

Note

Syntheses of daidzein-7-yl β -D-glucopyranosiduronic acid and daidzein-4',7-yl di- β -D-glucopyranosiduronic acid

Paul W. Needs,* Gary Williamson

Institute of Food Research, Norwich Research Park, Colney, Norwich NR4 7UA, UK

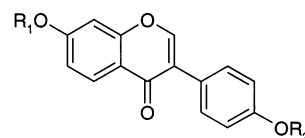
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Abstract

Syntheses of the title compounds — commonly known as ‘daidzein 7-glucuronide’ and ‘daidzein 4',7-diglucuronide’ — are described. Selective 7-deacetylation of 4',7-di-*O*-acetyldaidzein is employed. © 2001 Elsevier Science Ltd. All rights reserved.

Keywords: Daidzein; Glucuronide; Daidzein 7-glucuronide; Daidzein 4',7-diglucuronide; Synthesis

Several classes of plant flavonoids form part of the human diet. They are of great current interest, as many appear to be protective against coronary heart disease and/or a variety of carcinomas.^{1,2} One type, the isoflavones, are additionally suspected of estrogenic activity.³ Daidzein **1**, occurs as its 7- β -glucopyranoside, daidzin **2**, in soya-based foods. After absorption from the intestinal tract, together with deglycosylation,⁴ the resultant daidzein is thought, in common with other absorbed non-nutrients, to be glucuronidated prior to circulation and excretion.⁵ We thus required a synthetic route to the title compounds daidzein-7-yl β -D-glucopyranosiduronic acid **3** and daidzein-4',7-yl di- β -D-glucopyranosiduronic acid **4**, in order to study their possible biological effects.



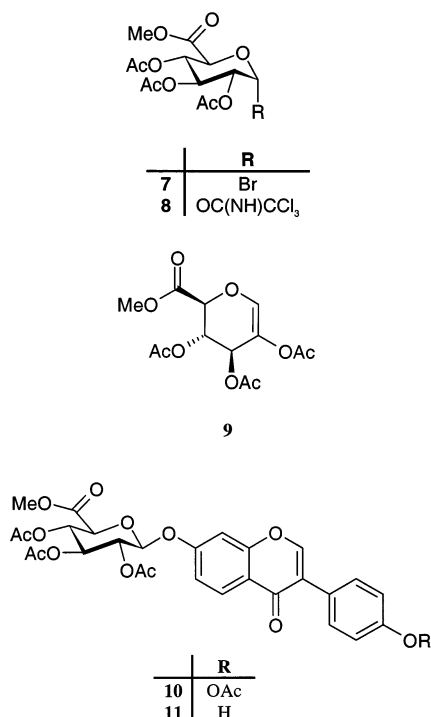
	R ₁	R ₂
1	H	H
2	β -D-Glcp	H
3	β -D-GlcpA	H
4	β -D-GlcpA	β -D-GlcpA
5	Ac	Ac
6	H	Ac

Glycosylation of daidzein with protected, simple aldopyranosyl bromides to give 7-glycosides directly has been reported, albeit in low yield.⁶ These reactions were performed in aqueous acetone with hydroxide catalysis. More recent work suggests that C-ring cleavage is, in fact, the predominant reaction under these conditions.⁷ We thus attempted glucuronidation of daidzein with **7** under non-aqueous conditions, which had successfully provided some other flavonoid derivatives.⁸ Thus **1** was treated with 1.1 equiv of **7** and silver carbonate in pyridine, or with 3 equiv of **7** and silver carbonate in pyridine or DMF.

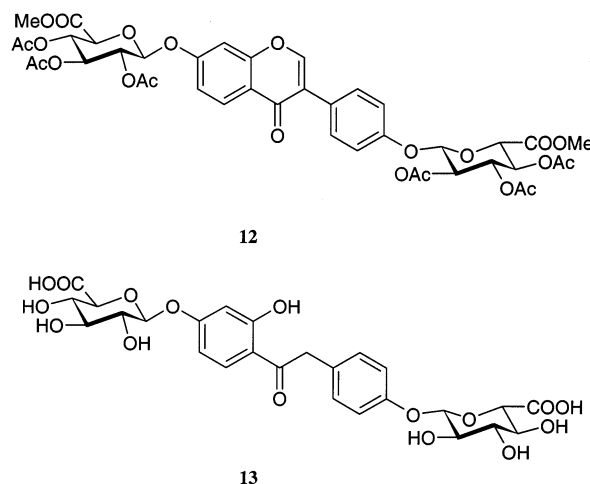
* Corresponding author. Tel.: +44-1603-255000; fax: +44-1603-507723.

These reactions were unsuccessful; they gave mainly the glycal **9**, recovered **7**, and daidzein. Only minor amounts of glucuronidated products were obtained. In no case was the expected **11** detected. NMR showed that all products were 4'-O-acetylated; some **10** was present, together with variable amounts of some other, unassigned products, apparently bearing 7-substituents other than the expected derivatised glucuronic acid group. A two-phase procedure,⁷ which has been reported to give good yields of neutral daizein-7-yl glycosides also failed to give **11**. These results were not entirely unexpected; **9** is often a major product of reactions of **7** with 'complex polyphenolics', and yields of conjugates are often low.^{9,10}

Compound **3** was synthesised as follows. Daidzein **1** was diacetylated under acidic conditions¹¹ to give **5**. Regioselective deacetylation of **5** with imidazole in aqueous THF at pH 7 gave the 4'-O-acetyl derivative **6**. Treatment of **6** in dichloromethane with methyl 2,3,4-tri-O-acetyl- α -D-glucopyranosyluronate bromide (**7**), silver carbonate and collidine gave the corresponding 7-glucuronic acid derivative **10** in 30% yield, together with the glycal **9**. Compound **10** was converted, with lithium hydroxide in aqueous methanol at 0 °C, to give **3** in 67% yield.



Compound **12**, the precursor of **4**, was obtained by treatment of daidzein with boron trifluoride and 2 equiv of the trichloroacetimidate **8**, but in only 14% yield. Traces of the 7-substituted product were also detected. Hydrolysis of **12** with LiOH, as above, gave not only **4**, but also the ring-opened product **13**. The decreased susceptibility of **10** to ring opening is presumably a consequence of its conversion, after cleavage of the 4'-O-acetyl group, to a phenolic anion which is resistant to nucleophilic attack. However, treatment of **12** with sodium carbonate in aqueous methanol gave **4**, with only traces of **13**, as shown by high-performance liquid chromatography–electrospray mass spectroscopy (HPLC–ESMS). Preparative HPLC gave pure **4**. No attempt has been made to optimise the synthesis of **12**, as the yield of **4** was sufficient for use as an HPLC standard and in biological assays.



1. Experimental

General methods.—Solvents were dried over freshly activated 3 Å molecular sieves. Evaporations were performed in vacuo at 40 °C. Solids were dried overnight in vacuo over P₂O₅ before use. TLC was performed on Macherey–Nagel Silica Gel 60/UV254 plates using 2 or 9% MeOH in CH₂Cl₂ as eluant, and UV light, or 50% sulphuric acid and charring, for visualisation. Medium pressure column chromatography (MPLC) was performed on a 2.5 × 60 cm column of Merck 109385 grade silica (40–63 μm mesh), eluted

with 1% MeOH–CH₂Cl₂. Analytical HPLC, and HPLC–ESMS, used a 5 μ m Prodigy ODS3 column (250 \times 4.4 mm) eluted at 1 mL/min. Eluant A 80% for 5 min (isocratic); to 10% A at 35 min (gradient). Eluate was monitored by UV detection at 205 and 280 nm. Preparative HPLC used a 5 μ m Prodigy ODS3 column (Phenomenex Inc., 250 \times 21.2 mm + 60 \times 21.2 mm guard) eluted at 5 mL/min. Eluant A, 0.1% CF₃COOH (TFA); eluant B, CH₃CN. 80% A for 15 min (isocratic); then to 10% A at 75 min (gradient). NMR spectra were run on a JEOL GX400 spectrometer. Sample temperature was 27 °C and chemical shifts were using the residual solvent's absorption for calibration. Atmospheric pressure chemical-ionisation mass spectroscopy (APCIMS), ESMS, and HPLC–ESMS analyses were performed on a Micromass Quattro II mass spectrometer (Manchester, UK). High resolution fast atom bombardment (FABMS) spectra were obtained on a Kratos MS890 mass spectrometer using glycerol or thioglycerol as a matrix.¹³ Elemental analyses were performed on a Carlo Erba 1108 CHN analyser by Satco Ltd. (Hatfield, UK). Melting points were determined on a Reichert Thermopan Koffler hot stage microscope, and are uncorrected.

4'-O-Acetyldaidzein (6).—4',7-Di-O-acetyldaidzein **5** (6.0 g, 9.0 mmol) (prepared by treatment of daidzein with concd H₂SO₄ and Ac₂O¹¹ in 87% yield) was dissolved, with gentle warming, in THF (750 mL). After cooling to 25–30 °C, a solution (750 mL) of imidazole in water (1 M, adjusted to pH 7 with concd HCl) was added, and the mixture stirred for 15 min at 25–30 °C, after which time an initial slight precipitate had redissolved. The mixture was adjusted to pH 4.5 with gl AcOH, water (1 L) was added, and the mixture extracted with CH₂Cl₂ (3 \times 400 mL). The combined organic extracts were washed with water (1 \times 500 mL), satd Na₂CO₃ (1 \times 500 mL) and water (1 \times 500 mL), dried (MgSO₄), and evaporated to dryness. The resultant solid was recrystallised from EtOH (500 mL). Evaporation of the filtrate, and recrystallisation of the solid from MeOH, yielded a further crop of product. The combined yield of white flakes of **6** was 5.28 g (94%), mp 225.5–226.5 °C. ¹H

NMR (CD₃)₂SO: 8.40 (s, 1 H, H-2), 7.98 (d, 1 H, *J*_{5,6} 8.8 Hz, H-5), 7.60 (dd, 2 H, AA'BB' system, *J*_{2',3'} 8.8 Hz, H-2', H-6'), 7.18 (AA'BB' system, 2 H, H-3', H-5'), 6.94 (dd, 1 H, *J*_{6,8} 2.0 Hz, H-6), 6.88 (d, 1 H, H-8), 2.28 (s, 3 H, 4'-COCH₃). APCIMS: *m/z* 295 [M – H][–]. Anal. Calcd for C₁₇H₁₂O₅: C, 68.92; H, 4.08. Found C, 68.42; H, 4.12.

Methyl (4' - O - acetyldaidzein - 7 - yl β - D - 2'',3'',4'' - tri - O - acetylglucopyranosid)uronate (10).—A suspension of **6** (3 g, 10.1 mmol), **7** (4.43 g, 1.1 equiv), Ag₂CO₃ (3.07 g, 1.1 equiv) and ground 3 Å molecular sieves (2 g) in CH₂Cl₂ (120 mL) was stirred under Ar at rt. Collidine (1.47 mL, 1.1 equiv) was added dropwise over 10 min. The mixture was stirred in the dark for 7 days, and filtered through celite. The latter was washed with 10% MeOH–CH₂Cl₂ (50 mL). The filtrates were combined and washed successively with 10% aq AcOH (2 \times 50 mL), water (1 \times 100 mL), 0.1 M Na₂S₂O₃ (1 \times 100 mL), water (1 \times 100 mL), satd NaHCO₃ (1 \times 100 mL), and water (1 \times 100 mL), and dried (MgSO₄). Evaporation gave 4.89 g of a solid, which was dissolved in 50 mL 1% MeOH–CH₂Cl₂ and cooled to –20 °C overnight. Precipitated **6** (0.27 g) was filtered off. Polar material, and further residual **6** were removed by step elution through a short silica column with 1, 2 and 10% MeOH–CH₂Cl₂, and pooling of appropriate fractions. This gave a crude product (3.7 g) virtually free of **6**. This was used to prepare **3** without further purification.

A sample of the crude product (0.5 g) was purified by MPLC to give 254 mg (30%) of **10**. A portion was recrystallised from MeOH to give white crystals mp 186–188 °C (after drying in vacuo at 70 °C). ¹H NMR (CDCl₃): 8.25 (d, 1 H, *J*_{5,6} 8.4 Hz, H-5), 7.97 (s, 1 H, H-2), 7.58 (AA'BB' system, 2 H, *J*_{2',3'} 8.4 Hz, H-2', H-6'), 7.16 (AA'BB' system, 2 H, H-3', H-5'), 7.08 (d, 1 H, *J*_{8,6} 2.4 Hz, H-8), 7.05 (dd, 1 H, H-6), 5.27–5.44 (m, 4 H, H-1'', H-2'', H-3'', H-4''), 4.29 (d, 1 H, *J*_{5'',4''} 9.2 Hz, H-5''), 3.73 (s, 3 H, COOCH₃), 2.32 (s, 3 H, 4'-OAc), 2.08, 2.07, 2.06 (3 s, 3 \times 3 H, 3 \times COCH₃). APCIMS: *m/z* 613 [M + H]⁺. Anal. Calcd for C₃₀H₂₈O₁₄: C, 58.83; H, 4.61. Found C, 58.66; H, 4.50.

Daidzein-7-yl β -D-glucopyranosiduronic acid (3).—Crude **10** (0.5 g, containing 254 mg,

0.41 mmol of pure **10**) was sonicated into suspension in MeOH (30 mL), and cooled to 1 °C. Aq LiOH (30 mL, 0.5 M), precooled to 1 °C was added, and the mixture stirred at 1 °C for 16 h. Dowex 50W resin (H⁺ form, 20 mL) was added; after stirring for a further 10 min, the solution was eluted through a 5 × 2 cm column of Dowex, and the latter was washed with MeOH. The combined eluate was evaporated to dryness, and the residue dissolved in MeOH (4 mL). The material (4 × 1 mL) was purified by preparative HPLC (see above), and purity was checked by analytical HPLC. Yield 120 mg, 67%. ¹H NMR (CD₃OD): δ 8.19 (s, 1 H, H-2), 8.15 (d, 1 H, *J*_{5,6} 9.0 Hz, H-5), 7.38 (AA'BB' system, 2 H, *J*_{2',3'} 8.4 Hz, H-2', H-6'), 7.25 (d, 1 H, *J*_{8,6} 2.4 Hz, H-8), 7.22 (dd, 1 H, H-6), 6.84 (AA'BB' system, 2 H, H-3', H-5'), 5.17 (d, 1 H, *J*_{1,2} 7.2 Hz, H-1'), 4.01 (d, 1 H, *J*_{5,4} 10.0 Hz, H-5''), 3.4–3.7 (m, 3 H, H-2'', H-3'', H-4''). ESMS: *m/z* 429 [M – H][–]. FABMS (glycerol matrix): *m/z* [M – H][–]. Anal. Calcd 429.0822; Found 429.0899.

Daidzein-4',7-yl di-[methyl(2'',3'',4''-tri-O-acetyl-β-D-glucopyranosiduronic acid)] (12).—The trichloroimidate **8** (0.24 g, 0.50 mmol) and daidzein **1** (62 mg, 0.24 mmol) were suspended with stirring in CH₂Cl₂ (5 mL) at –15 °C under Ar. BF₃·Et₂O (8 μL) was added in one portion. The mixture was allowed to warm to rt over 1 h, after which a further portion of BF₃·Et₂O (8 μL) was added; stirring was continued for 60 h. EtOAc (100 mL) was added, and the mixture was washed with satd NaHCO₃ (2 × 50 mL), and water (1 × 50 mL), and dried (MgSO₄). The solution was evaporated to dryness, the resulting solid was triturated with CH₂Cl₂, and the insoluble material filtered off to yield recovered daidzein (40 mg, 65%). The solution was evaporated, and MPLC of the crude product, using 1% MeOH–CH₂Cl₂ as eluant, gave **12**, (29 mg, 14%) as a colourless glass. ¹H NMR (CDCl₃): 8.24 (d, 1 H, *J*_{5,6} 8.8 Hz, H-5), 7.94 (s, 1 H, H-2), 7.49 (AA'BB' system, 2 H, *J*_{2',3'} 8.6 Hz, H-2', H-6'), 7.07 (AA'BB' system, 2 H, H-3', H-5'), 7.07 (d, 1 H, *J*_{8,6} 2.4 Hz, H-8), 7.05 (dd, 1 H, H-6), 5.27–5.44 (m, 7 H, 1 × H-1'' (7-substituent)), 2 × H-2'', 2 × H-3'', 2 × H-4''), 5.17 (d, 1 H, *J*_{1,2} 6.8 Hz, H-1'' (4'-substituent)),

4.27 (d, 1 H, *J*_{5'',4''} 9.2 Hz, H-5'' (7-substituent)), 4.19 (d, 1 H, *J*_{5'',4''} 9.6 Hz, H-5'' (4'-substituent)), 3.74 and 3.73 (2 × s, 2 × 3 H, 2 × COOCH₃), 2.10, 2.08, 2.07, 2 × 2.06, 2.05 (6 s, 6 × 3 H, 6 × COCH₃). APIMS: *m/z* 887 [M + H]⁺.

Daidzein-4',7-yl di-β-D-glucopyranosiduronic acid (4).—Complex **12** (2.34 mg, 2.64 μmol) was sonicated into suspension in MeOH (300 μL) and stirred, and water (100 μL) and aq Na₂CO₃ (30 μL, 0.5 M, 15 μmol) were added at 20 °C. After 90 min, further MeOH (300 μL) and water (100 μL) were added. After 150 min, all solids had dissolved. The solution was desalted (Dowex 50W, H⁺) and evaporated. Preparative HPLC gave **4**; purity was checked by analytical HPLC. ¹H NMR ((CD₃)₂SO): δ 8.42 (s, 1 H, H-2), 8.03 (d, 1 H, *J*_{5,6} 8.9 Hz, H-5), 7.50 (AA'BB' system, 2 H, *J*_{2',3'} 8.4 Hz, H-2', H-6'), 7.23 (d, 1 H, *J*_{8,6} 2.4 Hz, H-8), 7.13 (dd, 1 H, H-6), 7.06 (AA'BB' system, 2 H, H-3', H-5'), 5.10 (d, 1 H, *J*_{1,2} 7.2 Hz, H-1'), 4.92 (d, 1 H, *J*_{1,2} 7.2 Hz, H-1''), 4.01 (m, 2 H, H-5'' and H-5'''), 3.3–3.7 (m, 6 H, H-2'', H-2''', H-3'', H-3''', H-4'' and H-4'''). ESMS: positive ion mode *m/z* 607 [M + H]⁺; negative ion mode *m/z* 605 [M – H][–], 719 [M + TFA – H][–]. FABMS (thioglycerol matrix): *m/z* [M – H][–]. Anal. Calcd 605.1143; Found 605.1105.

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