



The Use of the Selective Imidazoline I₁ Receptor Agonist Carbophenylene as a Strategy for Neuropathic Pain Relief: Preclinical Evaluation in a Mouse Model of Oxaliplatin-Induced Neurotoxicity

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Abstract

Anti-cancer therapy based on the repeated administration of oxaliplatin is limited by the development of a disabling neuropathic syndrome with detrimental effects on the patient's quality of life. The lack of effective pharmacological approaches calls for the identification of innovative therapeutic strategies based on new targets. We focused our attention on the imidazoline I₁ receptor (I₁-R) and in particular on the selective I₁-R agonist 2-(1-([1,1'-biphenyl]-2-yl)propan-2-yl)-4,5-dihydro-1H-imidazole (carbophenylene). The purpose of this work was the preclinical evaluation of the efficacy of carbophenylene on oxaliplatin-induced neuropathic pain in mice. Carbophenylene, acutely per os administered (0.1–10 mg kg⁻¹), induced a dose-dependent anti-hyperalgesic effect that was completely blocked by the pre-treatment with the I₁-R antagonist **3** or the I₁/α₂ receptor antagonist efaroxan, confirming the I₁-R-dependent mechanism. Conversely, pre-treatment with the I₂-R antagonist BU224 did not block the anti-nociceptive effect evoked by carbophenylene. Repeated oral administrations of carbophenylene (1 mg kg⁻¹) for 14 days, starting from the first day of oxaliplatin injection, counteracted the development of neuropathic pain in all behavioral tests (cold plate, Von Frey, and paw pressure tests) carried out 24 h after the last carbophenylene treatment on days 7 and 14. In the dorsal horn of the spinal cord, carbophenylene significantly decreased the oxaliplatin-induced astrocyte activation detected by immunofluorescence staining by the specific labelling with GFAP antibody. In conclusion, carbophenylene showed anti-neuropathic properties both after acute and chronic treatment with preventive effect against oxaliplatin-induced astrocyte activation in the spinal cord. Therefore, I₁-R agonists emerge as a new class of candidates for the management of oxaliplatin-induced neuropathic pain.

Key Words Imidazoline I₁ receptor agonist · oxaliplatin · chemotherapy-induced neuropathic pain · astrocytes · carbophenylene

Dedicated to Prof. Maria Pigini

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Introduction

Neuropathic pain is now defined by the International Association for the Study of Pain (IASP) as “pain caused by a lesion or disease of the somatosensory nervous system” [1]. Among different factors, nervous lesions can be induced by neurotoxic drugs like several anticancer agents able to evoke painful syndromes defined chemotherapy-induced neuropathies. Oxaliplatin, one of the lead treatments for colorectal cancers [2, 3], is associated with the development of a severe persistent neuropathic pain which remains the dose-limiting toxicity of this treatment [4]. Chronologically, oxaliplatin-induced acute and transient hyperexcitability followed by a chronic cumulative neuropathy experienced by almost the 50% of patients [5]. Signs and symptoms of chronic neuropathy include pain, paresthesia, hypoesthesia, and changes in

proprioception that persist during cycles [6]. In the long term, this syndrome has a deleterious impact on cancer survivors, often being associated with sleep disturbance, depressive symptoms, and impaired health-related quality of life [7]. Several molecular mechanisms participate to oxaliplatin neurotoxicity involving early targets in the peripheral nervous system as well as direct and indirect alterations of the neuronal and glial compartments of the central nervous system [8–13].

No treatments are currently available to reduce oxaliplatin-dependent neuropathic pain, further pain-relieving drugs used against neuropathic pain are not satisfactory due to limited efficacy and side effects [14, 15]. For these reasons, efforts are necessary to explore novel molecular targets for the pharmacological management of neuropathic pain.

Recent studies have highlighted that the imidazoline I_2 receptor (I_2 -R) activation by selective ligands such as 2-BFI [16] or phenzoline (1) (Fig. 1) [17], alone or in combination with opioids, triggers a significant anti-hyperalgesic effect. Therefore, this receptor system has been suggested as a promising target for the management of chronic pain [18–20]. The I_2 -R is a member of a wider family of imidazoline receptors, also including I_1 -R and I_3 -R, which play important roles in central cardiovascular regulation and in insulin secretion, respectively [21, 22]. In particular, I_1 -R is involved in the hypotensive activity of clonidine and related compounds supporting the idea that the I_1 -Rs are upstream from the α_2 -adrenergic receptors and work in tandem for its effect on blood pressure [23]. Additionally, I_1 -R activation may suppress sympathetic outflow at postganglionic sympathetic neurons through modulation of several signal transduction systems including N-type calcium channels, G protein inwardly rectifying potassium channel, adenosine receptors, phosphatidyl-choline-specific phospholipase C, and nicotinic receptors [24–29]. On this basis, a possible role of I_1 -R in pain

modulation can be theoretically suggested although so far unexplored. In the present work we studied the effects of I_1 -R modulation against oxaliplatin-induced neuropathic pain by using the selective I_1 -R agonist compound 2 (in this paper named carbophenylene, highly selective over I_2 , α_2 , and 5-HT $_{1A}$ receptors [30, 31]) (Fig. 1). Its effects on pain threshold were evaluated after single or repeated administrations in the absence or presence of the I_1 -R antagonist 3 [2-(1-methyl-2-phenylethyl)-4,5-dihydro-1H-imidazole] (Fig. 1) [32], highly selective over I_2 , α_2 and 5-HT $_{1A}$ receptor and the well-known I_1/α_2 receptor antagonist efaroan [33] which proved to be selective over several other biological targets [33; <https://pubchem.ncbi.nlm.nih.gov/compound/Efaroan#section=BioAssay-Results>]. Moreover, the I_2 antagonist BU224 was also used to exclude the involvement of I_2 -R [34]. Finally, to analyze the impact of I_1 -R stimulation on the maladaptive plasticity of the neuropathic central nervous system, the effect of carbophenylene on oxaliplatin-induced astrocyte activation in the dorsal horn of the spinal cord was assessed.

Materials and Methods

Drugs

The I_1 -R agonist 2-(1-([1,1'-biphenyl]-2-yl)propan-2-yl)-4,5-dihydro-1H-imidazole (2, carbophenylene) and the I_1 -R antagonist 2-(1-phenylpropan-2-yl)-4,5-dihydro-1H-imidazole (3) were obtained treating methyl 3-([1,1'-biphenyl]-2-yl)-2-methylpropanoate and methyl 2-methyl-3-phenylpropanoate, respectively, with ethylenediamine and trimethyl aluminum at 110 °C in toluene [30, 32]. Efaroxan hydrochloride and BU224 hydrochloride were purchased by Tocris. Oxaliplatin

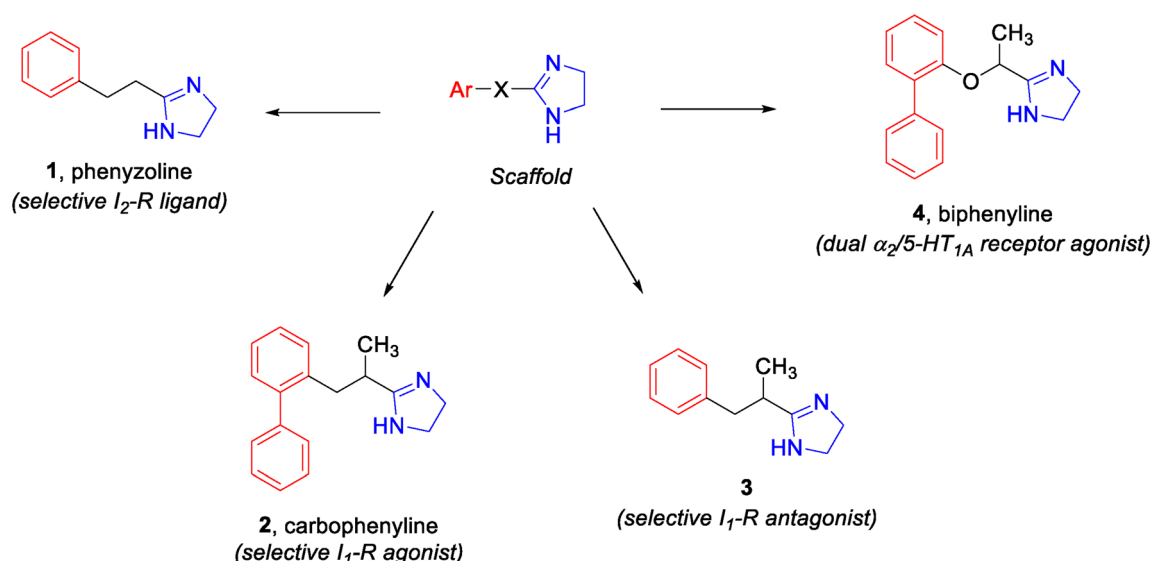


Fig. 1 Chemical structures of compounds 1–4, sharing a common bioversatile scaffold

and pregabalin were purchased by Carbosynth (Compton, UK).

Cell Cultures

The HT-29 human colon cancer cell line were acquired from the American Type Culture Collection. HT-29 were cultured in Complete Medium (CM) composed by high-glucose DMEM with 10% FBS, 2 mM L-glutamine, 100 IU/mL penicillin, and 100 µg mL⁻¹ streptomycin (Sigma-Aldrich, Italy).

Cell Viability Assay

To plate HT-29 cells (1×10^4 cells/well), 96-well cell culture plates (Corning) were used. After 48 h, the experiments were performed. Cells were incubated in serum-free DMEM with 0.3–100 µM oxaliplatin (Carbosynth, Compton, Berkshire, UK) in the absence or presence of carbophenylamine 10 µM for 48 h. Cell viability was evaluated by the reduction of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT; Sigma-Aldrich, Italy) as an index of mitochondrial compartment functionality. After 48 h incubation, 1 mg mL⁻¹ MTT in serum-free DMEM without phenol red was added into each well and incubated for 30 min at 37 °C. After washing the formazan crystals were dissolved in 150 µL dimethyl sulfoxide. The absorbance was measured at 550 nm. Experiments were performed in triplicate.

Animals

CD-1 mice (Envigo, Varese, Italy) weighing 20–25 g at the beginning of the experimental procedure were used. Animals were housed in the Centro Stabulazione Animali da Laboratorio (University of Florence) and used at least 1 week after their arrival. Ten mice were housed per cage (size 26 cm × 41 cm); animals were fed a standard laboratory diet and tap water ad libitum and kept at 23 ± 1 °C with a 12 h light/dark cycle (light at 7 A.M.). All animal manipulations were carried out according to the Directive 2010/63/EU of the European parliament and of the European Union council (22 September 2010) on the protection of animals used for scientific purposes. The ethical policy of the University of Florence complies with the Guide for the Care and Use of Laboratory Animals of the US National Institutes of Health (NIH Publication No. 85–23, revised 1996; University of Florence assurance number: A5278–01). Formal approval to conduct the experiments described was obtained from the Italian Ministry of Health (No. 54/2014-B) and from the Animal Subjects Review Board of the University of Florence. Experiments involving animals have been reported according to ARRIVE guidelines [35]. All efforts were made to minimize animal suffering and to reduce the number of animals used.

Oxaliplatin-Induced Neuropathic Pain

2.4 mg kg⁻¹ oxaliplatin was dissolved in 5% glucose solution and administered intraperitoneally (i.p.) for 5 consecutive days every week for 2 weeks (10 i.p. injections) [36 with minor modification, 37]. Control animals received an equivalent volume of 5% glucose i.p. (vehicle).

Compound Administration and Study of the Pharmacodynamic Mechanisms

When neuropathic pain was fully established (starting from day 14), carbophenylamine (0.1–10 mg kg⁻¹) was suspended in 1% solution of carboxymethylcellulose sodium salt (CMC) and per os (p.o.) acutely administered ranging from 0.1 mg kg⁻¹ to 10 mg kg⁻¹ to evaluate its symptomatic efficacy. Pregabalin (30 mg kg⁻¹, p.o.) was used as reference drug [38]. Afterwards, to highlight a preventive effect, repeated per os administrations of 1 mg kg⁻¹ of carbophenylamine were carried out daily from the beginning of oxaliplatin administration (day 1) to the end of the experiment (day 14). Control animals were treated with vehicles (5% glucose and CMC 1%).

For the study of the pharmacodynamic mechanisms, the selective I₁-R antagonist **3**, I₁/α₂ receptor antagonist efaroxan and selective I₂-R antagonist BU224 were used. All antagonists were suspended in CMC 1% and p.o. administered 15 min before carbophenylamine. For **3** and BU224 antagonists we used the first dose non hyperalgesic as shown in Supplementary table 1. Control animal were treated with vehicles (5% glucose and CMC 1%).

Paw Pressure Test

Mechanical hyperalgesia was determined by measuring the latency in seconds to withdraw the paw away from a constant mechanical pressure exerted onto the dorsal surface [39]. A 15 g calibrated glass cylindrical rod (diameter = 10 mm) chamfered to a conical point (diameter = 3 mm) was used to exert the mechanical force. The weight was suspended vertically between two rings attached to a stand and was free to move vertically. A single measure was made per animal. A cutoff time of 40 s was used.

Cold Plate

Thermal allodynia was assessed using the cold plate test. With minimal animal–handler interaction, mice were taken from home-cages, and placed onto the surface of the cold plate (Ugo Basile, Varese, Italy) maintained at a constant temperature of 4 °C ± 1 °C. Ambulation was restricted by a cylindrical Plexiglas chamber (diameter, 10 cm; height, 15 cm), with open top. A timer controlled by foot peddle began timing response latency from the moment the mouse was placed onto

the cold plate. Pain-related behavior (licking of the hind paw) was observed, and the time (seconds) of the first sign was recorded. The cutoff time of the latency of paw lifting or licking was set at 30 s [40].

Von Frey

The animals were placed in 20 × 20 cm Plexiglas boxes equipped with a metallic meshy floor, 20 cm above the bench. A habituation of 15 min was allowed before the test. An electronic Von Frey hair unit (Ugo Basile, Varese, Italy) was used: the withdrawal threshold was evaluated by applying force ranging from 0 to 5 g with 0.2 g accuracy. Punctuate stimulus was delivered to the mid-plantar area of each anterior paw from below the meshy floor through a plastic tip and the withdrawal threshold was automatically displayed on the screen. Paw sensitivity threshold was defined as the minimum pressure required to elicit a robust and immediate withdrawal reflex of the paw. Voluntary movements associated with locomotion were not taken as a withdrawal response. Stimuli were applied on each anterior paw with an interval of 5 s. The measure was repeated 5 times and the final value was obtained by averaging the 5 measures [41, 42].

Immunohistochemistry

On day 14, after behavioral measurements, mice were sacrificed and the lumbar spinal cord segments were removed, postfixed in 4% paraformaldehyde, and then cryoprotected in 30% sucrose solution at 4 °C. Slide-mounted cryostat sections (5 µm) were processed for indirect immunofluorescence histochemistry.

Formalin-fixed cryostat sections (5 µm) were incubated for 30 min in ready-to-go blocking solution (Bio-Optica, Milan, Italy) at room temperature to block unspecific binding. The primary antibodies, overnight incubated at 4 °C, were directed against Iba1 (rabbit, 1:200; Wako Chemicals, Richmond, USA) for microglial staining or against glial fibrillary acidic protein (GFAP; rabbit, 1:200; Dako, USA) for astrocyte staining. After rinsing in PBST, sections were incubated in donkey anti-rabbit IgG secondary antibody labelled with Alexa Fluor 488 (1:500, Invitrogen, Milan, Italy) at room temperature for 1 h. Nuclei were stained with DAPI (4',6-diamidin-2-fenilindolo; 1:2000; Invitrogen, Milan, Italy).

Negative control sections (no exposure to the primary antisera) were processed concurrently with the other sections for all immunohistochemical studies. We obtained a single optical density value for the dorsal horns by averaging the two sides in each mouse, and these values were compared to the homologous average values from the vehicle-treated animals.

Images were acquired by a motorized Leica DM6000B microscope equipped with a DFC350FX camera (Leica,

Mannheim, Germany). Microglia and astrocyte morphology were assessed by inspection of at least three fields (× 40 0.75NA objective) in the dorsal horn.

Quantitative analysis of GFAP and Iba1-positive cells was performed by collecting at least three independent fields through a × 20 0.5NA objective. Immunofluorescent staining was measured as mean fluorescence intensity using ImageJ software (ImageJ, National Institute of Health, USA, <https://imagej.nih.gov/>) by automatic thresholding algorithm. Furthermore, GFAP or Iba1 -positive cells were quantified by thresholding automatic count. Results, given as mean fluorescent intensity (arbitrary units) by the thresholded fluorescent signal, revealed a common trend between GFAP/Iba-1 expression and astrocyte/microglial cell number (data not shown). Five spinal cord sections were analyzed for each animal.

Statistical Analysis

Behavioral measurements were performed on ten mice for each treatment carried out in 2 different experimental sets. All assessments were made by researchers blinded to animal treatments. Results were expressed as mean ± (S.E.M.) with one-way analysis of variance. A Bonferroni's significant difference procedure was used as a post hoc comparison. *P* values < 0.05 or < 0.01 were considered significant. Data were analyzed using the Origin 9 software (OriginLab, Northampton, MA, USA).

Results

Behavioral Evaluations

The effects of carbophenyl line against neuropathic pain were studied in a mouse model of chemotherapy-induced neuropathic pain. The neurotoxicity of oxaliplatin was used to evoke a neuropathic syndrome characterized by thermal and mechanical hypersensitivity and allodynia after repeated administrations (2.4 mg kg⁻¹, 10 intraperitoneal injections) (Fig. 2). On day 14, the response to a cold non-noxious stimulus (allodynia-like symptom) was measured by the cold plate test. The pain threshold of oxaliplatin-treated animals decreased to 9.8 ± 0.2 s in comparison to 18.2 ± 0.4 s of the control group (vehicle + vehicle-treated animals). Increasing doses of carbophenyl line (0.1–10 mg kg⁻¹), acutely per os administered, reduced thermal allodynia in a dose-dependent manner. The single administration of compound at 10 mg kg⁻¹ was able to fully counteract oxaliplatin-induced neuropathic pain 30 min after injection with the onset 15 min after treatment that lasted up to 45 min. A dose of 1 mg kg⁻¹ increased the pain threshold of the animals starting 15 min after treatment with a peak at 45 min. The anti-

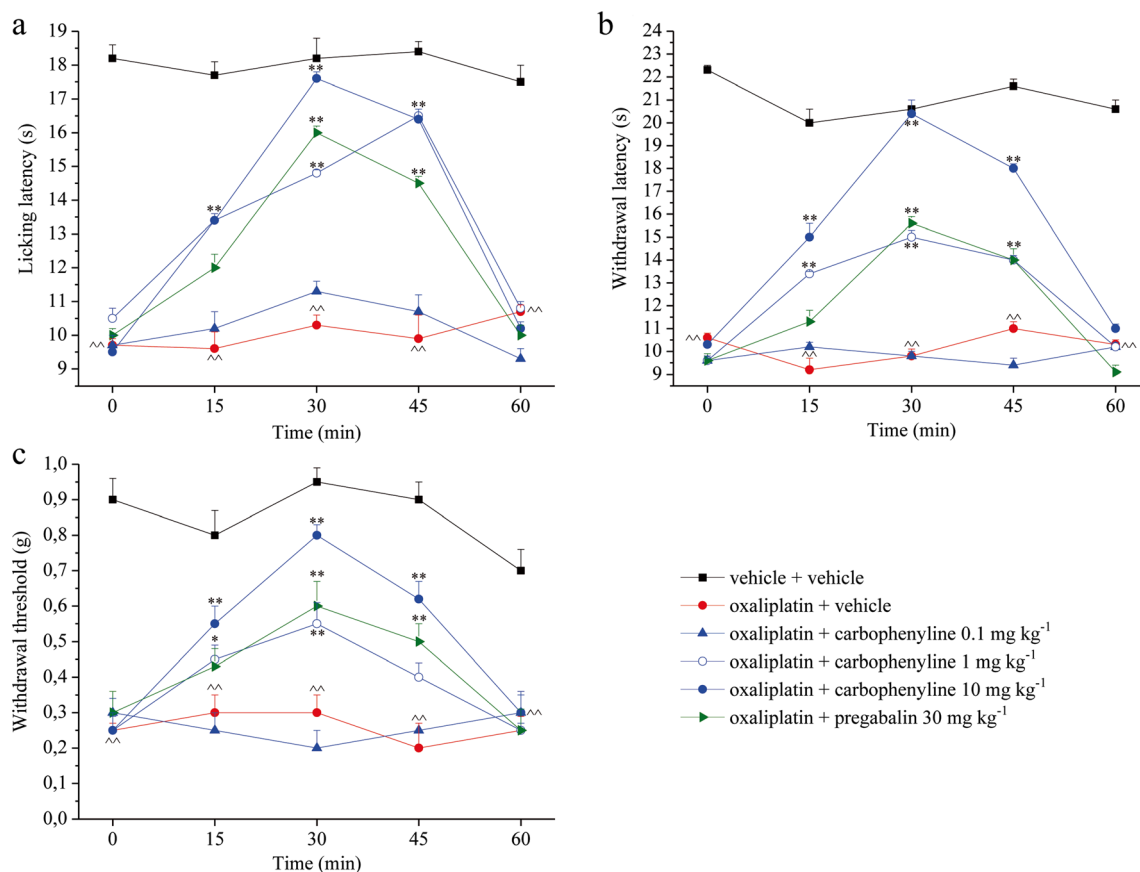


Fig. 2 Effect of acute carbophenyline administrations on pain behavior induced by oxaliplatin. Pain threshold to (a) a non-noxious thermal stimulus (cold plate test), (b) to a noxious mechanical stimulus (paw pressure test) and (c) to a non-noxious mechanical stimulus (Von Frey test) were measured. Oxaliplatin (2.4 mg kg^{-1} , i.p.) was administered for 2 weeks (10 injections). Starting from day 14, carbophenyline was acutely per os

administered ($0.1\text{--}10 \text{ mg kg}^{-1}$) and measurements assessed before and 15, 30, 45, 60 min after injection. Control animals were treated with vehicles. Each value represents the mean \pm S.E.M. of 10 animals per group performed in 2 experimental sets. Statistical analysis is one-way ANOVA followed by Bonferroni's post hoc comparison. $^{\wedge}P < 0.01$ vs vehicle + vehicle; $^{**}P < 0.01$ vs oxaliplatin + vehicle

hypersensitivity effect vanished at 60 min. The lowest dose (0.1 mg kg^{-1}) was inactive. Pregabalin was used as reference drug. The administration of the drug at the dose of 30 mg kg^{-1} significantly increased the pain threshold 30 min after treatment with the effect that remained active up 45 min without completely counteract oxaliplatin-induced thermal allodynia (Fig. 2a). The acute anti-hyperalgesic and anti-allodynic effects of carbophenyline were also evaluated by paw pressure test and Von Frey test, respectively (Fig. 2 b and c). Results obtained were comparable to that achieved by the cold plate test. Carbophenyline evoked a dose-dependent anti-neuropathic effect that reached the higher effect with the dose of 10 mg kg^{-1} , 30 min after the administration.

To confirm the pharmacodynamics of carbophenyline, three specific antagonists were used: **3** (selective I₁-R antagonist), efaroxan (I/ α_2 receptor antagonist), and BU224 (selective I₂-R antagonist). As shown in Fig. 3, the pre-treatment (15 min before carbophenyline administration) with **3** 10 (mg kg^{-1} , p.o.) was able to completely abolish the pain-relieving effect of carbophenyline till the end of the

experiment. The same result was obtained with the pre-treatment with efaroxan (10 mg kg^{-1} , p.o., 15 min before carbophenyline administration). BU224 (3 mg kg^{-1} p.o., 15 min before carbophenyline administration) was not able to modify the efficacy of carbophenyline over the time of observation (Fig. 3).

Repeated treatments of carbophenyline (1 mg kg^{-1} p.o.) were performed to evaluate the preventive effect of the compound in the same model of oxaliplatin-induced neuropathic pain. Oxaliplatin-treated animals were daily injected with carbophenyline starting from the same day of antitumor administration. The response to a thermal (cold plate test) and mechanical (paw pressure and Von Frey tests) stimuli was measured on days 7 and 14, 24 h after the last treatment (Fig. 4). Oxaliplatin-induced neuropathic pain increased during time reaching the plateau on week 2 (day 14) as shown by Fig. 4. Repeated daily administration of carbophenyline increased the pain threshold of oxaliplatin-treated animals at all time points without development of tolerance to the anti-hypersensitivity effect. The effect evoked by carbophenyline

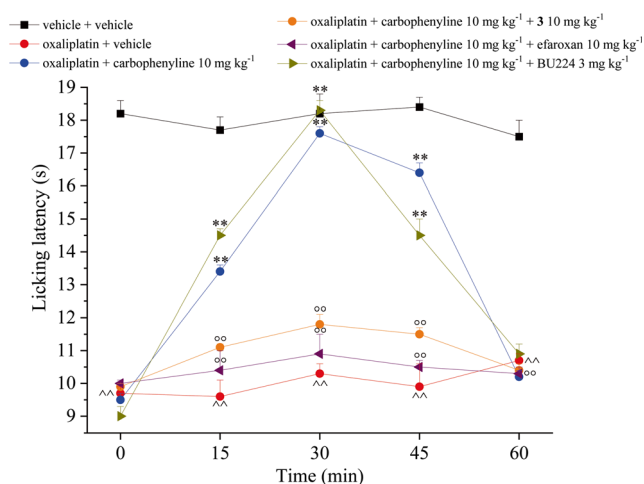


Fig. 3 Study of carbophenylamine pharmacodynamic profile. Pain was induced by repeated treatment with oxaliplatin. The hypersensitivity to a cold stimulus was measured by the cold plate test. Carbophenylamine was administered per os at 10 mg kg^{-1} . The I_1 receptor antagonist **3** (10 mg kg^{-1}), the I_1/α_2 receptor antagonist efaroan (10 mg kg^{-1}) and the I_2 receptor antagonist BU224 (3 mg kg^{-1}) were p.o. administered 15 min before carbophenylamine injection. Control animals were treated with vehicles. Each value represents the mean \pm S.E.M. of 10 animals per group performed in 2 experimental sets. Statistical analysis is one-way ANOVA followed by Bonferroni's post hoc comparison. $^{\wedge\wedge\wedge}P < 0.01$ vs vehicle + vehicle; $^{**}P < 0.01$ vs oxaliplatin + vehicle; $^{\circ\circ}P < 0.01$ vs oxaliplatin + carbophenylamine

remained stable on days 7 and 14 as highlighted by the cold plate and paw pressure tests (Fig. 4 a and b, respectively) counteracting oxaliplatin-induced neuropathic pain. Similar results were obtained measuring the response to a mechanical non-noxious stimulus (Von Frey test, Fig. 4c), carbophenylamine increased its efficacy during time completely abolishing mechanical allodynia on day 14.

Analysis of Glial Cells in the Spinal Cord

The central nervous system was analyzed to assess the capability of carbophenylamine to intervene against the maladaptive plasticity induced by oxaliplatin treatment. In the spinal cord, repeated oxaliplatin injections induced astrocyte activation, measured as increased GFAP fluorescence intensity (day 14) on the entire surface and, particularly, in the superficial laminae (Fig. 5b) with respect to the control animals (Fig. 5a). The repeated administration of carbophenylamine significantly prevented the astrocytic response to oxaliplatin as shown in Fig. 5c. As previously reported [9], spinal microglia was not altered by oxaliplatin on day 14 (Supplementary Figure S1); the co-treatment with carbophenylamine did not modify this condition (Fig. S1).

HT-29 Cell Line

Aimed to evaluate a possible interaction between the treatment with carbophenylamine and the therapeutic properties of oxaliplatin, we analyzed the vitality of the human colorectal cancer cell line HT-29. Table 1 shows oxaliplatin lethality after 48 h incubation ($0.3\text{--}100 \mu\text{M}$) in the absence or presence of carbophenylamine. Ten μM carbophenylamine did not significantly alter the oxaliplatin-dependent decrease of cell viability. Carbophenylamine *per se* did not influence HT-29 viability up to $100 \mu\text{M}$ as shown in Table 2.

Discussion

The present data show for the first time the involvement of the I_1 -R in the signaling of neuropathic pain, highlighting the properties of the selective I_1 -R agonist carbophenylamine in relieving pain alterations in a mouse model of oxaliplatin-induced neurotoxicity. Moreover, repeated administration of carbophenylamine prevented the maladaptive changes that occur to the astrocyte population in the central nervous system. A new class of potential neuropathic pain relievers has been outlined.

Many different pharmacological actions have been associated with imidazoline compounds [21] and our research on them has highlighted that the 2-substituted imidazoline ring linked to an aromatic moiety (Ar) by a biatomic bridge (X) is a bioversatile scaffold to obtain agents interacting with different targets [43–46] (Fig. 1). Some of these imidazoline ligands demonstrated to be efficacious as anti-neuropathic agents. In particular, in addition to the already mentioned I_2 ligand phenylzoline (**1**), characterized by a $-\text{CH}_2\text{CH}_2-$ bridge, the dual $\alpha_2/5\text{-HT}_{1A}$ receptor agonist *S*-(-)-biphenylamine (**4**) [47, 48], bearing an $-\text{OCH}(\text{CH}_3)-$ bridge and an ortho phenyl substituent in the aromatic ring, showed a significant pain-relieving effect in a rat model of neuropathic pain [49]. The isosteric substitution of the oxygen atom of the bridge with a CH_2 group produced derivative carbophenylamine, characterized by an interesting modulation of the biological profile. Indeed, carbophenylamine behaves as an I_1 -R agonist with high selectivity over I_2 , α_2 , and 5-HT_{1A} receptors [30, 31, 48]. The profitable activation of I_1 -R makes carbophenylamine a potent anti-hypertensive agent [30, 31].

Although imidazoline binding sites attracted attention in nociception, only the I_2 -R was studied. A possible role of I_1 -R can be hypothesized analyzing the efficacy of clonidine (a molecule possessing I_1 -R affinity in addition to the well-known adrenergic α_2 -mediated effects) against neuropathic and cancer pain [50–53], even if a possible I_1 -R component in clonidine anti-nociceptive effects was not described. Stone et al. [54] showed that there is a possible synergy between I_1 -R and opioid receptors at the spinal level based on the use of the

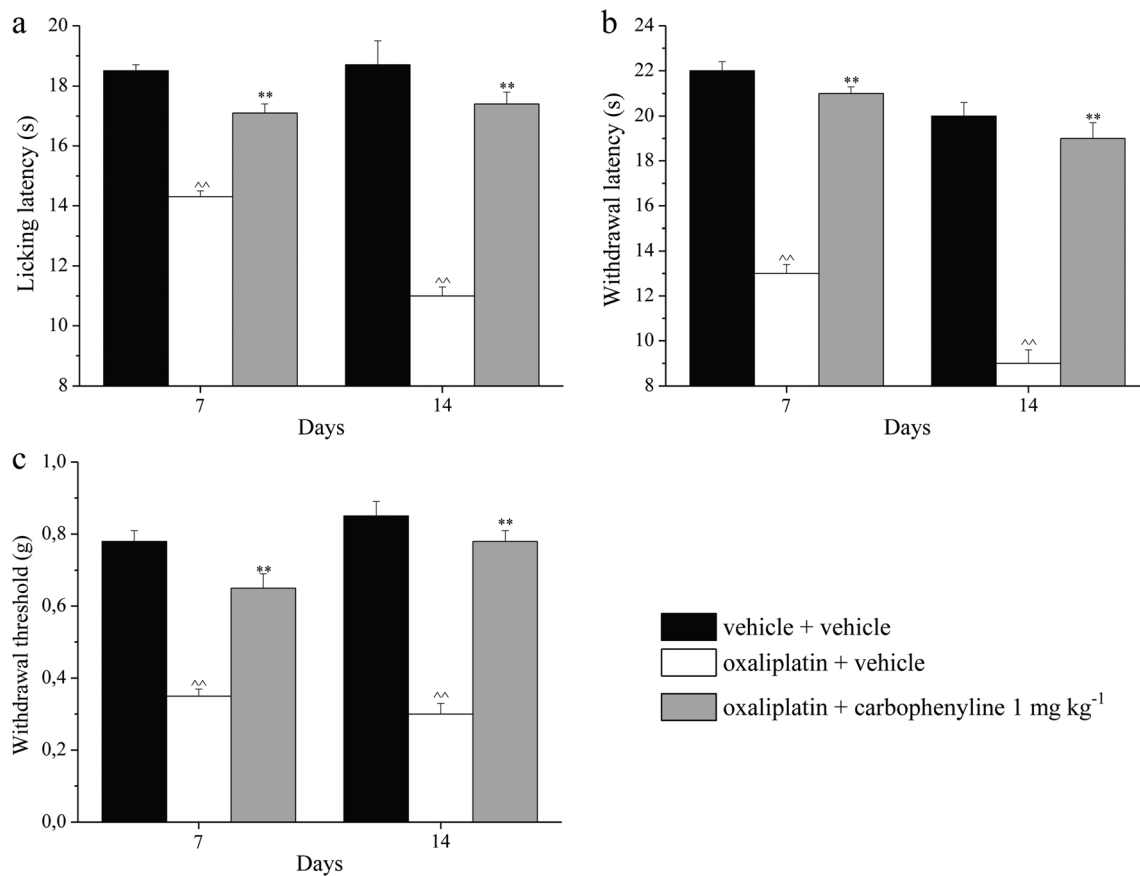


Fig. 4 Effects of repeated administration of carbophenylene against pain induced by oxaliplatin. Pain threshold to a non-noxious thermal stimulus as measured by the cold plate test (a). Sensitivity to a noxious mechanical stimulus as measured by the paw pressure test (b). Pain threshold to a non-noxious mechanical stimulus as measured by the Von Frey test (c). Behavioral tests were performed on days 7 and 14 after the beginning of oxaliplatin and carbophenylene administrations, 24 h after the last

treatment. Oxaliplatin (2.4 mg kg⁻¹, i.p.) was administered for 2 weeks (10 injections) whereas carbophenylene (1 mg kg⁻¹, p.o.) was daily administered, starting from day 1 of oxaliplatin injection. Control animals were treated with vehicles. Each value represents the mean \pm S.E.M. of 10 animals per group performed in 2 experimental sets. Statistical analysis is one-way ANOVA followed by Bonferroni's post hoc comparison. ^{^^} $P < 0.01$ vs vehicle + vehicle; ^{**} $P < 0.01$ vs oxaliplatin + vehicle

selective I₁-R agonist, diethyl-phenyl-amino-imidazoline, ST91. Exploring the possible anatomical and molecular impact of I₁-R in the nervous system for justifying the pain-relieving effect, some interesting data emerge. I₁ binding sites were found in bovine, rabbit, rat, and human brainstem and brain [55]. In the brainstem, I₁-Rs were detected predominantly on non-mitochondrial membranes [56]; in the brain, they are particularly expressed by synaptic membranes, and most likely from presynaptic terminals [55]. Among molecular mechanisms mediated by I₁-R, Edwards and Ernsberger [57] showed that the receptor activation can abolish the nerve growth factor-activated signalling pathway by increasing the levels of a specific phosphatase through ERK dephosphorylation. This effect could be particularly relevant in pain modulation since the nerve growth factor (as others growth factors) is a potent pain inducer [58] that seems unable to separate the neuroprotective effect from the algic one probably following the evolutionary positive alarm role of physiological pain. Moreover, moxonidine, an anti-hypertensive drug, inhibited

voltage-dependent N-type Ca²⁺ current in rat ganglion neurons via activating I₁-R attenuating repetitive firing [29]. Neuronal N-type Ca²⁺ current is closely related to nociception and the pivotal role of its modulation in the pharmacodynamic of anti-neuropathic drugs was recently described by our group [59]. On the contrary, other authors correlated the I₁-R activation in the striatum with pro-excitatory signals [60] that cannot fit with pain relief indicating that the nervous I₁-R signaling is complex and dependent on the anatomical localization.

Furthermore, a few data indicate a I₁-R role in neuroprotection. The stimulation of this receptor participates in the improvement of locomotor recovery in mice subjected to spinal cord injury treated with agmatine [61]. Treatment with moxonidine significantly attenuated locomotor activity, grip strength, learning, and memory impairments in a model of Huntington's disease [62].

Considering the wide expression of I₁-R in several areas of the CNS [55] and that no studies reported on the anti-neuropathic efficacy of selective I₁-R ligands, we thought it

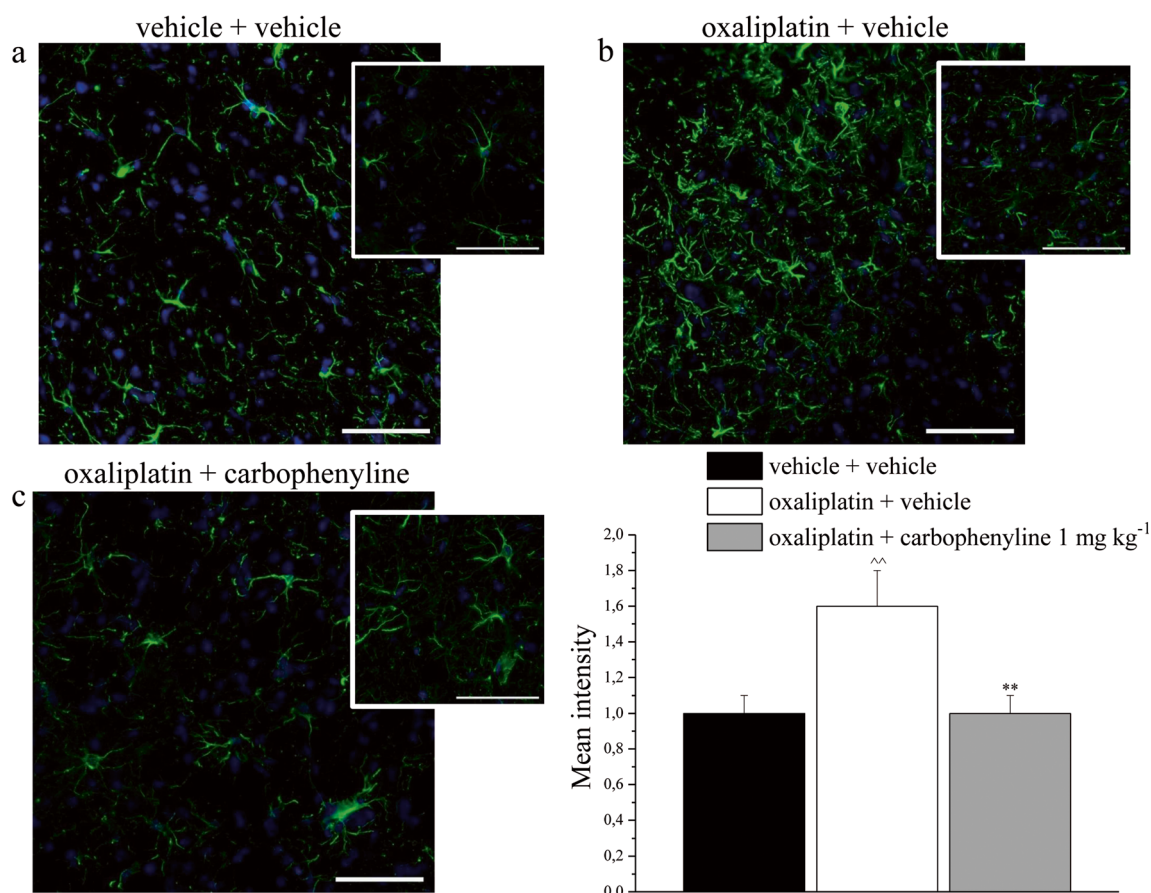


Fig. 5 Astrocytic profile in the spinal cord. The effect of repeated treatment with carbophenylne (1 mg kg^{-1}) was evaluated in oxaliplatin-treated mice on day 14. The number of GFAP-positive cells was measured in the dorsal horn of the L4–L5 spinal cord. Transverse sections of spinal cord imaged with $\times 20$ objective (scale bar = $50 \mu\text{m}$); insert shows morphological characteristics of a representative astrocytic

cell. Histograms show the quantitative analysis of GFAP fluorescence intensity. Each value represents the mean of 6 mice, performed by analyzing 4 independent fields for both sides of the dorsal horn of the lumbar spinal cord. Statistical analysis is one-way ANOVA followed by Bonferroni's post hoc comparison. $^{\wedge\wedge\wedge}P < 0.05$ vs vehicle + vehicle; $^{**}P < 0.01$ vs oxaliplatin + vehicle

Table 1 HT-29 cell viability after oxaliplatin alone or in combination with carbophenylne 48 h incubation

Oxaliplatin concentration (μM)	Cell viability %	
	48-h incubation	
	Control	Carbophenylne $10 \mu\text{M}$
0	100.0 ± 1.3	98.2 ± 2.2
0.3	91.8 ± 1.7	91.1 ± 2.4
1	89.0 ± 2.2	91.6 ± 1.1
3	79.7 ± 1.4	79.3 ± 0.9
10	72.5 ± 1.1	68.2 ± 0.7
30	64.5 ± 1.2	63.4 ± 0.9
100	31.8 ± 0.3	31.4 ± 1.7

HT-29 cells (cells/well) were treated with increasing concentrations of oxaliplatin (1 – $100 \mu\text{M}$) in the presence or absence of $10 \mu\text{M}$ carbophenylne. Incubation was allowed for 48 h. Cell viability was measured by the MTT assay. The control condition was arbitrarily set as 100% and values are expressed as the mean \pm S.E.M. of three experiments. *t* test was used to compare carbophenylne treated samples in combination with oxaliplatin to the oxaliplatin-treated samples

Table 2 HT-29 cell viability after carbophenylne 48 h incubation

Cell viability %	
Carbophenylne (μM)	48 h incubation
0	100.0 ± 1.9
1	98.0 ± 1.4
10	96.3 ± 3.3
30	97.2 ± 1.9
100	94.2 ± 1.4
300	$41.7 \pm 1.9^{***}$

HT-29 cells (cells/well) were treated with increasing concentrations of carbophenylne (1 – $300 \mu\text{M}$) for 48 h. Cell viability was measured by the MTT assay. The control condition was arbitrarily set as 100% and values are expressed as the mean \pm S.E.M. of three experiments. *t* test was used to compare the treated samples to the control. $^{***}P < 0.001$ vs the control condition

interesting to investigate the role played by I₁-R in the oxaliplatin-induced neuropathic pain using the selective I₁-R agonist carbophenylene. The selection of carbophenylene is rationally justified on the basis of its structural analogy with other anti-neuropathic imidazoline compounds at our disposal, in particular derivative **4** showing a significant pain-relieving effect in a rat model of neuropathic pain [49].

In vivo experiments demonstrated the symptomatic properties of carbophenylene in relieving oxaliplatin-induced allodynia and hyperalgesia after a single acute injection. The relevance of the I₁-R in the pain-relieving pharmacodynamic of carbophenylene has been validated through the use of **3**, efaroxan, and BU224 (I₁-, I₁/α₂-, and I₂-R antagonists, respectively) demonstrating the selective involvement of type I₁-R. Moreover, carbophenylene prevented the development of oxaliplatin-induced neuropathic pain after a repeated administration, and the effect remained stable both 7 and 14 days over treatment. The described preventive profile allows us to exclude the development of tolerance to the anti-hypersensitivity effects. Further, it suggests possible neuroprotective properties of carbophenylene able to modify the nervous system alterations leading the establishment and persistence of neuropathic pain.

This aspect was studied in the present research analyzing the possible impact of I₁-R stimulation on spinal cord alterations evoked by oxaliplatin. The repeated treatment with carbophenylene prevented the activation of astrocytes in the dorsal horns. Glia cells contribute to the persistence of pain, an effect that in neuropathic conditions prevails on the usual homeostatic functions [63]. The metabolic activation of microglia and astrocytes participates in the maladaptive plasticity of the central nervous system, facilitating nociceptive processes, generating clinical pain hypersensitivity, and making neuropathic pain an autonomous disease state [64]. Despite the limited ability of oxaliplatin to cross the blood–brain barrier [65], in our previous study we demonstrated that microglia and astrocytes cooperated to promote the initial phase of spinal cord sensitization but, although microglia conclude the task in the early phase of oxaliplatin administration, astrocytes maintain their activation contributing to the pain persistence [10]. These data are supported by the information that the spinal intrathecal infusion of fluorocitrate, an astrocyte selective inhibitor, fully prevented oxaliplatin-evoked pain alterations [10]. The present results revealed the modulatory role of carbophenylene on oxaliplatin-induced spinal astrocyte activation suggesting its ability to protect from the maladaptive plasticity of the central nervous system, a mechanism that could offer an explanation for the prevention of pain development demonstrated after a repeated treatment. The present first demonstration regarding the potential of I₁-R agonists in pain relief adds a fragment to the very limited information regarding the role of this receptor in nervous signals.

A crucial point in the study of novel treatments against chemotherapy-induced neuropathic pain is the evaluation

of possible interactions with the anticancer effect of the drug as the primary treatment outcome. As a preliminary evaluation we analyzed the effect of carbophenylene on oxaliplatin lethality against the human colon adenocarcinoma cell line HT-29, representing a main clinical target for this platin derivative [2, 3]. No interferences with the concentration-dependent effect of oxaliplatin were registered after 48 h treatment with 10 μM of carbophenylene, a concentration about 65-fold higher in comparison to the I₁-R affinity [30, 31]. Carbophenylene per se did not alter HT-29 cell viability suggesting the lack of a direct effect against cancer cells. On the base of pharmacophore modelling and virtual docking study, I₁-R agonists were suggested as possible apoptotic agents in cancer cells [66]. In the pheochromocytoma-derived cell line PC12, not specific I₁-R ligands showed antiproliferative effect [55]. Generally, imidazolines can sensitize cancer cells to antineoplastic DNA damaging agents [67]. The role of I₁-R in cancer and the effects of specific ligands, like carbophenylene, in cancer mechanisms as well as in the possible pharmacological interactions with antitumor drugs deserve deep investigation.

The development of oxaliplatin neuropathy is a clinical issue in patients receiving a cumulative dose exceed 780–850 mg/m², approximately two-thirds will have symptoms 1 year after treatment or beyond [4, 68]. The lack of effective therapies claims for new pharmacological approaches: in this context I₁-R stimulation rises to a very interesting potential.

In conclusion, for the first time the I₁-R has been demonstrated to be a valuable and effective target for the management of oxaliplatin-induced neuropathic pain. The potent and selective I₁-R agonist carbophenylene is a lead compound for the development of an innovative class of pain killers. The novelty of the obtained results stimulates the continuation of the research both from chemical and biological point of view. In fact, the presence in carbophenylene of an asymmetric carbon atom suggests the examination of the influence of stereochemistry on its anti-neuropathic effect. Moreover, it might be of interest to investigate whether I₁-R ligands can be active also in other type of neuropathic pain as well as to highlight the molecular pharmacodynamic of pain relief.

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Compliance with Ethical Standards

Conflict of Interest The authors declare that they have no conflict of interest.

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