LETTERS

Stereoselective Synthesis and Evaluation of C6"-Substituted 5a-Carbasugar Analogues of SL0101 as Inhibitors of RSK1/2

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Supporting Information

ABSTRACT: A convergent synthesis of 5a-carbasugar analogues of the *n*-Pr-variant of SL0101 is described. The analogues were synthesized in an effort to find compounds with potent *in vivo* efficacy in the inhibition of p90 ribosomal s6 kinase (RSK1/2). The synthesis derived the desired C-4 L-rhamnose stereochemistry from quinic acid and used a highly selective cuprate addition, NaBH₄ reduction, Mitsunobu inversion, and alkene dihydroxylation to install the remaining stereochemistry. A Pd-catalyzed cyclitolization stereoselectively installed the aglycon at the anomeric position. The analogues were evaluated as PSK1/2 inhibitors and found t



position. The analogues were evaluated as RSK1/2 inhibitors and found to have 3- to 6-fold improved activity.

The p90 ribosomal s6 kinases (RSK) are a family of Ser/Thr protein kinases.¹ Two isoforms (RSK1/2) from this family are involved in the etiology of a number of different cancers.² In an effort aimed at identifying RSK1/2 inhibitors, the flavonoid glycoside natural product SL0101 (1) was discovered as a relatively selective inhibitor of the *N*-terminal kinase domain (NTKD) of RSK.³ RSK has two kinase domains where the *N*terminal domain (NTKD) is responsible for phosphorylation of target substrates.⁴ Based on the crystal structure of the RSK2 NTKD complexed with SL0101, a major conformational rearrangement of the *N*-lobe of the kinase domain generates the inhibitor-binding pocket.⁵

Inspired by its unique activity and selectivity, we have been exploring structure–activity relationship (SAR) requirements associated with SL0101 (1).^{3,6} As part of these studies, we have developed a *de novo* asymmetric synthesis⁷ of SL0101, its enantiomer, and several congeners.⁸ Our studies have identified several analogues with improved activity and have emphasized the importance of the rhamnose sugar and its C-3 and C-4 acetates. In addition, we have found that substitution at the C-6 position^{8,9} and the ring oxygen (2 and 3) of the sugar gives improved efficacy in the *in vitro* kinase assays and cell-based studies.^{8,10}

SL0101 (1) has a short biological half-life *in vivo*,⁵ which is presumably due to the hydrolyzable *C*-3/*C*-4-acetates on the sugar, as well as an *O*-glycosidic bond. To identify less labile groups that could replace the ester without loss of affinity, we have investigated replacing the rhamnose *C*-4-acetate (e.g., **5a**–**f** with a *C*-4 acetamide), the *C*-3/*C*-4-acetates (e.g., **4h** with a *C*-3/*C*-4-n-Pr-carbamates), and the ring oxygen with a methylene group (i.e., **3a**–**d** carbasugars).⁸ As part of our ongoing effort to identify RSK1/2-inhibitors as potential therapeutics, we decided to test the effects of combining these three substitutions and

Scheme 1. SAR for SL0101 RSK Inhibition⁸



targeted two analogues 2a-b (Scheme 1). Herein we disclose the synthesis of cyclitol analogues 2a and 2b as well as the relative RSK2 inhibitory activity.¹¹

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Retrosynthetically, we envisioned preparing 2a and 2b in a route analogous to our previously established routes to SL0101 analogues 1-5 (e.g., 6 + 7, Scheme 2). Specifically, we expected

Scheme 2. Retrosynthesis of n-Pr Carbasugar Analogues



that 2a/b would arise from the Pd-catalyzed cyclitolization of 7 with enone 8.^{8,12} Base on our previous success, we viewed the cyclitol donor 8 would arise from quinic acid 10 via enone 9,¹³ where the quinic acid tertiary alcohol would become the *C*-4 ketone and the central alcohol of the triol would become the *C*-1 anomeric position.

At the outset, our efforts to extend our previous cyclitol synthesis (R = H) encountered difficulty, associated primarily with achieving high stereocontrol in the conversion of **9** into **12b**. Specifically, we explored the possibility of installing the sugar L-stereochemistry at C-5 by means of a selective hydrogenation of the enone **11** from the exoface to selectively provide **12b** over **12a**. To our surprise, when we exposed **11** to typical hydrogenation conditions (1 atm of H₂ with Pd/C), we found **12a** was the major isomer, but with poor diastereoselectivity (3:1) (Scheme 3). This problem was exacerbated by the fact that isomerization of the acetonide.¹² These two compounding factors prompted a search for an alternative approach.¹⁴

Scheme 3. Initial Attempt to α-L-Cyclitol Donors



Our redesigned retrosynthesis turned the original design 180° (Scheme 4), with the quinic acid tertiary alcohol becoming the





carbasugar C-1 anomeric position and the central alcohol of the triol becoming the C-4 enone 8. Because of complications with its conversion to enone 8 this effort turned to the synthesis of allylic acetate 17. Specifically, the allylic benzoate 17 already had a Pd- π -allyl leaving group at the pseudoanomeric position for the cyclitolization reaction. In addition, benzoate 17 also had the desired carbasugar C-4 acetate in the correct rhamno-stereochemistry. The question that remained was could we find conditions to selectively ionize the axial C-1 allylic p-NO₂Bz group without touching the allylic C-4 acetate. In addition, we were concerned that the C-4 acetate may not control the regiochemistry of nucleophilic attack to the π -allyl intermediate (*i.e.*, **TS-1**) as well as the C-4 ketone (*i.e.*, **TS-2**).¹⁵ In general, we found a C-4 ketone both improved the electrophilicity of the Pd- π -allyl intermediate and helped direct nucleophilic addition to the C-1 position. In this regard, we were hopeful that the regiocontrol issues could also be controlled by the C-5 n-propyl substituent.¹⁶ Altenatively, chiral ligands on the Pd- π -allyl could be used if the C-4 acetate is not sufficient for controlling the regiochemistry.

Our redesigned synthesis returned to enone 9, which underwent a highly stereoselective cuprate-promoted addition of *n*-propyl anion to furnish ketone 13 in an excellent yield and as a single diastereomer (89%).¹⁷ Exposure of 13 to DBU in benzene gave allylic alcohol 14 in good yield (80%). The required C-4 acetate was then installed by an acylation of 14 with acetic anhydride/DMAP with Et₃N to deliver allylic acetate 15 (74%). A stereoselective 1,2-reduction of enone 15 under Luche conditions¹⁸ gave the allylic alcohol **16** in good yield (89%) and diastereoselectivity (>10:1). Analysis of the allylic coupling constants in the ¹H NMR for 16 indicated that the Luche reduction occurred via axial attack to install an equatorial alcohol,¹⁹ which corresponds to β -anomeric stereochemistry in the resulting carbasugar. Thus, a Mitsunobu reaction was preformed on 16 with DIAD/PPh₃ and *p*-nitrobenzoic acid to yield the desired cyclitol donor 17 in excellent yield (90%).

Unfortunately, the conversion of the bis-allylic ester 17 to enone 8 proved to be untenable, as conditions for the selective hydrolysis of either of the two esters were not found. In an effort to find a viable alternative, we decided to explore the use of allylic p-NO₂-benzoate 17 as a cyclitol donor, in the cyclitolization reactions. The allylic p-nitrobenzoate 17 had some potential advantages to 8 in our planned synthesis of SL0101 analogue 2a. Specifically, the *C*-1 allylic-benzoate 17 already had a Pd- π -allyl leaving group at the anomeric position for the cyclitolization reaction. In addition, the *p*-nitrobenzoate 17 also had the desired *C*-4 acetate with the correct stereochemistry installed, and thus, the aglycon portion would go through two less transformations (Figure 1).



Figure 1. Competing Pd-*π*-allyl mechanisms.

To our delight, when a mixture of *p*-nitrobenzoate 17 and flavonol 7 was exposed to our typical glycosylation conditions $(2.5 \text{ mol } \% \text{ Pd}_2(\text{dba})_3 \cdot \text{CHCl}_3 \text{ and } 10 \text{ mol } \% \text{ of PPh}_3 \text{ in CH}_2\text{Cl}_2 \text{ at}$ 0 °C) the reaction proceeded smoothly to provide the desired product in good yield (80%), in excellent regio- and stereoselectivity (Scheme 5). Exposure of the allylic acetate 18 to the





Upjohn conditions²¹ (OsO₄/NMO; 70%) stereoselectively converted it into the *rhamno*-diol **19** which, when acylated with acetyl chloride and Hunig's base, gave a mixture of the *C*-3 and *C*-2 acetates **20** and **21** (1:1.6) in 38% yield. Fortunately, the undesired *C*-2 acetate **21** could be isomerized to a mixture of acetates which favored the desired *C*-3 acetate **20** (3:1). Finally, the desired regioisomer **20** was globally deprotected by an exhaustive hydrogenolysis (1 atm of hydrogen with Pd/C), which produced the target *C*-3/*C*-4 diacetate **2a** in good yield (70%).

With access to the desired diacetate **2a** having been established, we turned our attention to the second target compound biscarbamate **2b** (Scheme 6). Our approach to biscarbamate **2b** returned us to allylic acetate **18**, which could be hydrolyzed into allylic alcohol **22** with K_2CO_3 in methanol (84%). The allylic alcohol **22** was converted into *n*-Pr-carbamate **23** by exposure of it to *n*-Pr-isocyanate and base (10% DBU/

Scheme 6. Synthesis of Bis-carbamate Analogues



Et₃N in MeCN). Exposure of the allylic carbamate **23** to the Upjohn conditions (OsO_4/NMO ; 73%) stereoselectively converted it into the *rhamno*-diol **24**. Re-exposure of the C-2/C-3 diol **24** to the carbamate forming conditions (*n*-Pr-isocyanate, 10% DBU/Et₃N in MeCN) gave a mixture of C-3 and C-2 carbamates **20** and **21** (1:1). The desired C-3/C-4 biscarbamate **25** could be isolated from that mixture in a 40% overall yield. Unfortunately, the undesired C-2/C-4 biscarbamate **26** could not be isomerized under basic conditions as was the case with the diacetate. Finally, the desired C-3/C-4 regioisomer **25** was globally deprotected by an exhaustive hydrogenolysis (1 atm of hydrogen with Pd/C), to afford the target C-3/C-4 biscarbamate **2b** in good overall yield (72%).

The efficacy of the two cyclitol analogues **2a** and **2b** to inhibit RSK2 activity was determined in *in vitro* kinase assays using purified recombinant RSK2 (Table 1).³ The data were fit using nonlinear regression analysis. Both SL0101 analogues showed improved RSK inhibition over the lead structure SL0101, the *n*-Pr cyclitol diacetate **2a** having a >6-fold decrease in IC₅₀ (54 nM), whereas the bis-carbamate **2b** had an ~3-fold decrease in

Table 1. In Vitro Potency of SL0101 (1) and Analogues



^{*a*}For procedure see Supporting Information. ^{*b*}See ref 11.

IC₅₀ (137 nM). In the parent series, cyclitol substitution (i.e., **1** to **3a**) led to a modest improvement in the inhibitory activity (345 to 270 nM). In the *n*-Pr-series, however, the effect of cyclitol substitution (i.e., **4b** to **2a**) trended toward reduced efficacy (20 to 56 nM). This loss in activity should be easily compensated for by the expected improved bioavailability that results from the hydrolysis-resistant cyclitol substitution. There was also a loss in inhibitory activity (345 to 870 nM) by the *n*-Pr-carbamate substitution in the parent series (**1** to **4h**). This negative effect was also observed with the cyclitol series (i.e., **2a** to **2b**: 56 to 137 nM).

In conclusion, using a highly stereoselective Pd-cyclitolization reaction, two new cyclitol analogues of the natural product SL0101 were synthesized and evaluated. These analogues **2a** and **2b** showed significant improvement in RSK inhibitory activity. Improved bioavailability has already been shown for **2a**.¹¹ The synthesis of these new SL0101 cyclitol analogues required the discovery of a novel synthesis of a new cyclitol donor, which demonstrated that the previously believed requirement of a C-4 ketone in the cyclitol donor is not necessary for the reaction with phenol-like nucleophiles. Further studies aimed at defining the requirements for a specific-RSK1/2 inhibition are ongoing and will be reported in due course.

ASSOCIATED CONTENT

Supporting Information

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Experimental procedures and spectral data (PDF)

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Notes

The authors declare the following competing financial interest(s): The coresponding authors Deborah A. Lannigan and George A. O'Doherty have applied for patent protection for this class of compounds.

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REFERENCES

(1) Eisinger-Mathason, T. S.; Andrade, J.; Lannigan, D. A. *Steroids* **2010**, 75, 191.

(2) (a) Lara, R.; Mauri, F. A.; Taylor, H.; Derua, R.; Shia, A.; Gray, C.; Nicols, A.; Shiner, R. J.; Schofield, E.; Bates, P. A.; Waelkens, E.; Dallman, M.; Lamb, J.; Zicha, D.; Downward, J.; Seckl, M. J.; Pardo, O. E. *Oncogene* **2011**, *30*, 3513. (b) Doehn, U.; Hauge, C.; Frank, S. R.; Jensen, C. J.; Duda, K.; Nielsen, J. V.; Cohen, M. S.; Johansen, J. V.; Winther, B. R.; Lund, L. R.; Winther, O.; Taunton, J.; Hansen, S. H.; Frodin, M. *Mol. Cell* **2009**, *35*, 511. (3) Smith, J. A.; Poteet-Smith, C. E.; Xu, Y.; Errington, T. M.; Hecht, S. M.; Lannigan, D. A. *Cancer Res.* **2005**, *65*, 1027.

(4) Anjum, R.; Blenis, J. Nat. Rev. Mol. Cell Biol. 2008, 9, 747.

(5) Utepbergenov, D.; Derewenda, U.; Olekhnovich, N.; Szukalska, G.; Banerjee, B.; Hilinski, M. K.; Lannigan, D. A.; Stukenberg, P. T.; Derewenda, Z. S. *Biochemistry* **2012**, *51*, 6499.

(6) (a) Smith, J. A.; Maloney, D. J.; Clark, D. E.; Xu, Y.; Hecht, S. M.; Lannigan, D. A. *Bioorg. Med. Chem.* **2006**, *14*, 6034. (b) Smith, J. A.; Maloney, D. J.; Hecht, S. M.; Lannigan, D. A. *Bioorg. Med. Chem.* **2007**, *15* (14), 5018.

(7) (a) Iyer, A.; Zhou, M.; Azad, N.; Elbaz, H.; Wang, L.; Rogalsky, D. K.; Rojanasakul, Y.; O'Doherty, G. A.; Langenhan, J. M. ACS Med. Chem. Lett. **2010**, *1*, 326. (b) Shi, P.; Silva, M.; Wu, B.; Wang, H. Y. L.; Akhmedov, N. G.; Li, M.; Beuning, P.; O'Doherty, G. A. ACS Med. Chem. Lett. **2012**, *3*, 1086. (c) Wu, B.; Li, M.; O'Doherty, G. A. Org. Lett. **2010**, *12*, 5466.

(8) (a) Li, M.; Li, Y.; Mrozowski, R. M.; Sandusky, Z. M.; Shan, M.; Song, X.; Wu, B.; Zhang, Q.; Lannigan, D. A.; O'Doherty, G. A. ACS Med. Chem. Lett. 2015, 6, 95. (b) Mrozowski, R. M.; Sandusky, Z. M.; Vemula, R.; Wu, B.; Zhang, Q.; Lannigan, D. A.; O'Doherty, G. A. Org. Lett. 2014, 16, 5996. (c) Mrozowski, R. M.; Vemula, R.; Wu, B.; Zhang, Q.; Schroeder, B. R.; Hilinski, M. K.; Clark, D. E.; Hecht, S. M.; O'Doherty, G. A.; Lannigan, D. A. ACS Med. Chem. Lett. 2013, 4, 175. (d) Shan, M.; O'Doherty, G. A. Org. Lett. 2010, 12, 2986.

(9) Wang, H. Y. L.; Xin, W.; Zhou, M.; Stueckle, T. A.; Rojanasakul, Y.; O'Doherty, G. A. *ACS Med. Chem. Lett.* **2011**, *2*, 73.

(10) Wang, H. Y.; Wu, B.; Zhang, Q.; Kang, S. W.; Rojanasakul, Y.; O'Doherty, G. A. *ACS Med. Chem. Lett.* **2011**, *2*, 259.

(11) For detailed *in vivo* studies of **2a**, see: Ludwik, K. A.; Campbell, J. P.; Li, M.; Li, Y.; Sandusky, Z. M.; Pasic, L.; Sowder, M. E.; Brenin, D. R.; Pietenpol, J. A.; O'Doherty, G. A.; Lannigan, D. A. *Mol. Cancer Ther.* **2016**, *15*, 2598.

(12) Shan, M.; O'Doherty, G. A. Synthesis 2008, 2008, 3171.

(13) (a) Federspiel, M.; Fischer, R.; Hennig, M.; Mair, H. J.; Oberhauser, T.; Rimmler, G.; Albiez, T.; Bruhin, J.; Estermann, H.; Gandert, C.; Gockel, V.; Gotzo, S.; Hoffmann, U.; Huber, G.; Janatsch, G.; Lauper, S.; Rockel-Stabler, O.; Trussardi, R.; Zwahlen, A. G. Org. *Process Res. Dev.* **1999**, *3*, 266. (b) Trost, B. M.; Romero, A. G. J. Org. *Chem.* **1986**, *51*, 2332. (c) Audia, J. E.; Boisvert, L.; Patten, A. D.; Villalobos, A.; Danishefsky, S. J. J. Org. Chem. **1989**, *54*, 3738.

(14) (a) Guo, H.; O'Doherty, G. A. *Tetrahedron* 2008, 64, 304.
(b) Abrams, J. N.; Babu, R. S.; Guo, H.; Le, D.; Le, J.; Osbourn, J. M.; O'Doherty, G. A. *J. Org. Chem.* 2008, 73, 1935. (c) Coral, J. A.; Guo, H.; Shan, M.; O'Doherty, G. A. *Heterocycles* 2009, 79, 521. (d) Borisova, S. A.; Guppi, S. R.; Kim, H. J.; Wu, B.; Penn, J. H.; Liu, H. W.; O'Doherty, G. A. *Org. Lett.* 2010, *12*, 5150.

(15) Bajaj, S. O.; Sharif, E. U.; Akhmedov, N. G.; O'Doherty, G. A. *Chem. Sci.* **2014**, *5*, 2230.

(16) Previously, we have found that 1,4-regiocontrol can be addressed with chiral ligands; see ref 8b.

- (17) Lipshutz, B. H.; Ellsworth, E. L. Tetrahedron Lett. 1988, 29, 893.
- (18) Luche, J. L. J. Am. Chem. Soc. 1978, 100, 2226.
- (19) Eliel, E. L.; Senda, Y. Tetrahedron 1970, 26, 2411.

(20) Mitsunobu, O. Synthesis 1981, 1981, 1.

(21) VanRheenen, V.; Kelly, R. C.; Cha, D. Y. Tetrahedron Lett. 1976, 17, 1973.