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Noteworthy observations accompanying synthesis of the apoptolidin disaccharide†

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A stereoselective synthesis of the apoptolidin disaccharide is reported. The key chemistry features a new transformation utilizing a highly selective tetramethylalkoxyalanate[v]-directed syn-methylation of a vinylogous ester, isolation of a hydrate of a 2-keto sugar, an eco-friendly radical cleavage of a bromomethyl group, and an efficient preparation of a fluorodisaccharide via the use of XtalFluor-E.

Apoptolidin A 1 is a complex natural product isolated from Nocardiopsis sp. by Seto and co-workers in 1997.^{1,2} Apoptolidin A 1 shows anti-proliferative activity towards various cancer cell lines studied by the National Cancer Institute. Several structural congeners were isolated by Wender,³ which demonstrated comparable biological activity. Three total syntheses⁴ and several preparations of the disaccharide units have been reported.⁵ Apoptolidin consists of a 20membered macrolactone coupled with a side chain containing a cyclic hemiketal. 6-Deoxy-4-O-methyl-l-glucose is attached to O9 and a disaccharide consisting of l-olivomycose and d-oleandrose is linked to O27. Studies by Khosla revealed F0F1-ATPase as the possible biological target of Apoptolidin. Although oleandrose and olivomycose sugars in apoptolidin are not absolutely essential to the binding of apoptolidin to this target, they are crucial for cellular activity. 6 Apoptolidin D^{3c} 2 is the C-6-des-methylated analogue of Apoptolidin A 1. Our synthetic efforts are focused on the synthesis of Apoptolidin A and D and the SAR of the three additional C19,20 diastereomers thereof.⁷

Apoptolidins A and D were to be prepared from glycosylation of 3 using sugars 4 and 5 (Fig. 1). Sugar 4 is a synthon for 6-deoxy-4-O-methyl-l-glucose moiety found at O-9, and glycosyl fluoride 5 is a synthon for the disaccharide containing l-olivomycose and d-oleandrose, found at O-27. Glycosidic donor 5 was to be prepared from disaccharide 6. The synthesis of **6** envisaged β-selective ester-directed glycosylation of sugars 7 and 8 (Fig. 2). Sugar 7 was to be prepared from L-rhamnose.

L-Rhamnose was converted to known intermediate 9 in 3 steps and 95% yield. Deacetylation provided diol 10 in 96% yield. Oxidation of 10 with pyridinium dichromate⁹ afforded key vinylogous ester 11 (Scheme 1). Treatment of 11 with methyllithium generated a mixture of syn- and anti-methyl adducts in a 1.5:1 ratio. The combination of dimethylzing and methyllithium improved the ratio to 3:1. Treating 11 with one equivalent of trimethylaluminium at -78 °C followed by one equivalent of methyllithium gave better selectivity (7.5:1 ratio). High yield and outstanding selectivity (50:1) for synthesis of 12-svn were obtained by treating 11 with one equivalent of trimethylaluminium at -78 °C followed by two equivalents of methyllithium (Table 1). This new strategy is superior to the alternative methylation protocols employed by various groups 10 and later adopted by Koert^{4a} and Roush^{5c} in their syntheses of apoptolidin. The structure of 12-syn was confirmed by X-ray crystallography.

Based upon the well-known delivery of hydride to proximal ketones via hydridoalkoxyalanate[IV] intermediates,11 reaction of Me₃Al with alcohol 12-syn to form the alkoxyalane[III] followed by addition one equivalent of MeLi to presumably formed trimethylalkoxyalanate[IV] 13a12,13 which intramolecularly transferred methyl to the ketone carbonyl from alpha face with 7.5:1 selectivity (Scheme 1). While pentaalkyl aluminium[v] species are known, 14 computer searches reveal

Retrosynthesis for apoptolidin A and D.

Sugar 8 was envisioned to arise from methyl-D-glucopyranose which has three of the five required stereogenic centers.

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Fig. 2 Acetate-directed glycosylation strategy for the synthesis of 5.

Scheme 1 Synthesis and methylation of vinylogous ester 11.

no examples of dianionic alkoxyteteraalkyl species such as 13b which is tentatively proposed to explain the exquisite > 50:112-syn/12-anti selectivity observed in the reaction adding two equivalents of methyl lithium to the alkoxyalane preformed

Diol 12-syn was converted to acetate 14 in quantitative yield. Tert-alcohol 14 was protected as silyl ether 15. Acetate 15 was reduced to alcohol 16 using DIBAL-H.^{4a} Finally, iodoglycosylation of 16 with benzyl alcohol provided 7 in 83% yield and excellent α -selectivity, completing the synthesis of olivomycose sugar in 24% yield over 10 steps from L-rhamnose (Scheme 2).15

Synthesis of d-oleandrose sugar began from methyl D-glucopyranoside. Protection of 1,3-diol followed by benzoylation provided 17 in 67% over 2 steps (Scheme 3). Methylation of free alcohol in 17 followed by the deprotection of benzoyl ester gave 18 in quantitative yield over 2 steps. Hanessian-Hullar reaction¹⁶ of benzylidene **18** provided bromide **19**¹⁷ along with ketone-hydrate 20 which is produced via hypobromide oxidation¹⁸ and hydration. The literature reveals that other 2-oxo-sugars are susceptible to forming hydrates. 19 The propensity of hydrate formation is likely due to ring strain relief in

Scheme 2 Synthesis and iodo-glycosylation of 16.

Scheme 3 Preparation of bromosugar 19

conjunction with hydrogen bonding to the C-1 oxygen mojety. The structure of 20 was characterized by ¹H, ¹³C-NMR (C-2 appearing at 108.9 ppm) and MS.

The dehalogenation of bromide 19 was explored next. Subjecting compound 19 to dehalogenation conditions developed by Fu,²⁰ led to only 25% conversion. Dehalogenation under typical AIBN/tributyltin hydride²¹ conditions improved the conversion to 66%. Reaction with triethylboron/tributyltin hydride²² led to complete conversion and 87% isolated yield. Parallel attempts with the eco-friendly Kim combination of peroxydisulfate mediated²³ conditions offered good conversions. Ideally, quantitative conversion and 90% isolated yield of the benzoyl ester 21 were obtained when ammonium formate and tetrabutylammonium peroxysulfate was used in DMF at 60 °C. It appears that this is the first application of the Kim reagent for radical reductive cleavage of a C-Br bond.

Sugar 21 was converted to glycosyl acceptor 8 in 3 steps and 74% yield. Acetylation of 21 provided anomeric acetates 22 in 5:1 dr. Regioselective hydrolysis of anomeric acetate in 22 provided lactol 23 as a 3:1 anomeric mixture. Treatment of lactol 23 with trichloroacetonitrile²⁴ and DBU afforded the chloroacetimidate 8 in excellent yield and high selectivity (Scheme 4). Gratifyingly, acetate-directed glycosylation of 8 with alcohol 7 afforded β -glycoside 6 in 89% yield and 9:1 selectivity.

The acetate of 6 was cleaved under aqueous hydrazine conditions, and the resulting alcohol 24 was converted to xanthate 25 in 72% yield over two steps (Scheme 5). The xanthate

Table 1 Methylation of 11

Reagent	12-syn/12-anti	Yield of 12-syn	Yield of 12-anti
MeLi (1.1 eq.)	1.5:1	55%	35%
Me ₂ Zn (1.1 eq.), then MeLi (1 eq.)	3:1	60%	20%
Me ₃ Al (1.1 eq.), then MeLi (1.1 eq.)	7.5:1	75%	10%
Me ₃ Al (1.1 eq.), then MeLi (2.2 eq.)	50:1	85%	Trace
Me_4Zr^{10c}	N/A	64%	N/A

Scheme 4 Preparation of 6.

Scheme 5 Preparation of sugar 26.

Scheme 6 Synthesis of glycosyl donor 5.

and the iodo groups were removed in one pot using tributyltin hydride to provide **26** in 88% yield. ²⁵

Reduction of 26 under Pd/H₂ conditions followed by the fluorination of lactol 27 with diethylamino sulfur trifluoride²⁶ (DAST) provided the glycosidic donor 5 in 81% yield but required chromatography (Scheme 6). A more effective fluorinating agent, XtalFluor-E[®] 28 was recently reported.²⁷ Fluorination of lactol 27 by this reagent provided the fluoride 5 in excellent yield with no side products, thus eliminating the need for chromatography.

The synthesis of **5** was completed in 10% yield over 16 linear steps. Studies on O-27 glycosylation of the glycosyl fluoride **5** are currently under investigation.

Notes and references

- J. W. Kim, H. Adachi, K. Shin-ya, Y. Hayakawa and H. Seto, J. Antibiot. (Tokyo), 1997, 50(7), 628.
- 2 For a recent review see: P. T. Daniel, U. Koert and J. Schuppan, Angew. Chem., Int. Ed., 2006, 45, 872.
- (a) P. A. Wender, M. Sukopp and K. Longcore, Org. Lett., 2005, 7, 3025; (b) P. A. Wender and K. E. Longcore, Org. Lett., 2009, 11, 5474; (c) P. A. Wender and K. E. Longcore, Org. Lett., 2007, 9, 691; (d) P. A. Wender, A. V. Gulledge, O. D. Jankowski and

- H. Seto, *Org. Lett.*, 2002, **4**, 3819; (*e*) P. A. Wender and K. E. Longcore, *Org. Lett.*, 2009, **11**, 5474.
- 4 (a) H. Wehlan, M. Dauber, M. T. Mujica Fernaud, J. Schuppan, R. Mahrwald, B. Ziemer, M. E. Juarez Garcia and U. Koert, Angew. Chem., Int. Ed., 2004, 43, 4597;
 (b) K. C. Nicolaou, Y. Li, K. Sugita, H. Monenschein, P. Guntupalli, H. J. Mitchell, K. C. Fylaktakidou, D. Vourloumis, P. Giannakakou and A. O'Brate, J. Am. Chem. Soc., 2003, 125, 15443; (c) M. T. Crimmins, H. S. Christie, A. Long and K. Chaudhary, Org. Lett., 2009, 11, 831.
- 5 (a) K. C. Nicolaou, Y. Li, K. C. Fylaktakidou, H. J. Mitchell, H.-X. Wei and B. Weyershausen, Angew. Chem., Int. Ed., 2001, 40, 3849; (b) M. T. Crimmins and A. Long, Org. Lett., 2005, 7, 4157; (c) M. Handa, W. J. Smith, III and W. R. Roush, J. Org. Chem., 2008, 73, 1036.
- 6 (a) A. R. Salomon, D. W. Voehringer, L. A. Herzenberg and C. Khosla, *Proc. Natl. Acad. Sci. U. S. A.*, 2000, 97, 14766; (b) A. R. Salomon, D. W. Voehringer, L. A. Herzenberg and C. Khosla, *Chem. Biol.*, 2001, 8, 71.
- 7 Y. Kim and P. L. Fuchs, Org. Lett., 2007, 9, 2445.
- D. Horton, W. Priebe and O. Varela, *Carbohydr. Res.*, 1985, **144**, 317.
- 9 S. Czernecki, K. Vijayakumaran and G. Ville, *J. Org. Chem.*, 1986, 51, 5472.
- (a) G. Jung and A. Klemer, *Chem. Ber.*, 1981, 114, 740;
 (b) K. A. Parker and S. M. Meschwitz, *Carbohydr. Res.*, 1988, 172, 319;
 (c) T. Kauffmann, W. Klaffke, C. Philipp and J. Thiem, *Carbohydr. Res.*, 1990, 207, 33.
- 11 (a) J. A. Katzenellenbogen and S. B. Bowlus, J. Org. Chem., 1973, 38, 627; (b) S. B. Bowlus and J. A. Katzenellenbogen, J. Org. Chem., 1974, 39, 3309; (c) T. Nakata, T. Tanaka and T. Oishi, Tetrahedron Lett., 1983, 24, 2653; (d) N. Bajwa and M. P. Jennings, J. Org. Chem., 2008, 73, 3638.
- 12 For a report on intramolecular methyl delivery to an adjacent epoxide via trimethylalkoxyalanate(iv), see: M. Sasaki, K. Tanino and M. Miyashita, Org. Lett., 2001, 3, 1765.
- 13 For diastereoselective 1,4-addition of methyl group to a quinol system via trimethylalkoxyalanate(iv), see: (a) M. C. Carrenō, M. Perez-Gonzalez, M. Ribagorda and K. N. Houk, J. Org. Chem., 1998, 63, 3687; (b) M. C. Carrenō, M. Perez-Gonzalez, M. Ribagorda and J. Fisher, J. Org. Chem., 1996, 61, 6758; (c) V. Rodeschini, J.-G. Boiteau, P. Van de Weghe, C. Tarnus and J. Eustache, J. Org. Chem., 2003, 69, 357.
- 14 T. S. Pritytskaya, S. V. Sviridov and T. K. Morozova, Zh. Obshch. Khim., 1987, 57, 839.
- 15 For a related glycosylation, see: D. Hou and T. L. Lowary, *J. Org. Chem.*, 2009, **74**, 2278.
- 16 (a) S. Hanessian, Carbohydr. Res., 1966, 4, 86; (b) D. L. Failia,
 T. L. Hullar and S. B. Siskin, Chem. Commun., 1966, 716;
 (c) S. Hanessian and N. R. Pleassas, J. Org. Chem., 1969, 34, 1035, 1045, 1053; (d) T. L. Hullar and S. B. J. Siskin, J. Org. Chem., 1970, 35, 225.
- 17 K. Tori, T. That Thang, M. Sangare and G. Lukacs, *Tetrahedron Lett.*, 1977, 18, 717.
- 18 R. Filler, Chem. Rev., 1963, 63, 21.
- (a) H.-M. Liu, Y. Sato and Y. Tsuda, Chem. Pharm. Bull., 1993, 41, 491; (b) F. W. Lichtenthaler and P. Jarglis, Chem. Ber., 1980, 113, 489; (c) C. D. Warren, C. Augé, M. L. Laver, S. Suzuki, D. Power and R. Jeanloz, Carbohydr. Res., 1980, 82, 71; (d) D. C. Baker, D. Horton and C. G. Tindall, Jr., Carbohydr. Res., 1972, 24, 192; (e) M. Mori and S. Tejima, Chem. Pharm. Bull., 1983, 31, 1593; (f) H. Tian, X. She and Y. Shi, Org. Lett., 2001, 3, 715; (g) M. A. Mortellaro, S. Hong, D. T. Winn and A. W. Czarnik, Bioorg. Med. Chem. Lett., 1994, 4, 2041.
- 20 J. Tormo and G. C. Fu, Org. Syn., 2002, 78, 239.
- 21 H. G. Kuivila, Acc. Chem. Res., 1968, 1, 299.
- 22 P. Renaud and M. Gerster, Angew. Chem., Int. Ed., 1998, 37, 2562
- 23 H. S. Park, H. Y. Lee and Y. H. Kim, Org. Lett., 2005, 7, 3187.
- 24 R. Preuss and R. R. Schmidt, Synthesis, 1988, 694.
- 25 D. H. R. Barton and S. W. McCombie, J. Chem. Soc., Perkin Trans. 1, 1975, 16, 1574.
- 26 W. J. Middleton, J. Org. Chem., 1975, 40, 574.
- 27 A. L'Heureux, F. Beaulieu, C. Bennett, D. R. Bill, S. Clayton, F. o. LaFlamme, M. Mirmehrabi, S. Tadayon, D. Tovell and M. Couturier, *J. Org. Chem.*, 2010, 75, 3401.