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Meloddy H. Manyeruke, Thendamudzimu Tshiwawa, Heinrich C.Hoppe, Michelle Isaacs, Ronnett Seldon, Digby F. Warner, Rui W.M. Krause, Perry T. Kaye

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Synthesis and Biological Evaluation of Bis-N²,N²'-(4-hydroxycoumarin-3-yl)ethylidene]-2,3dihydroxysuccinodihydrazides

Meloddy H. Manyeruke^a, Thendamudzimu Tshiwawa^a, Heinrich C.Hoppe^{b,c}, Michelle Isaacs^c, Ronnett Seldon^d, Digby F. Warner,^eRui W.M. Krause^{a,c*} and Perry T. Kaye^{a,c*}

 ^aDepartment of Chemistry, ^b Department of Biochemistry and Microbiology and ^cCentre for Chemico- and Biomedicinal Research, Rhodes University, Grahamstown, 6140, South Africa.
^eDrug Discovery and Development Centre (H3-D), Department of Chemistry, University of Cape Town, Rondebosch 7701, South Africa
^fMolecular Mycobacteriology Research Unit, Department of Pathology and Institute of Infectious Disease and Molecular Medicine, University of Cape Town, Cape Town, South Africa.

Abstract. A series of N^2 , N^2 '-bis[4-hydroxycoumarin-3-yl)ethylidene]-2,3-dihydroxysuccinohydrazides, containing 4-hydroxycoumarin, hydrazine and tartaric acid moieties, have been prepared and examined for possible biological activity. Several of these compounds exhibit promising HIV-1 integrase inhibition (IC₅₀=3.5µM), and anti-*T.brucei* (32% viability) and anti-mycobacterial (Visual MIC90 = 15.63 µM)

Keywords. 4-Hydroxycoumarins, succinodihydrazides, synthesis, HIV-1 integrase, anti-trypanosomal, anti-mycobacterial.



Corresponding authors: Prof Perry Kaye. E-mail: <u>P.Kaye@ru.ac.za</u> and Prof Rui Krause. E-mail: <u>R.Krause@ru.ac.za</u>

Numerous coumarin derivatives have been found to be present as secondary metabolites in bacteria, fungi and plants, thus prompting research into their isolation and the study of their biological activities.¹ Such derivatives exhibit wide varieties of medicinal properties, including anti-parasitic, anti-HIV, anti-bacterial, as well as serine protease, cholinesterase and lipoxygenase monoamine oxidase inhibitory activities.²⁻⁴ The antibiotics, novobiocin (isolated from *Streptomyces niveus* and *Streptomycesspheroides*) and chartreusin (isolated from *Streptomyces chartreusis*) are examples of coumarin derivatives isolated from bacteria,⁵ while aflatoxins are highly toxic, coumarin-containing, fungal metabolites found in *Aspergillus* species.⁶

Various biologically active coumarin derivatives have also been synthesised,⁷ including: coumarin-2-carboxamides,⁸ which exhibit ant-cancer activity; the 4-hydroxycoumarin derivative **1**, which exhibits HIV-1 protease (PR) inhibition activity;⁹ and coumarin-containing hydrazine derivatives, such as compound **2**, which exhibit minimum inhibitory concentrations (MIC) of 15-17 μ M against *M.tuberculosis*.¹⁰ In their highly cited paper, Zhao *et al.*,¹¹ discuss coumarin-based HIV Integrase (IN) inhibitors. More recently, 4-hydroxycoumarin dimers¹² and coumarin-3-carbohydrazide derivatives¹³ have been reported to exhibit HIV-1 IN inhibitory activities with IC₅₀ values in the low micromolar and nanomolar ranges, respectively. In earlier studies, we have explored the synthesis and biological activity of various coumarin-AZT conjugates **3** as potential dual-action HIV-1 PR and reverse transcriptase (RT) inhibitors.¹⁷ We now report the synthesis and bioactivity of a series of 4-hydroxy-coumarin derivatives generated by the conjunction of 4-hydroxycoumarin, hydrazine and tartaric acid moieties.



Figure 1. Structures of synthetic, biologically active coumarin derivatives.

The wide spectrum of applications of coumarin derivatives has led to considerable interest in their synthesis and numerous methods have been developed. These include the classic Perkin, Knoevenagel and Pechmann syntheses as well applications of Morita-Baylis-Hillman (MBH) methodology developed by our group.¹⁸ However, our attempts to use MBH methodology to access the 4-hydroxycoumarins derivatives required in this study proved unsatisfactory and, consequently, the 4-hydroxycoumarins **5a-g** (Scheme 1) were prepared using the method reported by Zhao *et al.*¹⁹ but with minor modifications, *viz.*, i) the use of dimethyl carbonate, in some cases, instead of diethyl carbonate; and ii) omitting recrystallisation from ethanol when NMR analysis indicated formation of the products in satisfactory purity. The 2-hydroxyacetophenones **4a-g** were reacted with either diethyl or dimethyl carbonate in the presence of sodium hydride to afford the corresponding 4-hydroxycoumarins **5a-g** in yields of 60-87%.



Scheme 1. Synthesis of bis-(4-hydroxycoumarinyl)succinohydrazides.

Acetylation of the 4-hydroxycoumarins **5a-g** was effected following the method described by Sukdolak *et al.*²⁰ It was observed, however, that the yields improved with longer reaction times (at least 1 hour rather than the reported 30 minutes); moreover, the purity of the compounds was satisfactory after washing the precipitate with methanol thereby permitting omission of a crystallisation step. Thus, the 4-hydroxycoumarins **5a-g** were each reacted with POCl₃ in glacial acetic acid under reflux for at least 1 hour, after which the resulting precipitates were filtered, washed with methanol and dried to give the respective products (**6**) in yields ranging from 41% to 90%.

The 2,3-dihydroxysuccinodihydrazide 7, required for the final step in the preparation of the title compounds **8a-g**, was obtained *via* acid (HCl or AcOH)-catalysed reaction of *racemic* diethyl tartrate with hydrazine hydrate in refluxing ethanol.²¹ The resulting precipitate was recrystallised from methanol to give the desired dihydrazide 7 as a white solid, which was then reacted with the 3-acetylated 4-hydroxycoumarins **6a-g** in ethanol under reflux for at least 2 hours (Scheme 1).²² The precipitates formed in each reaction were filtered, washed with methanol and dried to give the required bis[4-hydroxycoumarin-3-yl)ethylidene]-2,3-dihydroxysuccinohydrazides **8a-g** in high purity and in yields of 41% to 58%.

The targeted products **8a-g** contain: i) the 4-hydroxycoumarin moiety present in the HIV-1 PR inhibitor **1**; ii) the hydroxyethylene peptide isosteres and overall C_2 symmetry characteristic of clinical HIV-1 PR inhibitors; and iii) the coumarin and hydrazine moieties present in the anti-mycobacterial derivative **2** and the recently reported coumarin-3-carbohydrazide HIV-1 IN inhibitors.¹³ Compounds **8a-g** were thus screened for HIV-1 IN and PR inhibition potential, anti-tuberculosis (*M. tuberculosis*) activity and cytotoxicity (against HeLa cells). The opportunity was also taken to screen the compounds for anti-*Plasmodium falciparum* (*P.falciparum*) and anti-trypanosome (*T.b. brucei*) activity. The results of the bioassays are summarised in Table 1.

With the exception of the brominated derivative **8d**, all of the title compounds (**8**) showed some inhibition of HIV-1 IN – particularly, the parent system **8a** with an IC₅₀ value of 3.5 μ M. It is interesting to note that sequential introduction of a halogen substituent (R¹ = F, Cl, Br) into the unsubstituted aromatic rings in **8a** (R¹ = H) resulted in graded decreases in the HIV-1 IN inhibition (**8a**: 59.4% > **8b**: 35.4% > **8c**: 27.6% > **8d**: 0%) which appear to follow their increasing bulk *inversely* and their electronegativity *directly*. The apparent advantage conferred by the absence of additional substituents in the benzene ring is reflected in the HIV-1 inhibition activity of other 4-hydroxycoumarin derivatives.¹¹ Only one of the compounds (**8e**; 8.3% inhibition) proved to be active, albeit very weakly, against HIV-1 PR at a concentration of 20 μ M.

Table 1. Bioassay data for compounds **8a-g**, showing % inhibition of HIV-1 IN and PR, % viability of *P. falciparum*, *T.b. brucei* and HeLa cells at 20 μ M and MIC₉₀ values against *M. tuberculosis*.



Compd.	R ¹	R ²	% Yield	% HIV-1 IN inhibition ^a	% HIV-1 PR inhibition ^b	% P. <i>falciparum</i> viability ^c	% <i>T.b.brucei</i> viability ^d	% HeLa cell viability ^e	Visual MIC90 (µM) ^{f g,}	Calc. MIC90 (µM) ^{f,b}
8a	Н	Н	58	59.4 ⁱ	0.0	90.7	42.7	78.4	>125	>125
8b	F	Н	58	35.4	0.0	92.8	32.8	84.6	>125	>125
8c	Cl	Н	45	27.6	0.0	93.8	44.6	100.0	31.25	54.25
8d	Br	Н	42	0.0	0.0	100.0	100.0	100.0	15.63	18.91
8e	Me	Н	41	26.4	8.3	100.0	100.0	85.3	>125	>125
8 f	MeO	Н	43	34.6	0.0	100.0	100.0	84.6	>125	>125
8g	5,6-Be	nzo	45	35.7	0.0	100.0	100.0	98.1	31.25	29.46
Control				100.0 ^a	99.8 ^b	_ c	d	e	0.019 ^f	0.007^{f}

Controls: ^achicoric acid; ^britonavir; ^cchloroquine: $IC_{50} = 0.012 \ \mu\text{M}$; ^dpentamidine: $IC_{50} = 0.039 \ \mu\text{M}$; ^eemetine: $IC_{50} = 0.013 \ \mu\text{M}$ and ^frifampicin. ^gVisual MIC90 7D 7H9 GLU CAS Tx . ^hCalculated MIC90 7D 7H9 GLU CAS Tx (μ M). ⁱ $IC_{50} = 3.5 \ \mu\text{M}$.

The products 8a-g were also evaluated for anti-parasitic activity at a concentration of 20 μ M against both *P.falciparum* and *T.b.brucei* (which is responsible for sleeping sickness). The products showed little, if any, significant activity in the *P.falciparum* screen; compounds **8a-c**, however, clearly exhibited anti-trypanosomal activity, decreasing residual parasite *T.b. brucei* viabilities appreciably (**8a**: 43%; **8b**: 33%; and **8c**: 45%). When tested at a concentration of 20 μ M against HeLa cells, none of the compounds **8a-g** exhibited significant cytotoxicity. As indicated earlier, coumarin-containing hydrazine derivatives (*e.g.*, compound **2**, Figure 1) have been reported by Arshad *et al.*¹³ to exhibit minimum inhibitory concentrations (MIC) in the 15-17 μ M range against *M.tuberculosis*, and three of our compounds (**8c**, **8d** and **8g**) have also shown promising anti-mycobacterial potential with relatively low visual MIC₉₀ (31.25, 15.63 and 31.25 μ M, respectively) and calculated MIC₉₀ values (54.25, 18.91 and 29.46 μ M, respectively).

In silico docking studies were undertaken to assess the binding affinity of compounds **8a-g** to selected HIV-1 IN and PR, P. falciparum, T. Brucei and M.tuberculosis enzyme receptors, the detailed results of which are summarised in the Supporting Information file. Clear correlations between the *in silico* binding affinities and the *in vitro* data generally proved to be limited, but several instances are noteworthy. Compound 8a, which exhibited the highest in vitro HIV-1 IN inhibition (Table 1) exhibited the best in silico binding affinity (-4.628 kcal/mol) in the noncatalytic active-site of HIV-1 IN crystal structure PDB 4E1M (illustrated in Figure 2a) and the second best binding affinity (-6.215 kcal/mol) in the HIV-1 IN crystal structure PDB 5FRN. Interestingly, compound 8g, which exhibited the second-highest in vitro HIV-1 IN inhibition, exhibited the best binding affinity (-6.794 kcal/mol) for 5FRN, but the lowest binding affinity (-0.163 kcal/mol) for 4E1M; the binding of this ligand in the active site of 5FRN is illustrated in Figure 2b. The strong in silico HIV-1 PR 1YT9 binding affinities for all of the ligands 8a-g (between -6.744 and -9.420 kcal/mol, compared with -7.309 kcal/mol for the control, ritonavir) are completely at odds with the in vitro data in Table 1! The corresponding in silico HIV-1 PR IZP8 binding affinities for the ligands **8a-g** are all significantly lower than that of ritonavir but are at least consistent with their general, in vitro inactivity.



Figure 2. Potential hydrogen-bonding interactions between amino acid residues in: a) the non-catalytic active-site of HIV-1 IN (PDB 4E1M) and compound **8a**; and b) the active-site of HIV-1 IN (PDB 5FRN) and compound **8g**.

Although compounds **8a-g** all exhibited very encouraging *in silico* binding affinities (relative to chloroquine) with three different *P.falciparum* enzyme structures, none of them exhibited

significant *in vitro* anti-parasitic activity. The three compounds with the strongest *in vitro* antitrypanosomal activity (**8a-c**) exhibit the weakest *in silico* binding affinities with the *T.b.brucei* enzyme structure! It is similarly difficult to draw any meaningful correlations between the *in vitro* anti-microbacterial and the *in silico* enzyme-binding affinities of the title compounds.

In summary, it is apparent that the title compounds are readily accessible, exhibit insignificant cytotoxicity and some of them show promising HIV-1 IN inhibition (IC₅₀=3.5 \square M), anti-*T.brucei* activity (32% viability at 20 mM) and anti-mycobacterial potential (Visual MIC90 = 15.63 μ M). Experimental data for the synthesised compounds, NMR spectra for all new compounds, the *in silico* docking protocols and the resulting binding affinities are provided in the Supporting Information file. Bioassay protocols have been reported previously.^{23,24}

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