ , $J = 7.0$), 1.29 d (3H, CH ₃ , $J = 7.2$), 3.99 q (1H, CH, $J = 7.2$), 4.10 q (2H, CH ₃ CH ₂ , $J = 7.0$), 7.49 t (1H, 6-H, $J = 7.5$), 7.55 d (1H, 8-H, $J = 8.3$), 7.78 t (1H, 7-H, $J = 7.8$), 7.98 d (1H, 5-H, $J = 7.8$), 8.74 s (1H, 4-H), 11.22 s (1H, NH)	13.6, 14.4, 48.1, 61.5, 116.7, 118.7, 120.2, 125.8, 130.8, 135.2, 148.5, 154.4, 160.1, 162.2, 171.1, 172.8
7c	1.19 t (3H, CH ₃ CH ₂ , J = 7.0), 1.30 d (3H, CH ₃ , J = 7.3), 4.00 q (1H, CH, J = 7.3), 4.11 q (2H, CH ₃ CH ₂ , J = 7.0), 7.42 d (1H, 8-H, J = 8.4), 7.59 d (1H, 7-H, J = 8.4), 7.70 s (1H, 5-H), 8.72 s (1H, 4-H), 11.32 s (1H, NH)	13.6, 14.4, 20.8, 48.2, 61.4, 116.6, 119.2, 120.8, 120.9, 132.4, 137.6, 146.3, 152.3, 159.7, 162.2, 170.8, 172.6
8a	6.97 d (1H, 3-H, <i>J</i> = 8.8), 7.53 d. d (1H, 4-H, <i>J</i> = 2.0, 8.0), 7.90 d (1H, 6-H, <i>J</i> = 2.0), 8.93 s (1H, CH=N), 11.18 br. s (1H, OH)	111.0, 119.4, 121.1, 132.0, 135.9, 158.2, 161.1
9a	1.49 d (3H, CH ₃ , $J = 7.3$), 3.92 q (1H, CH, $J = 7.3$), 7.45 d (1H, 8- H, $J = 8.8$), 7.82 d.d (1H, 7-H, $J = 2.3$, 8.8), 8.22 d (1H, 5-H, $J = 2.3$), 8.76 s (1H, 4-H), 12.24 br. s (1H, NH), 13.87 br.s (1H, COOH)	16.4, 39.3, 117.0, 117.1, 118.8, 121.1, 131.9, 135.6, 141.1, 151.1, 152.7, 158.1, 161.5, 173.8
9b	1.48 d (3H, CH ₃ , $J = 7.1$), 3.90 q (1H, CH, $J = 7.1$), 7.43 t (1H, 6-H, $J = 7.4$), 7.50 d (1H, 8-H, $J = 8.2$), 7.70 t (1H, 7-H, $J = 7.8$), 7.99 d (1H, 5-H, $J = 7.8$), 8.81 s (1H, 2-H), 12.85 br.s (2H, NH, COOH)	16.4, 39.3, 115.8, 116.6, 119.2, 125.4, 130.1, 133.4, 142.5, 150.4, 153.7, 158.6, 161.9, 173.9
9c	1.48 d (3H, CH ₃ , <i>J</i> = 7.3), 3.91 q (1H, CH, <i>J</i> = 7.3), 7.39 d (1H, 8- H, <i>J</i> = 8.3), 7.51 d.d (1H, 7-H, <i>J</i> = 1.5, 8.3), 7.75 d (1H, 5-H, <i>J</i> = 1.5), 8.72 s (1H, 4-H), 12.33 br.s (1H, NH), 13.90 br.s (1H, COOH)	16.4, 20.8, 39.4, 115.7, 116.4, 118.9, 129.6, 134.4, 134.7, 142.5, 150.0, 151.9, 158.8, 162.0, 173.8

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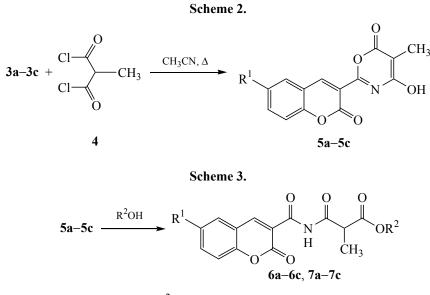
Comp. no.	¹ H NMR spectrum, δ , ppm (<i>J</i> , Hz)	13 C NMR spectrum, δ_{C} , ppm
10a	1.54 d (3H, CH ₃ , <i>J</i> = 7.3), 4.07 q (1H, CH, <i>J</i> = 7.3), 7.42 d (1H, 8- H, <i>J</i> = 8.9), 7.62 m (5H, Ph), 7.80 d. d (1H, 7-H, <i>J</i> = 2.4, 8.9), 8.23 d (1H, 5-H, <i>J</i> = 2.4), 8.73 s (1H, 4-H), 12.96 br.s (1H, COOH)	
10b	1.55 d (3H, CH ₃ , J = 7.3), 4.07 q (1H, CH, J = 7.3), 7.40 t (1H, 6-H, J = 7.5), 7.45 d (1H, 8-H, J = 8.3), 7.63 m (6H, Ph, 7-H), 7.96 d (1H, 5-H, J = 7.5), 8.75 s (1H, 4-H), 12.81 br.s (1H, COOH)	16.2, 37.3, 116.4, 119.1, 119.2, 125.2, 126.1, 130.0, 130.1, 130.3, 133.2, 137.0, 143.0, 153.5, 155.8, 156.3, 156.8, 172.4
10c	1.54 d (3H, CH ₃ , $J = 7.3$), 4.06 q (1H, CH, $J = 7.3$), 7.36 d (1H, 8- H, $J = 8.2$), 7.50 d.d (1H, 7-H, $J = 1.5$, 8.2), 7.63 m (5H, Ph), 7.73 d (1H, 5-H, $J = 1.5$), 8.68 s (1H, 4-H), 12.78 br.s (1H, COOH)	

Table 2. (Contd.)

5-methyl group, and a broadened singlet of the hydroxy proton at δ 12.65–12.66 ppm. In the ¹³C NMR spectra of these compounds, the 5-CH₃ and C⁵ carbon nuclei resonated at δ_C 8.4–8.6 and 90.5–91.6 ppm, respectively, and signals of C², C⁴, and C⁶ of the oxazine ring were located in the downfield region (δ_C 156.1–165.6 ppm). The structure of **5a–5c** was also confirmed by elemental analyses (Tables 1, 2) and by the high-resolution mass spectrum of **5b**.

While determining the structure of we have noticed their anomalously low hydrolytic stability. Compounds **5a–5c** underwent partial or complete hydrolysis via cleavage of the C⁶–O bond during recording their NMR spectra in DMSO containing traces of water. The hydrolysis products were *N*-acylmalonamic acids **6a**– **6c**. Complete hydrolysis of **5a–5c** with quantitative formation of **6a–6c** was also observed on exposure to atmospheric moisture for 10–12 days. Further studies showed that oxazines **5a–5c** equally readily (at room temperature) reacted with another O-nucleophile, ethanol, to form malonamic acid esters **7a–7c** which were isolated in 87–94% yield (Scheme 3).

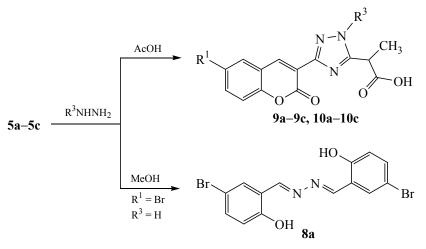
It should be noted that such hydrolytic stability is not typical of 2-aryl- and 2-styryl-substituted 4hydroxy-5-methyl-6H-1,3-oxazin-6-ones [6]. The latter do react with O-nucleophiles to give products of oxazine ring cleavage at the C⁶–O bond [5, 6], the corresponding malonamic acids or their esters analogous to **6** and **7**. However, these reactions occur under more severe conditions, on heating in aqueous



 $R^2 = H$ (6a–6c), Et (7a–7c).

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 $R^3 = H(9), Ph(10).$

acetonitrile or in alcohols at the boiling point for 1-2 h.

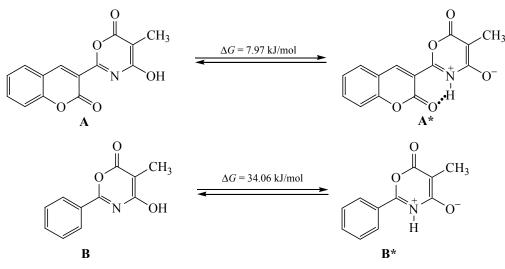
As shown previously [5, 6], reactions of 4-hydroxy-6H-1,3-oxazin-6-ones react with binucleophiles, such as hydrazines and guanidine, in methanol involve opening of the oxazine ring via cleavage of the C^2 –O bond usually with subsequent recyclization to azole or azine heterocyclic system. Analogous reaction of 5a with hydrazine in methanol resulted in decomposition of the pyran ring with formation of aldehyde azine 8a as the major product (yield 69%; Scheme 4). Compound 8a was identified by comparing its melting point and ¹H NMR data with those reported in [10] for 5-bromosalicylaldehyde azine. According to published data [10, 11], such behavior is typical of 3-acetylcoumarin and coumarin-3-carboxylic acid derivatives. Oxazines 5a-5c can also be regarded as analogs of the latter. When the reactions of 5a–5c with hydrazine and phenylhydrazine were carried out in glacial acetic acid, the nucleophilic attack was directed mainly at C^2 of the oxazine ring, and the final products were coumarincontaining 1,2,4-triazole derivatives 9a-9c and 10a-10c (yield 72-89%; Scheme 4).

The structure of **6a–6c**, **7a–7c**, **9a–9c**, and **10a–10c** was confirmed by elemental analyses and ¹H and ¹³C NMR spectra (Tables 1, 2). Compound **9b** was also characterized by the high-resolution mass spectrum. In the ¹H NMR spectra of **6**, **7**, **9**, and **10** we observed two signals belonging to a CH₃CH fragment (a doublet at δ 1.29–1.54 ppm and a quartet at δ 3.90–4.07 ppm) instead of 5-CH₃ singlet typical of initial oxazines **5a**–

5c: the corresponding carbon nuclei resonated in the ¹³C NMR spectra at $\delta_{\rm C}$ 13.4–16.2 (CH₃) and 37.5–48.2 ppm (CH). Malonamic acid derivatives **6** and **7** showed a singlet at δ 11.22–11.33 ppm in the ¹H NMR spectra due to the NH proton, and triazoles **9** and **10** showed a broadened singlet at δ 12.85–13.87 ppm due to hydroxy proton of the COOH group). Signals in the region $\delta_{\rm C}$ 155.9–158.8 ppm were assigned to carbon atoms of the triazole ring.

Our results led us to conclude that the presence of a coumarin fragment in the 2-position strongly affects the chemical behavior of the 1,3-oxazine system. Presumably, the 2-substituent affects geometric parameters of the oxazine ring in 5a-5c, which largely determine their reactivity toward O-nucleophiles. In order to evaluate the effect of the 2-substituent we performed B3LYP/6-31+G(d,p) quantum chemical calculations (GAUSSIAN 09 A.02 [12]) of the geometric parameters and free energies of tautomeric forms of 2-coumarinyloxazine (A and A*) and 2phenyloxazine (B and B*). While optimizing the geometric parameters, stationary points on the potential energy surfaces were identified as energy minima by the absence of imaginary vibration frequencies in the corresponding Hessian matrices.

We previously showed [13] by both theoretical and experimental methods that 4-hydroxy-1,3-oxazin-6ones exist preferentially as 4-hydroxy-6-oxo tautomers (**A** and **B**) among four theoretically possible ones. One of the minor tautomers which could exist in equilibrium with the major form is dipolar ion A^* or



Scheme 4.

B^{*}. On the basis of the calculated Gibbs energies we estimated the compositions of tautomeric mixtures A/A* and **B**/**B*** (χ , %) by the equation $\Delta G = -R T \ln K$ (Table 3). It is seen that the transformation of tautomer A of 5b into structure A* should be relatively facile and that analogous transformation of 2-phenyloxazine is hindered. The equilibrium concentration of A* is 3.85%, whereas zwitterionic tautomer B* of 2phenyloxazine is almost absent in the equilibrium mixture. This difference in the tautomeric compositions may be related to the effect of the oxo group in the coumarin fragment of 5b, which stabilizes structure A* due to formation of hydrogen bond C=O···H-N (d = 1.962 Å). The formation of this hydrogen bond was confirmed by topological analysis of electron density distribution in tautomer A* by the atoms-inmolecules method [14] (AIMAll [15]) (see figure). A critical point of the (3, -1) type was found for tautomer A*, which corresponded to $N-H\cdots O$ hydrogen bonding. The bond energy was determined from the potential energy density at the critical point according to the procedure described in [16], $E_{\rm HB} = 28.4$ kJ/mol.

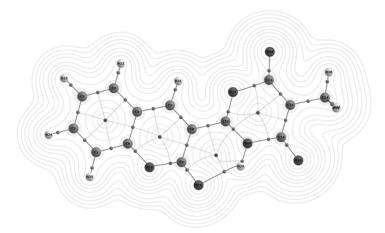
Assuming that reactions of 1,3-oxazines **5a–5c** with O-nucleophiles are favored by anomalously high concentration of dipolar structure A^* , we compared the reactivities of tautomers **A** and A^* . As we already noted, the hydrolysis and alcoholysis of 1,3-oxazines involves cleavage of the C⁶–O bond. Its strength was quantitatively assessed by its length and order in structures **A**/**A*** and **B**/**B*** (Table 3). The bond orders were calculated according to Wiberg using the NBO (natural bond orbital) module implemented in Gaussian 09. The results showed that the transforma-

tion of **A** into **A**^{*} is accompanied by appreciable elongation of C⁶–O in **5b** (from 1.433 to 1.618 Å); simultaneously, the C⁶–O bond order decreases. Analogous pattern was observed for tautomers **B** and **B**^{*} of 2-phenyloxazine. Thus, dissociation of the C⁶–O bond in dipolar tautomers **A**^{*} and **B**^{*} is much easier than in major 4-hydroxy-6-oxo tautomers **A** and **B**, and the low stability of **5a–5c** toward hydrolysis and alcoholysis is determined by the presence of dipolar tautomer **A**^{*} which readily undergoes ring opening because of low strength of the C⁶–O bond.

In summary, we have synthesized for the first time 4-hydroxy-6*H*-1,3-oxazin-6-one derivatives containing a coumarin fragment in the 2-position of the oxazine ring. These compounds react with with nucleophiles to give previously unknown coumarin derivatives possessing malonamic acid and 1,2,4-triazole fragments. Taking into account published data [1–4], the synthesized compounds attract interest as potential biologically active substances with antitumor, antitubercular, and other valuable therapeutic properties.

Table 3. Characterisitc of tautomers A/A* and B/B*

Parameter	Oxaz	in 5b	2-Phenyloxazine		
Falainetei	Α	A*	В	B*	
Gibbs energy ΔG , kJ/ mol	7.973		34.059		
Equilibrium concentration χ , %	96.15	3.85	99.9999	0.0001	
Bond length C ⁶ –O, Å Bond order C ⁶ –O	1.433 0.840	1.618 0.579	1.435 0.838	1.613 0.587	



Topological analysis of tautomer A*.

According to quantum chemical calculations, high reactivity of coumarin-containing 1,3-oxazines toward O-nucleophiles is related to their partial existence as reactive zwitterionic tautomers.

EXPERIMENTAL

The ¹H and ¹³C NMR spectra were recorded from solutions in DMSO- d_6 on a Bruker Avance III spectrometer (400 and 100 MHz, respectively), using the residual proton and carbon signals of the solvent as reference. The elemental analyses were obtained on a LECO CHNS-932 analyzer. The mass spectra were recorded on a Bruker micrOTOF instrument (electrospray ionization, positive ion detection). The purity of the isolated compounds was checked, and the progress of reactions was monitored, by thin-layer chromatography on Silica gel 60 F254 plates (Merck) using ethyl acetate as eluent; spots were visualized under UV light. The solvents were purified and dried by standard methods. The yields, melting points, and analytical data of compound 5-10 are given in Table 1, and their spectral parameters are presented in Table 2.

6-Bromo-2-oxo-2*H***-chromene-3-carboxamide (3a).** A mixture of 20 g (10 mmol) of 5-bromosalicylaldehyde (**1a**), 12.6 g (15 mmol) of cyanoacetamide (**2**), 2.3 g (1 mmol) of benzyl(triethyl)ammonium chloride, 5 g of sodium hydrogen carbonate, 450 mL of water, and 50 mL of methylene chloride was stirred for 3 h at 18–20°C. The mixture was acidified with 1 M aqueous HCl to pH 2–3 and heated for 2 h at 60–65°C. The mixture was then cooled, and the white solid was filtered off, washed with water, and dried at 50°C. Yield 23.0 g (86%). Coumarin-3-carboxamides **3b** and **3c** were synthesized in a similar way.

4-Hydroxy-5-methyl-2-(2-oxo-2*H***-chromen-3-yl)-6***H***-1,3-oxazin-6-one (5b). A mixture of 1.5 g (8 mmol) of amide 3b and 1.8 g (12 mmol) of methylmalonyl chloride (4) in 20 mL of anhydrous acetonitrile was refluxed for 2–3 h (until the initial amide disappeared according to the TLC data). The mixture was cooled, and the yellow precipitate was filtered off, washed with acetonitrile, and dried in a vacuum desiccator over phosphoric anhydride. Yield 1.7 g (76%). Mass spectrum (HRMS ESI): m/z 294.0377 [M + Na]^+; calculated for C₁₄H₉NNaO₅: 294.0374.**

Oxazines **5a** and **5c** were synthesized in a similar way.

3-[(6-Bromo-2-oxo-2*H***-chromen-3-ylcarbonyl)amino]-2-methyl-3-oxopropanoic acid (6a).** Oxazine **5a**, 1.4 g (4 mmol), was kept on exposure to air for 10– 12 days (TLC). Yield 1.4 g (95%).

N-Acylmalonamic acids **6b** and **6c** were synthesized in a similar way.

Ethyl 3-[(6-bromo-2-oxo-2*H***-chromen-3-ylcarbonyl)amino]-2-methyl-3-oxopropanoate (7a).** A suspension of 1.4 g (4 mmol) of oxazine **5a** in 10 mL of ethanol was stirred for 3–4 h. When the reaction was complete, the precipitate was filtered off and dried in air. Yield 1.4 g (87%).

Esters 7b and 7c were synthesized in a similar way.

5-Bromosalicylaldehyde azine (8a). A mixture of 1.4 g (4 mmol) of oxazine **5a** and 200 μ L (200 mg, 4 mmol) of hydrazine hydrate in 10 mL of methanol was stirred for 12 h until the initial oxazine disappeared (TLC). The precipitate was filtered off,

washed with methanol, and dried in air. Yield 0.55 g (69%), mp 288–290°C [10].

2-[3-(2-Oxo-2*H***-chromen-3-yl)-1***H***-1,2,4-triazol-5-yl]propanoic acid (9b).** A mixture of 1.1 g (4 mmol) of oxazine **5b** and 200 μ L (200 mg, 4 mmol) of hydrazine hydrate in 10 mL of glacial acetic acid was stirred for 2–3 days (TLC). The white solid was filtered off, washed with acetic acid, and dried under reduced pressure. Yield 0.9 g (72%). Mass spectrum (HRMS ESI): *m/z* 308.0640 [*M* + Na]⁺; calculated for C₁₄H₁₁N₃NaO₄: 308.0642.

Triazoles **9a**, **9c** and **10a–10c** were synthesized in a similar way. In the synthesis of 1-phenyl-1,2,4-triazoles **10a–10c** phenylhydrazine was used instead of hydrazine hydrate.

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