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PET chemosensors





ICT chemosensor



Highlights

1,8-Naphthalimides containing aza-15-crown-5 group at C-4 and N-imide are prepared.

The sensors have been shown to bind Ca^{2+} ions in MeCN with different affinities.

Ion induced optical changes are strongly dependent on the position of receptor unit.

Results have been rationalised in terms of PET and ICT mechanisms.

Comparative analysis of the PET and ICT sensor properties of 1,8-naphthalimides containing aza-15-crown-5 ether moiety

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Abstract

Novel 4-amino- and 4-(acetyl)amino-*N*-aryl-1,8-naphthalimides containing aza-15crown-5 ether receptor unit in the *N*-aryl fragment and at C-4 of the naphthalimide residue were designed and prepared. Significant internal charge transfer from electron donating amino or amido group at C-4 of the naphthalene ring to the acceptor carboxyimide moiety as well as photoinduced electron transfer between *N*-aryl receptor and the naphthalimide fragment was revealed by the UV/Vis absorption spectroscopy and considerable fluorescence quenching. The addition of calcium perchlorate to an acetonitrile solution of naphthalimides with the receptor at imide nitrogen hindered the photoinduced electron transfer process and accordingly restored the quenched fluorescence of the free ligands. In the case of the compound in which the aza-15crown-5 receptor is located at C-4, the coordination with Ca²⁺ reduced the internal charge transfer interaction in the chromophore and caused a significant blue-shift of the absorption and emission peak. The observed spectral effects were analyzed using PM6 semiempirical calculations. Formation of complexes was also confirmed by ¹H NMR spectroscopy.

Keywords: 1,8-naphthalimide, crown compounds, sensors, photoinduced electron transfer (PET), internal charge transfer (ICT), optical spectroscopy.

1. Introduction

Detecting cations is of great interest to many scientists, including chemists, biologists, clinical biochemists and environmentalists. With this regard, the development of fluorescent and colorimetric chemosensors seems to be advantageous, since the optical spectroscopy has turned to be one the most effective and reliable analytical methods [1]. Optical sensor devices are generally multi-component systems comprising a signaling moiety (chromophore) and cation binding site (usually termed as receptor). The components are chosen so that the complexation

event triggers the modulation of photophysical characteristics of the chromophore such as the position of emission / absorption band maxima or the fluorescence intensity. Noteworthy, the mechanism of binding signal transduction between the sensor components may differ depending on the sensor design and may involve the photoinduced electron transfer (PET), the internal charge transfer (ICT), the resonance energy transfer or the formation of excimer / exciplex [2–5].

1,8-Naphthalimides bearing electron donating substituents in the naphthalene moiety are well-known organic fluorophores showing good photostability, large Stokes shifts and strong absorption and emission in the UV/Vis region. Such compounds have been used in a large variety of areas, including coloration of polymers and optical brightening [6–8], laser active media [9,10], luminescent crack detection [11], electroluminescent materials [12–14], anticancer agents [15], liquid crystal displays [16,17] and fluorescent markers in biology [18–20]. The ability of 1,8-naphthalimide derivatives to be very sensitive to their surrounding render them as solvatochromic probes where the effects of solvent polarity on the electronic emission and absorption spectra are studied [21,22].

Owing to the unique feature of the naphthalimide chromophore that consists in its electron-accepting properties, and thus possibility of photoinduced electron / energy transfer process or excimer / exciplex formation in the excited state, a significant number of naphthalimides have been developed as fluorescent chemosensors for metal cations and protons. pH-Dependent fluorescence was found for piperazine substituted 4-aminonaphthalimides, which were configured as «fluorophore–spacer–receptor» systems playing on the PET ON-OFF response [23,24]. Wang et al. synthesized a fluorescent naphthalimide based PET-sensor for Zn^{2+} with di-2-picolylamine as a recognizing group [25]. Typically to ICT-sensors, 4-piperidine-naphthalimide derivatives containing 2-imino-oxazolidin (thiazolidin) side chain at the imide moiety exhibited strong fluorescence quenching and some red shift in weakly acidic conditions, caused by the formation of an intramolecular H-bond [26]. A selective and ratiometric Cu²⁺ fluorescent probe based on naphthalimide excimer–monomer switching was designed by Xu et al. [27].

Even though crown ether receptors have been extensively used in the construction of fluorescent probes, there have been only few examples of naphthalimide chemosensors bearing crown ether groups as a receptor unit [28–33]. Recently, we studied the cation-dependent fluorescent properties of benzo-15-crown-5-containg 4-amino- and 4-(acetyl)amino-*N*-aryl-1,8-naphthalimides **1c** and **2c** (Scheme 1) [34]. It was shown that the complex formation with the alkaline earth metal cations (Mg^{2+} , Ba^{2+}) results in enhancement of emission intensity in the case of 4-(acetyl)aminonaphthalimide **2c** due to inhibition of the PET process, which is originally

operated in the free ligand. In contrast, **2a** demonstrated the lack of fluorescence PET switching upon the coordination with metal ions.

Herein, we report on the synthesis and investigation of complexation properties of aza-15-crown-5-substituted naphthalimides **1b**, **2b** and **3a**. Considering the general principles used in the design of fluorescent probes [2–4], there exist two opportunities for combination of azacrown ether receptor and naphthalimide chromophore. In the compounds **1b** and **2b**, the receptor is attached to the *N*-aryl moiety. Alternatively, the aza-15-crown-5 may be presented in the molecule as a substituent at C-4 of the naphthalimide residue in a manner ensuring the conjugation between the receptor's amino nitrogen and carbonyl groups of the carboxyimide functionality, the feature implemented by us in the design of sensor **3a**. In such a case cation detection is possible not only by monitoring of emission intensity but also position of absorption and fluorescence maxima, which may be of interest for the development of ratiometric probes.

2. Experimental

2.1. Materials and general methods

4-Nitro-1,8-naphthalic anhydride [35,36] **4**, 4-bromo-*N*-phenyl-1,8-naphthalimide **9** [37] and *N*-(4-aminophenyl)aza-15-crown-5 ether **7** [38] were prepared by literature procedures. All other reagents were purchased from commercial sources and were of the highest grade. Solvents were purified and dried according to standard procedures. Calcium perchlorate was dried in vacuum (7–8 mm Hg) at 240 °C and kept anhydrous over P_2O_5 in desiccator (Caution! Calcium perchlorate may explode when heating. It decomposes at 270 °C [39]). Raney nickel catalyst was prepared from nickel aluminum alloy (weight percentage of nickel is 50%) according to the known procedure [40].

Melting points were measured on Melt-temp melting point electrothermal apparatus and were uncorrected. The reaction course and purity of the final products was followed by TLC on silica gel (DC-Alufolien Kieselgel 60 F_{254} , Merck). Column chromatography was conducted over silica gel (Kieselgel 60, particle size 0.063-0.200 mm, Merck) using as an eluent the gradient solvent mixtures benzene – ethanol and hexane – ethylacetate. FT-IR spectra were recorded on Magna-IR 750 Nicolet spectrometer and were obtained with potassium bromide pellets. Elemental analyses were carried out in the Microanalysis Laboratory of the A.N. Nesmeyanov Institute of Organoelement Compounds.

NMR measurements. ¹H and ¹³C NMR spectra were recorded on an Avance 300, Avance 400 and Avance 600 spectrometers (Bruker) operating at 300.13, 400.13, 600.22 MHz (for ¹H) and 75.47, 100.61, 150.93 MHz (for ¹³C) respectively. The measurements were performed in DMSO- d_6 , CDCl₃ and CD₃CN solutions. The chemical shifts (given as δ) were

determined with an accuracy of 0.01 ppm relative to the signals corresponding to the residual solvents and recalculated to the internal standard (TMS); the spin-spin coupling constants (*J*) were measured with an accuracy of 0.1 Hz. The numbering of carbon atoms in the naphthalimide moiety, *N*-aryl ring and crown ether fragment of compounds **1b**, **2b**, **3a**, **5** and **6** commonly used by us for the description of ¹H and ¹³C NMR spectra is shown on Fig. 2, 5 and 8. The assignment of ¹H and ¹³C signals is based on 2D NMR experiments (HMBC, HSQC, ¹H COSY), which were performed using standard pulse sequences from the Bruker library. 2D NMR spectra are presented in Supplementary data.

Mass spectrometry. LC-ESI-MS analyses were performed on a Finnigan LCQ Advantage mass spectrometer equipped with octopole ion-trap mass-analyzer, MS Surveyor pump, Surveyor auto sampler, Schmidlin-Lab nitrogen generator (Germany) and Finnigan X-Calibur 1.3 software for data collecting and processing. Acetonitrile (Panreac, HPLC-gradient grade) was used as the mobile phase. Dilute MeCN solutions of compounds **1b**, **2b**, **3a** and **5** were studied in the presence of 1 eq. of HClO₄. The isocratic elution was maintained at a flow rate of 50 μ L · min⁻¹ without column. The effluent from LC was passed directly into the electrospray ion source without a split. Positive electrospray ionization was achieved using a ionization voltage at 3 kV with temperature at 200 °C. Isotope patterns were calculated with Molecular Weight Calculator, Version 6.37 (Matthew Monroe).

Electron impact (EI) (70 eV) mass spectra were obtained from Finnigan Polaris Q instrument (ion-trap) in standard conditions.

Optical measurements. The absorption spectra were taken on a Varian-Cary 5G spectrophotometer. The fluorescence quantum yield measurements were performed using a Varian-Cary 5G spectrophotometer and a FluoroMax-3 spectrofluorimeter. Spectral measurements were carried out in air-saturated acetonitrile solutions (acetonitrile of spectrophotometric grade, water content <0.005%, Aldrich) at 20 ± 1 °C; the concentrations of studied compounds were of about $0.5-2.0 \cdot 10^{-5}$ M. All measured fluorescence spectra were corrected for the nonuniformity of detector spectral sensitivity. Coumarin 481 in acetonitrile (ϕ^{fl} = 0.08 [41] was used as reference for the fluorescence quantum yield measurements. The fluorescence quantum yields were calculated by equation the (1) [42]: wherein φ_{i}^{fl} and φ_{0}^{fl} are the fluorescence quantum yields of the studied solution and the standard compound, respectively; D_i and D_0 are the absorptions of the studied solution and the standard, respectively; S_i and S_0 are the areas underneath the curves of the fluorescence spectra of the studied solution and the standard, respectively; and n_i and n_0 are the refraction indices of the solvents for the substance under study and the standard compound ($n_i = n_0 = 1.342$, acetonitrile).

Equilibrium constant determination. Complex formation of dyes 1b, 2b and 3a with Ca^{2+} in acetonitrile at 20 ± 1 °C was studied by spectrofluorometric (for 1b, 2b and 3a) and spectrophotometric (for 3a) titration [43,44]. The ratio of dye to Ca^{2+} was varied by adding aliquots of a solution of $Ca(ClO_4)_2$ of known concentration to a solution of ligands 1b, 2b or 3a of known concentration. The fluorescence (absorption) spectrum of each solution was recorded, and the stability constants of the complexes were determined using the SPECFIT/32 program (Spectrum Software Associates, West Marlborough, MA). The following equilibria were considered in the fitting (equations (1) and (2), L = 1b, 2b or 3a; M = Ca^{2+}): In doing so, it was found that the experimental data corresponded to the theoretical ones if only the equation (2) was taken into account and the formation of the complexes with composition of 2 : 1 was not observed.

The equilibrium constants for protonation of ligands **1b** and **2b** were not determined by this method because of high stability ($K > 10^7 \text{ M}^{-1}$) of protonated forms (**1b**)·H⁺ and (**2b**)·H⁺.

Determination of fluorescence quantum yields of complexes. The fluorescence quantum yields of complexes (1b)·Ca²⁺ and (2b)·Ca²⁺ were determined using solutions of ligands 1-2b in CH₃CN containing an excess of Ca(ClO₄)₂ in order to obtain 90-95% of ligand bound with the cation. The required Ca(ClO₄)₂ excess was calculated from the known stability constants using the SPECFIT/32 program. The measurements of φ^{fl} for (1b)·H⁺ and (2b)·H⁺ were done in the presence of 2 eq. HClO₄ in ligand solutions, which can be understood from the fact that the further addition of HClO₄ did not result in fluorescence enhancement and complex formation had already been finished. In the case of complex (3a)·Ca²⁺, the direct quantum yield determination was impossible because the presence of considerable amount of the free ligand 3a which appreciably contributes the measured total absorption at λ_{ex} and integrated emission signal might not be avoided even at high excess of calcium cation. The quantum yield of (3a)·Ca²⁺.

X-ray diffraction analysis. Orange crystals of compound **1b** were prepared by slow evaporation of solution of **1b** in the mixture of benzene and ethanol (v/v = 1/1) at room temperature. Crystals of **1b** (C₂₈H₃₁N₃O₆) at 100 K are monoclinic, space group Cc, a = 13.801(2), b = 14.290(2), c = 12.990(2) Å, $\beta = 98.634(4)^{\circ}$, V = 2532.9(5) Å³, Z = 4, M = 505.56, $d_{calc} = 1.326$ g cm⁻³, μ (MoK α) = 0.94 cm⁻¹, F(000) = 1072. Intensities of 7584 reflections were measured with a Smart APEX2 CCD diffractometer at 100 K (λ (MoK α) = 0.71072 Å, ω -scans with 0.3° step, $2\theta < 58^{\circ}$), and 3356 independent reflections ($R_{int} = 0.0425$) were used in further refinement. The structure was solved by direct method and refined by the full-matrix least-squares technique against F^2 in the anisotropic-isotropic approximation. Hydrogen atoms of NH₂

group were located from the Fourier synthesis while all others were placed in calculated positions and refined in the riding model. The refinement converged to $wR_2 = 0.0950$ and GOF = 1.009 for all independent reflections ($R_1 = 0.0460$ was calculated against *F* for 2595 observed reflections with $I > 2\sigma(I)$). All calculations were performed using SHELX software package [45]. Crystallographic data for the structure of **1b** has been deposited with the Cambridge Crystallographic Data Centre as supplemental publication CCDC 916216. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre through www.ccdc.cam.ac.uk/data_request/cif.

Computational details. Quantum chemical calculations were carried out by the MOPAC 2009 program package using the PM6 semiempirical method [46]. CI calculations were performed at optimized geometries, which reached gradient variations less than 0.01 kcal/mol. The solvent effect was included in geometry optimizations following the "COnductorlike Screening MOdel" (COSMO) implemented in MOPAC 2009. A dielectric constant of $\varepsilon = 40$ and a refraction index of solvent (*n*) such that $n^2 = 2$ were used. The CI included eight occupied and eight unoccupied MOs.

2.2. Synthesis of the compounds

2-(4-(1,4,7,10-Tetraoxa-13-azacyclopentadecan-13-yl)phenyl)-6-amino-1Hbenzo[d,e]isoquinoline-1,3(2H)-dione (1b). Method A. To a solution of compound 5 (242 mg, 0.452 mmol) in 1,4-dioxane (10.3 mL) hydrazine-hydrate (1.8 mL, 100 wt. %) was added at 90 °C. Raney nickel catalyst prepared from 1.5 g of nickel aluminum alloy was added portionwise to the reaction mass over a period of 30 min. The resulting mixture was stirred for 1.5 h at 90 °C (TCL control in a solvent system benzene – ethanol, v/v = 5/1) and then was filtered hot. The filtrate was evaporated under vacuum. The crude product was purified by column chromatography on silica gel using the gradient solvent mixture benzene – ethanol and then recrystallized from ethanol. Yield 112 mg (49%). M. p. 282 - 284 °C. ¹H NMR (600.22 MHz, DMSO- d_6 , 31 °C): $\delta = 3.51 - 3.60$ (m, 16H; CH₂(15), CH₂(17), CH₂(18), CH₂(19), $CH_2(20), CH_2(21), CH_2(22), CH_2(24)), 3.67 - 3.72$ (m, 4H; $CH_2(16), CH_2(23)), 6.70$ (d, ³J(H,H) = 8.9, 2H; H(11), H(13)), 6.87 (d, ${}^{3}J$ (H,H)= 8.5, 1H; H(3)), 7.01 (d, ${}^{3}J$ = 8.9, 2H; H(10), H(14)), 7.40 (br s, 2H; NH₂), 7.67 (dd, ${}^{3}J(H,H) = 7.3$, ${}^{3}J(H,H) = 8.6$, 1H; H(6)), 8.18 (d, ${}^{3}J(H,H) = 8.5$, 1H, H(2)), 8.41 (d, ${}^{3}J(H,H) = 7.3$, 1H; H(7)), 8.64 (d, ${}^{3}J(H,H) = 8.6$, 1H; H(5)). ${}^{13}C$ NMR (100.61 MHz, DMSO- d_6 , 27 °C): $\delta = 52.10$ (CH₂(15), CH₂(24)), 67.93 (CH₂(16), CH₂(23)), 69.12 (CH₂(19), CH₂(20)), 69.45 (CH₂(17), CH₂(22)), 70.38 (CH₂(21), CH₂(18)), 108.05 (C(1)), 108.16 (C(3)), 110.86 (C(11), C(13)), 119.49 (C(4a)), 122.40 (C(8)), 124.00 (C(6)), 124.18 (C(9)), 129.31 (C(5)), 129.56 (C(10), C(14)), 130.08 (C(8a)), 131.07 (C(7)), 133.95 (C(2)), (C(2))

146.70 (C(12)), 152.65 (C(4)), 163.51 (C(8b)), 164.35 (C(8c)). FT-IR (KBr), cm⁻¹: 3355, 3415, 3255 (v_{NH2}); 3080, 2865 (v_{CH}), 1680, 1635 ($v_{N-C=0}$). ESI-MS in MeCN, calculated for [M+H]⁺, m/z: 506.23; found: 506.19. Elemental analysis: calculated (%) for C₂₈H₃₁N₃O₆ (MW 505.56): C 66.53, H 6.14, N 8.32; found C 66.56, H 6.10, N 8.27. Method B. To a suspension of 4nitronaphthalimide 5 (68 mg, 0.127 mmol) in ethanol (1.0 mL) a solution of SnCl₂·2H₂O (100 mg) in concd hydrochloric acid (0.5 mL, $\rho = 1.18$ g/mL) was added dropwise and the mixture was stirred at ambient temperature for 2 h. The reaction mixture was poured into NaOH water solution (7.0 mL, 5 wt. %). The brown precipitate was filtered off, washed with distilled water, dried at 60 °C and then chromatographed on silica gel using gradient benzene – ethanol mixture as an eluent. Consequently, two fractions were isolated: fraction 1 (10 mg) and fraction 2 (14 mg). Fraction 1 was found to be 3-chloro-4-aminonaphthalimide 6 (the data for the compound 6are listed below), fraction 2 - target product 1b (yield 22%). M. p. 264 - 272 °C. ¹H NMR (300.13 MHz, DMSO- d_6 , 23 °C): $\delta = 3.46 - 3.63$ (m, 16H; CH₂(15), CH₂(17), CH₂(18), CH₂(19), CH₂(20), CH₂(21), CH₂(22), CH₂(24)), 3.64 - 3.75 (m, 4H; CH₂(16), CH₂(23)), 6.69 $(d, {}^{3}J(H,H) = 8.9, 2H; H(11), H(13)), 6.86 (d, {}^{3}J(H,H) = 8.5, 1H; H(3)), 7.01 (d, {}^{3}J(H,H) = 8.9, 2H; H(11), H(13)), 6.86 (d, {}^{3}J(H,H) = 8.9, 2H; H(13)), 7.01 (d, {}^{3}J(H,H) = 8.9, 2H; H(11), H(13)), 6.86 (d, {}^{3}J(H,H) = 8.9, 2H; H(13)), 7.01 (d, {}^{3}J(H,H) = 8.9, 2H; H(11), H(13)), 7.01 (d$ 2H; H(10), H(14)), 7.44 (br s, 2H; NH₂), 7.67 (dd, 1H, ${}^{3}J$ (H,H) = 7.3, ${}^{3}J$ (H,H) = 8.6; H(6)), 8.18 $(d, {}^{3}J(H,H) = 8.5, 1H; H(2)), 8.41 (dd, {}^{4}J(H,H) = 0.7, {}^{3}J(H,H) = 7.3, 1H; H(7)), 8.64 (dd, {}^{4}J(H,H))$ $= 0.7, {}^{3}J(H,H) = 8.6, 1H; H(5))$. EI-MS, m/z (I, %): 506 (19), 505 (59) [M]⁺, 449 (29), 386 (34), 360 (46), 330 (54), 316 (100), 303 (34), 196 (36), 195 (81).

N-(2-(4-(1,4,7,10-Tetraoxa-13-azacvclopentadecan-13-vl)phenvl)-1,3-dioxo-2,3dihydro-1H-benzo[d,e]isoquinolin-6-yl)acetamide (2b). To a suspension of compound 1b (40 mg, 0.079 mmol) in an anhydrous dichloromethane (1.0 mL) acetylchloride (60 µL, 0.844 mmol) was added. The reaction mass was refluxed and stirred for 3 h to afford a clear solution. After the completion of the reaction, the mixture was diluted with ethanol (1.0 mL). The solvents were removed under vacuum and Na₂CO₃ water solution (5.0 mL, 10 wt. %) was added to the residual solid to neutralize HCl. Product 2b was extracted with dichloromethane, the extract was washed with distilled water and evaporated under vacuum. The resulting solid (38 mg) was recrystallized from ethanol to afford 33 mg of pure **2b** (yield 76%). M. p. 114 - 117 °C. ¹H NMR (300.13) MHz, DMSO- d_6 , 21 °C): $\delta = 2.30$ (s, 3H; CH₃), 3.48 – 3.64 (m, 16H; CH₂(15), CH₂(17), CH₂(18), CH₂(19), CH₂(20), CH₂(21), CH₂(22), CH₂(24)), 3.64 - 3.75 (m, 4H; CH₂(16), CH₂(23)), 6.71 (d, 2H, ${}^{3}J(H,H) = 8.9$; H(11), H(13)), 7.09 (d, ${}^{3}J(H,H) = 8.9$, 2H; H(10), H(14)), 7.85 - 7.97 (m, 1H; H(6)), 8.33 (d, 1H, ${}^{3}J(H,H) = 8.2$; H(3)), 8.47 (d, 1H, ${}^{3}J(H,H) = 8.2$; H(2)), 8.52 (d, ${}^{3}J(H,H) = 7.3$, 1H; H(7)), 8.74 (d, ${}^{3}J(H,H) = 8.6$, 1H; H(5)), 10.44 (br s, 1H; NHCO). ¹³C NMR (150.93 MHz, DMSO- d_6 , 21 °C): $\delta = 24.09$ (CH₃), 52.12 (CH₂(15), CH₂(24)), 67.91 (CH₂(16), CH₂(23)), 69.12 (CH₂(19), CH₂(20)), 69.47 (CH₂(17), CH₂(22)), 70.38 (CH₂(18), CH₂(21)), 110.93 (C(11), C(13)), 118.12 (C(1)), 119.44 (C(3)), 122.96 (C(8)), 123.54 (C(9)), 124.16 (C(4a)), 126.40 (C(6)), 128.71 (C(8a)), 129.23 (C(5)), 129.47 (C(10), C(14)), 130.88 (C(7)), 131.61 (C(2)), 140.26 (C(4)), 146.96 (C(12)), 163.51 (C(8b)), 164.07 (C(8c)), 169.62 (CH₃<u>C</u>O). FT-IR (KBr), cm⁻¹: 3350 (v_{NH}); 2920, 2855 (v_{CH}); 1705, 1660 (v_{N-C=O}); 1520 (δ_{NH}). ESI-MS in MeCN, calculated for [M+H]⁺, m/z: 548.24; found: 548.18. Elemental analysis: calculated (%) for C₃₀H₃₃N₃O₇ (MW 547.60): C 65.80, H 6.07, N 7.67; found C 65.71, H 6.06, N 7.62.

2-Phenyl-6-(1,4,7,10-tetraoxa-13-azacyclopentadecan-13-yl)-1H-benzo-[d,e]isoquinoline-1,3(2H)-dione (3a). A mixture of 4-bromo-N-phenyl-1.8-naphthalimide 9 (422 mg, 1.199 mmol) and aza-15-crown-5 ether (988 mg, 4.511 mmol) was heated to reflux in methoxylethanol (3 mL) under argon atmosphere for 8 h (TCL control in a solvent system benzene – ethanol, v/v = 5/1). After removal of methoxylethanol, the solid was purified by silica gel column chromatography using benzene – ethanol gradient mixture as an eluent. The weight of isolated product **3a** was 220 mg (yield 37%). M. p. 149 - 150 °C. ¹H NMR (300.13 MHz, DMSO- d_6 , 21 °C), $\delta = 3.52 - 3.67$ (m, 16H; CH₂(15), CH₂(17), CH₂(18), CH₂(19), CH₂(20), CH₂(21), CH₂(22), CH₂(24)), 3.68 – 3.77 (m, 4H; CH₂(16), CH₂(23)), 7.30 – 7.38 (m, 2H; H(10), H(14), 7.41 – 7.58 (m, 4H; H(3), H(11), H(12), H(13)), 7.79 (dd, ${}^{3}J(H,H) = 7.3$, ${}^{3}J(H,H) = 8.5$, 1H; H(6)), 8.36 (d, ${}^{3}J(H,H) = 8.2$, 1H; H(2)), 8.46 (dd, ${}^{4}J(H,H) = 0.7$, ${}^{3}J(H,H) = 7.3$, 1H; H(7)), 8.81 (dd, ${}^{4}J(H,H) = 0.7$, ${}^{3}J(H,H) = 8.5$, 1H; H(5)). ${}^{13}C$ NMR (100.61 MHz, DMSO- d_{6} , 27 °C): δ = 52.76 (CH₂(15), CH₂(24)), 68.66 (CH₂(16), CH₂(23)), 69.85 (2×CH₂), 69.97 (2×CH₂), 70.42 (2×CH₂), 114.93 (C(1)), 116.59 (C(3)), 122.86 (C(8)), 125.42 (C(6)), 126.22 (C(4a)), 128.04 (C(12)), 128.81 (C(11), C(13)), 129.18 (C(10), C(14)), 130.09 (C(8a)), 130.69 (C(7)), 131.67 (C(2) or C(5)), 131.76 (C(5) or C(2)), 136.27 (C(9)), 155.41 (C(4)), 163.28 (C(8b)), 163.91 (C(8c)). ESI-MS in MeCN, calculated for $[M+H]^+$, m/z: 491.22; found: 491.29. EI-MS, m/z (I, %): 490 (32) [M]⁺, 357 (34), 341 (37), 316 (49), 315 (91), 314 (100), 313 (66), 301 (59), 299 (52), 288 (47), 208 (34), 196 (34). Elemental analysis: calculated (%) for C₂₈H₃₀N₂O₆ (MW 490.21): C 68.56, H 6.16, N 5.71; found C 68.49, H 6.19, N 5.68.

2-(4-(1,4,7,10-Tetraoxa-13-azacyclopentadecan-13-yl)phenyl)-6-nitro-1*H*-benzo[d,e]isoquinoline-1,3(2*H*)-dione (5). A solution of a mixture of 4-nitro-1,8-naphthalic anhydride 4 (304 mg, 1.251 mmol) and *N*-(4-aminophenyl)aza-15-crown-5-ether 7 (554 mg) in 80 % acetic acid (1.3 mL) was kept at reflux for 1.5 h. The reaction mass was cooled down; the crystals precipitated upon cooling were filtered off and washed on a filter with distilled water. The product was dried at 80°C and purified by column chromatography on silica gel using the mixture of benzene – ethanol (v/v = 50/1) as an eluent. The weight of the obtained product **5** was 401 mg (yield 60 %). M. p. 227 – 229 °C. ¹H NMR spectra (400.13 MHz, DMSO- d_6 , 50 °C):

2-(4-(1,4,7,10-Tetraoxa-13-azacyclopentadecan-13-yl)phenyl)-6-amino-5-chloro-1Hbenzo[d,e]isoquinoline-1,3(2H)-dione (6). According to the Method B for the reduction of nitro group in the compound 5 (see above), 10 mg of 4-amino-3-chloronaphthalimide 6 (yield 15%) was obtained from 68 mg of 5, 100 mg of SnCl₂·2H₂O, 0.5 mL of concd HCl and 1.0 mL of ethanol. M. p. 288 – 291 °C. ¹H NMR (300.13 MHz, DMSO- d_6 , 23 °C): $\delta = 3.49 - 3.63$ (m, 16H; CH₂(15), CH₂(17), CH₂(18), CH₂(19), CH₂(20), CH₂(21), CH₂(22), CH₂(24)), 3.64 - 3.75 (m, 4H; CH₂(16), CH₂(23)), 6.69 (d, ${}^{3}J(H,H) = 8.9$, 2H; H(11), H(13)), 7.03 (d, ${}^{3}J(H,H) = 8.9$, 2H; H(10), H(14)), 7.55 (br s, 2H; NH₂), 7.76 (dd, 1H, ${}^{3}J(H,H) = 7.3$, ${}^{3}J(H,H) = 8.6$; H(6)), 8.21 (s, 1H; H(2)), 8.44 (dd, ${}^{4}J(H,H) = 0.7$, ${}^{3}J(H,H) = 7.3$, 1H; H(7)), 8.78 (dd, ${}^{4}J(H,H) = 0.7$, ${}^{3}J(H,H) = 0.7$ 8.6, 1H; H(5)). ¹³C NMR (150.93 MHz, DMSO- d_6 , 25 °C): $\delta = 52.10$ (CH₂(15), CH₂(24)), 67.91 (CH₂(16), CH₂(23)), 69.12 (CH₂(19), CH₂(20)), 69.47 (CH₂(17), CH₂(22)), 70.38 (CH₂(18), CH₂(21)), 109.48 (C(1)), 110.89 (C(11), C(13)), 111.71 (C(3)), 120.29 (C(4a)), 122.74 (C(8)), 123.76 (C(9)), 125.40 (C(6)), 128.35 (C(8a)), 129.37 (C(5)), 129.48 (C(10), C(14)), 131.04 (C(7)), 132.65 (C(2)), 146.82 (C(12)), 147.51 (C(4)), 162.85 (C(8b)), 164.03 (C(8c)). EI-MS, m/z (*I*, %): 541 (20) $[M+2]^+$, 539 ($[M]^+$) (55), 486 (26), 420 (29), 394 (49), 380 (37), 364 (59), 350 (100), 337 (38), 231 (28), 229 (75). Elemental analysis: calculated (%) for C₂₈H₃₀ClN₃O₆ (MW 540.01): C 62.28, H 5.60, N 7.78; found C 62.34, H 5.65, N 7.74.

3. NMR data for the free dyes and their complexes with Ca^{2+} in CD_3CN .

Compound 1b. $C_{\rm L} = 6.8 \cdot 10^{-3}$ M. ¹H NMR (300.13 MHz, CD₃CN, 21 °C): $\delta = 3.50 - 6.64$ (m, 16H; CH₂(15), CH₂(24), CH₂(17), CH₂(22), CH₂(18), CH₂(21), CH₂(19), CH₂(20)), 3.69 - 3.78 (m, 4H; CH₂(16), CH₂(23)), 5.90 (br s, 2H; NH₂), 6.77 (d, ³*J*(H,H) = 8.9, 2H; H(11), H(13)), 6.93 (d, ³*J*(H,H) = 8.3, 1H; H(3)), 7.04 (d, ³*J*(H,H) = 8.9, 2H; H(10), H(14)), 7.69 (dd,

 ${}^{3}J(H,H) = 7.3$, ${}^{3}J(H,H) = 8.3$, 1H; H(6)), 8.27 (d, ${}^{3}J(H,H) = 8.3$, 1H; H(2)), 8.35 (d, ${}^{3}J(H,H) = 8.3$, 1H; H(7)), 8.49 (d, ${}^{3}J(H,H) = 7.3$, 1H; H(5)).

Compound 2b. $C_{\rm L} = 1.0 \cdot 10^{-2}$ M. ¹H NMR (300.13 MHz, CD₃CN, 21 °C): $\delta = 2.30$ (s, 3H; CH₃CO), 3.53 – 6.65 (m, 16H; CH₂(15), CH₂(24), CH₂(17), CH₂(22), CH₂(18), CH₂(21), CH₂(19), CH₂(20)), 3.70 – 3.78 (m, 4H; CH₂(16), CH₂(23)), 6.79 (d, ³*J*(H,H) = 8.9, 2H, H(11), H(13)), 7.08 (d, ³*J*(H,H) = 8.9, 2H; H(10), H(14)), 7.83 (dd, ³*J*(H,H) = 7.2, ³*J*(H,H) = 8.5, 1H; H(6)), 8.31 (d, ³*J*(H,H) = 8.2, 1H; H(3)), 8.49 (d, ³*J*(H,H) = 8.2, 1H; H(2)), 8.52 (d, ³*J*(H,H) = 8.5, 1H; H(7)), 8.55 (d, ³*J*(H,H) = 7.2, 1H; H(5)); 8.88 (br s, 1H; NHCO).

Compound 3a. $C_{\rm L} = 1.0 \cdot 10^{-2}$ M. ¹H NMR (300.13 MHz, CD₃CN, 21 °C): $\delta = 3.55 - 3.70$ (m, 16H; CH₂(15), CH₂(24), CH₂(17), CH₂(22), CH₂(18), CH₂(21), CH₂(19), CH₂(20)), 3.71 - 3.80 (m, 4H; CH₂(16), CH₂(23)), 7.27 - 7.38 (m, 2H; H(10), H(14)), 7.42 - 7.59 (m, 4H; H(3), H(11), H(12), H(13)), 7.74 (dd, ³*J*(H,H) = 7.2, ³*J*(H,H) = 8.6, 1H; H(6)), 8.41 (d, ³*J*(H,H) = 8.3, 1H; H(2)), 8.50 (dd, ⁴*J*(H,H) = 1.1, ³*J*(H,H) = 7.2, 1H; H(7)), 8.85 (dd, ⁴*J*(H,H) = 1.1, ³*J*(H,H) = 8.6, 1H; H(5)).

Complex (1b)·Ca²⁺. $C_{\rm L} = 1.0 \cdot 10^{-2}$ M. Metal to ligand ratio M : L = 3 : 1. ¹H NMR (300.13 MHz, CD₃CN, 21 °C): $\delta = 3.39 - 3.49$ (m, 4H; CH₂(15), CH₂(24)), 3.82 - 3.98 (m, 16H; CH₂(16), CH₂(17), CH₂(18), CH₂(19), CH₂(20), CH₂(21), CH₂(22), CH₂(23)), 6.02 (br s, 2H; NH₂), 6.94 (d, ³*J*(H,H) = 8.3, 1H; H(3)), 7.34 (d, ³*J*(H,H) = 8.9, 2H; H(11), H(13)), 7.45 (d, ³*J*(H,H) = 8.9, 2H; H(10), H(14)), 7.70 (dd, ³*J*(H,H) = 7.3, ³*J*(H,H) = 8.4, 1H; H(6)), 8.28 (d, ³*J*(H,H) = 8.3, 1H; H(2)), 8.39 (dd, ⁴*J*(H,H) = 1.0, ³*J*(H,H) = 8.4, 1H; H(7)), 8.50 (dd, ⁴*J*(H,H) = 1.0, ³*J*(H,H) = 7.3, 1H; H(5)).

Complex (2b)·Ca²⁺. $C_{\rm L} = 1.0 \cdot 10^{-2}$ M. Metal to ligand ratio M : L = 3 : 1. ¹H NMR (300.13 MHz, CD₃CN, 21 °C): $\delta = 2.33$ (s, 3H; CH₃CO), 3.41 – 3.50 (m, 4H; CH₂(15), CH₂(24)), 3.84 – 3.99 (m, 16H; CH₂(16), CH₂(17), CH₂(18), CH₂(19), CH₂(20), CH₂(21), CH₂(22), CH₂(23)), 7.36 (d, ³*J*(H,H) = 8.9, 2H; H(11), H(13)), 7.47 (d, ³*J*(H,H) = 8.9, 2H; H(10), H(14)), 7.88 (dd, ³*J*(H,H) = 7.4, ³*J*(H,H) = 8.4, 1H; H(6)), 8.30 (d, ³*J*(H,H) = 8.1, 1H; H(3)), 8.48 – 8.64 (m, 3H; H(2), H(5), H(7)), 9.06 (br s, 1H; NHCO).

Complex (3a)·Ca²⁺. $C_{\rm L} = 1.0 \cdot 10^{-2}$ M. Metal to ligand ratio M : L = 5 : 1. ¹H NMR (300.13 MHz, CD₃CN, 21 °C): $\delta = 3.64 - 3.73$ (m, 4H; CH₂(15), CH₂(24)), 3.76 - 3.91 (m, 16H; CH₂(16), CH₂(17), CH₂(18), CH₂(19), CH₂(20), CH₂(21), CH₂(22), CH₂(23)), 7.30 - 7.39 (m, 2H; H(10), H(14)), 7.45 - 7.60 (m, 4H; H(3), H(11), H(12), H(13)), 7.86 (dd, ³J(H,H) = 7.2, ³J(H,H) = 8.6, 1H; H(6), 8.51 (d, ³J(H,H) = 8.2, 1H; H(2)), 8.58 (dd, ⁴J(H,H) = 1.0, ³J(H,H) = 7.2, 1H; H(7)), 8.78 (dd, ⁴J(H,H) = 1.0, ³J(H,H) = 8.6, 1H; H(5)).

3. Results and Discussion

3.1. Synthesis of crown containing naphthalimides

Compounds 1b and 2b containing crown a ether fragment in the N-aryl moiety were prepared as shown in Scheme 2. Starting from 4-nitronaphthalic anhydride 4, the intermediate 4-nitro-1,8-napthalimide 5 was afforded by the reaction with N-(4-aminophenyl)aza-15-crown-5 ether 7 in AcOH media. The reduction of nitro group in 5 to yield 1b was performed by the hydrazine hydrate over Raney nickel catalyst in 1,4-dioxane solution. Unexpectedly, the use of tin (II) chloride with hydrochloric acid for the reduction didn't allow us to isolate compound 1b with high yield unlike the synthesis of 4-aminonaphthalides containing benzo-15-crown-5 ether groups we have described previously [47]. Moreover, we observed simultaneous introduction of chlorine atom in *ortho*-position towards the amino group in this case. At the final step, the treatment of **1b** with acetylchloride under anhydrous conditions resulted in 4-(acetyl)aminonaphthalimide 2b.

Scheme 2 also shows synthesis of azacrown ether derivative 3a. According to the routine procedure [37], 4-bromonaphthalic anhydride 8 first was refluxed in glacial acetic acid with aniline. Subsequent nucleophilic substitution of the bromine atom in the isolated *N*-phenylimide 9 with aza-15-crown-5 ether yielded target aminonaphthalimide 3a.

Amine **7** used in Scheme 2 is a known compound; it was obtained from commercially available *N*-(phenyl)diethanolamine and triethylene glycol di(*p*-toluenesulfonate) in a three-step synthesis [38]. 4-Amino- and 4-(acetyl)amino-*N*-phenyl-1,8-naphthalimides **1a** and **2a** have been previously synthesized in our laboratory [47].

3.2. Photophysical properties of the compounds

Photophysical characteristics of the compounds **1a**, **1b**, **2a**, **2b** and **3a** were measured in acetonitrile solution and the data are presented in Table 1. As an example, Fig. 1 shows absorption and emission spectra of *N*-phenylnaphthalimides **1a**, **2a** and **3a**. Similarly to other 4-amino- and 4-amidonaphthalimide derivatives [48,49], the broad and structureless absorption band in the long wavelength region arises from the polarization of chromophore naphthalimide system due to the intramolecular charge transfer (ICT) interaction occurring between the electron-donating substituents at C(4) and the acceptor carboxyimide fragment. Substitution of amino hydrogen in **1a**, **1b** by the acetyl group to produce **2a**, **2b** causes a blue shift of λ_{max}^{abs} and λ_{max}^{fl} with a concomitant changes in fluorescence quantum yields, which is a result of decreased donor ability of NHAc group relative to NH₂.

The introduction of aza-15-crown-5 ether substituent at C-4 of the naphthalene ring when going from **1a** to **3a** is expected to enhance ICT interaction in the molecule because of inductive

effect of two alkyl groups. Surprisingly, we observed only slight bathochromic shift (3 nm) in the absorption spectra (Table 1, Fig. 1). As compared with compound **1a**, the fluorescence maximum of **3a** is 22 nm red-shifted with the emission intensity dramatically decreased. This is very similar to the observations of Saha and Samanta [50] who studied the influence the structure of the amino group on the photophysical properties of 4-aminonaphthalimide derivatives. The small red shift in the absorption spectra can be understood if it is assumed that the dialkylamino group is twisted with respect to the naphthalimide plane in a manner ensuring the partial loss of steric repulsion between the azacrown ether substituent and the *peri* hydrogen atoms of naphthalene residue. Such repulsion disfavors charge separation and reduces the donor properties of azacrown ether substituent. In fact, the PM6 optimized ground state geometry of the compound **3a** depicted in Fig. S1 (see Supplementary data) confirms this assumption.

The relatively low fluorescence quantum yield of compound 3a with respect to 1a is also a result of conformational changes at the 4-amino moiety. As it has been proposed by Saha and [50], the increase in steric strain in the excited Samanta **ICT**-state of 4-dialkylaminonaphthalimides, more planar in nature in comparison with the ground state, enhances the nitrogen inversion rate responded for nonradiative deactivation pathway in these systems.

To predict the cation-induced spectral effects in the case of crown-containing derivatives **1b** and **2b** we studied the influence of the nature of the *N*-aryl group on photophysical properties. *N*-Phenyl substituted naphthalimides **1a** and **2a** were used for comparison. The introduction of electron-donating dialkylamino group in the *N*-aryl fragment doesn't virtually change the position of absorption and emission maxima, whereas the fluorescence quantum yield drops sharply by more than two orders of magnitude (Table 1). The former observation is consistent with the X-ray crystallography data of the compound **1b** (Fig. 2) showing the dihedral angle between the naphthalimide moiety and *N*-aryl group to be close to 90°. Thus, one can conclude that the π -electronic interaction between these two fragments is very weak and negligible in the ground state.

The effect of decrease in emission intensity induced by the presence of azacrown ether groups in the compounds **1b** and **2b** can be explained in terms of PET quenching mechanism commonly used in the design of fluorophore–spacer–receptor signaling systems [2–5]. With this point of view, fast electron transfer process could be suggested between the electron rich *N*-aryl receptor in **1b**, **2b** molecules and the excited naphthalimide fluorophore, which acts as an acceptor. Apparently, the *N*-aryl σ -bond is used as a spacer in this case. According to common principles in the construction of fluorescent PET sensors [2], such spacer enables the required

separation of donor and acceptor units due to their nearly orthogonal disposition in space and also links them together in a shortest way facilitating effective PET.

A striking evidence for existence of PET quenching pathway is also provided by the PM6 semiempirical calculations of the highest occupied and lowest unoccupied molecular orbitals (HOMO–LUMO) for the compounds **1a**, **2a** and **1b**, **2b**. The calculation results are collected in Fig. 3 and Table 2. Fig. 3 shows the energy levels of the frontier orbitals of the naphthalimide fluorophore and local MOs of *N*-aryl groups (marked with spot) in the crown ether containing derivatives **1b**, **2b**. The assignment of local orbitals is based on their images, which are also depicted on Fig. 3 (see Fig. S2 in Supplementary data for orbital images of other molecules and complexes presented in Table 2). As it can be seen, both molecules exhibit the HOMO localized over the benzene ring. The highest occupied MOs of naphthalimide fragments appear to be the HOMO(-1) in whole molecules **1b**, **2b** and therefore possess lower energy. Such situation corresponds to high driving force for PET process occurring between HOMO of *N*-aryl receptor and half occupied HOMO(-1) in the excited state. In contrast, the energy level of the *N*-phenyl group HOMO in the compounds **1a**, **2a** is lower relative to 4-amino- or 4-amidonaphthalimide frontier orbitals (Table 2). Thus, in this case PET is hampered and a high fluorescence quantum yield value should be anticipated.

3.3. Complex formation studies of naphthalimides 1b, 2b and 3a

Having established the spectral effects induced by the introduction of aza-15-crown-5 fragment in *N*-phenyl-1,8-naphthalimides **1a**, **2a**, we examined the ability of crown ether derivatives **1b**, **2b** and **3a** to change their photophysical properties as a result of Ca^{2+} binding, the cation we used for the complexation studies. This cation was chosen because it has high affinity for aza-15-crown-5 ether in acetonitrile solution [51], which is convenient when studying cation-induced optical effects.

Typically to fluorescent PET sensors, the addition of calcium perchlorate to the acetonitrile solutions of ligands **1b**, **2b** does not lead to significant changes in the positions of absorption and fluorescence bands but results in pronounced increase in fluorescence intensity (Table 1). The fluorescence enhancement was assigned to coordination of Ca²⁺ with crown ether moiety in **1b**, **2b** molecules. Such coordination should weaken the electron donor strength of *N*-phenylazacrown ether receptor thus decelerating the PET quenching process. This is consistent with the fact that in complexes (**1b**)·Ca²⁺ and (**2b**)·Ca²⁺ the HOMOs of *N*-aryl groups possess lower energy relative to the free ligands (Table 2). Nearly identical position of λ_{max}^{abs} and λ_{max}^{fl} for **1b**, **2b** in comparison with (**1b**)·Ca²⁺, (**2b**)·Ca²⁺ is obviously due to efficient separation of *N*-arylazacrown ether receptor and naphthalimide chromophore by the *N*–Ar σ -bond. The

fluorescence emission spectra of compounds **1b** and **2b** were recorded in the presence of graduated amount of $Ca(ClO_4)_2$ in acetonitrile solution (see Fig. 4 for **1b** and Fig. S3 in Supplementary data for **2b**) and the titration data were applied for the calculation of stability constants (Table 1).

The NMR analysis clearly confirmed the formation of complexes (Table 3). The addition of Ca^{2+} cations causes the downfield shift of the resonance proton signals of *N*-phenylazacrown ether part. The positions of the other proton signals stay unchanged. As an example, Fig. 5 shows cation induced changes in the ¹H NMR spectrum of **1b**. These results are in good agreement with the optical data demonstrating only small changes in the position of absorption and emission bands of ligands **1b** and **2b** upon the addition of Ca(ClO₄)₂.

As is evident from Table 1, the fluorescence quantum yield value for complex $(1b) \cdot Ca^{2+}$ is very close to that for *N*-phenylnaphthalimide **1a**. In contrast, $(2b) \cdot Ca^{2+}$ demonstrates considerable reduction of emission intensity as compared with **2a**. To explain this fact we proposed that in the case of $(2b) \cdot Ca^{2+}$ PET interaction is only partially blocked due to the relatively low energy level of the highest occupied MO of 4-amidonaphthalimide fluorophore (– 9.50 eV, Table 2) in comparison with 4-aminonaphthalimide fragment (–9.04 eV) in (**1b**) $\cdot Ca^{2+}$. Thus, the decrease in energy of the *N*-aryl localized MO in **1b** occurring under complex formation with Ca^{2+} is high enough to ensure efficient quenching of PET interaction, whereas in (**2b**) $\cdot Ca^{2+}$ PET inhibition is lower because the same changes in the donor ability of *N*-aryl group lead to more proximate disposition of HOMO and HOMO(-1) on energy scale (Table 2).

With the aim to confirm our assumption concerning partial inhibition of PET in the $(2b)\cdot Ca^{2+}$, we measured photophysical characteristics of protonated forms of ligands 1b, 2b (Table 1). The addition of HClO₄ in acetonitrile solutions of 1b, 2b results in formation of highly fluorescent complexes $(1b)\cdot H^+$ and $(2b)\cdot H^+$ similar in φ^{fl} values to 1a, 2a. In these complexes the lone electron pair of receptor's anilino nitrogen is fully engaged in coordination with the cation due to formation of N–H σ -bond, which breaks the conjugation, significantly lowers the potential energy of *N*-aryl group and thus completely quenches PET process in both cases. It should be also noted that in spite of the presence of two amino groups in the compound 1b only the anilino nitrogen of crown ether is subjected to protonation. This results from the observation that the position of λ_{max}^{abs} and λ_{max}^{fl} remains virtually unchanged upon addition of HClO₄.

Unlike the typical PET sensors **1b** and **2b**, the mechanism for binding signal transduction in compound **3a** is supposed to be completely different. In this case the crown ether receptor's nitrogen is part of the π -system of naphthalimide signaling moiety. Thus, the complexation should give rise to dual response in both absorption and fluorescence emission spectra. Fig. 6 shows the influence of gradually increasing amounts of Ca²⁺ on the electronic absorption spectra

of **3a** in acetonitrile solution. As it can be seen, the binding of metal ion causes an enhancement in the absorbance at 352 nm which corresponds to complex $(3a) \cdot Ca^{2+}$ with decreasing the absorbance of the free ligand **3a** centered at 420 nm. Obviously, the blue shift of the long wavelength band is due to strong interaction of the crown ether nitrogen lone pair with the cation and reduced displacement of π -electron density from donor to acceptor unit in naphthalimide chromophore as a result.

The spectrophotometric titration data of **3a** were applied for calculation of complex $(3a) \cdot Ca^{2+}$ stability constant (Table 1) as well as its absorption spectra (marked with dash-dot line on Fig. 6). Considering the higher intensity at 352 nm of the calculated spectra as compared with that of the spectra recorded in the presence of 1000 equivalents of Ca²⁺, one could propose that ligand **3a** is not fully engaged in a calcium complex even at such high excess of metal cation. Indeed, the constant stability value for $(3a) \cdot Ca^{2+}$ appears to be lower by more than two orders of magnitude than in the case of $(1b) \cdot Ca^{2+}$ and $(2b) \cdot Ca^{2+}$, which is a consequence of conjugation between receptor's nitrogen with electron acceptor carbonyl groups of carboxyimide moiety.

The fluorescence spectra of the compound **3a** exhibited pronounced increase in emission intensity with the fluorescence band maxima blue shifted up to 27 nm in the presence of ascending amounts of Ca²⁺ (Fig. 7). Interestingly, the blue shift of the long wavelength absorption band was considerably higher ($\Delta \lambda_{max}^{abs} = \lambda_{max}^{abs}$ [**3a**] – λ_{max}^{abs} [**3a**] · Ca²⁺] = 68 nm). This result indicates that, following the excitation, the nitrogen lone pair breaks its bond with the cation (as already reported for other fluoroionophores [33,52,53]), but the cation is not excluded from the macrocyclic cavity since it is still bonded with the oxygenated part of the crown ether. Such cation – nitrogen atom decoordination leads to ICT excited state in which the emission characteristics of naphthalimide chromophore fairly resemble that of the free ligand **3a**. However, the proximity of positively charged metal ion reduces to some extent the electron donor properties of nitrogen atom due to electrostatic interaction, thereby decreasing the pushpull character of ICT state. This explains the moderate hypsofluoric shift and enhancement of emission intensity. The data presented on Fig. 7 were also used for the calculation of complex stability constant and the obtained lg *K* value was essentially the same to the value derived from spectrophotometric titration experiment.

Upon addition of $Ca(ClO_4)_2$, the changes in ¹H NMR spectrum of compound **3a** in CD_3CN were related only to downfield shift of naphthalimide and crown ether proton signals (Fig. 8), which indicates the receptor's nitrogen lone pair is engaged in the cation coordination. Simultaneously, the position of *N*-phenyl group proton signals remains virtually unchanged.

4. Conclusions

In this work the synthesis and detailed investigation of optical characteristics and complexation ability of naphthalimide derivatives bearing aza-15-crown-5 ether receptor in the N-aryl moiety and as a C(4)-substituent of naphthalene ring were done. It was shown that the presence of crown ether group in the N-aryl moiety of ligands 1b, 2b leads to dramatic decrease of emission intensity with respect to N-phenylnaphthalimides 1a, 2a due to efficient electron transfer between naphthalimide chromophore and receptor unit. Upon the addition of Ca²⁺ the lone pair of electrons of the crown ether receptor's nitrogen atom is engaged in cation binding; this suppresses PET process and causes fluorescence enhancement occurring at constant λ_{max}^{abs} and λ_{max}^{fl} values. In the case of compound **3a** receptor's nitrogen atom is a part of naphthalimide chromophore system, which is a structural feature of the ICT chemosensor. In contrast to PET sensors 1b, 2b, the coordination of Ca^{2+} with 3a reduces ICT interaction and causes significant hypsochromic and hypsofluoric shift. It was also shown that the various positions of the azacrown ether fragment in naphthalimide residue result not only in substantial difference in optical responses but also influence dramatically the affinity of the aza-15-crown-5 ether receptor for calcium cations. Noteworthy, the response of the compound 3a is useful both in absorption and emission spectroscopy. The strong wavelength shifts observed in the presence of Ca^{2+} make this system suitable for dual wavelength analysis in self-calibrating measurements.

The presented results have shown that compounds **1b**, **2b** and **3a** can be of interest for the development of fluorescent and colorimetric chemosensors for different kinds of cationic analysis. Due to versatile structural modification on the crown ether moiety, selective probes based on aza-15-crown-5-containing 4-amino- and 4-amidonaphthalimides for various metal cations can be prepared. Related research is being carried out in our laboratory.

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Appendix A. Supplementary data

Supplementary data associated with this article can be found in the online version at doi

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Schemes



Scheme 1.



Scheme 2.

















Figure and scheme captions

Fig. 1. Steady-state absorption (1, 2, 3) and normalized emission (1', 2', 3') spectra of the compounds 1a (1, 1'), 2a (2, 2') and 3a (3, 3') in acetonitrile $(C \sim 10^{-5} \text{ M})$.

Fig. 2. X-Ray crystallography data for the compound **1b**. C(8b)–N(2)–C(9)–C(10) Dihedral angle value is 84.3 °.

Fig. 3. Energy levels of frontier MOs of **1b** and **2b** molecules. *N*-Aryl group local orbitals are marked with spot. Orbitals in brackets correspond to the excited state of naphthalimide fluorophore. Vertical arrows describe lowest energy electronic transition (ICT transition) in naphthalimide fluorophore.

Fig. 4. The changes in the fluorescence spectrum of 1b in the presence of graduated amount of Ca(ClO₄)₂ in MeCN solution. $C_{\rm L} = 5.0 \cdot 10^{-6}$ M, $\lambda_{\rm ex} = 420$ nm. The inserts show the fluorescence intensity at $\lambda_{\rm max}$ ($I_{\rm max}$) versus equivalents of Ca²⁺.

Fig. 5. ¹H NMR spectra (300.13 MHz) of compound **1b** in the presence (b) and absence (a) of Ca(ClO₄)₂ in CD₃CN solution at 21 °C. a) $C_{\rm L} = 6.8 \cdot 10^{-3}$ M; b) $C_{\rm L} = 1.0 \cdot 10^{-2}$ M, L : M = 1 : 3. The proton signals of the benzene ring and crown ether moiety are marked with red color.

Fig. 6. The changes in absorption spectra of compound **3a** in the presence of graduated amount of Ca(ClO₄)₂ in MeCN. $C_{\rm L} = 2.0 \cdot 10^{-5}$ M. The insert shows absorption at 420 nm ($A_{\rm max}$) versus equivalents of Ca²⁺.

Fig. 7. The changes in emission spectra of compound 3a in the presence of graduated amount of Ca(ClO₄)₂ in MeCN. $C_{\rm L} = 5.0 \cdot 10^{-6}$ M, $\lambda_{\rm ex} = 355$ nm. The insert shows fluorescence intensity at $\lambda_{\rm max}$ ($I_{\rm max}$) versus equivalents of Ca²⁺.

Fig. 8. ¹H NMR spectra (300.13 MHz) of compound **3a** in the presence (b) and absence (a) of Ca(ClO₄)₂ in CD₃CN solution at 21 °C. a) $C_{\rm L} = 1.0 \cdot 10^{-2}$ M; b) $C_{\rm L} = 1.0 \cdot 10^{-2}$ M, L : M = 1 : 5. The proton signals of the naphthalimide moiety and crown ether fragment are marked with red color.

Scheme 1. Chemical structure of the compounds 1a-c, 2a-c and 3a

Scheme 2. Synthesis of the compounds 1b, 2b and 3a

Tables

		$\varepsilon_{\lambda} \cdot 10^{-3}$ /			<u> </u>
	λ_{\max}^{abs} /	$L \cdot mol^{-1} \cdot cm^{-1}$	$\lambda_{\max}^{\text{fl}}(\lambda_{ex}) / \text{nm}$	$\pmb{arphi}^{ ext{fl}}$	lg K ^a
1a	417	12.9	518 (420)	0.55	
1b	415	14.8	518 (420)	0.0024	-
$(\mathbf{1b}) \cdot \mathbf{Ca}^{2+}$	420	14.4	520 (420)	0.43	4.94 ± 0.02
$(\mathbf{1b}) \cdot \mathbf{H}^+$	420	13.1	520 (420)	0.53	>7
2a	367	17.3	454 (365)	0.90	
2b	366	15.9	454 (365)	0.0017	+
$(\mathbf{2b}) \cdot \mathbf{Ca}^{2+}$	369	15.4	456 (365)	0.067	4.85 ± 0.02
$(2b) \cdot H^+$	369	15.3	457 (365)	0.89	>7
3 a	420	12.1	540 (420)	0.027	
$(3a) \cdot Ca^{2+}$	352	11.5	513 (340)	0.26	2.38 ± 0.03

Table 1. Optical properties and stability constants of 1a, 1b, 2a, 2b, 3a and their complexes in acetonitrile at 20 °C.

^aThe dimension of K is M^{-1}

Table 2. Energies (eV) of HOMO(-1), HOMO and LUMO of compounds **1–2a,b** and their complexes^a.

	1a	1b	$(\mathbf{1b}) \cdot \mathbf{Ca}^{2+}$	$(\mathbf{1b}) \cdot \mathbf{H}^+$	2a	2b	$(\mathbf{2b}) \cdot \mathbf{Ca}^{2+}$	$(\mathbf{1b}) \cdot \mathbf{H}^+$
LUMO	-5.34	-5.32	-5.84	-5.51	-6.05	-5.75	-6.17	-5.82
НОМО	-8.85	-7.97	-9.04	-9.00	-9.34	-8.02	-9.50	-9.46
HOMO(-1)	-9.86	-8.85	-9.73	-10.01	-9.96	-9.32	-9.74	-10.14

^a*N*-Aryl group local orbitals are marked with gray color.

Table 3. The changes in ¹H NMR spectra of compounds **1b**, **2b** induced by the presence of $Ca(ClO_4)_2$ in CD₃CN solution.

Hydrogen designation		H(10,14)	H(11,13)	H(15,24)	H(16,23)	H(17–22)
$\Delta\delta$ / ppm ^a	Compound 1b	0.41	0.57	-0.14	0.19	0.31
	Compound 2b	0.39	0.57	-0.13	0.20	0.32

 $^{a}\Delta\delta = \delta_{\rm LM} - \delta_{\rm L}$

Equations

$$\varphi_{i}^{fl} = \varphi_{0}^{fl} \frac{S_{i}(1 - 10^{-D_{0}})n_{i}^{2}}{S_{0}(1 - 10^{-D_{i}})n_{0}^{2}}$$

Equation 1.

 $L + M \implies LM$ Equation 2.

 $2L + M \iff L_2M$ Equation 3.

ACCEPTED MANUSCRIPT Electronic Supplementary Information

Comparative analysis of the PET and ICT sensor properties of 1,8-naphthalimides containing aza-15-crown-5 ether moiety

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4.	2D NMR spectra of the compounds 1b, 2b, 3a, 5 and 6.	S5

ACCEPTED MANUSCRIPT 1. Ground state geometry of the compound **3a**



Fig. S1. Optimized ground state geometry of the compound **3a** (Method PM6). C(3)–C(4)–N–C(15) Dihedral angle value is 21.5 °. The numbering of carbon atoms in the naphthalimide moiety, *N*-phenyl ring and crown ether fragment in **3a** is presented in Figure 8.

2. Frontier orbitals for the compounds 1b, 2b and their complexes



Fig. S2. Frontier orbitals (HOMO(-1), HOMO, LUMO) for compounds **1b**, **2b** and their complexes with, Ca^{2+} , H⁺ cations.

ACCEPTED MANUSCRIPT 3. Spectrofluorometric titration data for the compound **2b**



Fig. S3. The changes in the fluorescence spectra of **2b** in the presence of graduated amount of Ca(ClO₄)₂ in MeCM solution. $C_{\rm L} = 5,0 \cdot 10^{-6}$ M, $\lambda_{\rm ex} = 365$ nm. The inserts show the fluorescence intensity at $\lambda_{\rm max}^{\rm fl}$ ($I_{\rm max}$) versus equivalents of Ca²⁺.



Fig. S5. Aromatic part of HSQC spectrum of compound 1b in DMSO- d_6



Fig. S7. Aromatic part of HMBC spectrum of compound 1b in DMSO-d₆



Fig. S9. Aromatic part of HSQC spectrum of compound 2b in DMSO-d₆



Fig. S11. Aromatic part of HMBC spectrum of compound 2b in DMSO-d₆



Fig. S13. Aromatic part of HSQC spectrum of compound 3a in DMSO-d₆



Fig. S15. Aromatic part of HMBC spectrum of compound 3a in DMSO- d_6



Fig. S17. HSQC spectrum of compound 5 in DMSO- d_6



Fig. S19. HMBC spectrum of compound 5 in DMSO- d_6



Fig. S21. ¹H COSY spectrum of compound 5 in DMSO- d_6



Fig. S23. Aromatic part of HSQC spectrum of compound 6 in DMSO-d₆



Fig. S25. Aromatic part of HMBC spectrum of compound 6 in DMSO-d₆

Novel 1,8-naphthalimide based chemosensors containing an aza-15-crown-5 ether receptor unit at the imide position and at C-4 have been designed and prepared. The sensors have been shown to bind Ca^{2+} ions in acetonitrile solution with different affinities. The cation induced changes in absorption and fluorescence spectra are strongly dependent on the position of crown ether group in the molecule. Results have been rationalised in terms of PET and ICT mechanisms.