Bioorganic & Medicinal Chemistry Letters 24 (2014) 2263-2266

Contents lists available at ScienceDirect

ELSEVIER



Bioorganic & Medicinal Chemistry Letters

Towards the discovery of drug-like epigallocatechin gallate analogs as Hsp90 inhibitors



Rohit Bhat^a, Amna T. Adam^a, Jungeun Jasmine Lee^a, Thomas A. Gasiewicz^b, Ellen C. Henry^b, David P. Rotella^{a,*}

^a Department of Chemistry and Biochemistry, Montclair State University, Montclair, NJ 07043, United States ^b Department of Environmental Medicine, University of Rochester Medical Center, Rochester, NY 14642, United States

ARTICLE INFO

Article history: Received 26 January 2014 Accepted 26 March 2014 Available online 4 April 2014

Keywords: Hsp90 inhibitor Natural product Epigallocatechin gallate

ABSTRACT

(–)-Epigallocatechin gallate (EGCG) is the major flavonoid of green tea and has been widely explored for a range of biological activities including anti-infective, anti-inflammatory, anti-cancer, and neuroprotection. Existing structure-activity data for EGCG has been largely limited to exploration of simple ethers and hydroxyl deletion. EGCG has poor drug-like properties because of multiple phenolic hydroxyl moieties and a metabolically labile ester. This work reports a substantial expansion of structure-activity understanding by exploring a range of semi-synthetic and synthetic derivatives with ester replacements and variously substituted aromatic and alicyclic groups containing more drug-like substituents. Structure-activity relationships for these molecules were obtained for Hsp90 inhibition. The results indicate that amide and sulfonamide linkers are suitable ester replacements. Hydroxylated aromatic rings and the *cis*-stereochemistry in EGCG are not essential for Hsp90 inhibition. Selected analogs in this series are more potent than EGCG in a luciferase refolding assay for Hsp90 activity.

© 2014 Elsevier Ltd. All rights reserved.

(-)-Epigallocatechin gallate (EGCG, 1, Fig. 1) is the primary flavonoid component of green tea and has been widely studied due to its diversity of biological properties including antiinflammatory, anti-oxidative, anti-cancer, anti-infective and neuroprotective activity.¹⁻³ Structure-activity studies on **1** reported to date have focused primarily on a limited spectrum of analogs involving methylation and/or deletion of phenolic hydroxvl moieties in 1-3.⁴⁻⁶ These compounds have been studied for biological activity in systems related to the therapeutic applications mentioned above. The resulting derivatives retain unfavorable characteristics of EGCG including chemical and metabolic instability. Our approach to modification of EGCG is focused on discovery of more drug-like derivatives in the context of an expanded structure-activity study that includes exploration of the influence of stereochemistry in the natural product on biological/target specific properties.

This Letter details our initial efforts toward the discovery of more drug-like analogs of EGCG using inhibition of Hsp90 as a representative biological target. Hsp90 is an intracellular chaperone protein essential for cell growth and division as well as protein trafficking.⁷ It is a target of interest for treatment of cancer and

^{*} Corresponding author. Tel.: +1 973 655 7204; fax: +1 973 655 7772. *E-mail address:* rotellad@mail.montclair.edu (D.P. Rotella).



potentially neurodegenerative disorders.⁸ We prepared a series of semisynthetic and synthetic EGCG analogs in which the metabolically labile ester is replaced by more stable linkers, aromatic rings



 R_1 = H, OH; n=0, 2 R_2 = H, OH; m=0, 2,3 X= O, NH Y= (C=O), SO₂ Z= aromatic, heteroaromatic, alicyclic

Figure 2.

are replaced by alicyclic systems and phenolic hydroxyl groups are removed and/or replaced with other substituents as outlined in Figure 2. Stereochemical effects on Hsp90 activity are evaluated using a commercially available derivative, gallocatechin gallate (GCG, **4**). EGCG was characterized previously as a C-terminal binding Hsp90 inhibitor.⁹ EGCG analogs were evaluated as Hsp90 inhibitors using a luciferase refolding assay because a direct Hsp90 binding assay for C-terminal inhibitors is not yet available. Since the recovery of luciferase enzymatic activity after heat-induced denaturation is dependent on Hsp90 chaperone function, this assay is commonly used to test compounds for their ability to inhibit Hsp90.^{9,10}

The semi-synthesis of EGCG analogs is outlined in Scheme 1 below.¹¹ Silyl protection of the phenolic hydroxyl groups in **1** followed by lithium aluminum hydride reduction furnished protected EGC derivative **6**. Esterification by DCC-mediated coupling provided silyl-protected intermediates in which silyl deprotection was best accomplished with aqueous HF in tetrahydrofuran, leading efficiently to targets **7a–e**. Oxidation of **6** with Dess–Martin reagent followed by reductive amination with benzylamine and hydrogenolytic debenzylation provided primary amine **8** that was treated with benzoyl chloride or phenylsulfonyl chloride to give the respective amide and sulfonamide which were de-silylated as above leading to amide **9a** and sulfonamide **9b** in good yield. Beginning with (+)-catechin (**10**), selective phenolic hydroxyl benzylation, esterification and hydrogenolytic debenzylation led to *trans* esters **11a–c**. Benzyl protection of **10** followed by



Scheme 1. Reagents and conditions: (a) TBDMS-CI, Et₃N, DMF, rt 95%; (b) LiAlH₄/THF, 0 °C 75%; (c) Z-CO₂H, DCC, DMAP, CH₂Cl₂, rt 70%; (d) 48% aq HF, THF, rt 80%; (e) Dess-Martin, CH₂Cl₂, rt 85%; (f) benzylamine, cat. HOAc, NaBH₃CN, THF, rt 75%; (g) H₂ (30 psi), 10% Pd-C, MeOH, rt 95%; (h) benzoyl chloride, Et₃N, CH₂Cl₂, rt 76%; (i) phenyl sulfonyl chloride, Et₃N, CH₂Cl₂, rt 68%; (j) BnBr, K₂CO₃, DMF, rt 30%; (k) L-Selectride, THF, -78 °C to rt, 76%; (l) EtOH/H^{*} reflux, 85%; (m) NaOH/EtOH rt, 89%; (n) methyl DAST, CH₂Cl₂ 0 °C to rt, 75%.



Scheme 2. Reagents and conditions: (a) BnBr, K₂CO₃, DMF, rt, 45%; (b) benzaldehyde, piperidine, ethanol, reflux, 1N HCl, 43%; (c) NaBH₄, EtOH/THF, reflux, rt 16 h, 15% HOAc, 40%; (d) BH₃-THF, THF, 0 °C, H₂O₂, NaOH, THF, 59%; (e) Z-CO₂H, DCC, DMAP, DCM, rt, 74%; (f) 30 psi H₂, 10% Pd/C, THF, 90%; (g) Dess-Martin, 0 °C to rt, DCM, 80%; (h) L-Selectride, THF, -78 °C to rt, 63%.

Dess-Martin oxidation and L-Selectride reduction efficiently gave an epimeric alcohol **12** that was esterified and debenzylated as before to give protected *cis* esters **18c** and **18f**. *cis*- and *trans*hydroxy-substituted cyclohexyl intermediates (**14** and **16**) necessary for esters **18d** and **18e** were prepared by standard methods and coupled with **12** then converted as indicated in Scheme 1. Fluoro derivatives **18a** and **18b** were prepared from their respective hydroxyl intermediates by methyl DAST.

Synthetic EGCG analogs lacking phenolic hydroxyl groups were prepared as outlined in Scheme 2. Chalcone synthesis using **19a** and benzyl protected acetophenone **20** furnished **21a** and **21b**, respectively, which underwent hydroboration/oxidation to give a mixture of alcohols in which the *trans* isomers **22a** and **22b** predominate.^{12,13} Esterification of **22b** and deprotection as above provided *trans* esters **26** and **27**. Dess–Martin oxidation of **22a/b** followed by stereoselective reduction by L-Selectride, esterification and debenzylation furnished *cis*-analogs **24** and **25**.

EGCG (1), GCG (4) and the new analogs were evaluated for functional inhibition of Hsp90 activity using a luciferase refolding assay as described elsewhere.^{9,10} The results are shown in Tables 1-3

Table 1

Semisynthetic D-ring and linker analogs



Compd	cis/trans ^a	Х	Y	Z	$IC_{50}^{b}(\mu M)$
1	cis	0	C=0	Gallate	155
4	trans	0	C=0	Gallate	34
7a	cis	0	C=0	Phenyl	24
7b	cis	0	C=0	4-F-Ph	41
7c	cis	0	C=0	3,4-Di-F-Ph	58
7d	cis	0	C=0	4-Pyridyl	64
7e	cis	0	C=0	Cyclohexyl	30
9a	cis	NH	C=0	Phenyl	75
9b	cis	NH	SO_2	Phenyl	44

^a Gallate B ring/X (absolute stereochemistry as in EGCG).

^b Average value based on at least two independent determinations.

Table 2

Semisynthetic catechol D-ring analogs



Compd	cis/trans ^a	Z	$IC_{50}^{b}(\mu M)$
11a	trans	Cyclohexyl	98
11b	trans	Phenyl	110
11c	trans	§ОН	280
18a	cis	≹ —∕─F	65
18b	cis	ξ −− √−−F	69
18c	cis	Cyclohexyl	81
18d	cis	§ —∕ −он	310
18e	cis	ξ √−−OH	440
18f	cis	Cyclopropyl	410

^a Catechol B ring/ester (absolute stereochemistry derived from GCG).

^b Avg based on at least two independent determinations.

below. The following structure–activity observations can be made: (1) The metabolically labile ester can be replaced by an amide (**9a**) or sulfonamide (**9b**) leading to more potent Hsp90 inhibitors. This further suggests that a poly-hydroxy D ring is not necessary for Hsp90 inhibition and the relative geometry of a pyramidal sulfonamide compared to the more planar amide or ester does not disrupt potential binding interactions in this region. The sulfonamide **9b** is approximately 3-fold more potent compared to EGCG. (2) Hsp90 inhibition associated with these and other des-hydroxy derivatives shows that there are a variety of potentially more drug-like options for replacement of the gallate ester including an electron deficient heterocycle (**7d**) and substituted alicyclics (**18a**, **18b**). Fluoro-substituted aromatic groups (**7b**, **7c**) also result in improved Hsp90 inhibition.

Table 3

Racemic, synthetic analogs



Compd	cis/trans ^a	R ₁	R_2	Z	$IC_{50}^{b}(\mu M)$
24	cis	OH	OH	Phenyl	63
25	cis	Н	Н	Gallate	35
26	trans	OH	OH	Phenyl	120
27	trans	OH	OH	Gallate	150

^a Relative phenyl/ester stereochemistry.

^b Avg based on at least two independent determinations.

are measurably more potent than EGCG, for example cyclohexyl ester **7e** and benzoate ester **7a**. (3) Deletion of both A- and B-ring hydroxyl groups as in **25** (a racemate) notably furnishes a more potent Hsp90 inhibitor (IC_{50} 35 μ M) compared to EGCG. (4) B-ring catechol esters are less active compared to analogous gallate analogs, cf. **11a** versus **7e** and **11b** versus **7a**. This observation suggests the possibility of more complex SAR in view of the improved potency of racemic des-hydroxy **25**. In this catechol series, fluoro substitution of the cyclohexane ring is preferable to hydroxyl (cf. **18b** vs **18e**) and the relative stereochemistry of fluorine has little effect on Hsp90 inhibition. It is also interesting to observe that the difference in potency between *cis*- and *trans*-isomers in the racemic, synthetic des-hydroxy analogs in Table 3 differs from that observed with EGCG and GCG. These results collectively suggest a number of avenues for future SAR studies.

This initial SAR survey of synthetic and semi-synthetic analogs was intended to highlight stereochemical and structural variations in rings A, B and D of EGCG that can serve as starting points for preparation and study of more drug like derivatives of this biologically interesting natural product. *trans*-Geometry on the C ring furnishes a more potent Hsp90 inhibitor compared to **1**. Selected compounds were more potent than EGCG in an Hsp90 functional assay. Furthermore our data show that multiple hydroxyl groups can be eliminated and/or replaced with other functional groups. This is especially important given the known tendency of the 4-hydroxy group in the B-ring to play a role in epimerization of EGCG.¹⁴ The metabolically labile ester can be replaced by amide and sulfonamide linkages and the aromatic D ring can be replaced by an alicyclic system. Ongoing work is focused on using this data to identify a combination of sites and analogs where significant improvements in potency of Hsp90 inhibition can be achieved. Other research in this laboratory is exploring connections between increased Hsp90 potency, structural variations and biological outcomes such as neuroprotection and cytotoxicity.

Acknowledgments

This research was supported by the Margaret and Herman Sokol Endowment, NIH Grant AT006366 and Center Grant ES01247, Montclair State University and the Sokol Institute for Pharmaceutical Life Sciences.

References and notes

- 1. Steinmann, J.; Buer, J.; Pietschmann, T.; Steinmann, E. Br. J. Pharmacol. 2013, 168, 1059.
- Huo, C.; Wan, S. B.; Lam, W. H.; Li, L.; Wang, Z.; Landis-Piwowar, K. R.; Chen, D.; Dou, Q. P.; Chen, T. H. Inflammopharmacology 2008, 16, 248.
- 3. Hugel, H. M.; Jackson, N. Mini-Rev. Med. Chem. 2012, 12, 380.
- 4. Khandelwal, A.; Hall, J. A.; Blagg, B. S. J. J. Org. Chem. 2013, 78, 7859.
- Kazi, A.; Wang, Z.; Kumar, N.; Falsetti, S. C.; Chang, T.-H.; Dou, Q. P. Anticancer Res. 2004, 24, 943.
- Osanai, K.; Landis-Piwowar, K. R.; Dou, Q. P.; Chan, T. K. Bioorg. Med. Chem. 2007, 15, 5076.
- Chiosis, G.; Rosen, N.; Mimnaugh, E.; Whitesell, L.; Neckers, L. Mol. Cancer Ther. 2003, 2, 123.
- Ernst, J. T.; Liu, M.; Zuccola, H.; Neubert, T.; Beaumont, K.; Turnbull, A.; Kallel, A.; Vought, B.; Stamos, D. Bioorg. Med. Chem. Lett. 2014, 24, 204.
- 9. Yin, Z.; Henry, E. C.; Gasiewicz, T. A. Biochemistry 2009, 48, 336.
- Turbyville, T. J.; Kithsiri Wijeratne, E. M.; Liu, M. X.; Burns, A. M.; Seliga, C. J.; Luevano, L. A.; David, C. L.; Faeth, S. H.; Whitesell, L.; Gunatilaka, A. A. L. J. Nat. Prod. 2006, 69, 178.
- 11. All final compounds were judged to be at least 95% pure and were characterized by NMR, HPLC and MS.
- 12. Zaveri, N.; Chao, W.-R. WO2003089423.
- 13. Devakaram, R.; Black, D. S.; Andrews, K. T.; Fisher, G. M.; Davis, R. A.; Kumar, N. Bioorg. Med. Chem. 2011, 19, 5199.
- 14. Suzuki, M.; Sano, M.; Yoshida, R.; Degawa, M.; Miyase, T. Agric. Food Chem. 2003, 51, 510.