



Conformationally restricted homotryptamines. Part 5: 3-(trans-2-aminomethylcyclopentyl)indoles as potent selective serotonin reuptake inhibitors

Derek J. Denhart^{*}, Jeffrey A. Deskus, Jonathan L. Ditta, Qi Gao, H. Dalton King, Edward S. Kozlowski, Zhaoxing Meng, Melissa A. LaPaglia, Gail K. Mattson, Thaddeus F. Molski, Matthew T. Taber, Nicholas J. Lodge, Ronald J. Mattson, John E. Macor

Bristol-Myers Squibb Pharmaceutical Research Institute, 5 Research Parkway, Wallingford, CT 06492-7660, USA

ARTICLE INFO

Article history:

Received 26 February 2009

Revised 5 June 2009

Accepted 8 June 2009

Available online 13 June 2009

Keywords:

Indoles

Homotryptamines

SSRI

Serotonin

ABSTRACT

A series of racemic 3-(trans-2-aminomethylcyclopentyl)indoles was synthesized and found to have potent binding to the human serotonin transporter (hSERT). The most active analog was synthesized stereospecifically and the active enantiomer was shown to have high affinity binding to hSERT.

© 2009 Elsevier Ltd. All rights reserved.

Serotonin (5-hydroxytryptamine or 5-HT) is an important neurotransmitter and alterations in its signaling are implicated in affective disorders. The selective serotonin reuptake inhibitors (SSRIs) block the serotonin transporter (SERT) and prevent reuptake into presynaptic neuron.¹

The SSRIs are widely used antidepressants, and are relatively safe despite some recognized issues.² Some recent SSRI research has sought to discover compounds with more rapid onset of action as well as discovering compounds that increase the number of patients who respond to treatment. Among other approaches, some investigations have attempted to combine SERT inhibition with 5-HT_{1A}, 5-HT_{1B}, or NK1 antagonism.³

The conformational restriction of ligands or inhibitors is a well-precedented way to improve binding and selectivity of these agents. We have previously reported SERT binding results with homotryptamines **1**⁴ and conformationally-restricted homotryptamine analogs **2**⁵ and **3**⁶ (Fig. 1).

In this report we discuss methods to vary the ring size from the cyclopropyl in **3** to cyclopentyl and the effect this modification has on SERT binding and the physical properties of the 3-cyclopentylindole derivatives. The 5-cyanoindole core (Fig. 1, X = CN) was maintained based on SAR from the indole cyclopropane series discovered previously.⁶

The synthesis of the racemic *cis* and *trans* 3-(2-aminomethylcyclopentyl)indoles is shown in Scheme 1 5-cyanoindole **4** is converted into iodoindole **5** and then subjected to a Stille coupling with **6** to provide olefin **7**.⁷ Tosyl removal and hydrogenation provided racemic *cis* compound **8**. Base equilibration followed by functional group conversion produced racemic *trans* aldehyde **9**

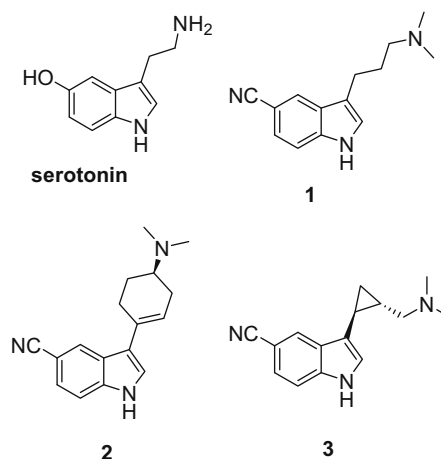
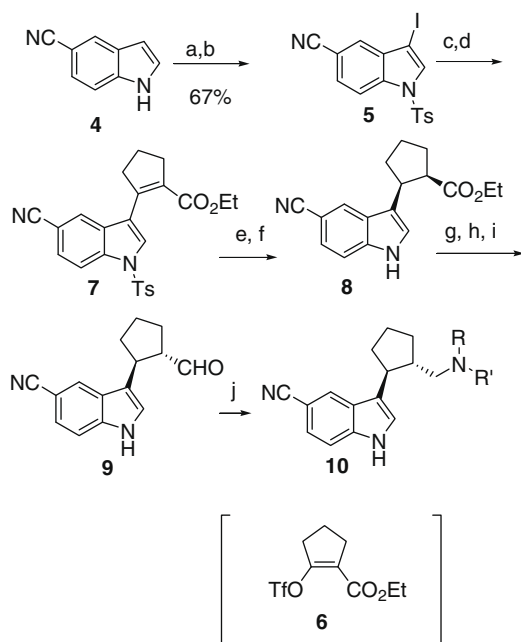


Figure 1. Previously reported indole SERT ligands.

^{*} Corresponding author. Tel.: +1 203 677 7117.

E-mail address: derek.denhart@bms.com (D.J. Denhart).



Scheme 1. Synthesis of *trans* racemic cyclopentanes. Reagents and conditions: (a) I_2 , KOH; (b) TsCl, DIPEA (67%); (c) $(Bu_3Sn)_2$, $Pd(OAc)_2$, PPh_3 (82%); (d) **6**, Pd_2dba_3 , Ph_3As (73%); (e) NaOH, H_2O (94%); (f) H_2 , Pd/C, (89%); (g) LiOH, H_2O , EtOH, reflux; (h) MeONHMe-HCl, EDCl, Et_3N (41%); (i) LAH, $-38^\circ C$ (68%); (j) R-NH-R', $NaBH(OAc)_3$.

which was then reductively aminated to provide a variety of amines **10**.

The data for the racemic *trans* compounds are shown in Table 1. This series provided several highly potent SERT⁸ inhibitors, most notably the dimethylamine **10b**. The data show a clear preference for small alkyl groups, exemplified by the potent binding of **10a–c** contrasted with the drop in activity for the diethylamine **10f**. These SAR trends are similar to what was observed for the previously reported 3-cyclopropylindole series.⁶

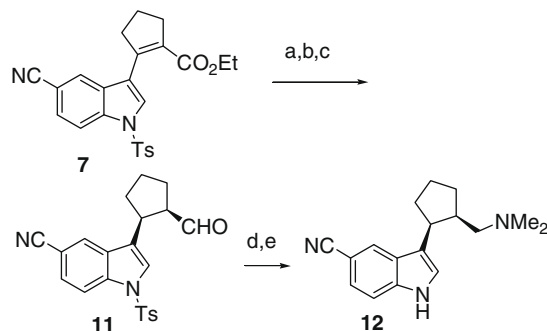
To test the effects of bond orientation about the cyclopentane ring, we desired to test the enantiomers of both *trans* and *cis* cyclopentane dimethylamines. The racemic *trans* **10b** could be separated into (1*S*,2*S*) (+)-**10b** and (1*R*,2*R*) (–)-**10b** using chiral HPLC. Alternatively, larger amounts of these *trans* enantiomers could be synthesized by an asymmetric Michael addition as previously reported.⁹ The synthesis of racemic *cis* compounds is shown in Scheme 2. The previously synthesized **7** was hydrogenated and subjected to careful ester reduction followed by Swern oxidation¹⁰ to provide racemic *cis* aldehyde **11**. Reductive amination followed by tosyl removal gave desired racemic *cis* amine **12**. This racemic mixture was separated into (+)-**12** and (–)-**12** using chiral HPLC.

As can be seen from the data in Table 2, the most potent compound was the *trans*-(1*S*,2*S*) enantiomer (+)-**10b** with a hSERT

Table 1
Binding affinities of racemic *trans* compounds **10a–g**^a

Compound	R	R'	SERT IC ₅₀ ^a (nM)
10a	Me	H	7.0 ± 1.1
10b	Me	Me	0.32 ± 0.04
10c	Me	Et	10 ± 0.50
10d	Me	CH ₂ Ph	85 ± 12
10e	Et	H	170
10f	Et	Et	240
10g	–(CH ₂) ₄ –		22 ± 4.2

^a Where indicated with ± SEM, $n \geq 3$ for the reported IC₅₀ values. Otherwise, $n = 2$.



Scheme 2. Synthesis of *cis* cyclopentane **10**. Reagents and conditions: (a) H_2 , Pd/C; (b) $LiAlH_4$, $-40^\circ C$; (c) $(COCl)_2$, DMSO, then ROH, then Et_3N ; (d) Me_2N , $NaBH(OAc)_3$; (e) NaOH, EtOH.

Table 2
hSERT binding affinities of stereoisomers of **10b** and **12**^a

Compound	<i>cis/trans</i>	SERT IC ₅₀ ^a (nM)
10b	<i>trans</i>	0.32 ± 0.04
(+)- 10b	<i>trans</i>	0.13
(–)- 10b	<i>trans</i>	58
12	<i>cis</i>	2.6 ± 0.34
(+)- 12	<i>cis</i>	130 ± 16
(–)- 12	<i>cis</i>	1.2 ± 0.21

^a Where indicated with ± SEM, $n \geq 3$ for the reported IC₅₀ values. Otherwise, $n = 2$.

Table 3
Binding affinities (IC₅₀ in nM) versus selected receptors for **2**, **3**, and (+)-**10b**

Compound	SERT	DAT	NET
2	0.72	290	390
3	0.36	4200	9200
(+)- 10b	0.13	690	7900

binding IC₅₀ of 0.13 nM. The *cis* compounds (+)-**12** and (–)-**12** were several fold less active than the *trans* analogs. The most active *cis* enantiomer was the (1*R*,2*S*) (+)-**12** with an IC₅₀ of 1.2 nM. The absolute configurations of active enantiomers were as expected based on the indole cyclopropane series reported previously.²

In Table 3 can be seen data on the binding potency of (+)-**10b** and the previously reported compounds **2** and **3** to the major biogenic amine transporters SERT, DAT (dopamine) and NET (norepinephrine).¹¹ The cyclopentyl compound (+)-**10b** is the most potent at binding to SERT, with the cyclohexene compound **2** being the least potent (although still subnanomolar in IC₅₀). Both compounds **2** and **3** show binding to DAT which is at least 5000 fold weaker than its binding to SERT, and binding to NET which is at least 25,000 fold weaker than SERT. Compound **2** is significantly less selective than **3** and (+)-**10b**.

In vivo microdialysis studies with (+)-**10b** are shown in Figure 2.¹² Compound (+)-**10b** given orally caused a robust increase in extracellular serotonin concentrations in the frontal cortex of awake, freely moving rats. The maximal response was achieved at 1 mg/kg (po), and this increased serotonin levels to greater than 300% of baseline. This effect was similar to that produced by the SSRI paroxetine given at 5 mg/kg.

In conclusion, the 3-(2-aminomethylcyclopentyl)indole series provided a compound (+)-**10b** which was a picomolar potent and selective inhibitor of the hSERT transporter. Future studies are ongoing to fully appreciate the therapeutic potential of such a compound.

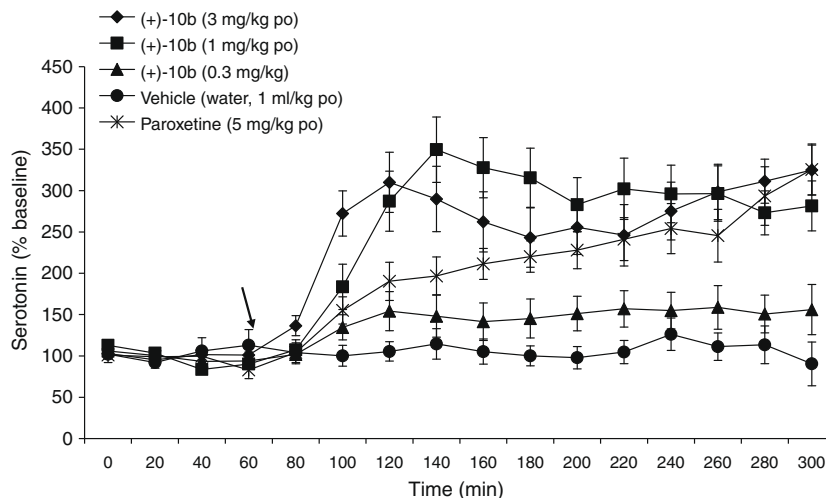


Figure 2. Dose-reponse curves for compound (+)-10b given po.

References and notes

- Spinks, D.; Spinks, G. *Curr. Med. Chem.* **2002**, *9*, 799.
- Ananth, J. *Psychother. Psychosom.* **1998**, *67*, 61.
- (a) Rocco, V. P.; Spinazze, P. G.; Kohn, T. J.; Honigschmidt, N. A.; Nelson, D. L.; Wainscott, D. B.; Ahmad, L. J.; Shaw, J.; Threlkeld, P. G.; Wong, D. T.; Takeguchi, K. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 2653; (b) Oficialdegui, A. M.; Martinez, J.; Perez, S.; Hears, B.; Irurzun, M.; Palop, J. A.; Tordera, R.; Lasheras, B.; del Rio, J.; Monge, A. *Farmaco* **2000**, *55*, 345; (c) Evrard, D. A.; Zhou, P.; Yi, S.; Zhou, D.; Smith, D.; Sullivan, K. M.; Hornby, G. A.; Scheckter, L. E.; Andree, T. H.; Mewshaw, R. E. *Bioorg. Med. Chem. Lett.* **2005**, *15*, 911; (d) Ryckmans, T.; Balancon, L.; Berton, O.; Genicot, C.; Lamberty, Y.; Lallemand, B.; Pasau, P.; Pirol, N.; Quere, L.; Talaga, P. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 261; (e) Ryckmans, T.; Berton, O.; Grimee, R.; Kogej, T.; Lamberty, Y.; Pasau, P.; Talaga, P.; Genicot, C. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 3195.
- (a) Schmitz, W. D.; Denhart, D. J.; Brenner, A. B.; Ditta, J. L.; Mattson, R. J.; Mattson, G. K.; Molski, T. F.; Macor, J. E. *Bioorg. Med. Chem. Lett.* **2005**, *15*, 1619; (b) Denhart, D. J.; Mattson, R. J.; Ditta, J. L.; Macor, J. E. *Tetrahedron Lett.* **2004**, *45*, 3803.
- Deskus, J. A.; Epperson, J. R.; Sloan, C. P.; Cipollina, J. A.; Dextraze, P.; Qian-Cutrone, J.; Gao, Q.; Ma, B.; Beno, B. R.; Mattson, G. K.; Molski, T. F.; Krause, R. G.; Taber, M. T.; Lodge, N. J.; Mattson, R. J. *Bioorg. Med. Chem. Lett.* **2007**, *17*, 3099.
- Mattson, R. J.; Catt, J. D.; Denhart, D. J.; Deskus, J. A.; Ditta, J. L.; Higgins, M. A.; Marcin, L. R.; Sloan, C. P.; Beno, B. R.; Gao, Q.; Cunningham, M. A.; Mattson, G. K.; Molski, T. F.; Taber, M. T.; Lodge, N. J. *J. Med. Chem.* **2005**, *48*, 6023.
- Ciattini, P. G.; Morera, E.; Ortar, G. *Tetrahedron Lett.* **1994**, *35*, 2405.
- The SERT binding affinities were determined using membrane homogenates from HEK-293 cells that stably expressed human serotonin transporters (HEK-hSERT cells). Membrane homogenates were incubated with 2 nM [³H]citalopram (specific activity = 85 Ci/mmol) and increasing concentrations of test compounds for 1 h at 25 °C in a total volume of 250 µL. Amount of radioligand bound in the presence and absence of a competitor was analyzed by plotting (–)log drug concentration versus the amount of radioligand specifically bound. Non-specific binding was defined with 10 µM fluoxetine. The midpoint of the displacement curve (IC₅₀ in nM) signified the potency.
- King, H. D.; Meng, Z.; Denhart, D.; Mattson, R.; Kimura, R.; Wu, D.; Gao, Q.; Macor, J. E. *Org. Lett.* **2005**, *7*, 3437.
- Omura, K.; Swern, D. *Tetrahedron Lett.* **1978**, *34*, 1651.
- SERT, DAT, and NET binding affinities were determined using membrane homogenates from stably transfected HEK-293 cell lines expressing the human form of the transporters. Membrane homogenates were incubated with ¹²⁵I labeled ligands. RTI-55 (Perkin–Elmer) was used for SERT (260 pM) and DAT (125 pM). A custom labeled ¹²⁵I Nisoxetine (Perkin Elmer) was used for NET (300 pM). The reactions were carried out in a 384-well format and harvested in 384-well filter plates. The IC₅₀ data was determined from a 20-point curve. Non-specific binding was defined using 10 µM fluoxetine for SERT, 100 µM Desipramine for NET, and 10 µM GBR-12935 for DAT.
- Taber, M. T.; Wright, R. N.; Molski, T. F.; Clarke, W. J.; Brassil, P. J.; Denhart, D. J.; Mattson, R. J.; Lodge, N. J. *Pharmacol., Biochem. Behav.* **2005**, *80*, 521.