Structure–activity relationship studies of CNS agents. Part 29. N-Methylpiperazino-substituted derivatives of quinazoline, phthalazine and quinoline as novel α_1 , 5-HT_{1A} and 5-HT_{2A} receptor ligands

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Summary — New *N*-methylpiperazino-substituted quinazolines 8 and 9, phthalazine 13, and quinoline 19 have been synthesized. The receptor binding profiles (α_1 , 5-HT_{1A}, 5-HT_{2A}) of these compounds and their analogs (7–22) have been determined. It has been demonstrated that orientation of a local dipole moment of the heteroaromatic ring system affects both the α_1 and 5-HT_{2A} affinity of the investigated class of ligands. Distortion of the coplanar unfused heteroaromatic ring system results in a decreased 5-HT_{2A} affinity 4-(4-Methylpiperazino)-2-(2-thienyl)quinoline 18 is the most active and selective α_1 ligand ($K_1 = 4.9$ nM) with a much lower affinity for 5-HT_{1A} ($K_1 = 3420$ nM) and 5-HT_{2A} ($K_1 = 211$ nM) receptors.

N-methylpiperazine / prazosin analog / α_1 receptor ligands / 5-HT_{1A} receptor ligands / 5-HT_{2A} receptor ligands / structure– affinity relationships

Introduction

Prazosin 1 and doxazosin 4 are regarded as parent compounds of a vast number of different α_1 -adrenergic receptor ligands [1]. Compounds 1 and 4 are highly potent and selective α_1 ligands ($K_1 = 0.19$ and 1.1 nM, respectively) [2, 3] and are classified as α_1 adrenoreceptor antagonists [1-3]. The basic structure of prazosin (2-4-diamino-6,7-dimethoxyquinazoline) has served as the core of a large number of derivatives (eg, 2-6, fig 1). Chern et al [4] showed that the 6,7dimethoxy substituents of the quinazoline nucleus are not necessary for the formation of a complex of derivatives 2 and 3 with α_1 receptors. They also found that various structural modifications of 2 and 3 affected the α_1 affinity of the particular derivatives. The highest α_1 affinity ($K_1 \sim 0.07$ nM) was observed for derivatives of 2 with $R^1 = CH_1$ and $R^2 = 2$ -OCH₃ and an arylpiperazine fragment attached to the heterocyclic ring system in position 4. and 3 with $X = OCH_3$ and $R = 2-OCH_3$. The lowest affinity ($K_1 > 10$ nM) was found for 2 with $R^{1} = CH_{3}$ and $R^{2} = H$, and the arylpiperazine fragment at position 5, and for 3 with $X = SCH_3$ and $R = H_2$. Campbell et al [3] reported on the structure-affinity relationships of doxazosin modified in the benzodioxane and piperazine portions (4 and 5, fig 1). The majority of derivatives of 5 showed very high affinity $(0.7 \le K_1 \le 7.8 \text{ nm})$ for α_1 receptors, except for a single derivative ($R^1 = 6.7$ -di-Cl, $R^2 = H$, X = Y = CH_2 , $K_1 = 13.3$ nM). Other structural modifications (X, Y, $\overline{R^1}$) had a small effect on affinity [3]. Alabaster et al [2] analyzed the α_1 -receptor affinity of a series of 1-amino-6,7-dimethoxyquinazolines substituted at position 2 with a complex piperidine fragment (6). All derivatives containing XCONR¹R² substituents at position 4 of the piperidine ring showed a high α_1 affinity (0.1 $\leq K_1 \leq$ 1.73 nM). Modifications of the XCONR¹R² substituent at position 3 of the piperidine ring resulted in a dramatic loss of affinity ($K_1 = 117$ nM for 3-CON(C₂H₅)₂) [2].

Many complex derivatives of prazosin show a high affinity for α_1 receptors ($K_1 < 10$ nM). The applied modifications of the structure include, in general, substituents at the 2-amino function of the 2,4-diamino-6,7-dimethoxyquinazoline skeleton. A 4-amino-2-(N-piperazino)-6,7-dimethoxyquinazoline core of the class

Abbreviations: 2-PP[.] 2-(*N*-piperazino)pyrimidine: 4-Me-2-PP: 2-(4-methylpiperazino)pyrimidine. NOE: nuclear Overhauser effect; 8-OH-DPAT: 8-hydroxy-2-(di-*n*-propylamino)tetralin.

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Fig 1. Structures of prazosin (1) and doxazosin (4), and their derivatives and analogs (2-6).

of α_1 ligands under discussion may also be regarded as an analog of 2-(*N*-piperazino)pyrimidine (2-PP). Also, it is well documented that a number of typical 5-HT_{1A} ligands of the 1-arylpiperazine class show a significant or even high α_1 receptor affinity [5–10]. Therefore, in the present paper we discuss the fundamental structural requirements responsible for the receptor binding profile (α_1 , 5-HT_{1A} and 5-HT_{2A}) of simple, model analogs of 2-(4-methylpiperazino)pyrimidine (4-Me-2-PP) which contain the quinazoline, phthalazine or quinoline ring system instead of a 2-pyrimidinyl moiety (fig 2).

Chemistry

The structures of compounds 7-22 used in this work are given in figure 2. We have shown previously that the reaction of aryllithium and heteroaryllithium reagents with 2.4-dichloroquinazoline is regioselective, resulting in the predominant substitution of the chlorine at position 4 [11]. This reaction is illustrated in scheme 1 by the synthesis of known quinazoline derivatives 23 and 24, and a new compound 25. Treatment of 23–25 with *N*-methylpiperazine furnished the corresponding 2-(*N*-methylpiperazino)quinazolines 7–9.



Fig 2. Structures of compounds 7–22.

A phthalazine 13 was obtained by a similar nucleophilic displacement of a chlorine atom in 1-chlorophthalazine (scheme 2). Synthesis of quinazolines 10–12 [12], quinolines 14–18 [13] and fused quinolines 20 [14], 21 [15], and 22 [16] has been reported by us previously. A new quinoline derivative 19 was prepared in a similar fashion (scheme 3). Thus, condensation of 2,5-bis(trifluoromethyl)aniline with acetophenone was followed by lithium *N*-methylpiperazide-mediated cyclization of the resultant Schiff base 26 to give the desired compound 19. The ketimine 26 and similar ketimines derived from aniline and aryl methyl ketones are thermodynamic mixtures of a major *E* dia-



Scheme 1.

Scheme 2.





stereomer, as shown for **26** in scheme 3, and a minor Z diastereomer [14, 17]. Analytically pure (*E*)-**26** was obtained by chromatography, and the suggested stereochemistry was fully consistent with the results of the proton NOE experiment. As expected, irradiation of the methyl singlet at δ 2.24 resulted in a strong singlet at δ 7.05 for H6 of the aniline proton and a two-proton doublet at δ 7.97 for H2 and H6 of the phenyl group.

Pharmacology

All compounds 7–22 as well as 2-PP (27) and 4-Me-2-PP (28) were evaluated for their receptor binding profile (α_1 , 5-HT_{1A}, 5-HT_{2A}). The receptor affinities of the investigated compounds were determined in the competition experiments using the following radioligands and the rat brain membranes: [³H]prazosin (cortex). [³H]-8-OH-DPAT (hippocampus). and [³H]ketanserin (cortex) for α_1 , 5-HT_{1A} and 5-HT_{2A} receptors, respectively. The affinities of prazosin for α_1 ($K_1 =$ 0.23 ± 0.03 nM), 8-OH-DPAT for 5-HT_{1A} ($K_1 =$ 1.43 ± 0.21 nM), and ritanserin for 5-HT_{2A} ($K_1 =$ 1.14 ± 0.13 nM) receptors were also determined and they serve as a standard in the conducted biding studies. The results are shown in table I.

Results and discussion

All investigated compounds show diverse α_1 , 5-HT_{1A} and 5-HT_{2A} receptor affinities which are within a range of 10⁻⁹ to 10⁻⁵ M, 10⁻⁸ to 10⁻⁵ M and 10⁻⁷ to 10⁻⁵ M, respectively (table I). Derivative **7** exhibits a significantly higher affinity than its parent compounds 2-PP (**27**) and 4-Me-2-PP (**28**) for α_1 , 5-HT_{1A} and 5-HT_{2A} receptors. 4-Phenyl- and 4-(2-thienyl)-2-(4-methyl-

piperazino)quinazolines 7 and 8 show the same α_1 affinity within experimental error (see table I), whereas extension of the 4-substituent results in the completely inactive 4-(2-benzo[b]) derivative 9. Permutation of substituents between positions 2 and 4 (cf. 10 vs 7) significantly enhances the α_1 affinity of 10 in relation to 7. Furthermore, replacement of the *N*-methylpiperazine fragment in compound **10** with a flexible N,N-dimethylethylenediamine chain (12) does not affect the α_1 affinity. The phthalazine derivative 13 has a low, micromolar α_1 affinity (table I). Thus, a comparison of the α_1 -binding data for 7–10, 12 and 13 may suggest that the orientation of a local dipole moment of the unfused heteroaromatic systems plays some role in stabilization of the ligand- α_1 receptor complex.

In order to verify the above hypothesis we have analyzed the α_1 affinities of a series of 4-(4-methylpiperazino)quinolines 14–21. The results obtained are meaningful. Replacement of the N3 atom of the quinazoline system in 10 ($K_1 = 290$ nM) by the C-sp² aro-

Table I. α_1 , 5-HT_{1A} and 5-HT_{2A} receptor affinities of compounds 7–22, 27 and 28.

Compound	$K_{r} \pm SEM(nM)^{a}$			
	α,	5-HT ₁₄	5-HT ₂₄	
7	1070 ± 160	545 ± 22	1190 ± 11	
8	1340 ± 220	290 ± 13	468 ± 5	
9	>50 000	43 ± 4	256 ± 3	
10	290 ± 26	924 ± 45	612 ± 29	
11	ND	409 ± 6	476 ± 7	
12	335 ± 33	4870 ± 70	2480 ± 40	
13	9700 ± 900	1545 ± 80	4470 ± 185	
14	17 ± 2	3710 ± 130	276 ± 37	
15	292 ± 22	3800 ± 210	1425 ± 85	
16	1280 ± 100	6140 ± 280	$10\ 300\pm 600$	
17	161 ± 14	3380 ± 210	280 ± 9	
18	4.9 ± 1.4	3420 ± 90	211 ± 11	
19	264 ± 15	9500 ± 1000	222 ± 7	
(4R)-(+)-20	$30\ 000 \pm 3300$	$43\;400\pm2600$	$45\ 000 \pm 6000$	
(4 <i>S</i>)-(–)- 20	6400 ± 350	>50 000	$30\ 800 \pm 1700$	
21	2140 ± 430	3650 ± 280	8090 ± 300	
22	446 ± 32	ND	2370 ± 140	
27 b	5970 ± 620	1430-	29 500°	
2 8 b	6100 ± 1150	2180°	19 700°	

^aMean values from at least three independent experiments; **b27**: 2-PP, **28**: 4-Me-2-PP; ^cdata taken from reference [18]; ND: not determined. matic atom strongly increases the α_1 affinity of the resultant quinoline 14 ($K_1 = 17.5$ nM). Further modifications of 14 culminated in a 4-(4-methylpiperazino)-2-(2-thienyl)quinoline (18), which is the most active α_1 ligand ($K_1 = 4.9$ nM) of all compounds under investigation (table I). Moreover, derivative 18 is the most selective α_1 ligand as its 5-HT_{1A} and 5-HT_{2A} affinities are 700- and 43-fold lower, respectively. The determined α_1 affinity of 15–17 is at least tenfold lower than that of 14. On the other hand, the 2-pyridyl derivative 15 shows the same α_1 affinity as quinazoline 10; the affinity of the 3-pyridyl isomer 16 is lower, whereas the K_1 values of the 4-pyridyl derivative 17 increase slightly in relation to 15 and 10 (table I). Again, it can be suggested that the orientation of a local dipole moment of the unfused heteroaromatic systems controls the α_1 affinity of the investigated compounds. A CF_3 group in position 7 of the quinoline skeleton significantly decreases the α_1 affinity of 19 as compared to the parent compound 14. Annelation of the quinoline skeleton with bicyclo[2.2.1]heptane yields enantiomers of 20, which bind to α_1 receptors in the micromolar range (10-6 to 10-5 M)only. Weak enantioselectivity, however, is observed, as the (4S)-(-)-20 enantiomer shows an α_1 affinity at least fourfold higher than its (4R)-(+)-20 counterpart.

Our earlier conformational analysis of unfused heteroaromatic ring systems clearly indicated that 4-(2-thienyl)- and 4-(2-furyl)pyrimidine exist predominantly in the opposite coplanar conformations, s-cis and s-trans, respectively [18-23]. In this work, we carried out a conformational analysis of several representatives of 7–21, using the recommended and previously applied semi-empirical PM3 method [22, 24, 25]. The results of the PM3 calculations for 2-phenyl-, 2-(2-pyridyl)-, 2-(3-pyridyl)-, and 2-(2-thienyl)-4-(4methylpiperazino)quinolines 14-16 and 18, respectively, are shown in figure 3. It was found that the PM3 method prefers coplanar conformations of the heteroaromatic fragments of 14, 16 and 18. The calculated differences between *s*-*cis* and *s*-*trans* conformers (τ_1 = 0° and 180° , respectively) for **16** and **18** are small and do not exceed 0.14 kcal/mol. Furthermore, the conformers s-cis and s-trans of 14 are equipopulated as their heats of formation are the same. As expected, two low-energy conformations of a 2-(4-pyridyl)quinoline 17 are only slightly deviated from coplanarity (data not shown). In marked contrast, 2-(2-pyridyl) derivative 15 exists predominantly in s-trans conformations, and the rotation barrier ($\Delta E = 1.88$ kcal/mol) is significantly higher than that calculated for other quinoline derivatives ($\Delta E = 0.63$, 0.92 and 0.87 kcal/mol for 14, 16 and 18, respectively). The *s*-*cis* conformations of 15 ($-30^{\circ} < \tau_1 < 30^{\circ}$) are apparently destabilized by an unfavorable orientation of dipole moment of the quinoline and pyridine subunits. On the other hand, our earlier studies have shown that the attractive $S \cdots N$ interactions additionally stabilize the *s*-*cis* conformation of the 4-(2-thienyl)pyrimidine fragment [19]. It appears that the same effect is responsible for stabilization of the calculated *s*-*cis* conformation of the unfused 2-(2-thienyl)quinoline ring system in **18**.

In order to verify our hypothesis that the orientation of the unfused heteroaromatic fragment controls the α_1 -receptor affinity of 14–18, we analyzed two additional derivatives, 21 and 22. These compounds contain a 2-(3-thienyl)quinoline fragment in the fixed s-trans conformation due to a rigid ethylene bridge in their structure, and they differ in the conformational freedom of the amino fragment. The PM3-optimized geometry of **21** is shown in figure 4. The α_1 affinity data (table I) clearly indicate that the s-trans conformation of 21 is unfavorable for interaction with the receptor. Furthermore, the flexible N.N-dimethylethylenediamine chain permits a slightly different orientation of 22 at the receptor, though its observed α_1 affinity $(K_1 = 446 \text{ nM})$ is considerably lower than that of 14 and 18, and is not more than two to three times different from the affinities of **15–17**.

The 5-HT_{1A} affinity of derivatives 7 and 8 (table I) is of the same order as that reported by Glennon et al for 2-(*N*-piperazino)naphthalene ($K_1 = 265$ nM) and 2-(*N*-piperazino)quinoline ($K_1 = 230$ nM) [26]. Surprisingly, derivative 9 is the most active 5-HT_{1A} ligand $(K_1 = 43 \text{ nM})$ of all investigated compounds. A comparison of the reported 5-HT_{1A} affinity of 1-(N-piperazino)naphthalene ($K_1 = 5$ nM) [26] with that of its phthalazine analog 13 ($K_1 = 9700$ nM) indicates that the presence of two adjacent nitrogen atoms in the ring structure is a highly undesirable feature. Derivatives 14–21 show a low or very low 5-HT_{1A} affinity (table I). These findings also agree with the results reported by Glennon, who suggested a region of limited bulk tolerance at the 5-HT_{1A} receptor [27, 28]. Indeed, relatively small R1-substituents of the investigated quinolines may reach that region at 5-HT_{1A} receptors, but these receptors do not tolerate large substituents.

The investigated compounds show a moderate (211 $\leq K_1 \leq 280$ nM for **9**, **14** and **17–19**). low (468 $\leq K_1 \leq$ 1425 nM for **7**, **8**, **10**, **11** and **15**) or even very low ($K_1 > 2000$ nM for **12**, **13**, **16** and **20–22**) affinity for 5-HT_{2A} receptors (table I). In previous studies we have proposed a pharmacophore which is responsible for the formation of a complex between 4,6-di(heteroaryl)-2-(4-methylpiperazino)pyrimidines and 5-HT_{2A} receptors [18, 22]. We have also defined three crucial distances, d_1 , d_2 and d_3 (for their definition see table II), and their optimal ranges necessary for high affinity ($K_1 = 10^{-9}$ to 10^{-8} M) of this class of 5-HT_{2A} ligands (table II). The d_1 , d_2 and d_3 parameters of the analyzed *N*-methylpiperazines **7–11** and **13–21** are within typical ranges, except for the $d_2 = 9.53$ Å value for derivative



Fig 3. Conformation energy profiles upon rotation (τ_1) of the inter-ring C2-C1'(2') bond calculated by the PM3 method.

9, which reaches the critical, upper limit of this distance (fig 5, table II) [22]. Compound **13** has a very low 5-HT_{2A} affinity, since its structure meets only in part the pharmacophore requirements (table II).

Our previous studies have clearly indicated that the most active 5-HT_{2A} ligands, such as 4.6-di(2-thienyl)-2-(4-methylpiperazino)pyrimidine and its 4,6-di(hetero-aryl) analogs, adopt favorable coplanar conformations of both the piperazinopyrimidine subunit and the unfused, tricyclic heteroaromatic system [18, 22]. We have also demonstrated that distortion of the coplanar heteroaromatic ring system significantly decreases the observed 5-HT_{2A} affinity [22]. The same effect is observed for derivatives **7–9**. Their unfused, heteroaromatic ring system exists predominantly in the twisted conformations shown in figure 6. Further-



Fig 4. The lowest-energy conformer of 21 calculated by the PM3 method.

more, an excellent qualitative relationship between the population of the twisted conformations and the 5-HT_{2A} affinity of **7–9** was observed: the lower the $\Delta E_{180^{\circ}-60^{\circ}}$, the higher the 5-HT_{2A} affinity (fig 6, table I).

The PM3 calculations show that arylpiperazine fragments of 7–11 and 13–21 adopt different conformations. While coplanar conformations are favored

Table II. The distances d_1 , d_2 and d_3 defining the 5-HT_{2A} receptor pharmacophore of the investigated compound.

Compound	$d_{i}(\mathring{A})^{a}$	<i>d</i> ₂ (Å) ^b	$d_{3}(\mathring{A})^{c}$
Optimal values ^d	5.2-8.4	5.7-8.5	4.6–7.3
7	7.83	8.47	5.03
8	7.83	8.37	4.91
9	7.83	9.53	5.87
11 ^e	5.98-6.61	8.00-8.66	6.29
13 ^e	5.98-6.48	-	_
14 ^e	5.97-6.58	8.21-8.94	6.44
15°	5.97-6.58	8.19-8.93	6.42
18 ^e	5.84-6.62	8.07-8.91	6.29
21°	5.95-6.58	8.11-8.78	6.25

^aDistance between N4-piperazine atom and a center of the fused benzene ring; ^bdistance between N4-piperazine atom and a center of the aromatic R or R¹ substituent; ^cdistance between centers of the fused benzene ring and the aromatic R or R¹ substituent; ^doptimal values of d_1 , d_2 and d_3 parameters taken for comparison from references [18] and [22]; ^eranges of d_1 and d_2 parameters measured for the opposite conformations of the *N*-methylpiperazine fragment as shown in figure 5.



Fig 5. Distances defining the 5-HT₂₄ receptor pharmacophore for 7 and 8 in the coplanar conformation of the arylpiperazine ring system, and for 11 and 21 (where a and b refer to either of the two energy minima of the arylpiperazine fragment in twisted conformations).

for 7–9, twisted or even orthogonal conformations are definitely more populated in the case of 10, 11 and 13–21 (fig 7). It should be stressed that our results for the conformational analysis of 7–9 are fully consistent with those reported by others for 2-(4-methylpiperazino)pyrimidine [18, 22, 25]. It may therefore be anticipated that the conformation of the arylpiperazine fragment is not critical for the ability to form a complex, as some derivatives with different conformations of this fragment have similar 5-HT_{2A} affinities (eg, 8 and 11). On the other hand, all analyzed 4-(4-methylpiperazino)quinolines 14–19 have basically the same conformation energy profiles as that shown for 14 in figure 7b, whereas their 5-HT_{2A} affinities are within a wide K_1 range for different R¹ substituents (table I).

It can be concluded that orientation of the local dipole moment and planarity of the unfused heteroaromatic ring system are the most important structural features of the investigated class of ligands, which are responsible for the observed 5-HT_{2A} affinity changes. Furthermore, it should be stressed again that the conclusions derived from the 5-HT_{2A} affinity studies are fully consistent with our previous findings and serve as an additional verification of our topographic model of 5-HT_{2A} sites [18, 22].

Experimental protocols

Chemistry

2,4-Dichloroquinoline [29] and 1-chlorophthalazine [30] were prepared as described Melting points (mp. Pyrex capillary) are not corrected. ¹H-NMR spectra were obtained on a Varian-400 (400 MHz) instrument at 23 °C with tetramethylsilane as an internal standard. Crude reaction mixtures were analyzed, and mass spectra of pure components were obtained on a Hewlett-Packard GC-MS instrument equipped with an on-column injector, a poly(dimethylsiloxane)-coated capillary, and a mass selective detector operating at 70 eV. Hydrobromide salts were obtained by using a general procedure [13] and the salts were crystallized from 95% ethanol. Elemental analyses indicated by the symbols of the elements were within $\pm 0.3\%$ of the theoretical values.

4-(2-Benzo[b]thienyl)-2-chloroquinazoline 25

This compound was obtained in the reaction of 2,4-dichloroquinazoline with 2-benzo[*b*]thienyllithium [31] by using a general procedure reported previously for the preparation of **23** and **24** [11]. After crystallization from dichloromethane/hexane



Fig 6. Conformation energy profiles upon rotation (τ_1) of the inter-ring C2-C1'(2') bond for 7 (a), 8 (b) and 9 (c) calculated by the PM3 method.

(1.9) the yield was 56%, mp 149–150 °C ¹H-NMR (CDCl₃) δ ⁷.46 (t, J = 8 Hz, 1H); 7.49 (t, J = 8 Hz, 1H); 7.74 (t, J = 8 Hz, 1H); 7.96 (m, 3H); 8.06 (d, J = 8 Hz, 1H); 8.15 (s, 1H); 8.61 (d, J = 8 Hz, 1H) MS (m/z); 296 (100, M⁺), 298 (35, M⁺). Anal C₁₆H₉ClN₂S (C, H, N).

A general method for the preparation of 7-9 and 13

A solution of 23–25 (1 mmol) or 1-chlorophthalazine (164 mg, 1 mmol) in *N*-methylpiperazine (2 mL) was heated under reflux for 2 h. After cooling the mixture was treated with water (3 mL) and extracted with ether (3 \times 25 mL). The extract was dried with Na₂SO₄, concentrated on a rotary evaporator, and the residue was subjected to chromatography on silica gel with hexane/triethylamine/ethanol (7.2:1) as an eluent

2-(4-Methylpiperazino)-4-phenylquinazoline 7. After crystallization from ethanol/hexane (1:9) this compound was obtained in a 90% yield; mp 98–100 °C, reported mp 97–98 °C [32].

2-(4-Methylpiperazuro)-4-(2-thienyl)quinazoline 8 This compound was obtained in an 85% yield; an oil ¹H-NMR (CDCl₄) δ : 2.37 (s, 3H); 2.54 (t, J = 5 Hz, 4H); 4.03 (t, J = 5 Hz, 4H); 7.23 (m, 2H); 7.58 (d, J = 5 Hz, 1H); 7.65 (m, 2H); 7.79 (d, J =4 Hz, 1H); 8.23 (d, J = 8 Hz, 1H). MS (*m*/₇): 240 (100), 310 (20, M⁺). 8-2HBr; mp >310 °C. Anal C₇H₁₈N₄S-2HBr (C, H, N).

4-(2-Benzo[b]thienyl)-2-(4-methylpiperazino)quinazoline 9. This compound was obtained in a 58% yield; an oil 9-2HBr·H₂O. mp >310 °C. ¹H-NMR (DMSO- $d_{\rm p}$) & 2.87 (s, 3H); 3.17 (m, 2H); 3.44 (m, 2H), 3.60 (m, 2H); 4.92 (m, 2H); 5.60 (br. exchangeable with D₂O); 7.51 (m, 3H); 7.71 (d, *J* = 8 Hz, 1H); 7.88 (d, *J* = 8 Hz, 1H); 8.09 (m, 2H). 8.50 (s, 1H); 8.54 (d, *J* = 8 Hz, 1H). Anal C₂₁H₂₀N₄S·2HBr·H₂O (C, H, N).

1-(4-Methylpiperazino)phthalazine **13**. This compound was obtained in a 73% yield; an oil. ¹H-NMR (CDCl₃) : 2.42 (s, 3H); 2.72 (t, J = 5 Hz, 4H); 3.60 (t, J = 5 Hz, 4H); 7.82 (m, 2H); 7.88 (m, 1H); 8.05 (m, 1H), 9.17 (d, J = 1 Hz, 1H). MS (*m/z*): 158 (100), 228 (3, M⁺). **13-**2HBr: mp 273–275 °C Anal C₁₃H₁₆N₄•2HBr (C, H, N).

(*E*)-*N*-(*1*-*Phenylethylidene*)-2,5-*bis*(*trifluoromethyl*)*aniline* **26** Condensation of 2.5-bis(trifluoromethyl)aniline with acetophenone was conducted by using a general procedure [13, 14]. Compound **26** was obtained in an 85% yield after distillation (128–130 °C/3 6 mmHg) on a Kugelrohr. ¹H-NMR (CDCl₄) : 2.24 (s, 3H): 7.05 (s, 1H). 7.42 (d, *J* = 8 Hz, 1H): 7.48 (m, 3H). 7.80 (d, *J* = 8 Hz, 1H): 7.97 (d, *J* = 8 Hz, 2H). MS (*m/z*): 316 (100), 331 (30, M⁺). Anal C₁₆H₁₁F₆N (C, H, N).

4-(4-Methylpiperazino)-2-phenyl-7-(trifluoromethyl)quanoline **19** Heterocyclization of ketimine **26** with hithium *N*-methylpiperazide was conducted by using a general procedure [13, 14]. Chromatography on silica gel with hexane/triethylamine/ethanol (7:2:1) as an eluent was followed by crystallization of **19** from hexane. Yield 67%: mp 141–142 °C. ¹H-NMR (CDCl₃) δ . 2 47 (s, 3H); 2 78 (m, 4H), 3.36 (m, 4H); 7 39 (s, 1H); 7 52 (m, 3H), 7 63 (d, J = 8 Hz, 1H); 8.11 (m, 3H), 8.44 (s, 1H) MS (*m*/z): 70 (100), 371 (40, M⁺) Anal C₂₁H₂₀F₃N₃ (C, H, N) **19-2HBr**-1.5H₂O; mp 268–270 °C. Anal C₂₁H₂₀F₃N₃•2HBr-1.5H₂O (C, H, N).



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Fig 7. Conformation energy profiles upon rotation ($\tau =$ lone pair-N1-C1' -N(or C)2') of the *N*-methylpiperazine fragment versus heteroaromatic moiety for **10** (a) and **14** (b).

Pharmacology

Radioligand binding experiments were conducted for 5-HT_{1A} receptors in the hippocampus of the rat brain, and in the cortex for 5-HT_{2A} receptors, according to the published procedure [33] [3H]-8-OH-DPAT (190 Ci/mmol, Amersham) and [3H]-ketanserin (60 Ci/mmol, NEN Chemicals) were used for labeling 5-HT_{1A} and 5-HT_{2A} receptors, respectively. The K_i values were determined on the basis of at least three competition binding experiments in which 10–14 drug concentrations (10⁻¹⁰ to 10⁻³ M), run in triplicate, were used.

α_i -Receptor binding experiments

[3H]Prazosin (26 Ci/mmol, NEN Chemicals) was used for labelling α_1 receptors. The membrane preparation and assay procedure were carried out according to the published procedures [34, 35] with slight modifications. The cortex tissue of the rat brain was homogenized in 20 vol (w/v) of ice-cold Tris-HCl buffer (50 mM, pH = 7.4) with an Ultra Turrax homogenızer The homogenate was centrifuged at $25\ 000 \times g$ for $10\ mmmode{min}$, and the resulting pellet was suspended in the same volume of Tris-HCl buffer, and was recentrifuged. The final pellet was resuspended in 170 vol (w/v) of Tris-HCl buffer (50 nM, pH =7.4). [3H]Prazosin in a volume of 100 µl was added to aliquots (1.7 mL) of the membrane suspension, and the samples were incubated at 25 °C for 30 min. The total incubation volume of 2 mL was filtered through Whatman GF/B glass filters, and was then washed with a cold buffer $(3 \times 5 \text{ mL})$ using a Brandel cell harvester. Non-specific binding of [³H]prazosin was obtained in the presence of phentolamine (200 μ L, final concentration 10-6 M). The final [3H]prazosin concentration was 3×10^{-10} M and the concentration of the analyzed compounds ranged from 10^{-10} to 10^{-3} M. K₁ values were determined from at least three independent experiments, run in triplicate.

Molecular modeling

All the molecular modeling experiments were conducted using a Sybyl 6.03 package (Tripos Associates, Inc), installed on an ESV 10/33 workstation. PM3 calculations were conducted using a Mopac 5.0 (QCPE) program implanted into the Sybyl 6.03. Full geometry optimization and gradient norm <0.1 kcal/mol/Å were setup during calculations of low-energy conformations. To investigate the rotational energy barriers, 12 conformations were generated by a step-wise rotation of 30° around the inter-ring bonds. Next, each of these conformations was optimized using a PM3 method over all internal coordinates except for those that define the relative orientation of the respective substituent (gradient norm <0.1 kcal/mol/Å).

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